

2

3 **Topology and enzymatic properties of a canonical PRC1**  
4 **isoform**

5

6 Matteo Colombo<sup>1</sup>, Ombeline Pessey<sup>1</sup>, and Marco Marcia<sup>1\*</sup>

7 <sup>1</sup>European Molecular Biology Laboratory, Grenoble Outstation, 71 Avenue des Martyrs, Grenoble  
8 38042, France

9

10 **\*Corresponding author:** Marco Marcia, European Molecular Biology Laboratory, Grenoble  
11 Outstation, 71 Avenue des Martyrs, Grenoble 38042, France, Tel: +33 04 76 20 7759, E-Mail:  
12 [mmarcia@embl.fr](mailto:mmarcia@embl.fr)

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

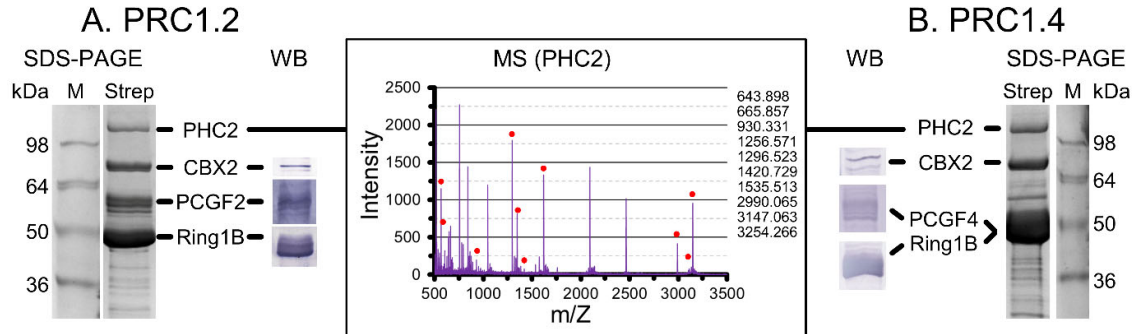
29

30 This file contains:

31 - Figures S1-S6

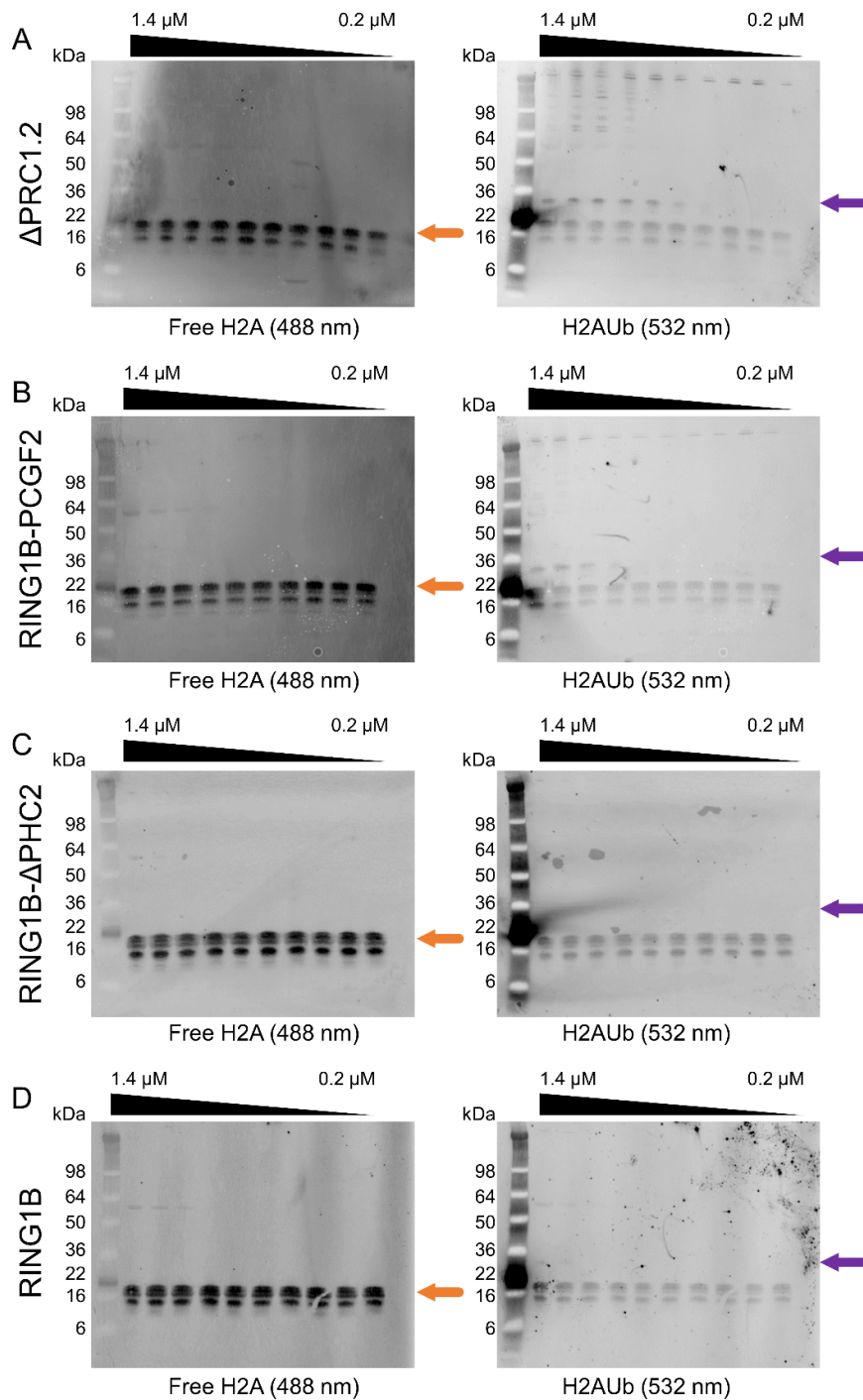
1 **Figure S1.** SDS-PAGE and biochemical characterization of PRC1.2 (A) and PRC1.4 (B). Both  
 2 complexes contain all four expected subunits after strep-tag affinity chromatography. All  
 3 subunits were identified by peptide mass fingerprinting mass spectrometry (here shown for  
 4 PHC2 only, with matched peptides indicated by red circles and corresponding m/Z values listed  
 5 on the right). CBX2, PCGF2, PCGF4, and RING1B were additionally identified by Western  
 6 blot (WB).

7



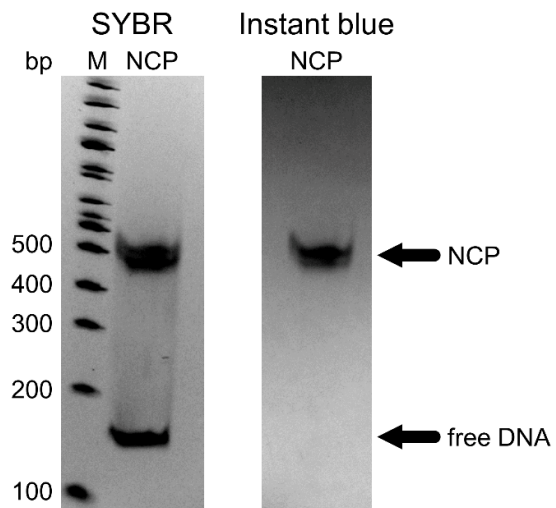
8

1 **Figure S2.** Representative Western blot membranes used for quantification of the H2A  
 2 monoubiquitination activity of  $\Delta$ PRC1.2 (**A**) and its subcomplexes (**B-D**) (quantification  
 3 reported in **Figure 5C**). The orange arrow indicates the free H2A histone (detected with an  
 4 antibody coupled to Alexa fluor 488 nm) while the purple arrow indicates ubiquitinated H2A  
 5 (detected with an antibody coupled to Alexa fluor 532 nm).  
 6



7

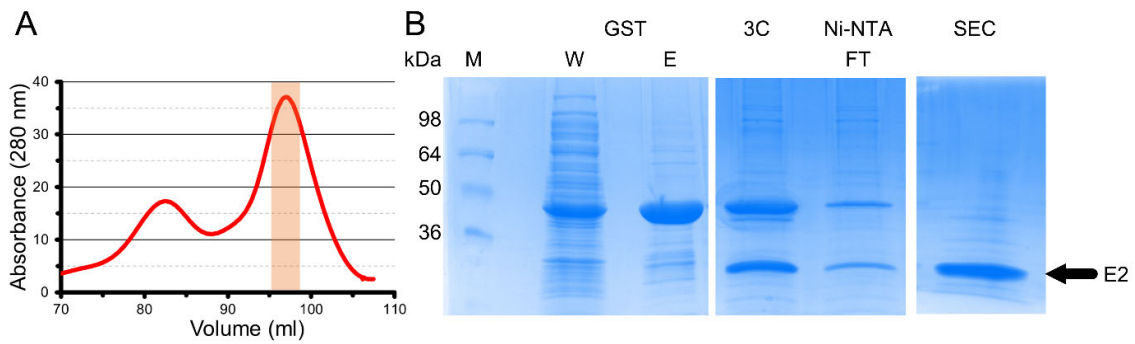
1 **Figure S3.** A) Representative native PAGE (6 % acrylamide) of nucleosome core particles  
2 (NCP) stained with SYBR safe (left) and Instant blue (right).  
3



4

1 **Figure S4.** Purification of the UbcH5c E2 enzyme. **A)** Representative SEC profile. The orange  
2 rectangle indicates the fractions of purified E2 used in this work. **B)** Representative SDS-PAGE  
3 gels showing the different steps of purification. M is a molecular size marker, W is the wash  
4 and E the elution fraction of the first GST-affinity chromatographic step, 3C indicates the  
5 digestion product of 3C precision protease, FT is the flow through of a Ni-NTA column used  
6 to remove undigested, His- and GST-tagged E2, and SEC is the eluate from the gel filtration  
7 column shown in panel A (orange rectangle). The black arrow on the right indicates the band  
8 corresponding to the E2 enzyme.

9

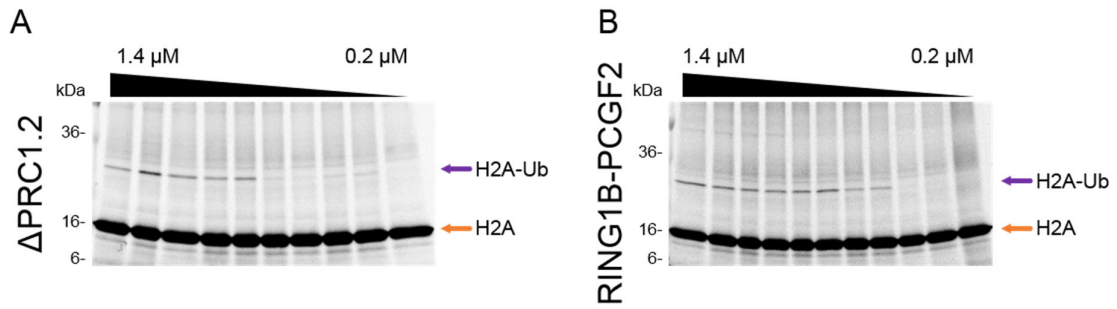


10

11

12

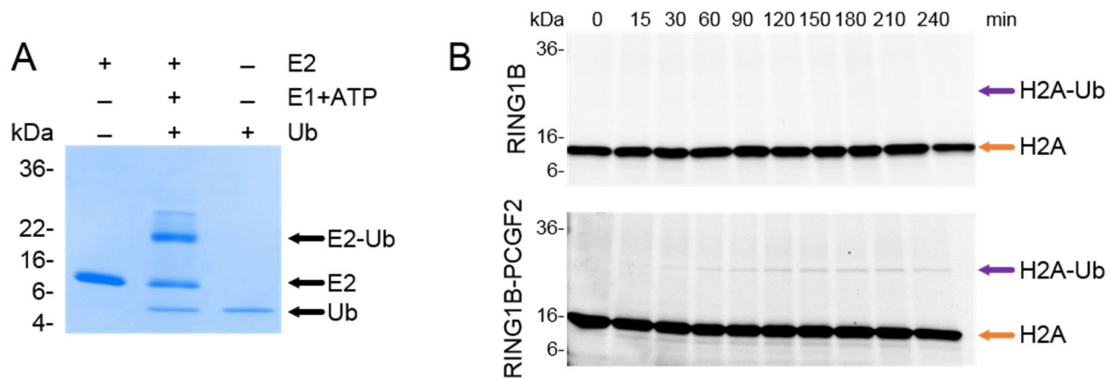
1 **Figure S5.** Representative SDS-PAGE gels used for quantification of the H2A  
2 monoubiquitination activity of  $\Delta$ PRC1.2 (**A**) and RING1B-PCGF2 (**B**) using Cy5-labelled  
3 nucleosomes (quantification reported in **Figure 5B**). The orange arrow indicates the free H2A,  
4 while the purple arrow indicates ubiquitinated H2A.  
5



6  
7  
8

1 **Figure S6.** E2-discharging assays. **(A)** Representative SDS-PAGE gel documenting ubiquitin  
 2 preloading on UbcH5c before apyrase treatment. Arrows on the right of the gel indicate free  
 3 ubiquitin (Ub), free UbcH5c (E2) and ubiquitinated UbcH5c (E2-Ub), respectively. **(B)**  
 4 Representative SDS-PAGE gels used for quantification of E2-discharging activity of RING1B  
 5 (top) and RING1B-PCGF2 (bottom; quantification reported in **Figure 5D**). Cy5-labelled free  
 6 and ubiquitinated H2A are indicated by the orange and purple arrows, respectively.

7



8

9