

Tervasmäki et al.

Supporting Information Figures S1–S5

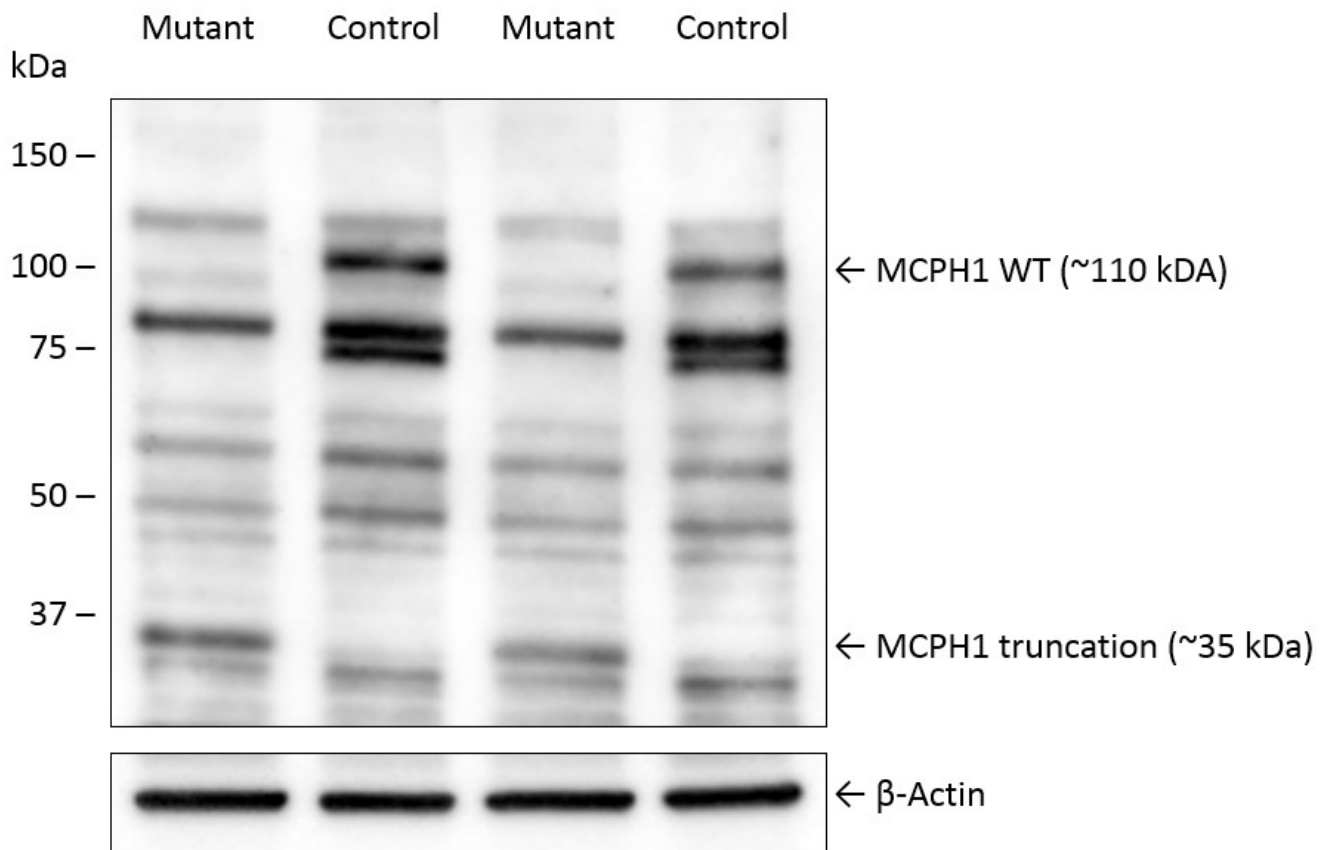


Fig. S1. Immunoblot of CRISPR/Cas9-edited *MCPH1* mutant and control MCF10A cell lines. Mutant cell line shows absence of wild type (~110 kDa) and presence of truncated (~35 kDa) *MCPH1* protein.

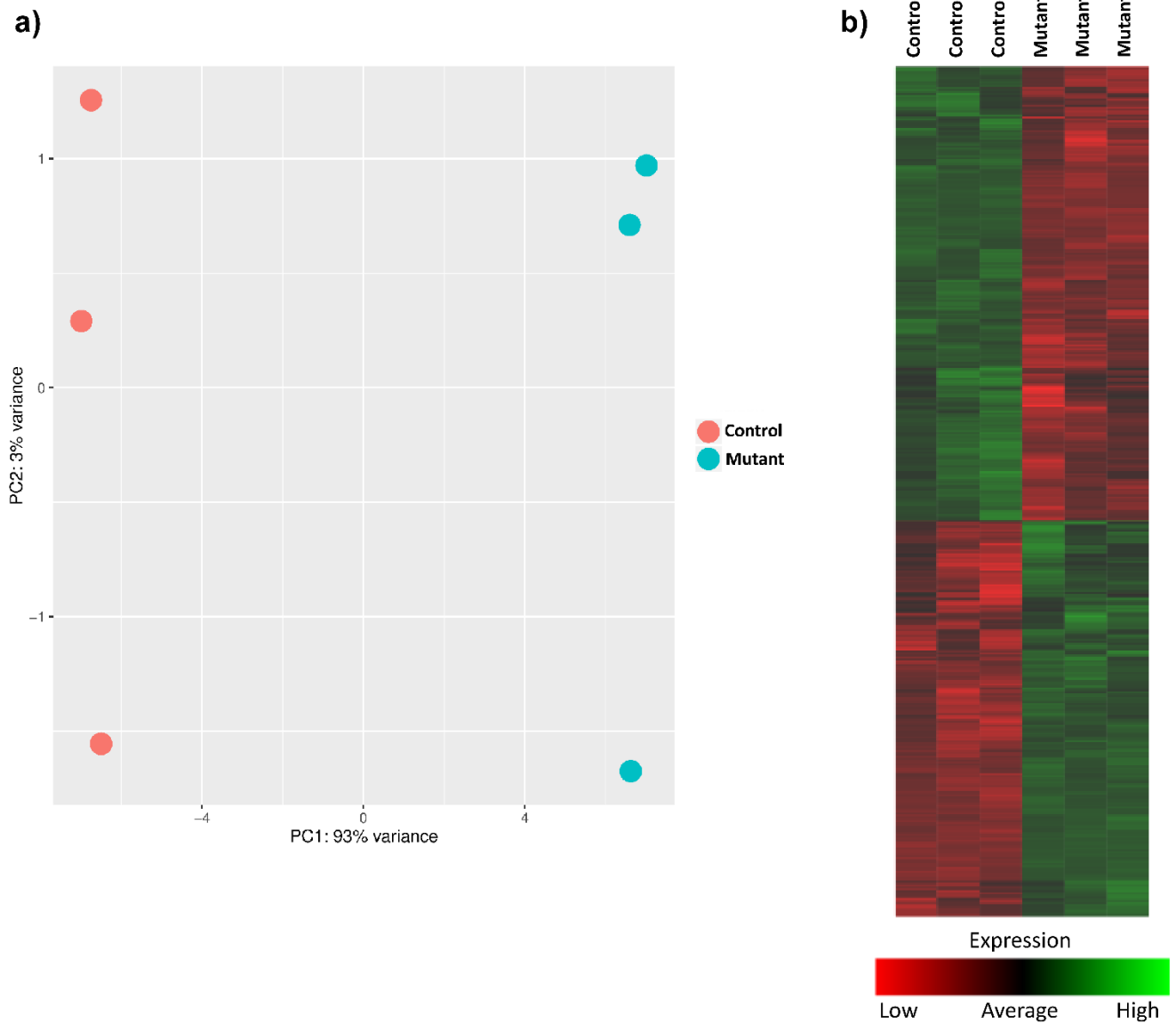


Fig. S2. *MCPH1* mutant and control cells have distinct gene expression profiles. **a)** Principal component analysis (PCA) shows gene expression patterns of *MCPH1* mutant and control cell lines being distinct from each other, principal component 1 explaining 93% of the variation between the samples. **b)** Differential gene expression patterns presented as a heatmap based on DESeq2 results. Red indicates decreased gene expression and green increased expression.

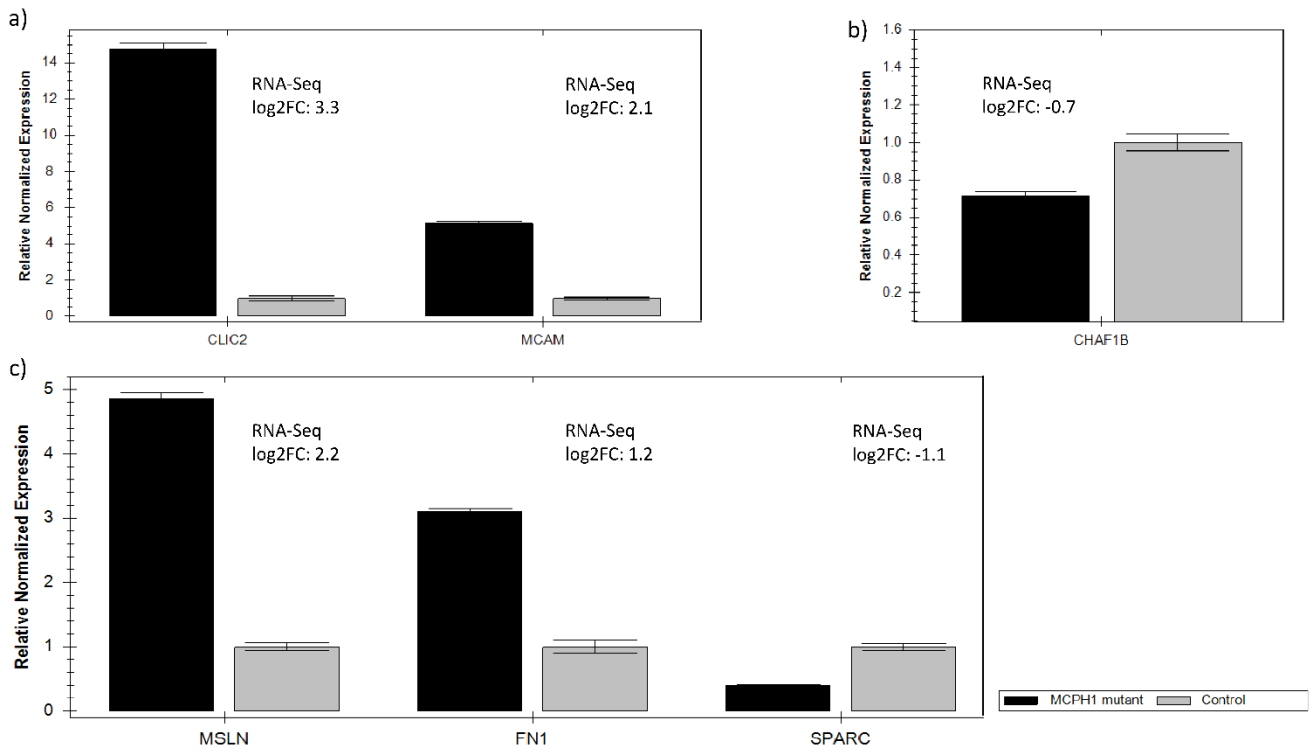


Fig. S3. Six of the differentially expressed genes in RNA-Seq were analyzed with RT-PCR. a) *CLIC2* and *MCAM*, b) *CHAF1B* and c) *MSLN*, *FN1* and *SPARC*. All analyzed genes showed similar differential expression in the cell lines with both methods. Results are mean (\pm SE) expression of 3 replicates.

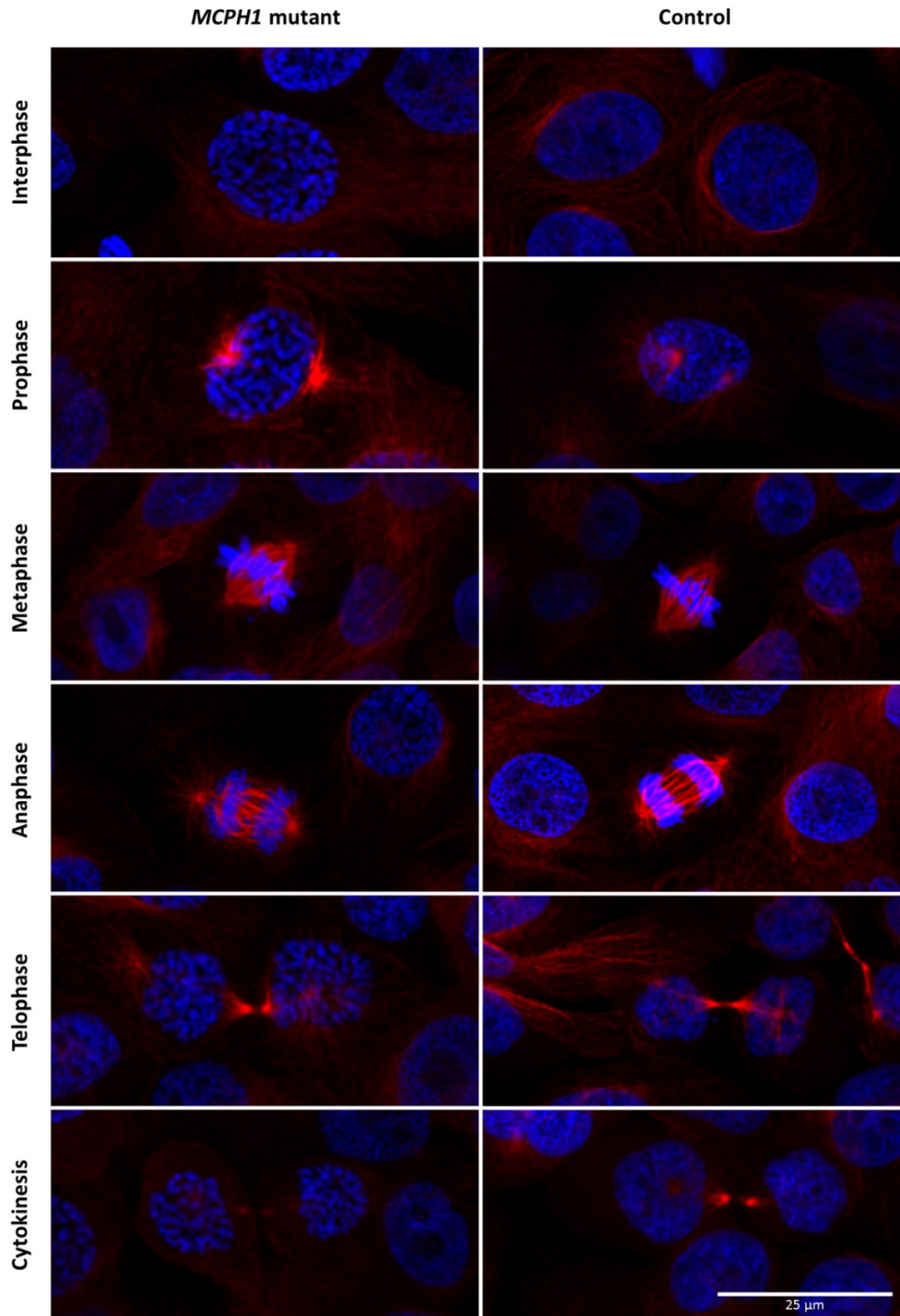


Fig. S4. Representative images of mitotic phases of *MCPH1* mutated and control MCF10 cells. Mutant cells show PCC phenotype and prolonged chromatin condensation during telophase and cytokinesis. Fixed cells were stained with DAPI (blue) and anti-tubulin (red). Scale bar 25 μ m.

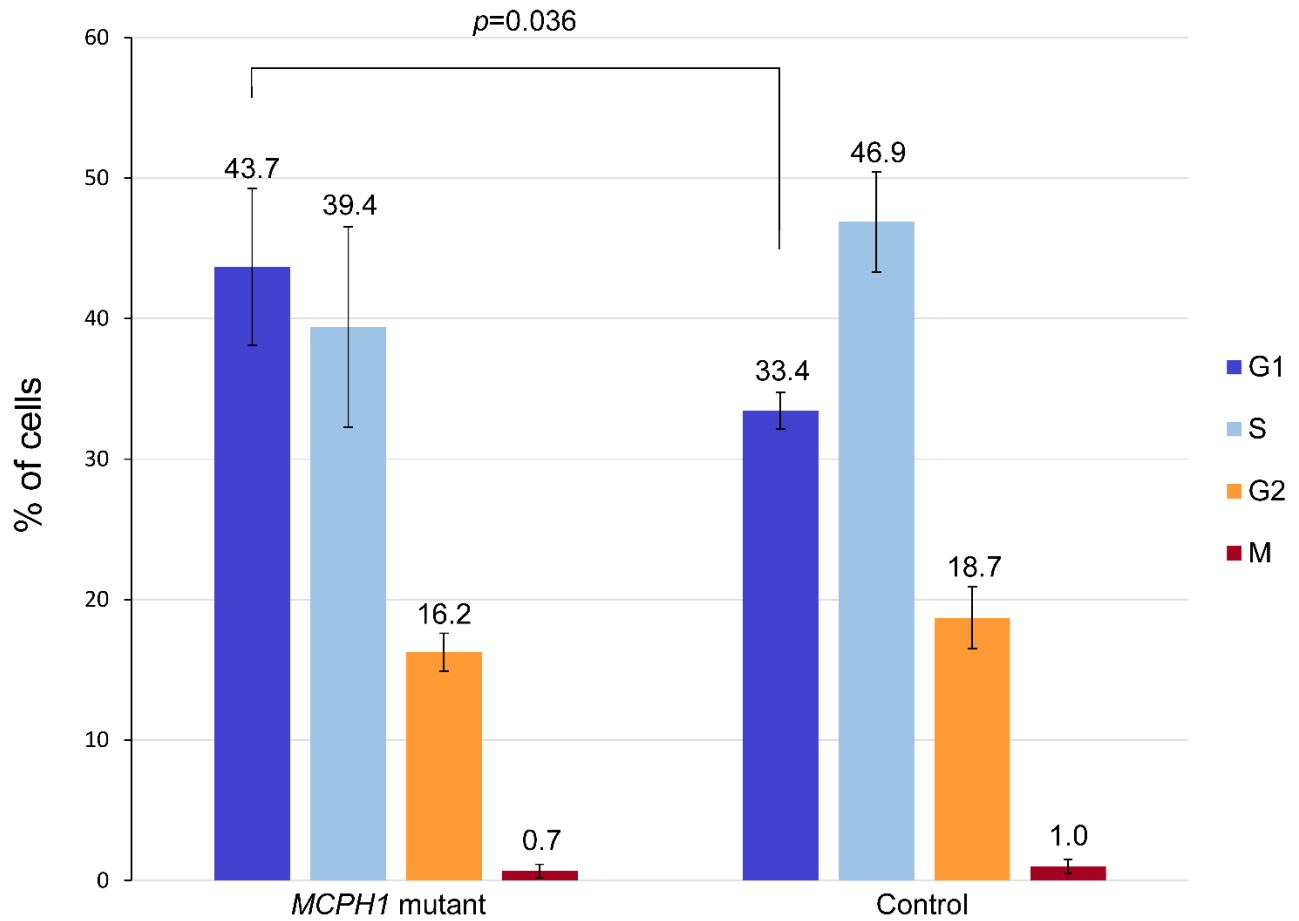


Fig. S5. *MCPH1* mutant cells have larger proportion of G1 phase cells. Cell cycle distribution analysis by flow cytometry shows *MCPH1* mutant cell line having more cells in G1 phase (43.7%) than controls (33.4%), $p=0.036$. Means of three independent experiments (\pm SD), Student's t-test.