## **Supplementary Information**

STAT6 acetylation curtails macrophage M2 polarization and potentiates anti-tumor immunity

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Supplementary Figure 1. Mass spectroscopy showing the potential acetylation sites of Stat6 after immunoprecipitation of Stat6 in 293T cells transfected with Stat6 and CBP.



Supplementary Figure 2. Trim24 mediated Lys63-linked ubiquitination of CBP

(a) Immunoblot of Trim24, Stat6 and Lamin B or Hsp60 in nuclear and cytosol extracts of peritoneal macrophages that stimulated with (+) or without (-) IL-4. (b) Luciferase assay of Stat6 transcriptional activity in 293T cells that transfected with the indicated expression vectors, then were stimulated with IL-4 for 4 h before detection. (c) Immunoblot of endogenous Stat6 ubiquitination in murine primary macrophages that were left unstimulated (-) or stimulated (+) with IL-4 for 30 min. (d and e) Ubiquitination of CBP in 293T cells transfected with the indicated expression vectors, assessed by immunoblot analysis with anti-HA after immunoprecipitation with anti-Flag (top) or by immunoblot analysis with input proteins in lysates without immunoprecipitation (below). (f) Co-immunoprecipitation analysis of the binding between Trim24 and WT or K119R mutant CBP in the 293T cells transfected with the indicated expression vectors, assessed by immunoblot with anti-Flag and immunoblot with anti-Flag. Data with error bars are represented as mean  $\pm$  SD. Each panel is a representative experiment of at least three independent biological replicates. \*\*p < 0.01 as determined by unpaired Student's t test. Source data are provided as a Source Data file.



Supplementary Figure 3. Scheme of the construction of Trim24 floxed mouse and verification of Trim24 deficiency in macrophages.

(a) Schematic picture of Trim24 gene targeting using an FRT-LoxP vector. Targeted mice were crossed with FRT deleter (Rosa26-FLPe) mice to generate Trim24-floxed mice, which were further crossed with Lysosome M (LysM) Cre mice to generate myeloid cells conditional knockout mice. (b) Genotyping PCR analysis of Trim24<sup>+/+</sup>LysM-cre<sup>-</sup> and Trim24<sup>#/f</sup>LysM-cre<sup>+</sup> mice. (c) Immunoblot analysis of Trim24 and Hsp60 (loading control) protein expression in macrophages and splenocytes that isolated from wild-type (WT) and *Trim24<sup>M-/-</sup>* mice. Source data are provided as a Source Data file.



## Supplementary Figure 4. Trim24 does not affect the development, maturation and activation of myeloid and lymphoid cells.

(**a-h**) Flow cytometry of the frequencies of the indicated lymphoid and myeloid immune cells in the bone marrow (**a** and **b**), spleen (**c** and **d**), thymus (**e** and **f**), and peripheral lymph nodes (**g** and **h**) of wild-type (WT) and *Trim24*<sup>M-/-</sup> mice. Data are presented as representative plots (**a**, **c**, **e** and **g**) and summary bar graph (**b**, **d**, **f** and **h**). Data are representative of three independent experiments. Data with error bars are represented as mean  $\pm$  SD. Each panel is a representative experiment of at least three independent biological replicates. Source data are provided as a Source Data file.



## Supplementary Figure 5. Trim24 does not affect macrophage M1 polarization.

(**a** and **b**) QPCR analysis of *Il1b* and *Tnf* mRNA level in wild-type (WT) and *Trim24*-deficient (*Trim24*<sup>M-/-</sup>) macrophage that were left non-treated or stimulated with TLR ligands LPS (100 ng/ml), Pam3CSK4 (100 ng/ml) and Poly (I:C) (20  $\mu$ g/ml) or with LPS plus IFN $\gamma$  (100 ng/ml). (**c**) ELISA showing the secretion of IL-6 in supernatants of WT and *Trim24*-deficient macrophage that were stimulated with LPS at the indicated time. (**d**) Immunoblot analysis of phosphorylated (P-) and total NF- $\kappa$ B, MAPKs, TBK1 and Stat1 signaling proteins, Trim24, or Hsp60 (loading control) in whole-cell lysates of WT and *Trim24*-deficient macrophages that were left unstimulated with LPS (100 ng/ml) at the indicated time. Data with error bars are represented as mean  $\pm$  SD. Each panel is a representative experiment of at least three independent biological replicates. Source data are provided as a Source Data file.



Supplementary Figure 6. STAT6 is required for the enhanced M2 gene induction in *Trim24*-deficient macrophages.

(**a** and **b**) Immunoblot analysis of phosphorylated (P-) and total Stat6 and Akt, Trim24, or Hsp60 (loading control) in whole-cell lysates (**a**) and immunoblot of Stat6, Lamin B or Gapdh in nuclear and cytoplasmic extracts (**b**) of WT and *Trim24*-deficient macrophages that were stimulated with IL-4 (20 ng/ml) for indicated time. (**c**) Immunoblot of Mrc1, Arg1, Trim24 and Hsp60 in WT and *Trim24*-deficient macrophages that pretreated with DMSO or Jak3 selective inhibitor tofacitinib, and then stimulated with (+) or without (-) IL-4 (20 ng/ml) for 48 h. (**d** and **e**) QPCR analysis of IL-4-induced *Arg1*, *Mrc1*, *Ym1* and *Ccl24* mRNA expression in immortalized BMDMs (iBMDMs) upon knockdown of *Trim24* or *Stat6* or *Trim24/Stat6* double knockdown (**d**). The knockdown efficiency of Stat6 or Trim24 in iBMDMs are showed by immunoblot analysis in (**e**). Data with error bars are represented as mean  $\pm$  SD. Each panel is a representative experiment of at least three independent biological replicates. \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001 as determined by unpaired Student's t test. Source data are provided as a Source Data file.



Supplementary Figure 7. MDSC is dispensable for the tumor growth in *Trim24*<sup>M-/-</sup> mice. (a) Flow cytometry analysis of the frequencies and absolute numbers of tumor-infiltrating CD45<sup>+</sup>CD11b<sup>+</sup>Gr-1<sup>+</sup> MDSC and CD45<sup>+</sup>CD11c<sup>+</sup>MHC II<sup>+</sup> dendritic cells in B16 melanoma-bearing wild-type (WT) and *Trim24<sup>M-/-</sup>* mice. Data are presented as representative plots (left panels) and summary bar graphs showing absolute cell numbers (right panels). (**b**, **c**) Flow cytometric analysis showing the depletion efficiency of CD45<sup>+</sup>CD11b<sup>+</sup>F4/80<sup>+</sup> macrophages in B16 melanoma of WT and Trim24<sup>M-/-</sup> mice. Data are presented as representative plots (b) and a statistical bar graph (c). (d and e) Tumor growth curve (d) and weight (e) of wild-type (WT) and  $Trim24^{M-/-}$  mice that were intravenously injected with anti-Ly6G antibody at a 3-day interval for 4 times starting from the one day before tumor inoculation, and then challenged with B16 tumor cells subcutaneously. (f and g) Flow cytometric analysis showing the depletion efficiency of MDSC in B16 melanoma of WT and  $Trim 24^{M-/-}$  mice. Data are presented as a representative plot (f) and summary graph (g). Data with error bars are represented as mean  $\pm$  SD. Each panel is a representative experiment of at least three independent biological replicates. \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001 as determined by unpaired Student's t test. Source data are provided as a Source Data file.



Supplementary Figure 8. The working model of Stat6 acetylation in regulating M2 polarization and anti-tumor response of macrophage.

Under healthy condition, Th2 cytokine IL-4 could induce the phosphorylation of Stat6, which were then translocated into the nucleus. Trim24 in nucleus was associated with CBP and mediated IL-4-induced Lys63-linked ubiquitination of CBP at Lys119. The ubiquitinated CBP promoted the recruitment of Stat6 for its acetylation at Lys383, which inhibited its DNA-binding activity, and thus restrains macrophage M2 polarization and potentiate anti-tumor immunity. However in tumor microenvironment, activated Stat6 directly bond into the promoter region, and mediated the transcriptional suppression of *Trim24* gene. Accordingly, decreased Trim24 failed to induce CBP ubiquitination, and thus impaired Stat6 acetylation, which in turn promoted TAM M2 polarization, suppressed anti-tumor immune function, and contributed to the immunosuppressive tumor niche.



Supplementary Figure 9. Gating strategies used for flow cytometric analysis or cell sorting. (a) Flow cytometric strategy to detect endogenous Stat6 acetylation at Lys383 in murine primary peritoneal macrophages (CD11b<sup>+</sup>) isolated from naïve C57BL/6 mice presented on Fig. 2d. (b) Flow cytometric strategy to detect endogenous Stat6 acetylation at Lys383 in control or Trim24 knockdown human PBMC-derived primary macrophages (CD11b<sup>+</sup>) presented on Fig. 6d. (c) Flow cytometric strategy of endogenous Stat6 acetylation detection at Lys383 in murine tumor-associated macrophage (TAM, CD45<sup>+</sup>CD11b<sup>+</sup>F4/80<sup>+</sup>) from B16 melanoma of WT and Trim24M- mice presented on Fig. 7e. The gating strategies used for sorting TAM in Fig. 7d and Fig. 7h-7j is the same as (c). (d-e) Flow cytometric strategy to analysis the frequencies of TAM (CD45<sup>+</sup>CD11b<sup>+</sup>F4/80<sup>+</sup>, Fig. 7k) and IFNγ-producing CD4<sup>+</sup>  $(CD45^+CD4^+IFN\gamma^+, Fig. 7l)$  and  $CD8^+(CD45^+CD8^+IFN\gamma^+, Fig. 7l)$  T cells in B16 melanoma of WT and  $Trim 24^{M-1}$  mice. (f-j) Flow cytometric strategy to analysis the frequencies of the indicated lymphoid and myeloid immune cells in the bone marrow ( $\mathbf{f}$ ), spleen ( $\mathbf{g}$ ), thymus ( $\mathbf{h}$ ) and peripheral lymph nodes (I, j) of WT and  $Trim24^{M/2}$  mice present on Supplementary Fig. 4. (k) Flow cytometric strategy to analysis the frequencies of the tumor-infiltrating MDSC (CD45<sup>+</sup>CD11b<sup>+</sup>Gr-1<sup>+</sup>) and dendritic cells (CD45<sup>+</sup>CD11c<sup>+</sup>MHCII<sup>+</sup>) in B16 melanoma of WT and Trim24<sup>M/-</sup> mice presented on supplementary Fig. 7a. (1) Flow cytometric strategy to analysis the frequencies of TAM (CD45<sup>+</sup>CD11b<sup>+</sup>F4/80<sup>+</sup>) in B16 melanoma of WT and *Trim24<sup>M-t</sup>* mice presented on supplementary Fig. 7b. (m) Flow cytometric strategy to analysis the frequencies of tumor-infiltrating MDSC (CD45<sup>+</sup>CD11b<sup>+</sup>Gr-1<sup>+</sup>) in B16 melanoma of WT and Trim24<sup>M-4</sup> mice presented on supplementary Fig. 7f.

Genes	Forward primers (5'-3')	Reverse primers (5'-3')
mActb	CGTGAAAAGATGACCCAGATCA	CACAGCCTGGATGGCTACGT
mIl1b	AAGCCTCGTGCTGTCGGACC	TGAGGCCCAAGGCCACAGGT
mTnf	CATCTTCTCAAAATTCGAGTGACAA	CCAGCTGCTCCTCCACTTG
mArg1	TTTTTCCAGCAGACCAGCTT	AGAGATTATCGGAGCGCCTT
mMrc1	CAGGTGTGGGGCTCAGGTAGT	TGGCATGTCCTGGAATGAT
mYm1	TTTCTCCAGTGTAGCCATCCTT	TCTGGGTACAAGATCCCTGAA
mFizz1	GGGATGACTGCTACTGGGTG	TCAACGAGTAAGCACAGGCA
mCcl24	ATTCTGTGACCATCCCCTCAT	TGTATGTGCCTCTGAACCCAC
m <i>Irf4</i>	GCCCAACAAGCTAGAAAG	TCTCTGAGGGTCTGGAAACT
m <i>Trim24</i>	GCAAGCGGCTGATTACATACA	TGGTCACAGGAGGAAGCATCAC
hACTB	ACTCTTCCAGCCTTCCTTCC	CGTACAGGTCTTTGCGGATG
h <i>TRIM24</i>	CCAGAACATACCACGACAAGCAA	GGAGTAGAGGATGTGCTGTTGG
hTGM2	TGTGGCACCAAGTACCTGCTCA	GCACCTTGATGAGGTTGGACTC
hCCL17	TTCTCTGCAGCACATCCACGCA	CTGGAGCAGTCCTCAGATGTCT
h <i>IRF4</i>	GAACGAGGAGAAGAGCATCTTCC	CGATGCCTTCTCGGAACTTTCC
mArg1	GGCACAACTCACGTACAGACA	TGAGGCATTGTTCAGACTTCC
(ChIP)		
mYm1	GTTCTTTGGAACTCTTAGGA	GCACACACCATTCTTCAGAA
(ChIP)		
m <i>Irf4</i>	GATCAAAGCAGAACAAGGCACAGACT	GGGAATCTACTTCCCAGACTCTTACA
(ChIP)		
h <i>TRIM24</i>	GGTTTATTTTTGCTTAATGATGCTATAT	AAAGGTGAAATTAATTTTTAAAGTCA
(ChIP)		

Supplementary Table 1. Primers used for real-time quantitative PCR

Supplementary Table 2. Sequences of shRNA/siRNA for specific genes knockdown

Target genes	Sense sequence (5'-3')	Antisense sequence (5'-3')
Mouse Stat6	AGCAGGAAGAACTCAAGTTTA	TAAACTTGAGTTCTTCCTGCT
Mouse Trim24	CCCAAGTTGGAGTCATTCGAT	ATCGAATGACTCCAACTTGGG
Mouse CBP	TAACTCTGGCCATAGCTTAAT	ATTAAGCTATGGCCAGAGTTA
Control (shRNA)	TTCTCCGAACGTGTCACGT	ACGTGACACGTTCGGAGAA
Human TRIM24	CCAUUUCUGUUAACCUCUUAU	AUAAGAGGUUAACAGAAAUGG
Control (siRNA)	UUCUCCGAACGUGUCACGUdTdT	ACGUGACACGUUCGGAGAAdTdT