Requirement	Please Include Requested Information
1.1. Purpose	Test whether or not a linear classifier can automatically
	and accurately classify different cell types in mass
	cytometry datasets, and be used to analyze large
	cohort studies
1.2. Keywords	Single cell, Mass cytometry, Cell type prediction, Machine learning
1.3. Experiment variables	We used five public mass cytometry datasets, including human blood samples, as well as human and mouse bone marrow (described in details in 'Sample description')
1.4. Organization name and address	Delft University of Technology Van Mourik Broekmanweg 6, 2628 XE Delft, The Netherlands
1.5. Primary contact name and email address	Dr. Ahmed Mahfouz a.mahfouz@lumc.nl
1.6. Date or time period of experiment	From 01-11-2017 to 30-09-2018
1.7. Conclusions	We showed that a linear classifier can be used to automatically assign labels to single cells in mass cytometry data. Using five different datasets, the linear classifier outperforms two state-of-the-art methods (1,2).
	 Lee H, Kosoy R, Becker CE, Dudley JT, Kidd BA. Automated cell type discovery and classification through knowledge transfer. Bioinformatics 2017;33:1689–1695. Li H, Shaham U, Stanton KP, Yao Y, Montgomery RR, Kluger Y. Gating mass cytometry data by deep learning. Bioinformatics 2017;33:3423–3430.
1.8. Quality control measures	We used all the published data from the public datasets, and QC is performed in the original studies.
2.1.1.1. (2.1.2.1., 2.1.3.1.) Sample description	 We used five publicly available mass cytometry datasets: AML is a healthy human bone marrow dataset (3). BMMC is also a healthy human bone marrow dataset (4). PANORAMA dataset consists of 10 replicates of mice bone marrow cells (5). Multi-Center study dataset is a collection of 16 samples drawn from a single subject (2). HMIS is a human mucosal immune system consists of 47 Peripheral Blood Mononuclear Cells (PBMC) samples from individuals with inflammatory bowel diseases (6). Samples are divided into control samples, samples with Crohn's disease, samples with Celiac disease, and samples with Refractory Celiac disease Type II.
	2. Li H, Shaham U, Stanton KP, Yao Y, Montgomery RR, Kluger Y. Gating mass cytometry data by deep learning.

Cytometry Part A Author Checklist: MIFlowCyt-Compliant Items

	Bioinformatics 2017:33:3423–3430
	3 Levine IH Simonds EF Bendall SC Downing IB Pe D
	Nolan GP Levine IH Simonds EF Bendall SC Davis KL Amir
	FD Tadmor MD Downing IR Pe D Nolan GP Data-Driven
	Phenotynic Dissection of AMI Reveals Progenitor-like Cells
	that Correlate with Prognosis Cell 2015:162:184–197
	Available at: $http://dx doi org/10 1016/i cell 2015 05 047$
	A Bendall SC Simonds EE Oiu P Amir ED Krutzik PO Einck
	R Bruggner R V Melamed R Treio A Ornatsky OI Balderas
	RS Plevritis SK Sachs K Pe'er D Tanner SD Nolan GP
	Single-Cell Mass Cytometry of Differential Immune and
	Drug Responses Across a Human Hematonoietic
	Continuum Science 2011:332:687–696
	5 Samusik N. Good 7 Spitzer MH. Davis KI. Nolan GP
	Automated manning of phenotype space with single-cell
	data Nat Methods 2016:13:493–496
	6 van Linen V. Li N. Molendijk I. Temurhan M. Höllt T. van
	der Meulen-de long AF Versnaget HW Mearin MI Mulder
	CL van Bergen L Lelieveldt BPE Koning E Mass Cytometry
	of the Human Mucosal Immune System Identifies Tissue-
	and Disease-Associated Immune Subsets Immunity
	2016-44-1227–1239
2.1.1.2. Biological sample source description	Bone marrow and blood (PBMC) samples
2.1.1.3. Biological sample source organism description	Human and mice samples
2.1.2.2. Environmental sample location	N.A
2.3. Sample treatment description	N.A
2.4. Fluorescence reagent(s) description	N.A
3.1. Instrument manufacturer	N.A
3.2. Instrument model	N.A
3.3. Instrument configuration and settings	N.A
4.1. List-mode data files	We used five public datasets, which are available through
	the original publications (references above), and through
	our GitHub link: https://github.com/tabdelaal/CyTOF-
	Linear-Classifier
4.2. Compensation description	N.A
4.3. Data transformation details	We applied hyperbolic arcsin transformation with a
	cofactor of 5 for all datasets.
4.4.1. Gate description	N.A
4.4.2. Gate statistics	N.A
4.4.3. Gate boundaries	N.A

Notes

Feel free to use more space than allocated.

You can embed graphics/figures in this document, if needed.

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