

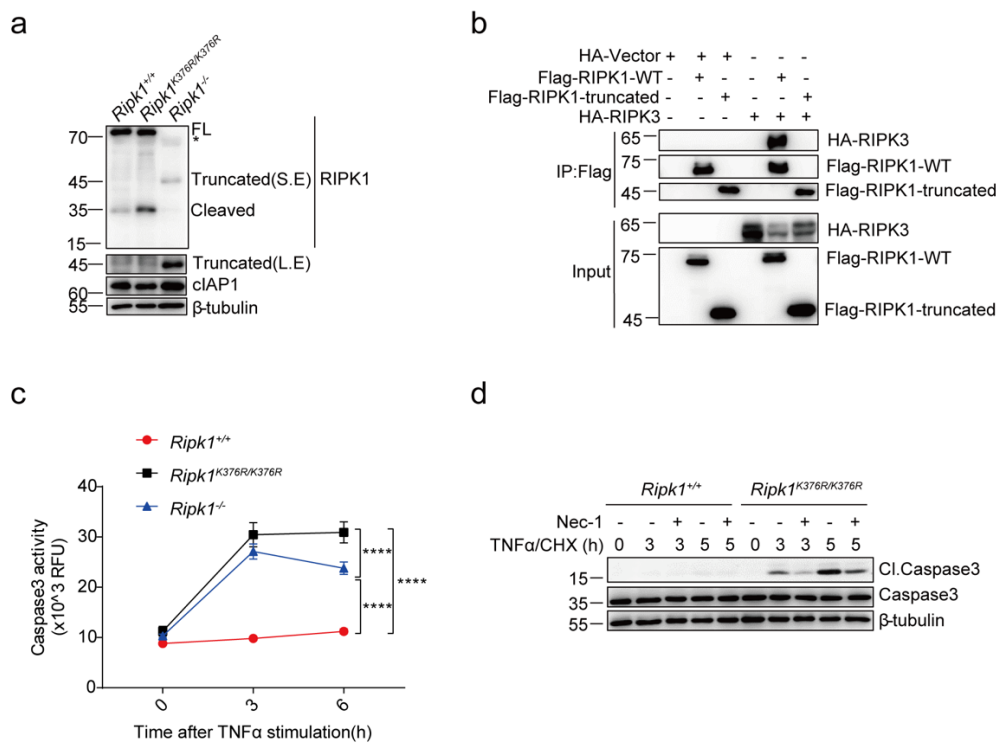
Tang et al, K63-linked ubiquitination regulates RIPK1 kinase activity to prevent cell death during embryogenesis and inflammation

Supplementary Figures

Genotypes/ Embryo Stage	E9.5	E10.5	E11.5	E12.5	E13.5	E14.5
<i>Ripk1</i> ^{+/+}	2	4	12	10	12	15
<i>Ripk1</i> ^{K376R/+}	5	8	24	18	22	28
<i>Ripk1</i> ^{K376R/K376R}	2	5	10	8	4	0
Total	9	17	46	36	38	43

Supplementary Figure 1. *Ripk1*^{K376R/K376R} mice are embryonic lethal at E13.5

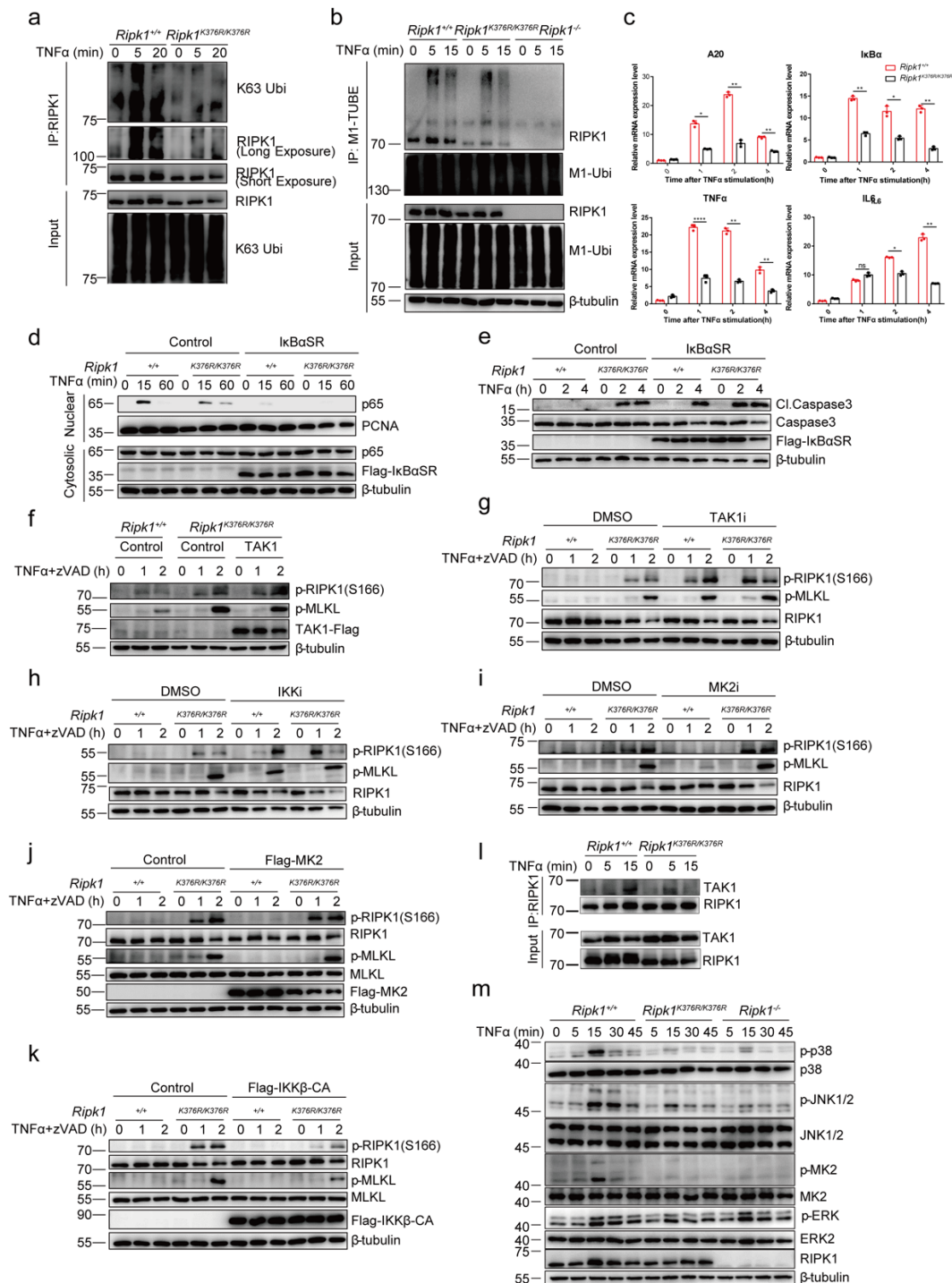
(a) Genotype analysis of offspring from *Ripk1*^{K376R/+} intercrosses mice at different embryonic stage.



Supplementary Figure 2. *Ripk1*^{K376R/K376R} mutation sensitizes cells to apoptosis and necroptosis

(a) Western-blotting detection of RIPK1 protein expression in primary MEFs from *Ripk1*^{+/+}, *Ripk1*^{K376R/K376R} and *Ripk1*^{-/-} mice (* refers to non-specific band). (b) Western-blotting analysis of immunoprecipitates using anti-Flag beads and total lysates of 293T cells transfected with the plasmids encoding Flag-RIPK1-WT, Flag-RIPK1-truncated and HA-RIPK3. (c) Caspase3 activity of *Ripk1*^{+/+}, *Ripk1*^{K376R/K376R} and *Ripk1*^{-/-} immortalized MEFs treated with TNFα (40ng/ml) for different time point were measured by DEVD-AMC fluorescence, the error bars

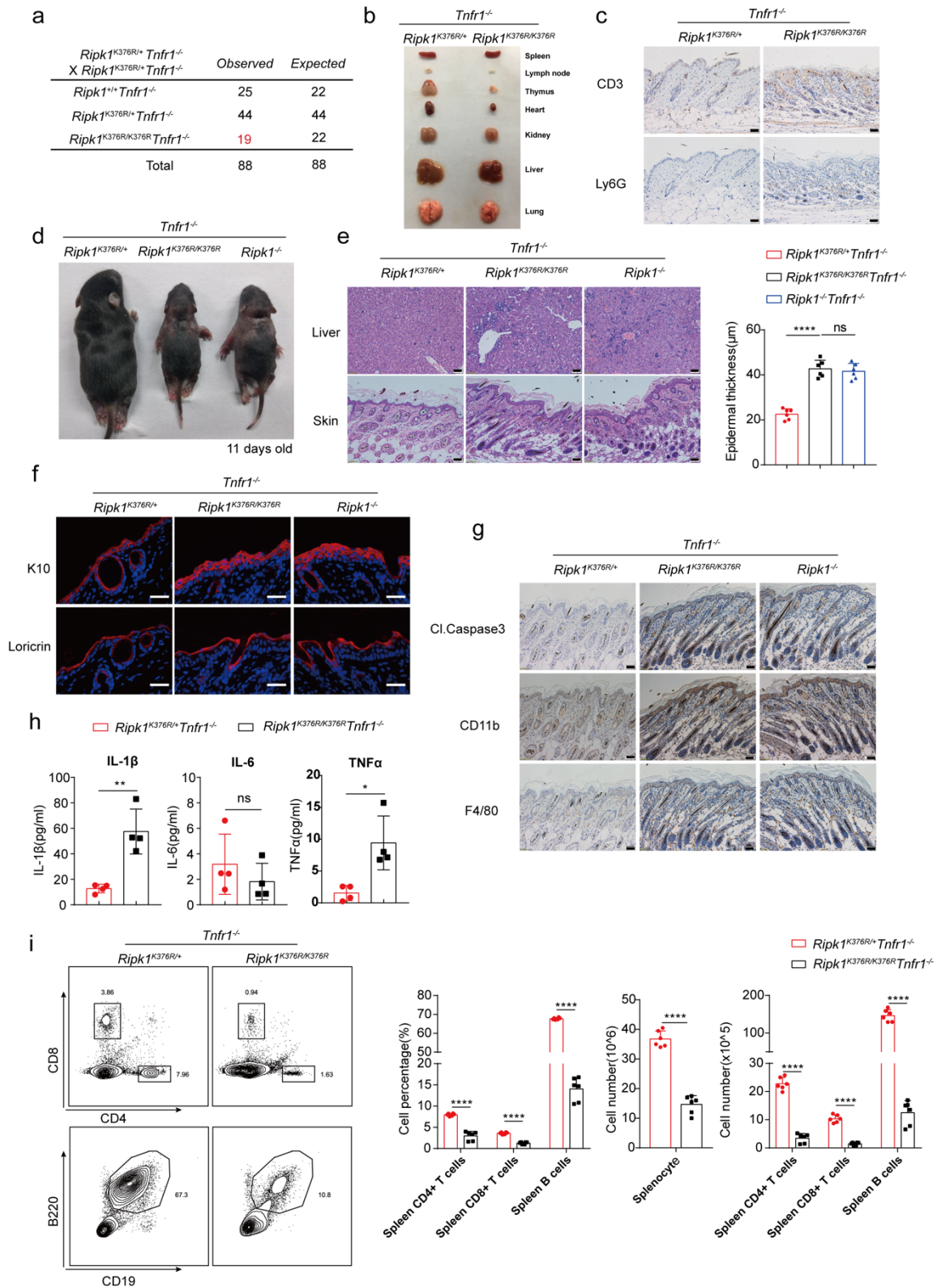
represent mean±s.e.m of data from three independent cell samples for each genotype. **(d)** *Ripk1*^{+/+} and *Ripk1*^{K376R/K376R} immortalized MEFs were treated with TNF α (10ng/ml) /CHX (10ug/ml) with or without pre-treatment of Necrostatin-1 for the indicated time, and the cell lysates were analyzed by western-blotting using the indicated antibodies. Statistical significance was determined using a two-tailed unpaired *t* test, *****P* < 0.0001.



Supplementary Figure 3. *Ripk1*^{K376R/K376R} mutation increases RIPK1 kinase activity

(a) Immunoprecipitation of RIPK1 antibody in *Ripk1*^{+/+} and *Ripk1*^{K376R/K376R} immortalized MEFs treated for the indicated time with TNF α (20 ng/ml). The immunocomplexes were

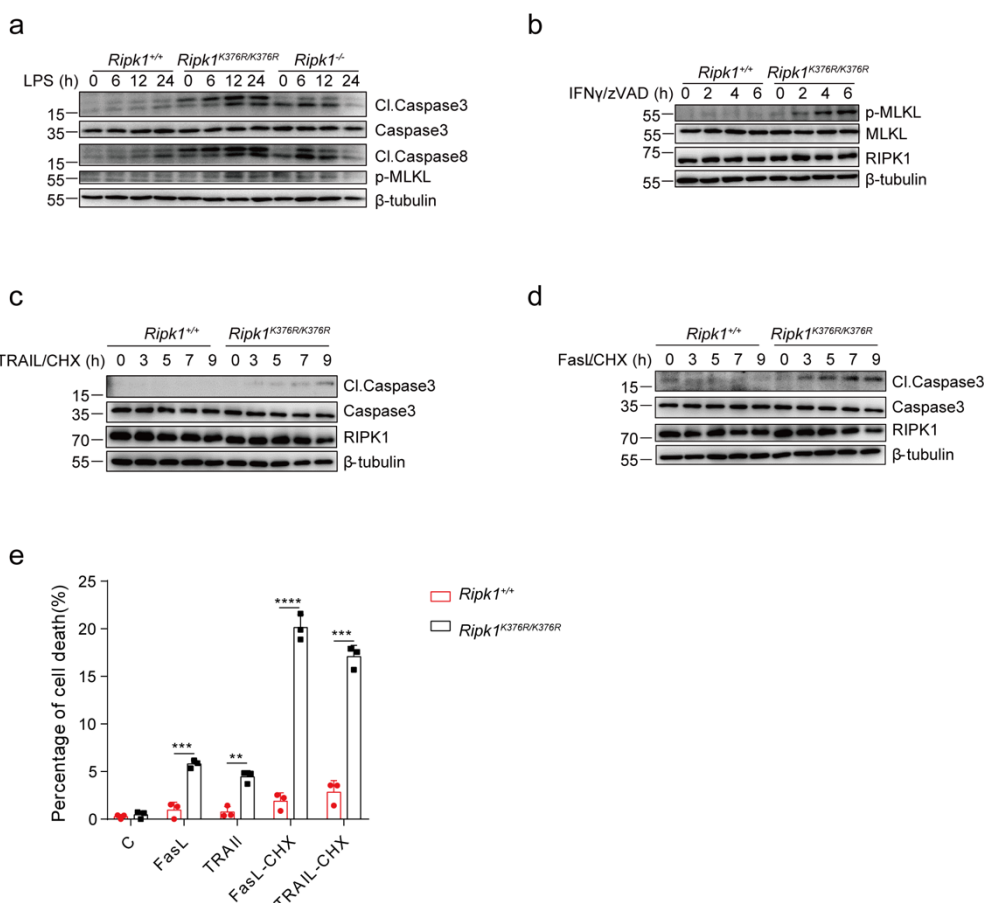
analyzed by western-blotting with antibody against K63-linked ubiquitination chains. **(b)** Immortalized *Ripk1*^{+/+}, *Ripk1*^{K376R/K376R} and *Ripk1*^{-/-} MEFs were treated for the indicated time with TNF α (20 ng/ml). The M1-ubiquitylated proteins were isolated by M1-TUBEs and analyzed by western-blotting. **(c)** RT-PCR analysis of NF- κ B-targeting genes expression in *Ripk1*^{+/+} or *Ripk1*^{K376R/K376R} MEFs treated with TNF α (20 ng/ml) for the indicated time, the error bars represent mean \pm s.e.m of data from three independent cell samples for each genotype. **(d)** Nuclear extracts were collected from *Ripk1*^{+/+} and *Ripk1*^{K376R/K376R} MEFs treated with TNF α (20 ng/ml) at indicated time with or without stably expressed Flag-tagged I κ B α -SR and analyzed by western-blotting with antibodies against p65 and PCNA. **(e)** *Ripk1*^{+/+} and *Ripk1*^{K376R/K376R} immortalized MEFs that stably expressed with the Flag-tagged I κ B α -SR were stimulated with TNF α (40 ng/ml) for different periods of time. The cell lysates were analyzed by western-blotting with indicated antibodies. **(f,j,k)** *Ripk1*^{+/+} and *Ripk1*^{K376R/K376R} immortalized MEFs that expressed with the Flag-tagged TAK1**(f)** , Flag-tagged MK2**(j)** or Flag-IKK β -CA**(k)** were stimulated with TNF α (20 ng/ml) plus zVAD.fmk (10 μ M) for different periods of time. The cell lysates were analyzed by western-blotting with indicated antibodies. **(g-i)** *Ripk1*^{+/+} and *Ripk1*^{K376R/K376R} immortalized MEFs were stimulated with TNF α (20 ng/ml) plus zVAD.fmk (10 μ M) for different periods of time with or without pre-treatment of TAK1 inhibitor**(g)**, IKK inhibitor**(h)**, MK2 inhibitor**(i)**, The cell lysates were analyzed by western-blotting with indicated antibodies. **(l)** Immunoprecipitation of RIPK1 antibody in *Ripk1*^{+/+} and *Ripk1*^{K376R/K376R} immortalized MEFs treated for the indicated time with TNF α (20 ng/ml). The immunocomplexes were analyzed by western-blotting with antibody against TAK1. **(m)** Immortalized *Ripk1*^{+/+}, *Ripk1*^{K376R/K376R} and *Ripk1*^{-/-} MEFs were treated for the indicated time with TNF α (20 ng/ml). The cell lysates were analyzed by western-blotting with indicated antibodies. Statistical significance was determined using a two-tailed unpaired *t* test, ns P > 0.05, *P < 0.05, **P < 0.01, ****P < 0.0001.



Supplementary Figure 4. TNFR1 deficiency partially delay the lethality of *Ripk1*^{K376R/K376R} mice

(a) Statistical analysis of the expected and observed offspring mice (11-days-old) from the intercrosses of *Ripk1*^{K376R/+} *Tnfr1*^{-/-} mice. (b) Representative macroscopic images of organs with

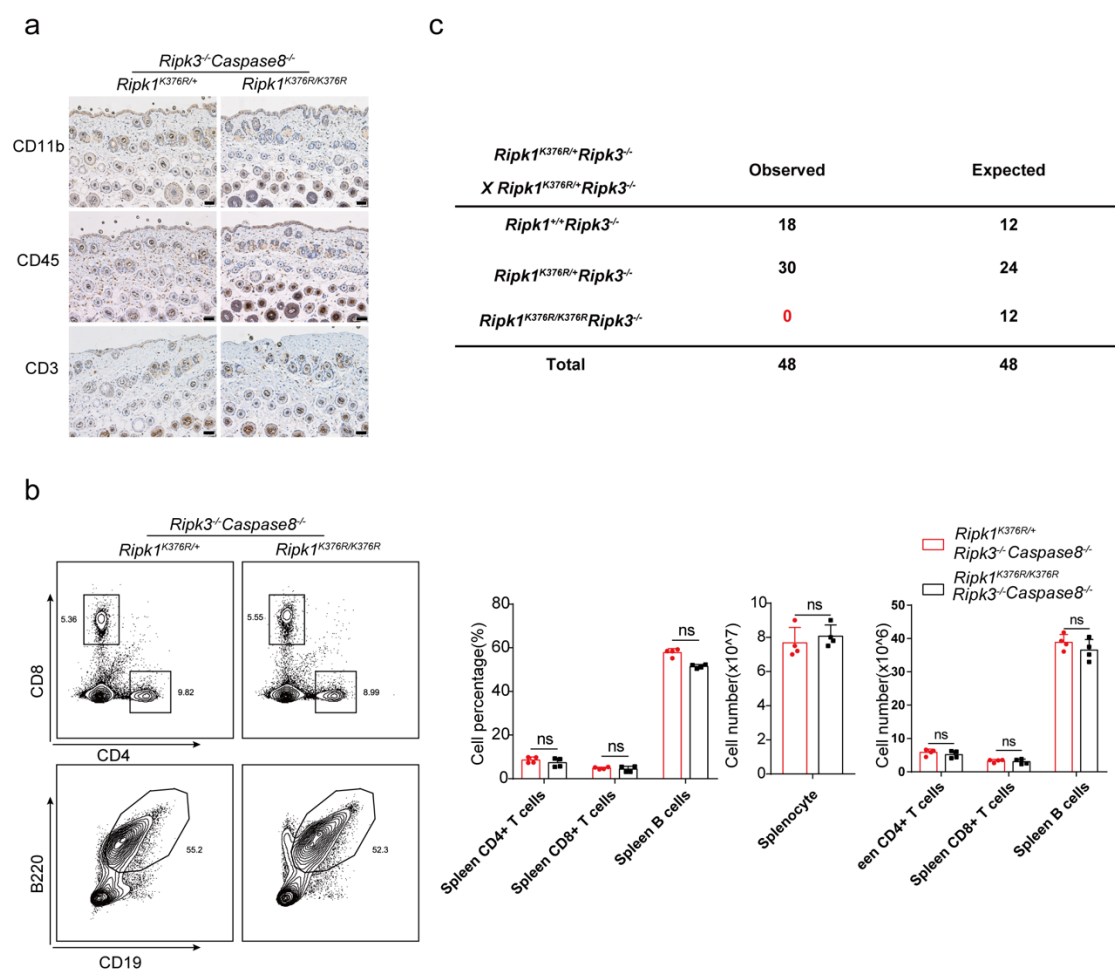
indicated genotypes at P11. (c) Immunohistochemical staining of CD3 and Ly6G in skin sections of *Ripk1*^{K376R/K376R}*Tnfr1*^{-/-} and *Ripk1*^{K376R/+}*Tnfr1*^{-/-} littermate mice at P11 (scale bar, 50 μ m). (d) Representative macroscopic images of *Ripk1*^{K376R/+}*Tnfr1*^{-/-}, *Ripk1*^{K376R/K376R}*Tnfr1*^{-/-} and *Ripk1*^{-/-}*Tnfr1*^{-/-} mice at P11. (e) H&E staining of liver and skin sections of *Ripk1*^{K376R/+}*Tnfr1*^{-/-}, *Ripk1*^{K376R/K376R}*Tnfr1*^{-/-} and *Ripk1*^{-/-}*Tnfr1*^{-/-} mice at P11 (scale bar, 50 μ m), and microscopic quantification of the epidermal thickness from H&E results (*Ripk1*^{K376R/+}*Tnfr1*^{-/-} mice: n=6; *Ripk1*^{K376R/K376R}*Tnfr1*^{-/-} mice: n=6; *Ripk1*^{-/-}*Tnfr1*^{-/-} mice: n=6). (f-g) Immunofluorescence staining of Loricrin and K10 (scale bar, 100 μ m) (f) or immunohistochemical staining of F4/80, CD11b, and cleaved Caspase3 (scale bar, 50 μ m) (g) in skin sections of *Ripk1*^{K376R/+}*Tnfr1*^{-/-}, *Ripk1*^{K376R/K376R}*Tnfr1*^{-/-} and *Ripk1*^{-/-}*Tnfr1*^{-/-} mice at P11. (h) Cytokines in lung homogenates were determined with the indicated genotypes at P11 (*Ripk1*^{K376R/+}*Tnfr1*^{-/-} mice: n=4; *Ripk1*^{K376R/K376R}*Tnfr1*^{-/-} mice: n=4). (i) Flow cytometry and statistical results of CD4⁺, CD8⁺ T cells and CD19⁺B220⁺ B cells in spleen from *Ripk1*^{K376R/K376R}*Tnfr1*^{-/-} and *Ripk1*^{K376R/+}*Tnfr1*^{-/-} littermate mice at P11 (*Ripk1*^{K376R/+}*Tnfr1*^{-/-} mice: n=6; *Ripk1*^{K376R/K376R}*Tnfr1*^{-/-} mice: n=6). In (e, h, i), data are mean \pm s.e.m. Statistical significance was determined using a two-tailed unpaired *t* test, ns *P* > 0.05, **P* < 0.05, ***P* < 0.01, *****P* < 0.0001.



Supplementary Figure 5. Other signaling such as IFNs also contributes to lethality of *Ripk1*^{K376R/K376R} mice

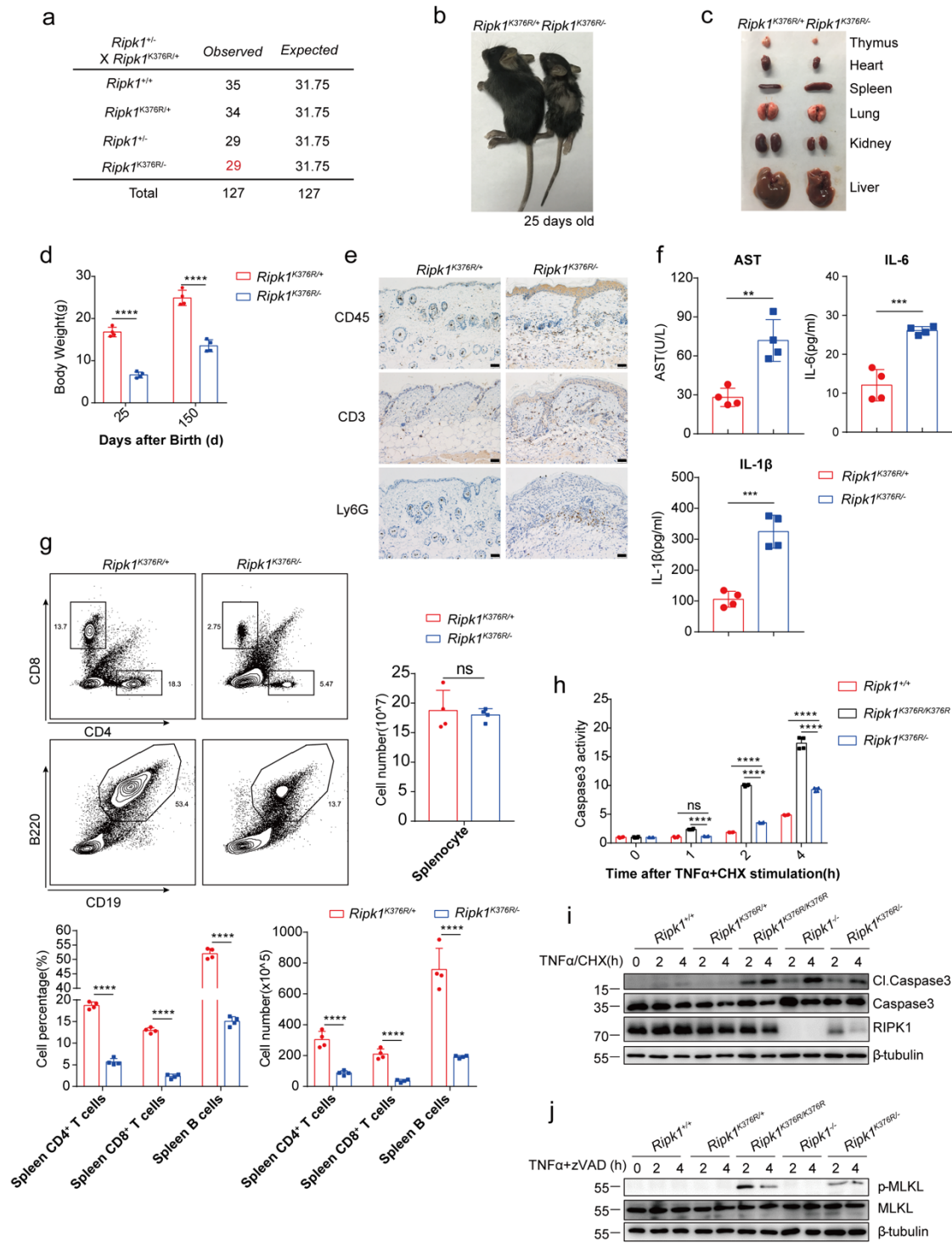
(a-d) *Ripk1*^{+/+}, *Ripk1*^{K376R/K376R} and *Ripk1*^{-/-} immortalized MEFs were treated with LPS (100 ng/ml) (a), IFN γ /zVAD (IFN γ : 10 μ g/ml; zVAD.fmk: 20 μ M) (b), TRAIL/CHX

(TRAIL:150ng/ml; CHX:10 ug/ml) (c) and FasL/CHX (FasL: 0.5ug/ml; CHX:10ug/ml) (d) for the indicated time. The cell lysates were analyzed by western-blotting using the indicated antibodies. (e) Cell death of *Ripk1*^{+/+} and *Ripk1*^{K376R/K376R} immortalized MEFs treated for 12h with different stimulators were measured by SytoxGreen positivity. FasL: 0.5ug/ml; TRAIL:150ng/ml; C: CHX (10ug/ml), the error bars represent mean±s.e.m of data from three independent cell samples for each genotype. Statistical significance was determined using a two-tailed unpaired *t* test, ***P* < 0.01, ****P* < 0.01, *****P* < 0.0001.



Supplementary Figure 6. Co-deletion of RIPK3 and Caspase8 fully rescue the phenotype of *Ripk1*^{K376R/K376R} mice

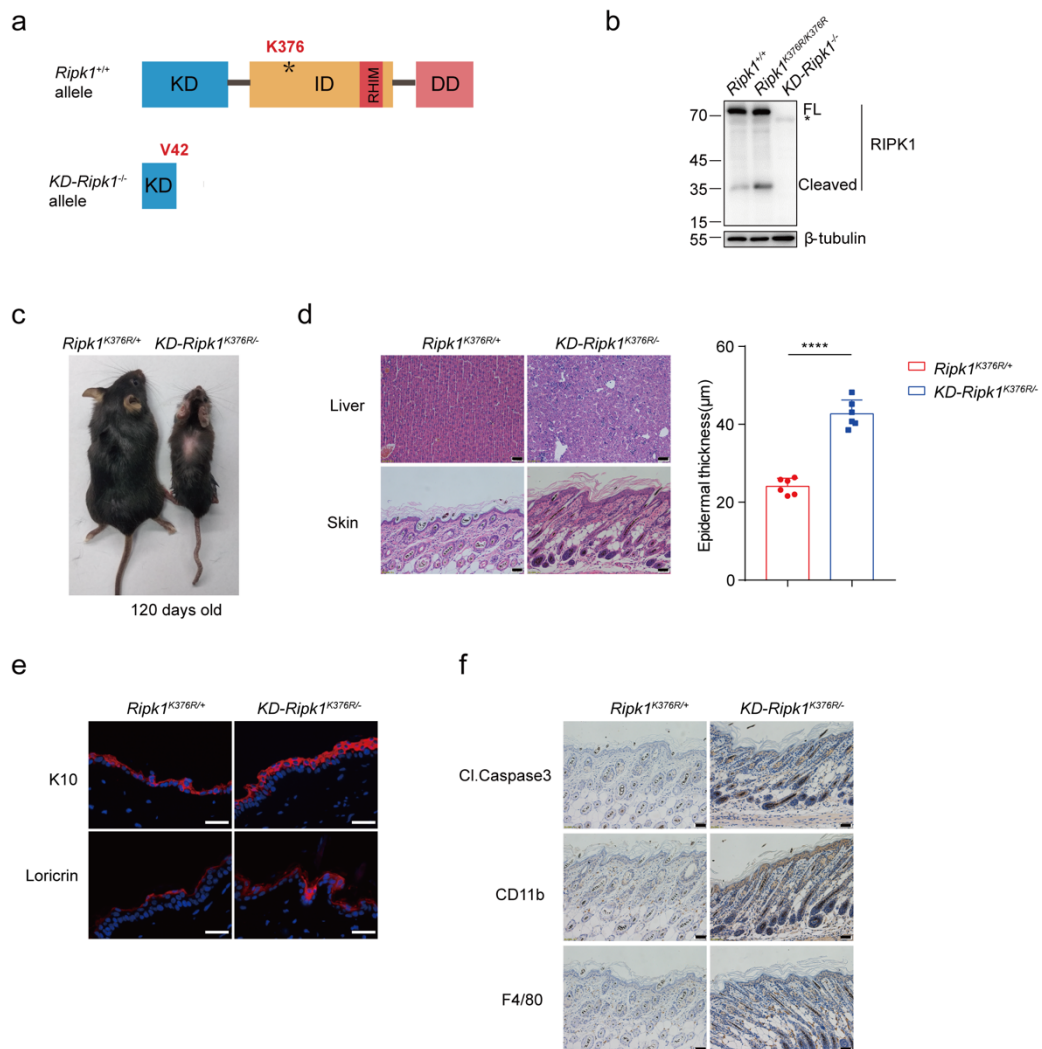
(a) Immunohistochemical staining of CD11b, CD45 and CD3 staining in skin sections of *Ripk1*^{K376R/K376R}*Ripk3*^{-/-}*Caspase8*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-}*Caspase8*^{-/-} littermate mice at P40 (scale bar, 50 μ m). (b) Flow cytometry and statistical results of CD4⁺, CD8⁺ T cells and CD19⁺B220⁺ B cells in spleen from *Ripk1*^{K376R/K376R}*Ripk3*^{-/-}*Caspase8*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-}*Caspase8*^{-/-} littermate mice at P40 (*Ripk1*^{K376R/+}*Ripk3*^{-/-}*Caspase8*^{-/-} mice: n=4; *Ripk1*^{K376R/K376R}*Ripk3*^{-/-}*Caspase8*^{-/-} mice: n=4). (c) Statistical analysis of the expected and observed offspring mice (11-days-old) from the intercrosses of *Ripk1*^{K376R/+}*Ripk3*^{-/-} mice. In (b), data are mean± s.e.m. Statistical significance was determined using a two-tailed unpaired *t* test, ns *P* > 0.05.



Supplementary Figure 7. *Ripk1*^{K376R/-} mice develops spontaneous inflammation

(a) Statistical analysis of the expected and observed offspring mice (21-days-old) from the intercrosses of *Ripk1*^{K376R/+} and *Ripk1*^{+/-} mice. (b) Representative macroscopic images of *Ripk1*^{K376R/-} and *Ripk1*^{K376R/+} littermate mice at P25. (c) Representative macroscopic images of organs with indicated genotypes at P150. (d) Statistical analysis of body weight of *Ripk1*^{K376R/+} and *Ripk1*^{K376R/-} littermate mice at P25 and P150 (*Ripk1*^{K376R/+} mice: n=4; *Ripk1*^{K376R/-} mice: n=4). (e) Immunohistochemical staining of CD45, CD3 and Ly6G in skin sections of *Ripk1*^{K376R/-} and

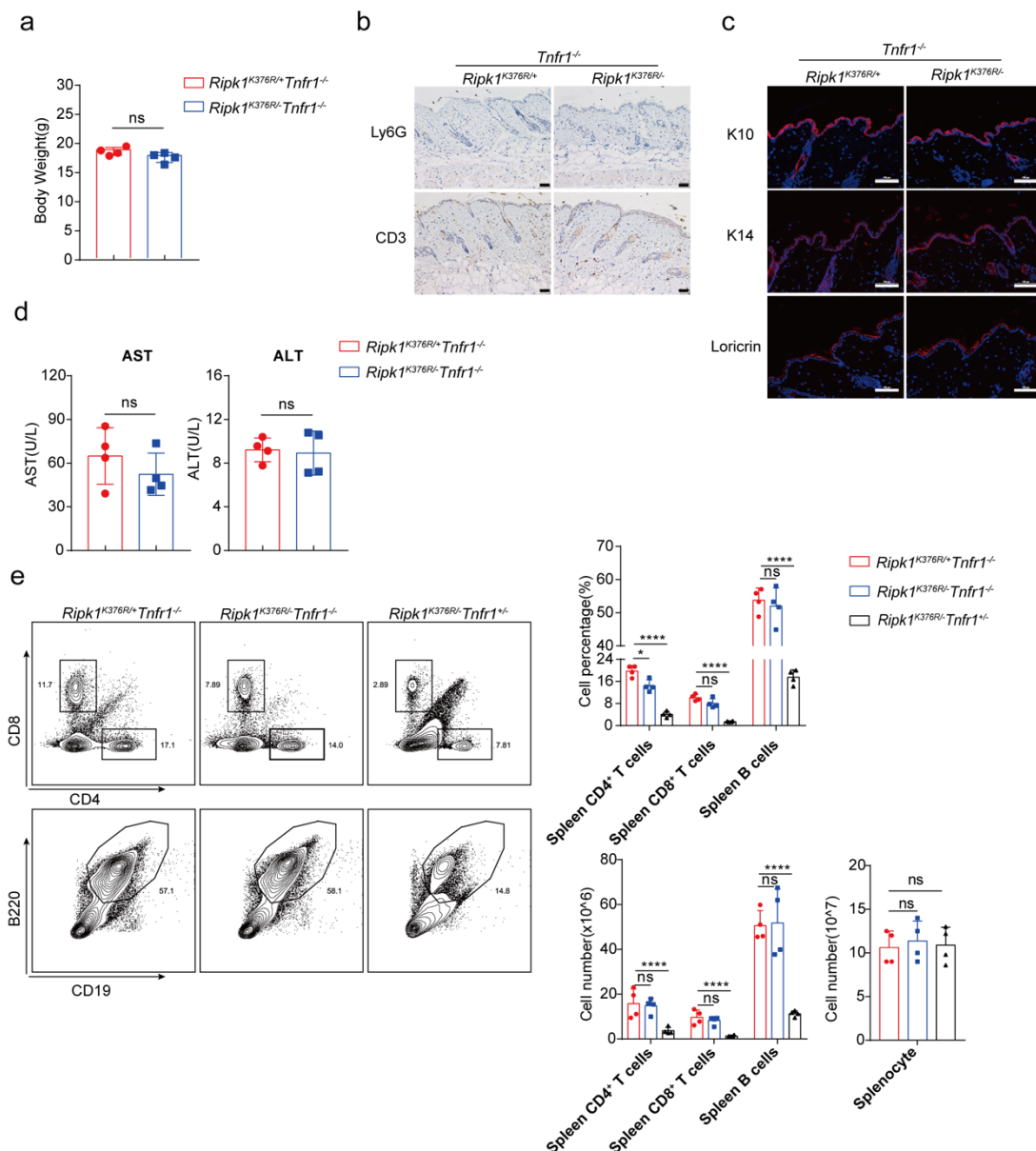
Ripk1^{K376R/+} littermate mice at P150 (scale bar, 50 μm). (f) AST in blood and cytokines in liver homogenates were determined with the indicated genotypes at P150 (*Ripk1*^{K376R/+} mice: n=4; *Ripk1*^{K376R/-} mice: n=4). (g) Flow cytometry and statistical results of CD4⁺, CD8⁺ T cells and CD19⁺B220⁺ B cells in spleen from *Ripk1*^{K376R/+} and *Ripk1*^{K376R/-} littermate mice at P150 (*Ripk1*^{K376R/+} mice: n=4; *Ripk1*^{K376R/-} mice: n=4). (h) Caspase3 activity of *Ripk1*^{+/+}, *Ripk1*^{K376R/-} and *Ripk1*^{K376R/K376R} immortalized MEFs treated with TNFα/CHX (TNFα: 20 ng/ml; CHX: 10 μg/ml) for the indicated time was measured by DEVD-AMC fluorescence, the error bars represent mean±s.e.m of data from three independent cell samples for each genotype. (i-j) *Ripk1*^{+/+}, *Ripk1*^{K376R/+}, *Ripk1*^{K376R/K376R}, *Ripk1*^{-/-} and *Ripk1*^{K376R/-} immortalized MEFs were treated with TNFα (20 ng/ml)/CHX (10 μg/ml) (i) and TNFα (20 ng/ml)/zVAD.fmk (10 μM) (j) for the indicated time, and the cell lysates were analyzed by western-blotting using the indicated antibodies. In (d-h), data are mean± s.e.m. Statistical significance was determined using a two-tailed unpaired *t* test, ns *P* > 0.05, ***P* < 0.01, ****P* < 0.001, *****P* < 0.0001.



Supplementary Figure 8. *KD-Ripk1*^{K376R/-} mice develop spontaneous inflammation

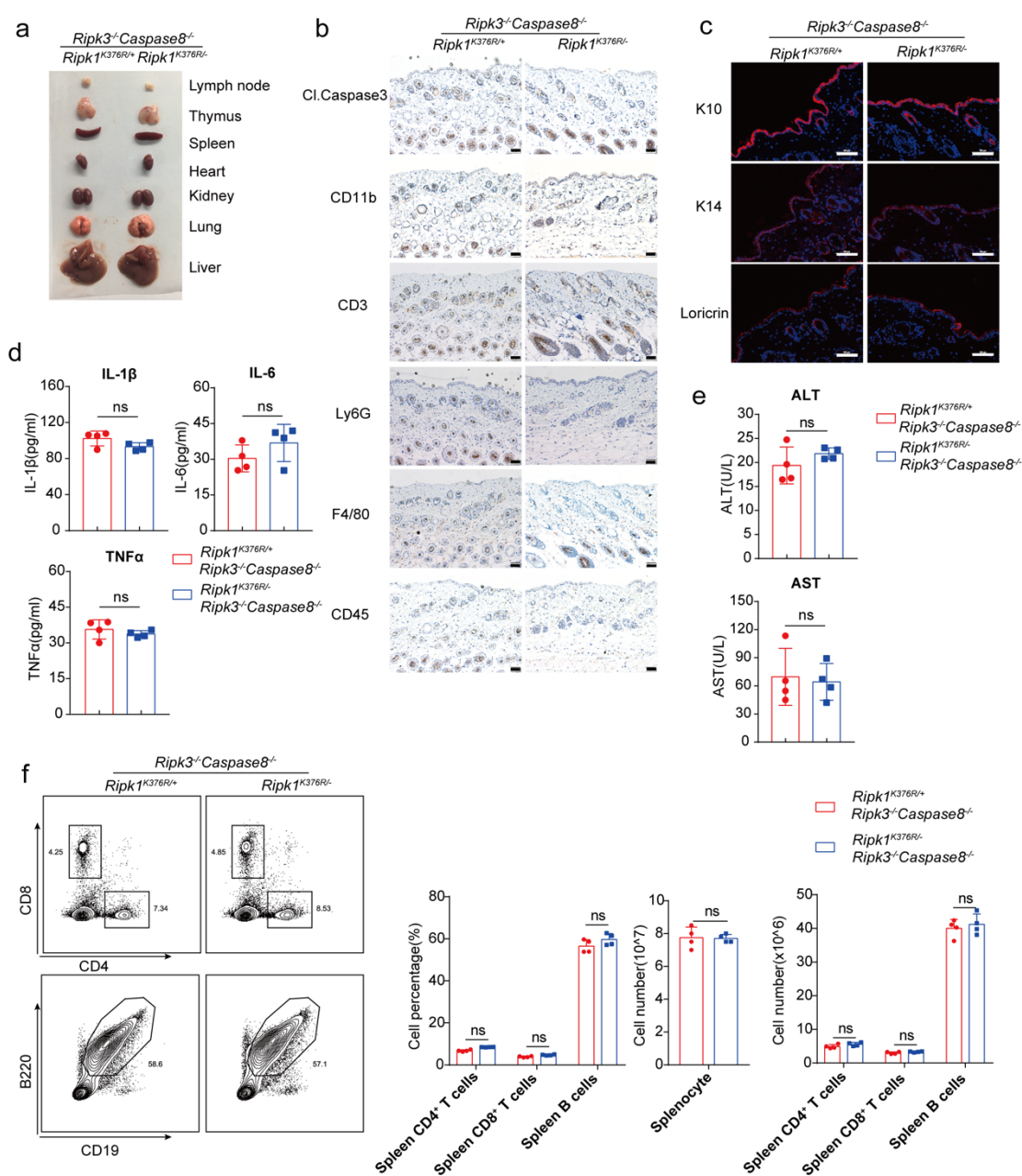
(a) Schematic overview of domain structure of *Ripk1*^{K376R/K376R} and *KD-Ripk1*^{-/-} mice. KD, kinase domain; ID, intermediate domain; DD, death domain; RHIM, RIP homotypic interaction motif. (b) Western-blotting detection of RIPK1 protein expression in primary MEFs from

Ripk1^{+/+} and *KD-Ripk1*^{-/-} mice (*refers to non-specific band). (c) Representative macroscopic images of *Ripk1*^{K376R/+} and *KD-Ripk1*^{K376R/-} littermate mice at P120. (d) H&E staining of liver and skin sections of *Ripk1*^{K376R/+} and *KD-Ripk1*^{K376R/-} littermate mice at P120 (scale bar, 50 μ m), and microscopic quantification of the epidermal thickness from H&E results (*Ripk1*^{K376R/+} mice: n=4; *KD-Ripk1*^{K376R/-} mice: n=4). (e-f) Immunofluorescence staining of Loricrin and K10 (scale bar, 100 μ m) (e) or immunohistochemical staining of F4/80, CD11b, and cleaved Caspase3 (scale bar, 50 μ m) (f) in skin sections of *Ripk1*^{K376R/+} and *KD-Ripk1*^{K376R/-} littermate mice at P120. In (d), data are mean \pm s.e.m. Statistical significance was determined using a two-tailed unpaired *t* test, ****P < 0.0001.



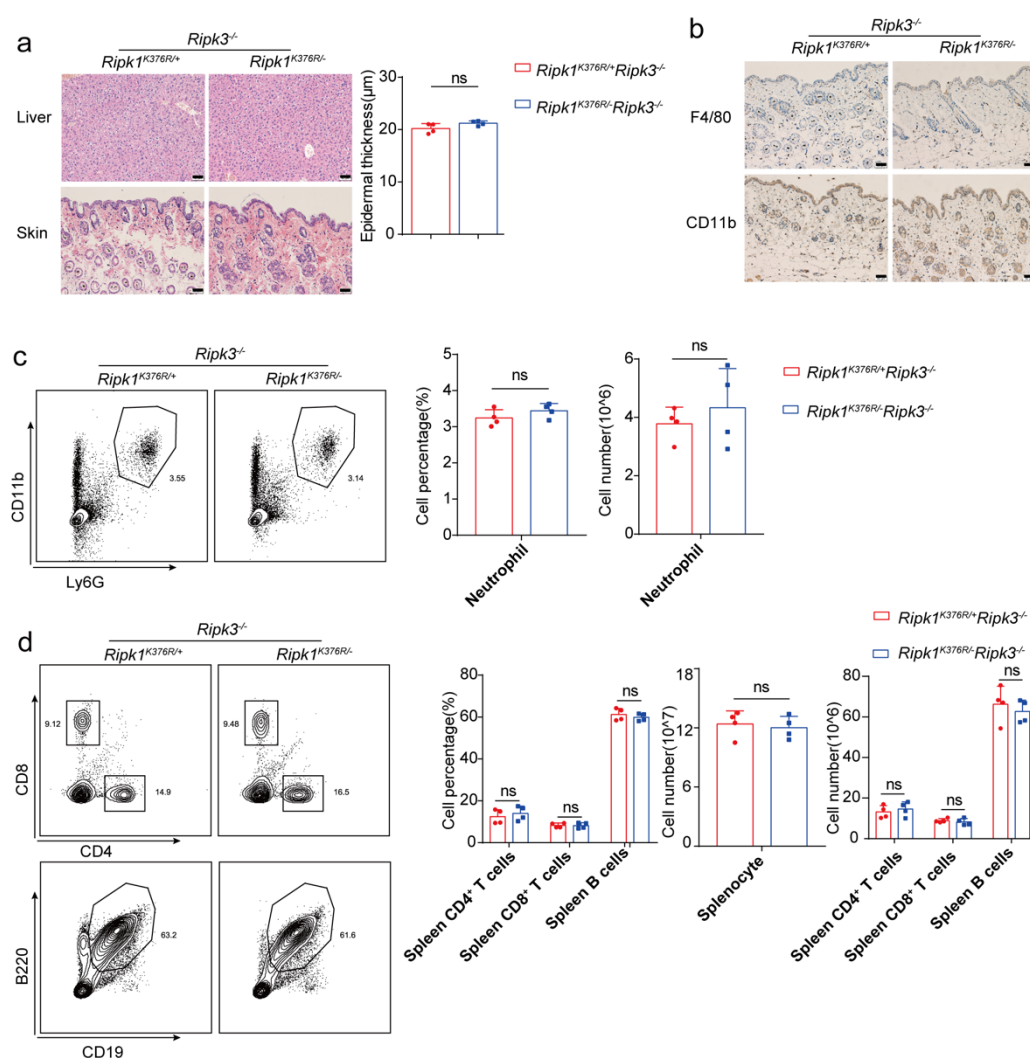
Supplementary Figure 9. TNFR1 deficiency suppress inflammation in *Ripk1*^{K376R/-} mice (a) Statistical analysis of body weight of *Ripk1*^{K376R/-}*Tnfr1*^{-/-} and *Ripk1*^{K376R/+}*Tnfr1*^{-/-} littermate mice at P40 (*Ripk1*^{K376R/+}*Tnfr1*^{-/-} mice: n=4; *Ripk1*^{K376R/-}*Tnfr1*^{-/-} mice: n=4). (b) Immunohistochemical staining of CD3 and Ly6G in skin sections of *Ripk1*^{K376R/-}*Tnfr1*^{-/-} and *Ripk1*^{K376R/+}*Tnfr1*^{-/-}

littermate mice at P40(scale bar,50 μm). (c) Immunofluorescence staining of Loricrin, K10, and K14 in skin sections of *Ripk1*^{K376R/-}*Tnfr1*^{-/-} and *Ripk1*^{K376R/+}*Tnfr1*^{-/-} littermate mice at P40(scale bar,100 μm). (d) AST and ALT in blood were determined with the indicated genotypes at P40(*Ripk1*^{K376R/+}*Tnfr1*^{-/-} mice: n=4; *Ripk1*^{K376R/-}*Tnfr1*^{-/-} mice: n=4). (e) Flow cytometry and statistical results of CD4⁺, CD8⁺ T cells and CD19⁺B220⁺ B cells in spleen from *Ripk1*^{K376R/-}*Tnfr1*^{-/-}, *Ripk1*^{K376R/-}*Tnfr1*^{+/-} and *Ripk1*^{K376R/+}*Tnfr1*^{-/-} littermate mice at P150(*Ripk1*^{K376R/+}*Tnfr1*^{-/-} mice: n=4; *Ripk1*^{K376R/-}*Tnfr1*^{+/-} mice: n=4; *Ripk1*^{K376R/-}*Tnfr1*^{-/-} mice: n=4). In (a, d, e), data are mean \pm s.e.m. Statistical significance was determined using a two-tailed unpaired *t* test, ns *P* > 0.05, **P* < 0.05, ***P* < 0.01, *****P* < 0.0001.



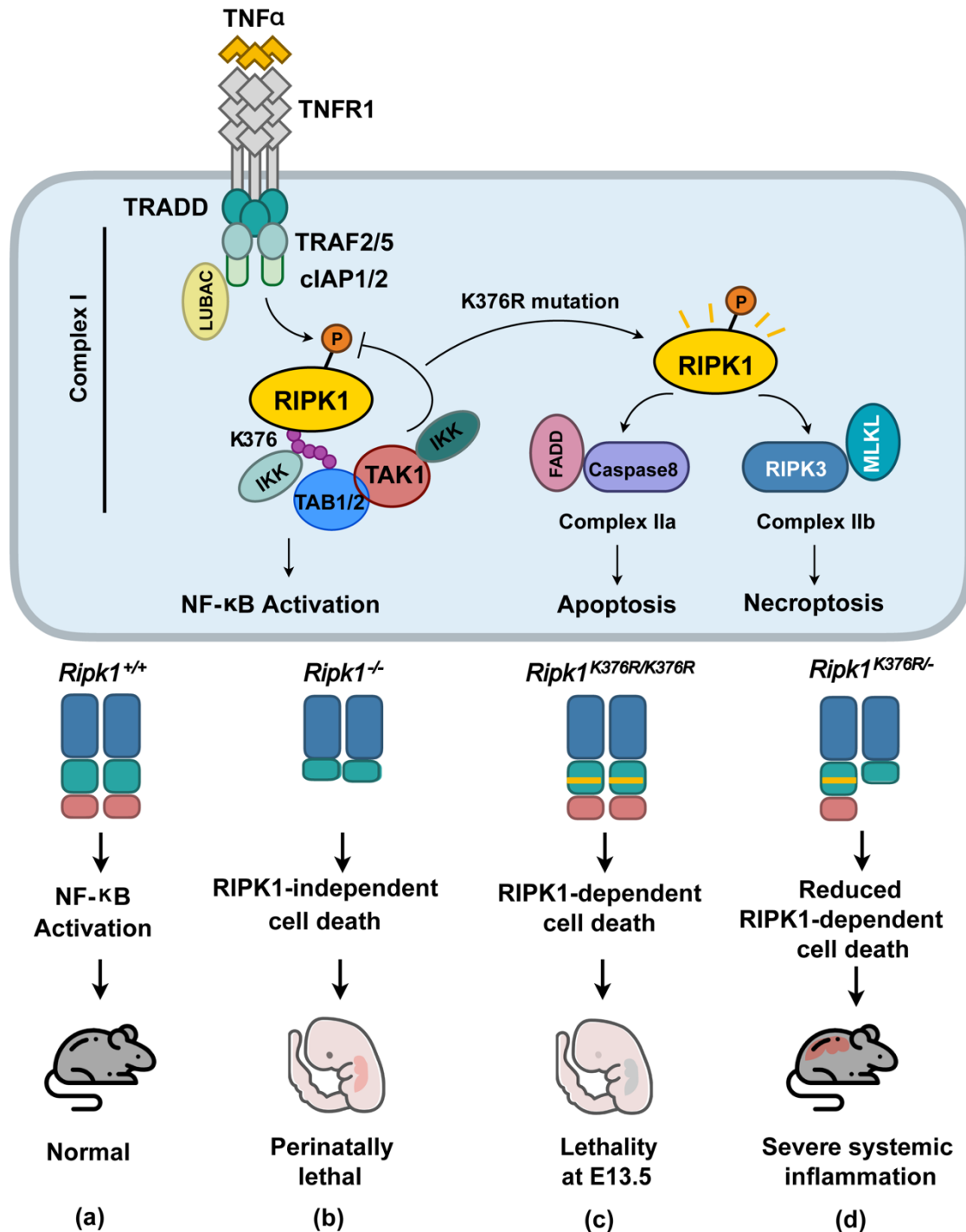
Supplementary Figure 10. Co-deletion of RIPK3 and Caspase8 suppress inflammation in *Ripk1*^{K376R/-} mice

(a) Representative macroscopic images of organs with indicated genotypes at P40. (b) Immunohistochemical staining of cleaved Caspase3, CD11b, CD3, Ly6G, F4/80 and CD45 in skin sections of *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} littermate mice at P40(scale bar,50 μ m). (c) Immunofluorescence staining of Loricrin, K10, and K14 in skin sections of *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} littermate mice at P40(scale bar,100 μ m). (d) Cytokines in liver homogenates were determined with the indicated genotypes at P40 (*Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} mice: n=4; *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} mice: n=4). (e) AST and ALT in blood were determined with the indicated genotypes at P40(*Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} mice: n=4; *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} mice: n=4). (f) Flow cytometry and statistical results of CD4⁺, CD8⁺ T cells and CD19⁺B220⁺ B cells in spleen from *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} littermate mice at P40(*Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} mice: n=4; *Ripk1*^{K376R/+}*Ripk3*^{-/-} *Caspase8*^{-/-} mice: n=4). In (d, e, f), data are mean \pm s.e.m. Statistical significance was determined using a two-tailed unpaired *t* test, ns *P* > 0.05.



Supplementary Figure 11. RIPK3 deletion suppress inflammation in *Ripk1*^{K376R/-} mice
 (a) H&E staining of liver and skin sections of *Ripk1*^{K376R/-}*Ripk3*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-} littermate mice at P40(scale bar,50 μ m), and microscopic quantification of the epidermal

thickness from H&E results (*Ripk1*^{K376R/+}*Ripk3*^{-/-} mice: n=4; *Ripk1*^{K376R/-}*Ripk3*^{-/-} mice: n=4). **(b)** Immunohistochemical staining of CD11b and F4/80 in skin sections of *Ripk1*^{K376R/-}*Ripk3*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-} littermate mice at P40 (scale bar, 50 μ m). **(c)** Flow cytometry and statistical results of splenocytes stained with Ly6G and CD11b from *Ripk1*^{K376R/-}*Ripk3*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-} littermate mice at P40 (*Ripk1*^{K376R/+}*Ripk3*^{-/-} mice: n=4; *Ripk1*^{K376R/-}*Ripk3*^{-/-} mice: n=4). CD11b⁺Ly6G⁺ cells were identified as neutrophils. **(d)** Flow cytometry and statistical results of CD4⁺, CD8⁺ T cells and CD19⁺B220⁺ B cells in spleen from *Ripk1*^{K376R/-}*Ripk3*^{-/-} and *Ripk1*^{K376R/+}*Ripk3*^{-/-} littermate mice at P40 (*Ripk1*^{K376R/+}*Ripk3*^{-/-} mice: n=4; *Ripk1*^{K376R/-}*Ripk3*^{-/-} mice: n=4). In **(a, c, d)**, data are mean \pm s.e.m. Statistical significance was determined using a two-tailed unpaired *t* test, ns *P* > 0.05.

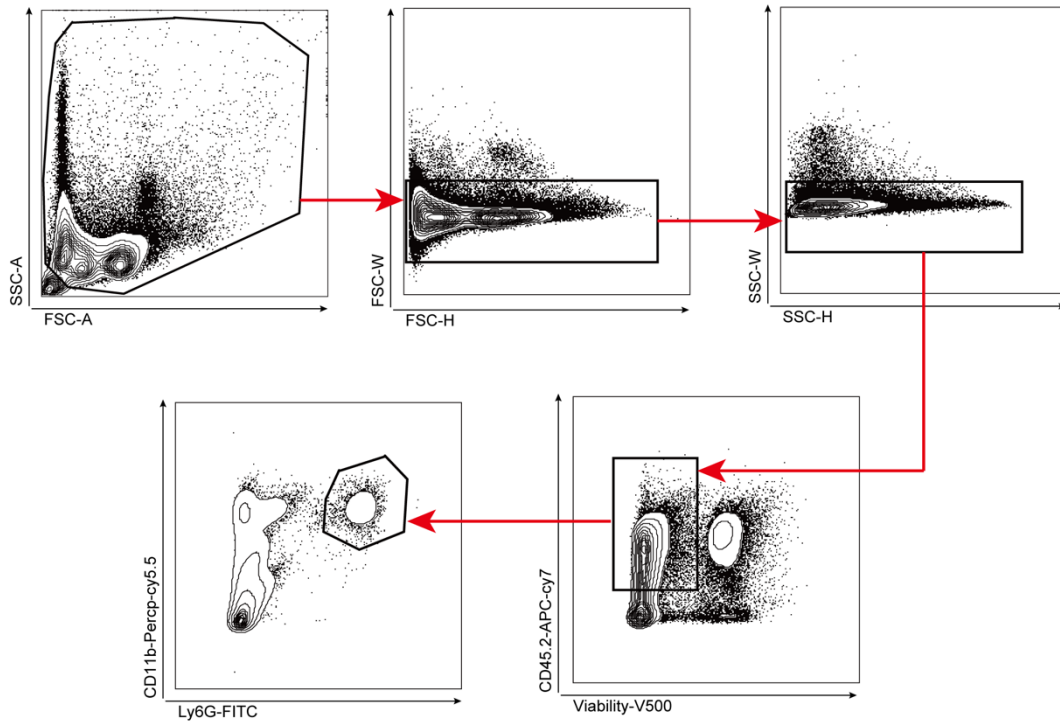


Supplementary Figure 12. Proposed model that K63-linked ubiquitination of RIPK1 on K376 regulating TNF α -induced signaling

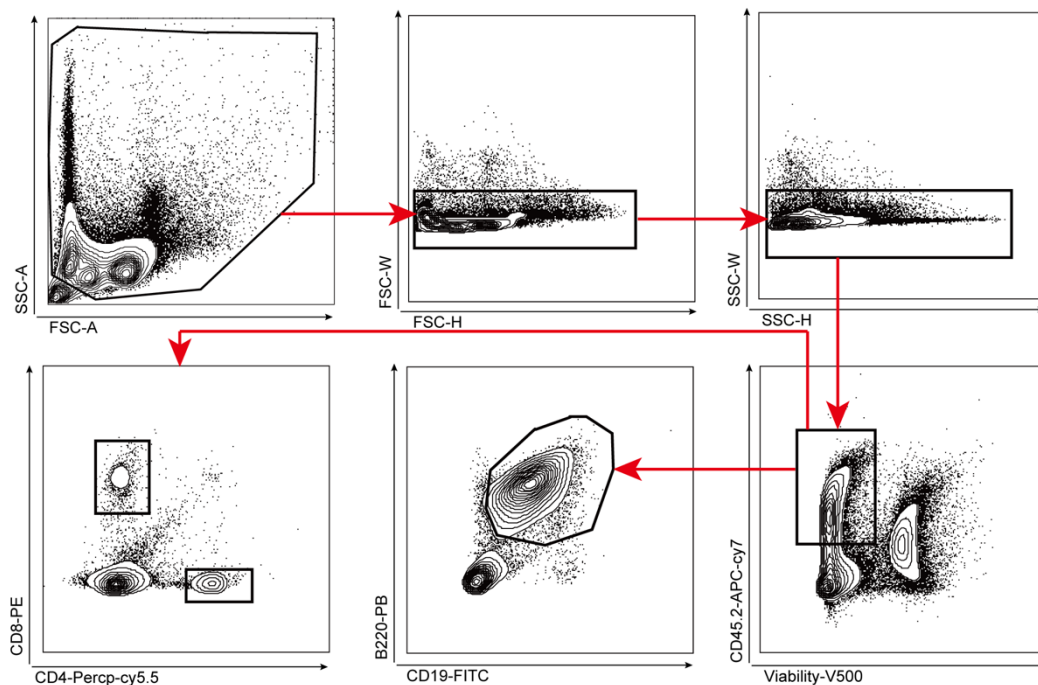
(a) In normal condition, K63-linked ubiquitination of RIPK1 on K376 can recruit TAK1/IKK to suppress RIPK1 kinase activity, and also recruit IKK complex to promote NF- κ B activation. These two signaling can both contribute to survival of *Ripk1*^{+/+} mice. (b) Deficiency of RIPK1 can induce RIPK1-independent cell death, resulting in perinatally lethal of *Ripk1*^{-/-} mice. (c) K376R mutation can enhance RIPK1 kinase activity to promote RIPK1-dependent cell death, resulting in embryonic lethality of *Ripk1*^{K376R/K376R} mice at E13.5. (d) In *Ripk1*^{K376R/-} cells,

RIPK1 kinase activity is relatively decreased comparing to *Ripk1*^{K376R/K376R} cells, resulting in reduced RIPK1-dependent cell death, and *Ripk1*^{K376R/-} could survive but develop severe systemic inflammation.

a



b



Supplementary Figure 13. Hematopoietic gating strategy

(a) For characterization of spleen neutrophils, the living cell fractions gated from preliminary FSC/SSC gates could be further divided into CD45⁺CD11b⁺Ly6G⁺ neutrophils. The

antibodies and fluorochrome used described as "Antibodies and reagents" in the methods.
 (b) For characterization of spleen T cells, B cells, the living cell fractions gated from preliminary FSC/SSC gates could be further divided into three cell types: CD45+CD4+ T cells, CD45+CD8+ T cells, CD45+CD19+B220+ B cells. The antibodies and fluorochrome used described as "Antibodies and reagents" in the methods.

Supplementary Table

Table 1. Primer sequences for generation of *Ripk1*^{K376R/K376R} and *KD-Ripk1*^{-/-} mice.

sgRNA(<i>Ripk1</i> ^{K376R/K376R})	tgtgcaggctaagctgcaag
Donor(<i>Ripk1</i> ^{K376R/K376R})	gtcctggtttctctccccagagtaccacaggacgagaatgatcgcagtggtgcaggccagactacag gaagaagccagctatcatgcttttggaatattgcagagaaacagacaaa
sgRNA(<i>KD-Ripk1</i> ^{-/-})	gccctgtgataactttttc

Table 2. The primer sequences for genotyping-PCR.

Gene	Forward Primer (5'--3')	Reverse Primer (5'--3')
Primer sequences for genotyping-PCR		
RIPK1 K376R WT	tgcaggctaagctgcaagag	gtgctgggatcagaatgacc
RIPK1 K376R KI	tgcaggccagactacaggaa	gtgctgggatcagaatgacc
RIPK1 ID-KO	ttctccccagagtaccac	gtgctgggatcagaatgacc
RIPK1 KD-KO	gtggagtacaagctagcctcagac	ctgtgttagccacacagatg

Table 3. The primer sequences for qRT-PCR.

Gene	Forward Primer (5'--3')	Reverse Primer (5'--3')
Primer sequences for qRT-PCR		
GAPDH	AACAGCAACTCCCCTCTTC	CCTGTTGCTGTAGCCGTATT
IL6	TCCATCCAGTTGCCTTCTTG	GGTCTGTTGGGAGTGGTATC
TNF α	CTACCTTGTTGCCTCCTCTTT	GAGCAGAGGTTTCAGTGATGTAG
I κ B α	TGAAGGACGAGGAGTACGAGC	TTCGTGGATGATTGCCAAGTG
CXCL1	CTGGGATTCACCTCAAGAACATC	CAGGGTCAAGGCAAGCCTC

CXCL2	CCAACCACCAGGCTACAGG	GCGTCACACTCAAGCTCTG
IFN β	CAGCTCCAAGAAAGGACGAAC	GGCAGTGTA ACTCTTCTGCAT
IFN γ	CTTTGGACCCTCTGACTTGAG	TCAATGACTGTGCCGTGG
A20	ACAGTGGACCTGGTAAGAAAACA	CCTCCGTGACTGATGACAAGAT