

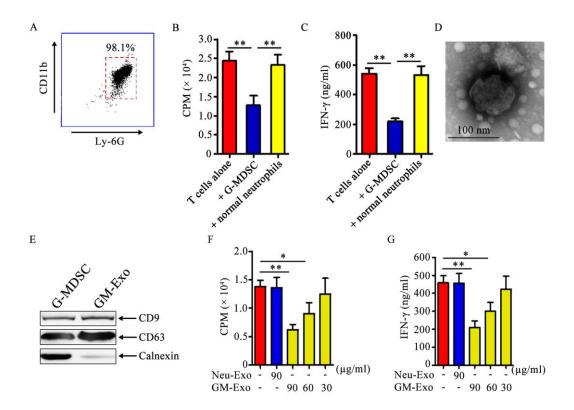
## **Supporting Information**

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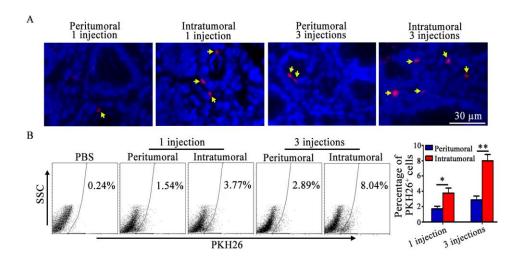
Granulocytic Myeloid-Derived Suppressor Cells Promote the Stemness of Colorectal Cancer Cells through Exosomal S100A9

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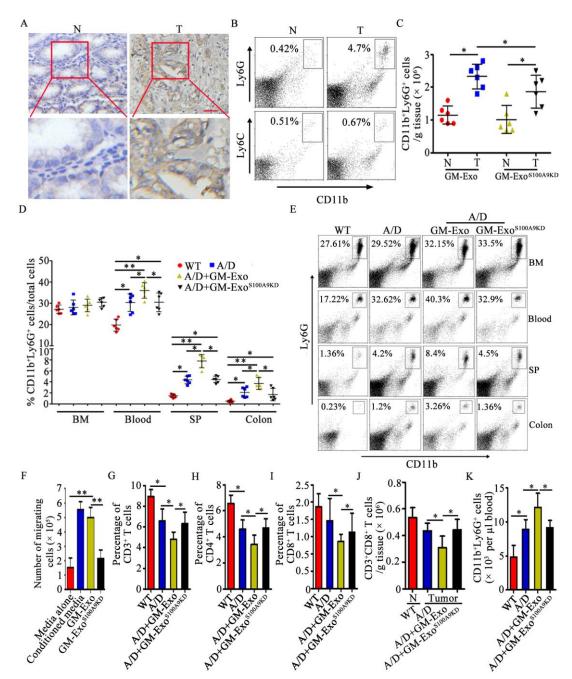
## **Supporting Information**



Supplementary figure 1. Extraction and identification of GM-Exo. A, Representative micrograph of CT-26 cells (magnification, 200×). **B**, Representative image of CT-26 tumor-bearing mice. C, Immunomagnetic beads were used to sort G-MDSCs from the spleens of CT-26-bearing mice. Ly-6G and CD11b were analyzed by FCM. **D**, G-MDSCs suppressed CD4<sup>+</sup> T cell proliferation. The counts per min (CPM) values of various wells were detected with a [<sup>3</sup>H]-thymidine incorporation experiment after the coculture of CD4<sup>+</sup> T cells and G-MDSCs. E, G-MDSCs suppressed IFN-γ secretion from CD4<sup>+</sup> T cells. The IFN-γ contents in the culture supernatants were detected by ELISA. F, Representative transmission electron micrograph of GM-Exo (magnification, 135,000×). G, The CD9, CD63, and calnexin expression levels of GM-Exo were detected by western blotting. **H**, GM-Exo suppressed CD4<sup>+</sup> T cell proliferation. The CPM values of various wells were detected with a [3H]-thymidine incorporation experiment after the culture of CD4<sup>+</sup> T cells in the presence of GM-Exo. **I,** GM-Exo suppress IFN-γ secretion from CD4<sup>+</sup> T cells. The IFN- $\gamma$  contents in culture supernatants were detected by ELISA. The data shown in A, B, C, F, and G were from one of three independent experiments. The data are shown as the mean  $\pm$  SEM of each group (n = 3) pooled from three independent experiments. \*p < 0.001, \*\*p < 0.01, analyzed by ANOVA.



**Supplementary figure 2.** Biodistribution of exogenous GM-Exo in colorectal tissues from CAC mice. **A,** Images of colorectal tissues from fluorescence microscopy. The arrows indicate exosomes foci. The immunofluorescence images are representative of six random fields. **B,** The percentage of GM-Exo-positive colorectal tissue cells was quantitated by FCM (n = 6). The data are shown as the mean  $\pm$  SEM of each group pooled from three independent experiments.\* p < 0.05; \*\* p < 0.01, analyzed by a t-test.



**Supplementary figure 3.** Exosomal S100A9 from G-MDSCs participates in immunosuppression through mobilizing G-MDSCs to the peripheral blood and colorectal tissues. **A,** Representative images of immunohistochemical staining for S100A9 in tumors (T) and adjacent tissues (N) are shown. Scale bar = 100 μm. **B,** Representative results for the percentages of MDSC subpopulations in peritumoral (N) and intratumoral (T) colorectal tissues from CAC mice. **C,** The quantity of G-MDSCs in colorectal tissues was represented as the number of G-MDSCs per gram. **D** and **E,** The percentages of G-MDSCs in the bone marrow (BM), peripheral blood (PB), spleen (SP), and colorectal tissues of mice in different groups were analyzed by FCM (**D**). Representative diagrams are shown (**E**). **F,** Exosomal S100A9 from G-MDSCs was chemotactic for G-MDSCs. G-MDSCs were placed in the upper compartment of a transwell, and tumor-conditioned medium, GM-Exo or GM-Exo<sup>S100A9KD</sup> were placed in the lower compartment. The number of G-MDSCs migrating to the lower compartment was determined. **G,** G-MDSC quantification in mouse blood

from different groups (n=6). **H-J,** The percentages of T cells and subsets in mouse blood (n = 6) from different groups were determined by FCM. **K,** The numbers of CD3<sup>+</sup>CD8<sup>+</sup>T cells in colorectal tissues from mice in different groups. The data are presented as the mean  $\pm$  SEM of each group pooled from three independent experiments. \*p < 0.05, \*\*p < 0.01, analyzed by ANOVA.