

Supplementary Materials for

Mechanical regulation of bone homeostasis through p130Cas-mediated alleviation of NF- κ B activity

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Published 25 September 2019, *Sci. Adv.* **5**, eaau7802 (2019)

DOI: 10.1126/sciadv.aau7802

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Figure S1

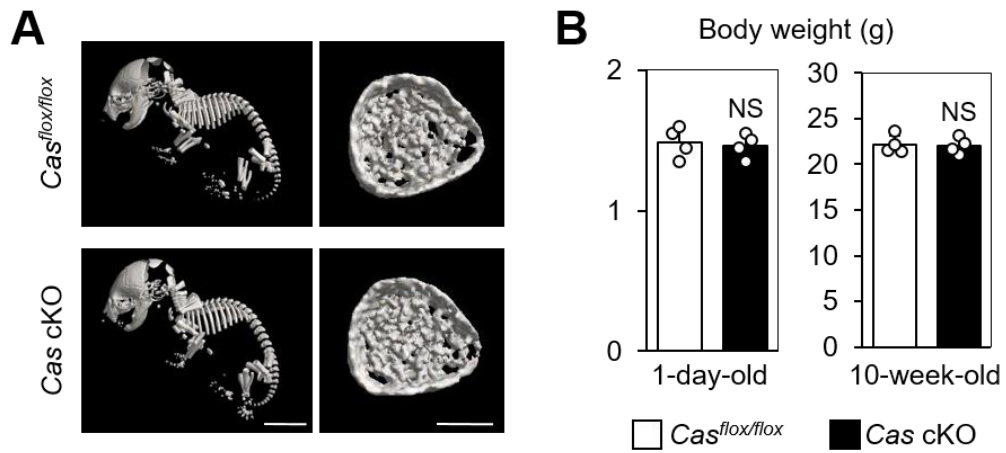


Fig. S1. Skeletal phenotype of *Cas* cKO mice is not distinct at birth. (A) μ CT analysis of *Cas* cKO

newborn mice and their normal *Cas^{flox/flox}* littermates (postnatal day 1, male or female). Left: whole skeleton, right: axial view of metaphyseal femur. Scale bars, 5 mm (left) and 250 μ m (right). (B) Body weight of *Cas* cKO mice and their normal *Cas^{flox/flox}* littermates at 1 day (male or female) or 10 weeks of age (male) (n = 4 mice per group).

Figure S2

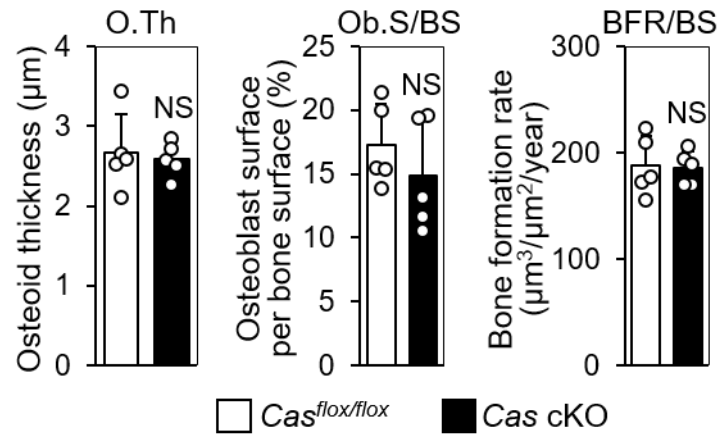


Fig. S2. Bone formation parameters are not significantly altered in *Cas cKO* mice. Parameters for osteoblastic bone formation in *Cas cKO* mice and their normal *Cas^{flox/flox}* littermates were determined by histomorphometric analysis (n = 5 mice per group).

Figure S3

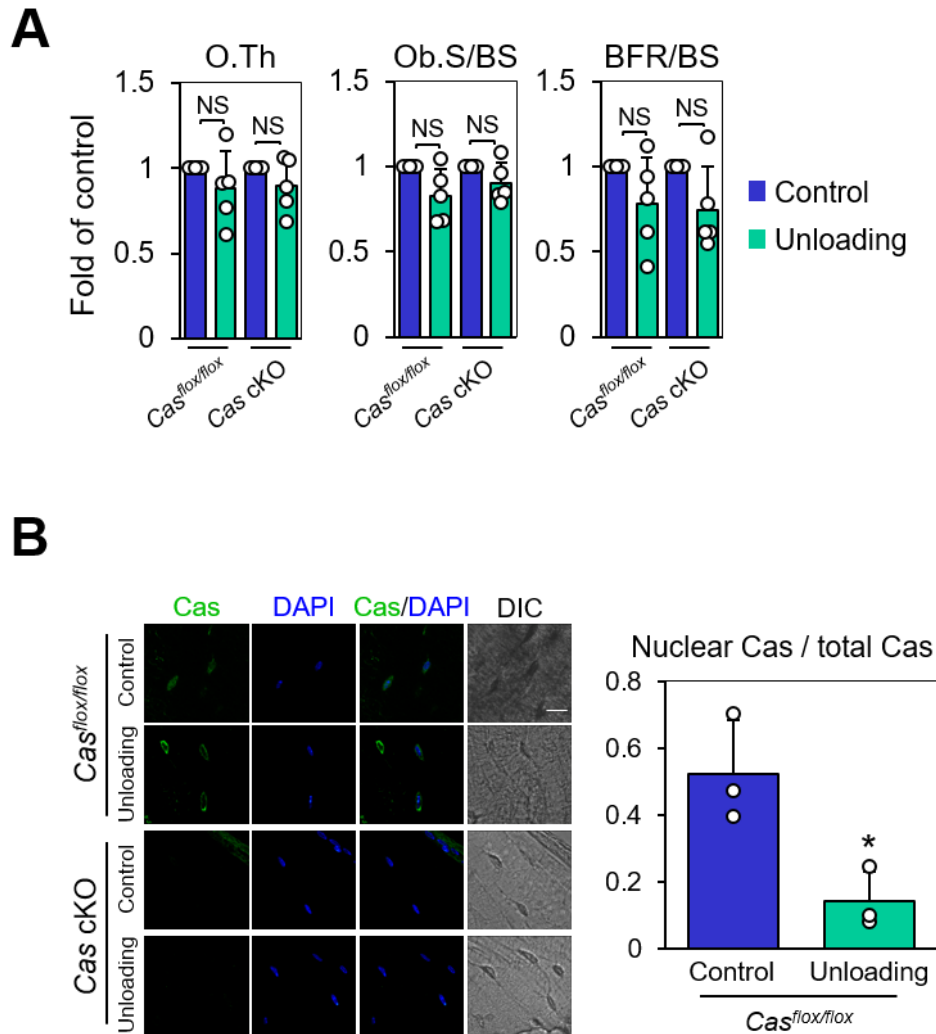


Fig. S3. Mechanical regulation of bone formation in control and *Cas cKO* mice and nuclear distribution

of Cas in their osteocytes. (A) Bone formation parameters of bones with and without unloading. Values from

unloaded bones were normalized against data from their contralateral controls as in Fig. 3B (n = 5 mice per

group). (B) Mechanical loading-dependent nuclear distribution of Cas in osteocytes. Nuclear distribution of

Cas was analyzed as in Fig. 1B (n = 3 mice per group, each value was averaged from 4 - 6 osteocytes analyzed in each bone).

Figure S4

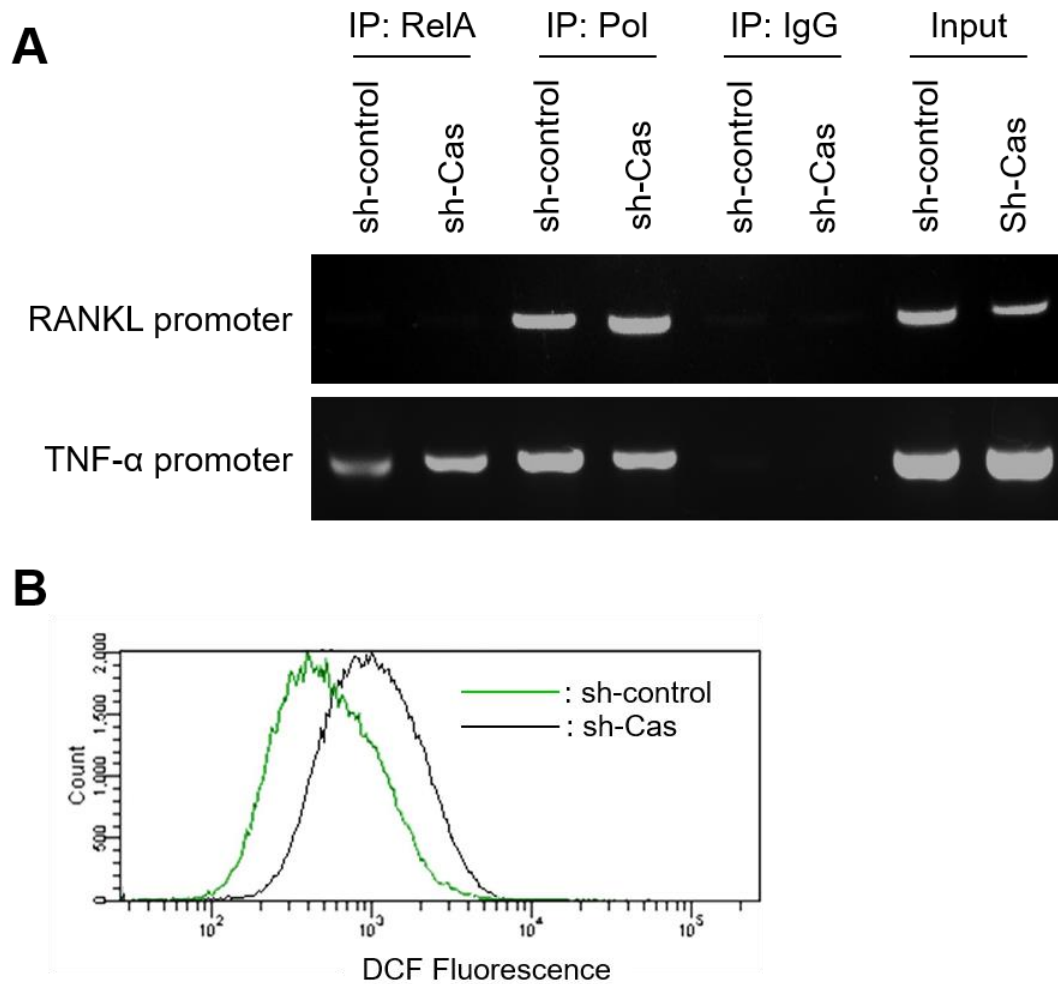


Fig. S4. ChIP analysis of RelA binding to the RANKL promoter region and measurement of intracellular ROS level. (A) Absence of detectable RelA binding to the RANKL promoter region in MLO-Y4 osteocytes. ChIP analysis of MLO-Y4 osteocytes was conducted using anti-RelA, anti-polymerase III (Pol, positive control) or anti-IgG (negative control) antibodies. DNA recovered from immunoprecipitates was amplified by PCR using the primers specific for RANKL and TNF- α promoters. (B) Measurement of

intracellular ROS level in MLO-Y4 osteocytes with (sh-Cas) and without (sh-control) Cas knockdown by flow cytometry using 2',7'-dichlorodihydrofluorescein (DCF).

Figure S5

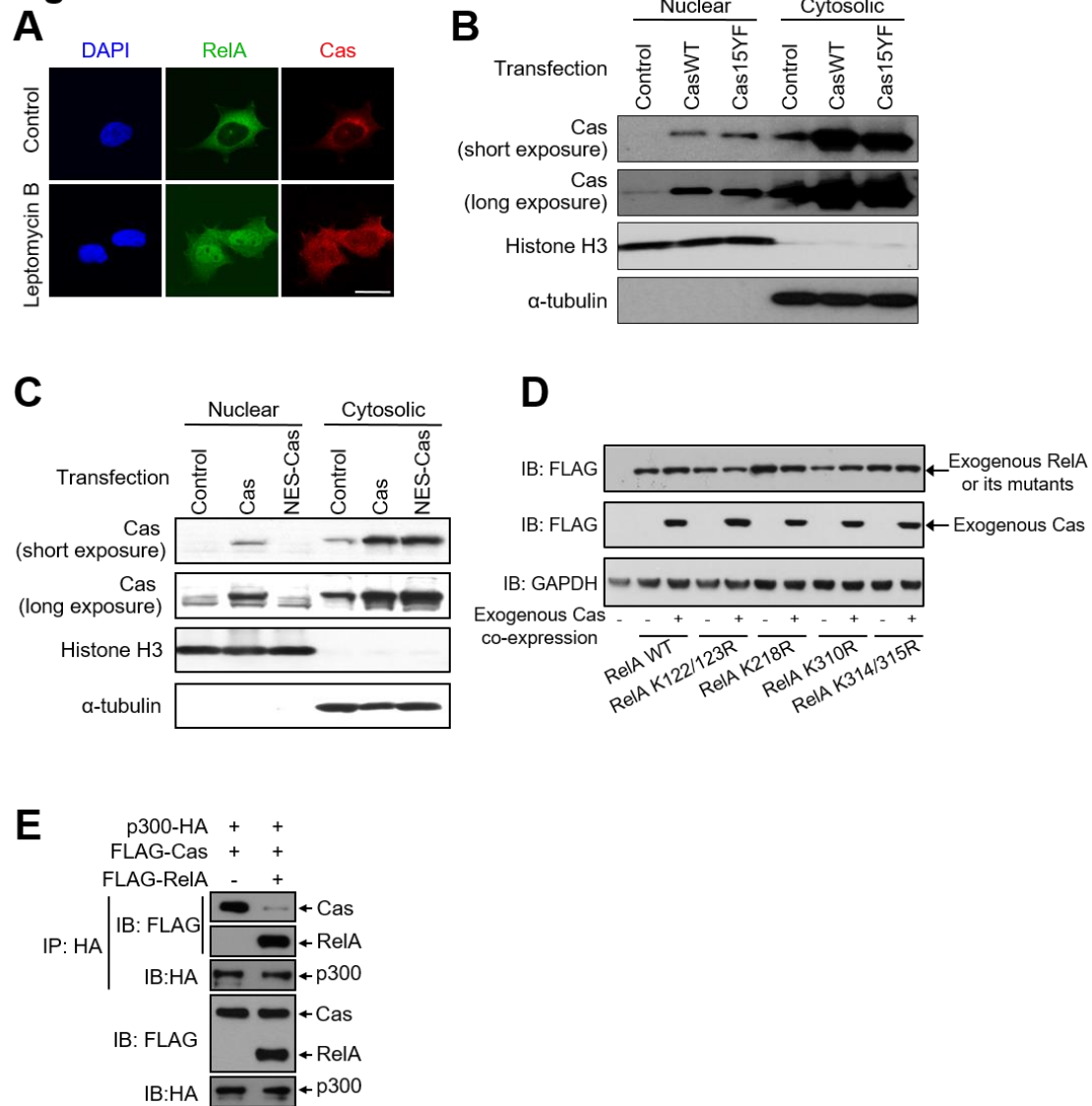


Fig. S5. Cas suppresses RelA acetylation at Lys³¹⁰ in the nucleus and alleviates NF-κB activity without

requiring phosphorylation of its substrate domain. (A) Cas accumulation in the nucleus after inhibition of

nuclear exportation. HEK293 cells, either left untreated or treated with Leptomycin B (10 ng/ml, 6 h), were

fixed and subjected to anti-RelA and anti-Cas immunostaining with nuclear staining (DAPI). Scale bar, 10

μm . **(B)** Phosphorylation-independent nuclear distribution of Cas. The nuclear and cytosolic fractions from transfected HEK293 cells were analyzed by immunoblotting. Endogenous Cas was detected in the nuclear fraction by the long exposure of the anti-Cas blot (second row, lane 1). **(C)** Hardly detectable recovery of nuclear export signal (NES)-attached Cas from the nuclear fraction. The nuclear and cytosolic fractions prepared from HEK293 cells transfected with Cas, NES-Cas or their control vector, were analyzed. Endogenous Cas was detected in the nuclear fraction by the long exposure of the anti-Cas blot (second row, lanes 1 and 3). **(D)** The expressions levels of transfected genes in Fig. 5J were evaluated by immunoblotting. **(E)** Attenuation of Cas-p300 interaction by exogenous RelA expression. HEK293 cells, co-transfected with p300-HA and FLAG-Cas together with FLAG-RelA or its control vector, were subjected to co-immunoprecipitation analysis as in Fig. 5K. Note the inverse correlation between RelA and Cas in the immunoprecipitates (see the top and second rows).

Figure S6

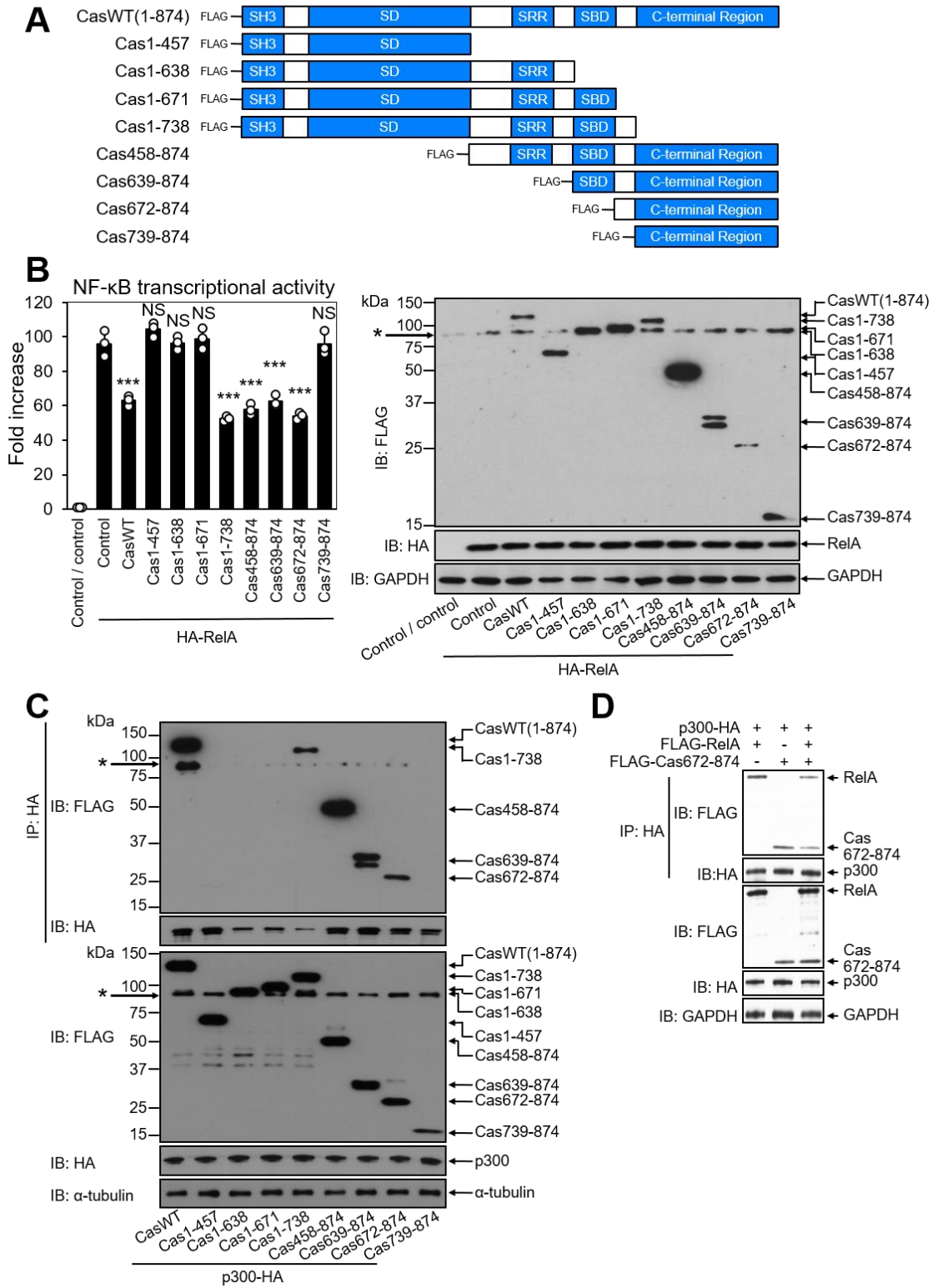


Fig. S6. Cas672-738 contains the region responsible for the inhibition of NF- κ B activity. (A) Schematic

representation of Cas truncation mutants tested. SH3: Src homology 3; SD: substrate domain; SRR:

serine-rich region; SBD: Src-binding domain. (B) Suppression of NF- κ B transcriptional activity by Cas

variants bearing the amino acids 672-738. Luciferase assay was conducted as in Fig. 5C with statistical

comparison with the RelA-transfected control sample (column 2) (n = 3) (left). The expression levels of

transfected HA-RelA and FLAG-tagged Cas variants were evaluated by immunoblotting. Non-specific

binding of the anti-FLAG antibody was observed at ~90 kDa (see an asterisk) (right). (C)

Co-immunoprecipitation of Cas variants bearing the amino acids 672-738 with p300. HEK293 cells

co-transfected with p300-HA and FLAG-tagged Cas variants were subjected to anti-HA immunoprecipitation

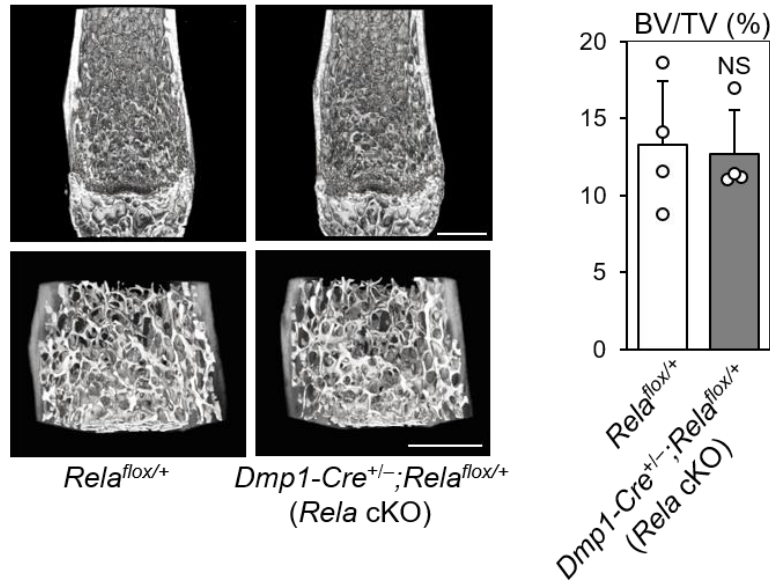
followed by immunoblot analysis. Non-specific binding of the anti-FLAG antibody was observed as in (B)

(see asterisks). (D) Inverse correlation between RelA-p300 and Cas672-874-p300 association. Co-expression

and co-immunoprecipitation experiments were conducted as in Fig. 5K.

Figure S7

A



B

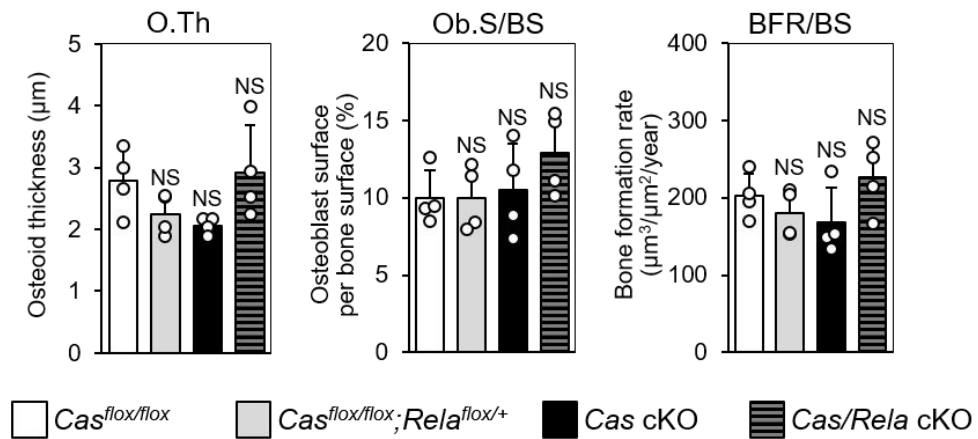


Fig. S7. Neither bone mass of mice with intact *Cas* alleles nor bone formation of *Cas* cKO mice is

significantly altered by heterozygous deletion of *Rela* in osteocytes (*Rela* cKO). (A) Representative μ CT

images of distal femurs of $Rela^{flox/+}$ (normal control mice) and $Dmp1-Cre^{+/-};Rela^{flox/+}$ mice (*Rela* cKO mice).

Scale bars, 1 mm (left). BV/TV was calculated from μ CT images (right, n = 4 mice per group). (B)

Histomorphometric parameters for bone formation at tibiae of mice with combinations of *Cas* and *Rela* cKO

(n = 4 mice per group, one-way ANOVA with post hoc Bonferroni test).