



Supplementary Material

Chemical Profiling of Polyphenolics in *Eucalyptus globulus* and Evaluation of Its Hepato-Renal Protective Potential Against Cyclophosphamide Induced Toxicity in Mice

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Annotation of The Polyphenolic Compounds Using HPLC-DAD-ESI-MS/MS (S1)

Phenolic Acids and Their Derivatives

Compound 3 was detected at *Rt* 1.41 min, it showed a deprotonated ion [M-H]⁻ at *m/z* 169 and a diagnostic fragment ion at *m/z* 125 corresponding to the loss of CO₂ moiety (*-m/z* 44 u) [M-H-CO₂ moiety]⁻, it could be identified as gallic acid [34]. **Compound 4** was detected at *Rt* 6.69, it showed a deprotonated ion [M-H]⁻ at *m/z* 183 and MSⁿ ions were observed at *m/z* 169, 168, 125, 109, and 93, it was tentatively identified as methylgallate [35].

Flavonoids

Compound 12 was detected at *Rt* 18.68 min, it showed a deprotonated molecule [M-H]⁻ at *m/z* 491 and MS fragment ion at *m/z* 315 [M-H-glucuronide moiety]⁻ corresponding to the loss of glucuronide moiety (*-m/z* 176 u) was assigned to isorhamnetin aglycone, and key fragment ions for aglycone were observed at *m/z* 271, 179 and 151. Thus, compound 12 could be identified as isorhamnetin 3-O-β-D-glucuronoside [36]. **Compound 18** was detected at *Rt* 25.31 min, it showed a deprotonated molecule [M-H]⁻ at *m/z* 329, and MSⁿ ions at *m/z* 314 [M-H-CH₃]⁻, 299 [M-H-2CH₃]⁻, 285 [M-H-CO₂]⁻ and 243 [M-H-C₂H₂O-CO₂]⁻, thus it could be identified as quercetin-3,4'-dimethyl ether [37,38]. Compound 22 was detected at *Rt* 40.34 min, it showed a deprotonated molecule [M-H]⁻

at m/z 303, and MS^n ions were obtained at m/z 301, 285, 179, 177, and 125, it could be identified as dihydroquercetin (taxifolin) [39].

Hydrolyzable Tannins (Gallotannins and Ellagitannins)

Compound 7 was detected at R_t 13.55 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 537 and MS^n fragment ions at m/z 385 $[M-H-152]^-$ due to the loss of galloyl moiety (-152 u), 313 $[M-H-224]^-$, a diagnostic ion was observed at m/z 271 $[M-H-266]^-$, this fragmentation pattern was typically assigned to 6S,9R-ionone 9-O-(6'-O-galloyl)- β -D-glucopyranoside (Mallophenol B) [40]. **Compound 9** was detected at R_t 16.29 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 483, producing daughter ion at m/z 271 $[M-H-212]^-$ due to the total loss of galloyl moiety (- m/z 152 u), water molecule (- m/z 18 u) and another fragment (- m/z 42 u), characteristic ions for galloylglucose derivatives were observed at m/z 331, 313, 169, 151 and 137, thus it could be assigned to digalloylglucose, this compound was previously identified in both of Algerian *Eucalyptus globulus* fruits [41] and Egyptian *Eucalyptus camaldulensis* leaves [42]. **Compound 10** was detected at R_t 17.16 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 421, producing daughter ion at m/z 169 $[M-H\text{-benzyl moiety-glucosyl moiety} (\Sigma\text{-}m/z\ 252\ \text{u})]^-$ due to the loss of benzyl and glucose moieties. Other MS^n fragment ions were observed at m/z 331 due to the neutral loss of benzyl moiety (- m/z 90 u) $[M-H\text{-benzyl moiety}]^-$, also a fragment ion was observed at m/z 313 due to the loss of water molecule (- m/z 18 u) from $[M-H\text{-benzyl moiety-H}_2\text{O}]^-$, it could be identified as benzyl-galloylglucose [42]. **Compound 11** was detected at R_t 17.50 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 481 and characteristic MS^n fragment ions for digalloylglucose nucleus at m/z 463 $[M-H\text{-H}_2\text{O} (-m/z\ 18\ \text{u})]^-$, 329 $[M-H\text{-galloyl moiety} (-m/z\ 152\ \text{u})]^-$, 301 $[M-H\text{-H}_2\text{O-glucose moiety} (-m/z\ 180\ \text{u})]^-$, 313 211, and 169; therefore, it could be assigned to hexahydroxydiphenyl-glucose (HHDP-glucose) [42]. **Compound 13** was detected at R_t 19.64 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 689 and it's a diagnostic fragment ion at m/z 537 $[M-H-152]^-$, which corresponds to the loss of a galloyl moiety, it can be interpreted as galloyl cypellocarpin B [42]. **Compound 14** was detected at R_t 19.82 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 939 and a daughter ion at m/z 769 corresponding to loss of gallic acid moiety $[M-H-170]^-$, in addition to the appearance of MS^n fragment ions at m/z 787 and 635 which corresponds to the loss of the first galloyl moiety $[M-H-152]^-$, and the second one $[M-H-304]^-$ respectively, accordingly this compound can be interpreted as pentagalloylglucose [41]. **Compound 15** was detected at R_t 20.12 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 635 and a daughter ion as a base peak at m/z 483 corresponding to the loss of galloyl moiety $[M-H-152]^-$, also diagnostic MS^n fragment ions were observed at m/z 465 $[M-H-170]^-$ and 423 $[M-H-212]^-$ due to the loss of gallic acid moiety, and the release of another galloyl moiety followed by cross ring fragmentation of glucose, 331 $[M-H-152-152]^-$, 313 $[M-H-152-152-18]^-$, 271 $[M-H-212-152]^-$ due to the loss of galloyl moieties, and water molecule; it could be interpreted as trigalloylglucose [41]. **Compound 16** was detected at R_t 21.54 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 625 and a daughter ion at m/z 473 corresponding to the loss of galloyl moiety $[M-H-152]^-$, furthermore diagnostic MS^n fragment ions were appeared at m/z 607 $[M-H\text{-H}_2\text{O}]^-$ due to the loss of water molecule (- m/z 18 u), 437 $[M-H-18-170]^-$ corresponding to the loss of water and gallic acid moieties, thus compound could be characterized as HHDP-diglucoside [43]. **Compound 17** was detected at R_t 24.91 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 629 and MS^n fragment ions at m/z 477 $[M-H\text{-galloyl moiety}]^-$ due to the loss of galloyl moiety (- m/z 152 u) and corresponding to methylellagic acid glucoside, m/z 315 $[M-H\text{-galloyl glucoside moiety}]^-$ due to the loss of galloyl glucoside moiety (- m/z 314 u) and corresponding to methyl ellagic acid, and a base peak at m/z 301 which corresponding to ellagic acid nucleus, this fragmentation pattern was typically assigned to galloyl ester of a methylellagic acid glucoside [41]. **Compound 19** was detected at R_t 27.17 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 1085 and a diagnostic fragment ions at m/z 765 $[M-H-320]^-$, 633 $[M-H-452]^-$ due to the loss of trigalloyl moieties (- m/z 452 u) which corresponding to HHDP moiety, so it could be identified as eucalbanin A or its isomer cornusiin B [41]. **Compound 20** was detected at R_t 28.52 min, it showed a deprotonated molecule $[M-H]^-$ at m/z 519 and fragment ions at m/z 353 $[M-H-166]^-$, 335 $[M-H-184]^-$ due to the loss of oleuropeic acid

moiety ($-m/z$ 184 u), and 233 [M-H-286]⁻, this compound could be identified as cypellocarpin C [41]. **Compound 21** was detected at R_t 29.61 min, it showed a deprotonated molecule [M-H]⁻ at m/z 1415 and fragment ions at m/z 1113 [M-H-302]⁻ due to the loss of one HHDP moiety ($-m/z$ 302 u), 933 [M-H-482]⁻, 783 [M-H-632]⁻ and 633 [M-H-782]⁻, this compound could be characterized as Di (HHDP-galloylglucose)-pentose [41]. **Compound 23** was detected at R_t 41.82 min, it showed a deprotonated molecule [M-H]⁻ at m/z 617 and fragment ions were appeared at m/z 465 [M-H-152]⁻, 393 [M-H-224]⁻, 317 [M-H-300]⁻, 241 [M-H-376]⁻ and 169 [M-H-448]⁻, these fragmentation pattern was assigned to trigalloyllevoglucosan [44]. **Compound 24** was detected at R_t 53.13 min, it showed a deprotonated molecule [M-H]⁻ at m/z 953 and MSⁿ fragment ions at m/z 635, 301 and 169, this fragmentation pattern was assigned to valoneoyl-digalloyl-glucopyranose. **Compound 26** was detected at R_t 56.04 min, it showed a deprotonated molecule [M-H]⁻ at m/z 469 as well as MSⁿ fragment ions at m/z 425 [M-H-CO₂ ($-m/z$ 44 u)]⁻, 301 [M-H-168]⁻ and 169 [M-H-300]⁻, it was tentatively identified as valoneic acid dilactone [41].

Structural Elucidation of The Isolated Phenolic Compounds (S2)

Compound 1 was isolated as off-white amorphous powder, observed as faint violet fluorescence under short UV light, which turned to deep blue colour with FeCl₃ spray reagent. Molisch test was positive indicating a gallic acid glycoside. Acid hydrolysis afforded gallic acid in organic layer and rhamnose in aqueous layer. ¹H-NMR spectral data (400 MHz, DMSO-*d*₆) δ_H ppm: two symmetrical aromatic protons were resonated at δ_H 6.99 ppm (2H, s, H-2 and H-6), anomeric proton of sugar moiety at δ_H 4.91 ppm (1H, d, $J=7.44$ Hz) and remaining sugar protons at δ_H 3.17-3.74 ppm (m). ¹³C-NMR spectral data (100 MHz, DMSO-*d*₆), carbon atoms of gallic acid nucleus were resonated at δ_C 167.9 (CO), 145.8 (C-3, C-5), 138.4 (C-4), 120.9 (C-1), 109.1 ppm (C-2, C-6). While, carbon atoms of *O*-β-xylopyranosyl moiety were resonated at δ_C 103.3 (C-1'), 76.8 (C-3'), 73.6 (C-2'), 69.8 (C-4') & 65.6 ppm (C-5'). Based on the above mentioned chromatographic properties, NMR data and literature [45], compound 3 was identified as 4-(*O*-β-D-xylopyranosyloxy)-3,5-di-hydroxy-benzoic acid (Gallic acid pentoside).

Compound 2 was isolated as off-white amorphous powder, observed as faint violet fluorescence under short UV light, which turned to deep blue colour with FeCl₃ spray reagent. Molisch test was positive indicating a gallic acid glycoside. Acid hydrolysis afforded gallic acid in organic layer and rhamnose in aqueous layer. ¹H-NMR spectral data (400 MHz, DMSO-*d*₆), two symmetrical aromatic protons were resonated at δ_H 6.97 ppm (2H, s, H-2 and H-6), anomeric proton of sugar moiety at δ_H 4.63 ppm (1H, brs, H-1'), remaining sugar protons were resonated at δ_H 3.25-3.55 ppm (m) and aliphatic methyl protons at δ_H 1.09 ppm (3H, d, $J=6.8$ Hz, CH₃-C-6'). ¹³C-NMR spectral data (100 MHz, DMSO-*d*₆), carbon atoms of gallic acid nucleus were resonated at δ_C 167.9 (COO), 145.8 (C-3, C-5), 138.4 (C-4), 120.8 (C-1), and 109.1 ppm (C-2 & C-6). On the other hand, carbon atoms of *O*-α-L-rhamnopyranosyl moiety were resonated at δ_C 104 (C-1'), 72.7 (C-4'), 70.3 (C-3 & 4), 69.4 (C-5'), and 18.9 ppm (CH₃-6'). Based on the above mentioned chromatographic properties, NMR data and literature [45], compound 4 was identified as 4-(*O*-α-L-rhamnopyranosyloxy)-3,5-di-hydroxy-benzoic acid (Gallic acid rhamnoside).

Compound 3 was isolated as off-white amorphous powder, observed as faint violet fluorescence under short UV light, which turned to deep blue colour with FeCl₃ spray reagent. UV spectral data showed a characteristic band at λ_{max} = 272 nm. ¹H-NMR spectral data (400 MHz, DMSO-*d*₆), revealed the presence of two symmetrical protons located in the aromatic region at δ_H 6.94 ppm (2H, s, H-2 and H-6). ¹³C-NMR spectral data (100 MHz, DMSO-*d*₆) showed a set of aromatic carbons were resonated at δ_C 109.1 ppm (C-2, C-6), 120.8 ppm (C-1), 138.4 ppm (C-4), 145.8 ppm (C-3, C-5) and carbonyl carbon at δ_C 167.9 ppm (-CO) [46]. Based on the above mentioned NMR data, comparison with authentic samples (CO-PC) and literature, compound 1 was identified as 3,4,5-trihydroxy-benzoic acid (gallic acid).

Compound 4 was isolated as off-white amorphous powder, observed as faint violet fluorescence under short UV light, which turned to deep blue colour with FeCl₃ spray reagent. UV spectral data showed a characteristic band at λ_{max} = 272 nm. ¹H-NMR spectral data (400 MHz,

DMSO- d_6), showed two symmetrical protons in the aromatic region at δ_H 6.91 ppm (2H, brs, H-2 and H-6), another characteristic signal for aliphatic methoxy protons was observed at δ_H 3.7 ppm (3H, s, -OCH₃). ¹³C-NMR spectral data (100 MHz, DMSO- d_6), showed three types of carbon signals including oxygenated aliphatic carbon at δ_C 51.68 ppm (OCH₃), a set of aromatic carbon signals were observed at δ_C 108.57 ppm (C-2, C-6), 119.36 ppm (C-1), 138.48 ppm (C-4), 145.56 ppm (C-3, C-5) and carbonyl carbon was detected at δ_C 166.41 ppm (-CO) [47,48]. Based on the above mentioned chromatographic properties, NMR data, comparison with authentic samples (CO-PC) and literature, compound 2 was identified as methyl 3,4,5-trihydroxybenzoate (methyl gallate).