1	Supplementary Information
2	Discovering genetic interactions bridging pathways in genome-wide association studies
3	Fang and Wang et.al.
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 $8 \qquad \ \ \, ^{\star}$ Binarization density is determined based on a pilot run (See method)

- 9 Supplementary Figure 1: Detailed view of the BridGE method for detecting genetic interactions from
- 10 GWAS data.



12 13 Supplementary Figure 2: Distribution of p-values from individual tests for pairwise SNP-SNP interactions for discovered Parkinson's disease BPM. SNP pairs supporting the pathway-pathway interaction between 14 the Golgi associated vesicle biogenesis gene set (Reactome) and Fc epsilon receptor I signaling pathway 15 16 (KEGG) discovered from the PD-NIA Parkinson's disease cohort were evaluated for association with PD based on a recessive and dominant disease model. The distribution of maximum -log₁₀ hypergeometric 17 18 test p-value of the two models for each SNP pair is plotted. None of the SNP pairs are significant after 19 multiple hypothesis correction (dashed line at the most significant SNP-SNP pair corresponds to 20 FDR=0.94).





interaction effect size, and sample size on the discovery of between-pathway interactions. The plot is
 same as Fig. 6, but the biological densities used are 2.5% (a) and 10% (b).



27



discovery of between-pathway interactions. The BPM significance $(-log_{10} \text{ p-value derived from 150,000})$

30 SNP permutations) is plotted for 100 embedded BPMs of different sizes and SNP-SNP interaction

31 densities (online method). The gray plane indicates the p-value cutoff corresponding to the average SNP

32 permutation p-values ($p = 3.0 \times 10^{-5}$) of the significant BPM discoveries across all GWAS cohorts

 $(FDR \le 0.25)$. Bars exceeding this plane represent BPMs that would have been discovered in this cohort

34 and provide an estimate of sensitivity of the approach.



37 Supplementary Figure 5: Distribution of sizes for discovered BPMs. The size of each candidate BPM was

38 measured as the total number of possible SNP-SNP pairs between the two pathways. The distribution of

39 sizes of all possible pathway-pathway pairs is plotted in (a) and only significant BPMs (FDR ≤ 0.25) from

40 the PD-NIA cohort are plotted in (b). BPMs discovered by BridGE span a large range of sizes.





43 Supplementary Figure 6: Comparison of false discovery rates derived from 10 sample permutations vs.

44 1000 sample permutations using PD-NIA dataset. BPMs that are significant (FDR \leq 0.25) based on either

- 45 10 sample permutations or 1000 permutations were plotted to show the agreement between two
- 46 permutations.





49 Supplementary Figure 7: Power simulation of the effect of sample size, interaction effect size (IE) and

50 minor allele frequency on the discovery of SNP-SNP interactions0, The discovery rates of 100 embedded

51 SNP-SNP interactions in the synthetic datasets with different sample sizes were plotted and colored with

52 corresponding interaction effect size. Each subplot is corresponded to a different minor allele frequency

53 assumption: (a) MAF=0.05, (b) MAF=0.1, (c) MAF=0.15, (d) MAF=0.2, (e) MAF=0.25.

54 Supplementary Discussion

55 BridGE results on hypertension and type2 diabetes

56 Although we did not conduct replication analyses for hypertension or type 2 diabetes, we found that many

of the pathways involved in interactions from the discovery cohorts were also highly relevant to the

corresponding disease. For example, in the hypertension cohort, we identified a risk-associated BPM

59 interaction involving hypoxia inducible factor (HIF) signaling, whose aberrant expression has been

60 previously associated with hypertension¹. Two BPMs and one WPM, all associated with increased risk,

61 involved the Rho cell motility signaling pathway, which has been previously implicated in the

62 pathogenesis of hypertension². For type 2 diabetes, we discovered BPMs associated with protective

63 effects involving an autoimmune thyroid disease gene set, glycosaminoglycan biosynthesis, and the

64 mTOR signaling pathway, all of which have strong links to diabetes^{3,4,5}.

65

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- 68 The genome-wide association datasets (PD-NIA, PD-NGRC, SZ-GAIN, BC-CGEMS-EUR, BC-MCS-
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- 70 were obtained from <u>https://www.ncbi.nlm.nih.gov/gap</u> through dbGaP accession numbers:
- 71 phs000089.v3.p2, phs000196.v3.p1, phs000021.v3.p2, phs000147.v3.p1, phs000517.v3.p1,
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- 80 **PD-NIA (phs000089.v3.p2):**
- 81 The genotyping of samples was provided by the National Institute of Neurological Disorders and Stroke
- 82 (NINDS). The dataset used for the analyses described in this manuscript were obtained from the NINDS
- 83 Database found at <u>https://www.ncbi.nlm.nih.gov/gap</u>

84 **PD-NGRC** (phs000196.v3.p1):

This work utilized in part data from the NINDS DbGaP database from the CIDR:NGRC PARKINSON'S
DISEASE STUDY.

87 SZ-GAIN (phs000021.v3.p2):

- 88 Funding support for the Genome-Wide Association of Schizophrenia Study was provided by the National
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- 91 MH81800, U01 MH46276, U01 MH46289 U01 MH46318, U01 MH79469, and U01 MH79470) and the
- 92 genotyping of samples was provided through the Genetic Association Information Network (GAIN). The
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- 94 and Phenotypes (dbGaP) found at http://www.ncbi.nlm.nih.gov/gap through dbGaP accession number
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98 BC-CGEMS-EUR (phs000147.v3.p1):

- 99 This dataset was from the Cancer Genetic Markers of Susceptibility (CGEMS) Breast Cancer Genome-
- 100 wide Association Study with dbGaP accession number phs000147.v3.p1.

101 BC-MCS-LTN, BC-MCS-JPN (phs000517.v3.p1):

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105 HT-eMERGE (phs000297.v1.p1):

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146 **ProC-CGEMS (phs000207.v1.p1):**

147 This data was from the Cancer Genetic Markers of Susceptibility (CGEMS) Prostate Cancer Genome-

148 Wide Association Study.

149 **ProC-BPC3** (phs000812.v1.p1):

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