

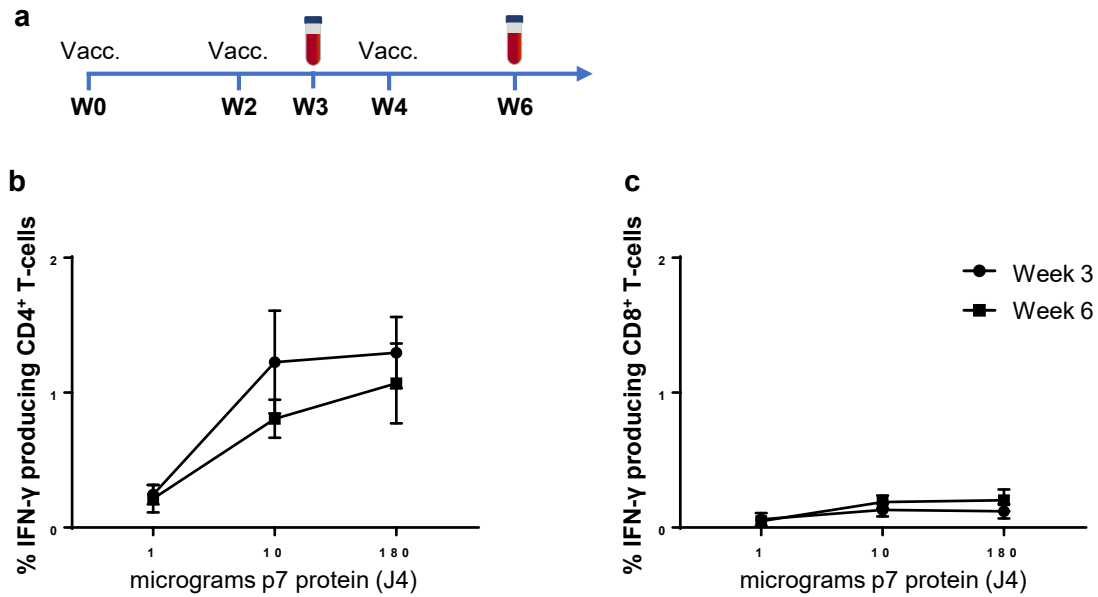
Supplementary Figures S1-8 and Table S1 for the manuscript entitled

“HCV p7 as a novel vaccine-target inducing multifunctional CD4⁺ and CD8⁺ T-cells targeting liver cells expressing the viral antigen”

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Supplementary Figure S1



Supplementary Figure S1

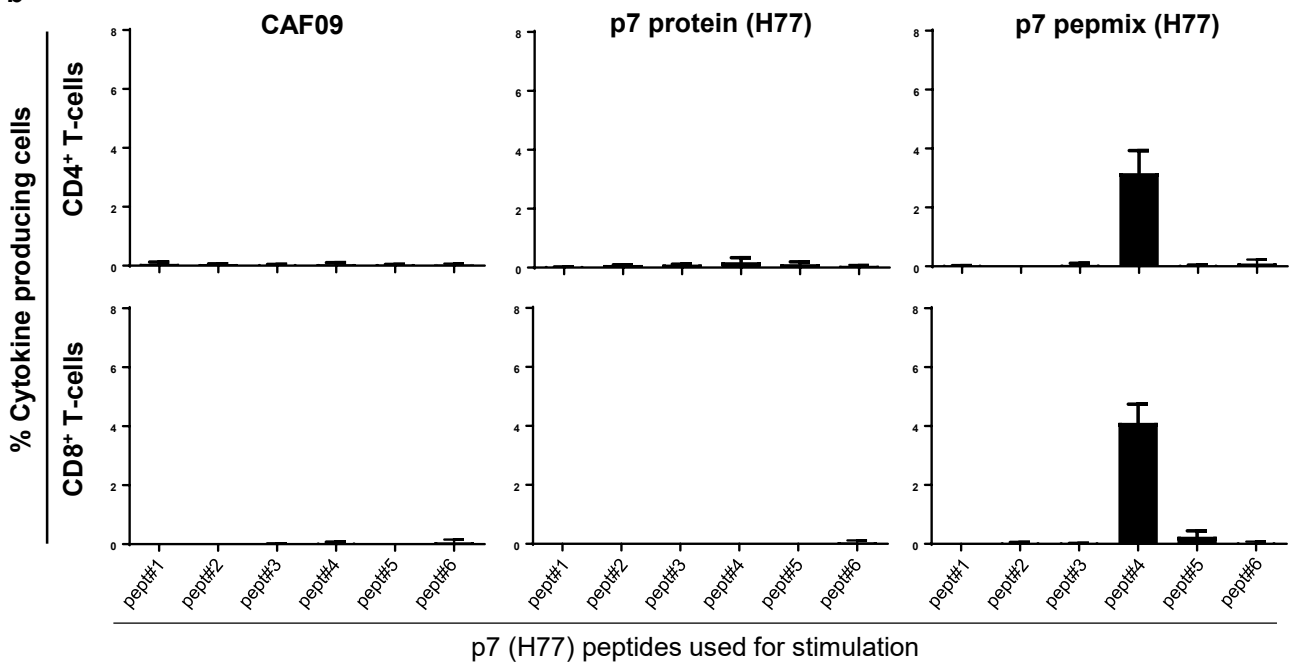
High antigen dose does not increase immunogenicity of the HCV p7 protein vaccine. CB6F1 mice were vaccinated three times at two-week intervals with 1 μ g, 10 μ g or 180 μ g of the HCV p7 protein (strain J4). (a) As illustrated on the time-line, PBMCs were harvested from individual mice three or six weeks after the priming vaccination and were restimulated with a pool of overlapping p7 (J4) peptides followed by FACS-analysis. Frequencies of antigen-specific IFN- γ producing CD44⁺ CD4⁺ T-cells (b) or CD44⁺ CD8⁺ T-cells (c) are shown as means and SEM on graphs (n = 5 to 6 mice in each vaccine group).

Supplementary Figure S2

a

p7 pepmix (H77)
Pept#1: ALENLVILNAASLAGTHGLV
Pept#2: ASLAGTHGLVSFLVFFCFAW
Pept#3: SFLVFFCFAWYLKGRWVPGA
Pept#4: YLKGRWVPGAVYAFYGMWPL
Pept#5: VYAFYGMWPLLLLLLALPQR
Pept#6: LLLLLALPQRAYA

b

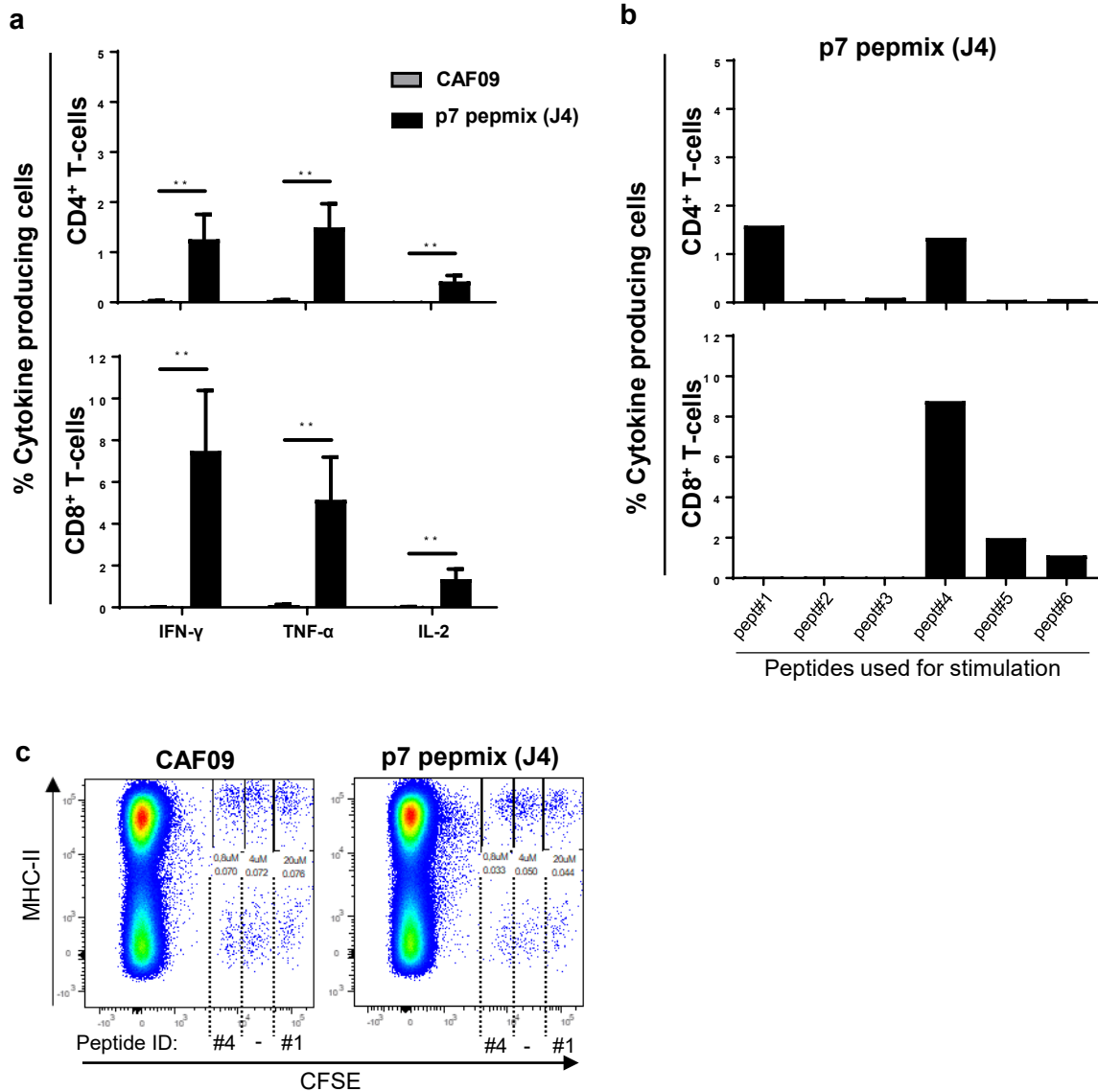


Supplementary Figure S2

Repertoires of p7 pepmix (H77) specific T-cells.

(a) The amino acid sequence of peptides comprising the p7 pepmix (H77) are shown. (b) Mice were vaccinated three times at 2-week intervals with the CAF09 adjuvant alone, HCV p7 protein or pepmix (H77) as indicated above the graphs. Splenocytes were isolated and pooled for each vaccine group and subsequently restimulated with each of the individual peptides (pept#1 to pept#6, indicated below the x-axis), to map the repertoire of epitope-specific responses. Bars show the total frequency of epitope-specific CD44⁺ CD4⁺ T-cells (upper panels) and CD44⁺ CD8⁺ T-cells (lower panels) able to produce IFN- γ , TNF- α or IL-2 in any combination. Data are shown as means and SEM from two independent experiments where splenocytes were pooled from 4-5 mice in each vaccine group.

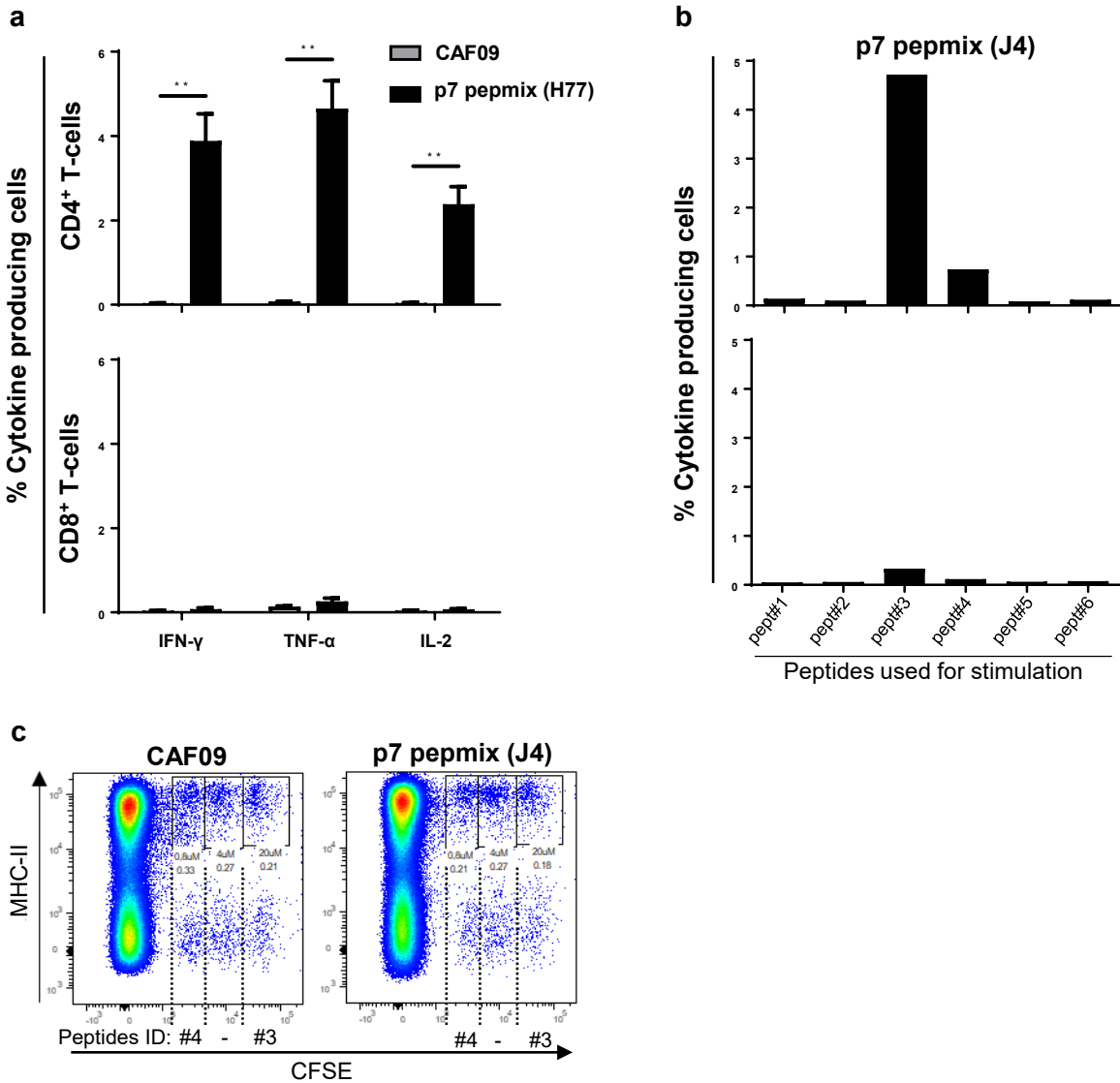
Supplementary Figure S3



Supplementary Figure S3

Cytotoxic potential of vaccine-induced T-cells in C57BL/6 mice. (a) Splenocytes from individual mice vaccinated with HCV p7 pepmix (J4) were isolated and restimulated with a pool of overlapping p7 (J4) peptides. Frequencies of antigen-specific IFN- γ , TNF- α and IL-2 producing CD4⁺ CD4⁺ T-cells (upper panel) and CD4⁺ CD8⁺ T-cells (lower panel) out of the total CD4⁺ or CD8⁺ T-cell population respectively, is shown in the bar charts as means and standard errors of the mean (SEM) (n = 6 mice in each vaccine group). ***P* < 0.01. (b) Splenocytes from p7 pepmix (J4) vaccinated mice were pooled (n = 6 mice) and restimulated with each of the individual p7 peptides to map the repertoire of epitope-specific CD4⁺ CD4⁺ T-cells (upper panel) and CD4⁺ CD8⁺ T-cells (lower panel) able to produce IFN- γ , TNF- α or IL-2 in any combination. (c) CFSE-labelled target cells from naïve mice were pulsed with single p7 peptides #1 or #4 or left untouched and subsequently injected i.v. into CAF09 and p7 pepmix (J4) vaccinated mice 12 days after the final immunization as indicated on the timeline in Fig. 2c. Eighteen hours after i.v. injection, splenocytes were isolated from individual mice and analyzed by FACS to determine their CFSE fluorescence. Representative plots of the three target cell populations in controls (left panel) and p7 pepmix vaccinated mice (right panel) were identified based on their CFSE-fluorescence and MHC-II expression (n = 5 mice in each vaccine group).

Supplementary Figure S4

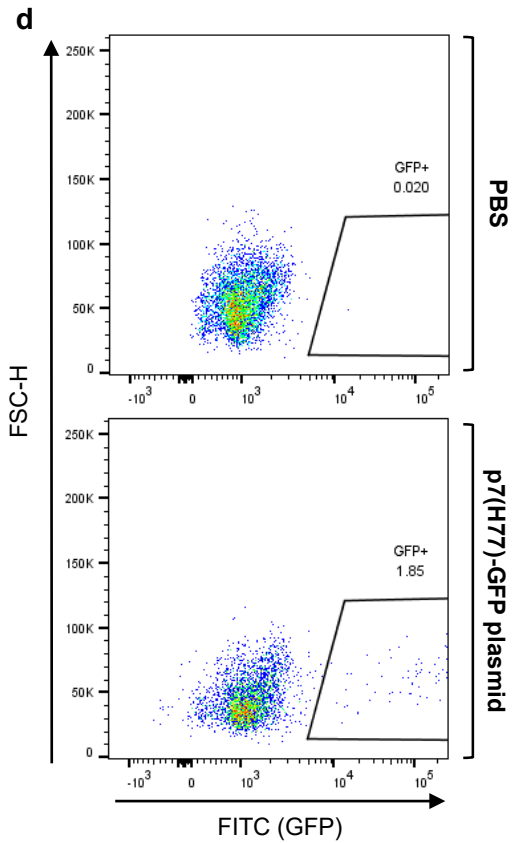
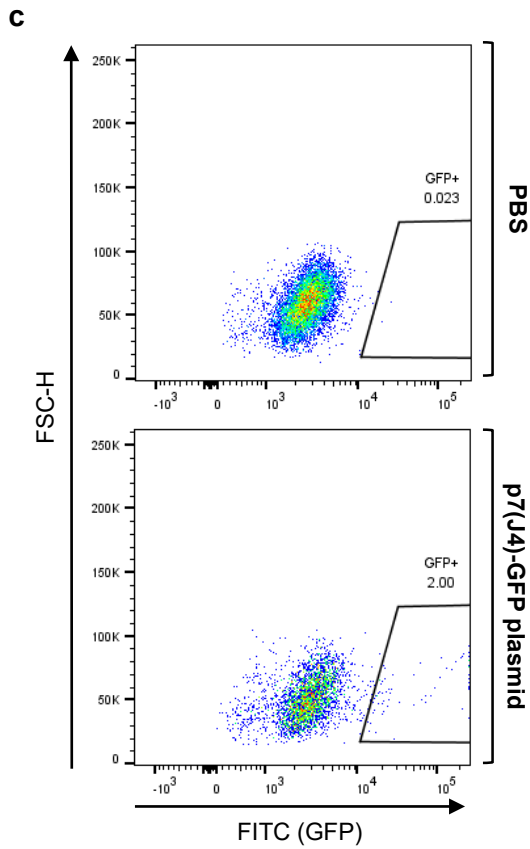
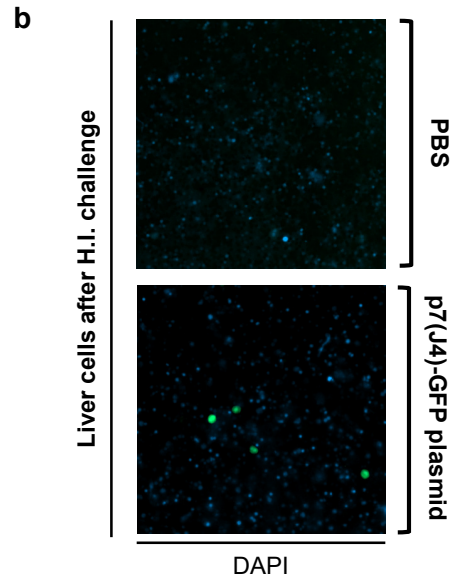
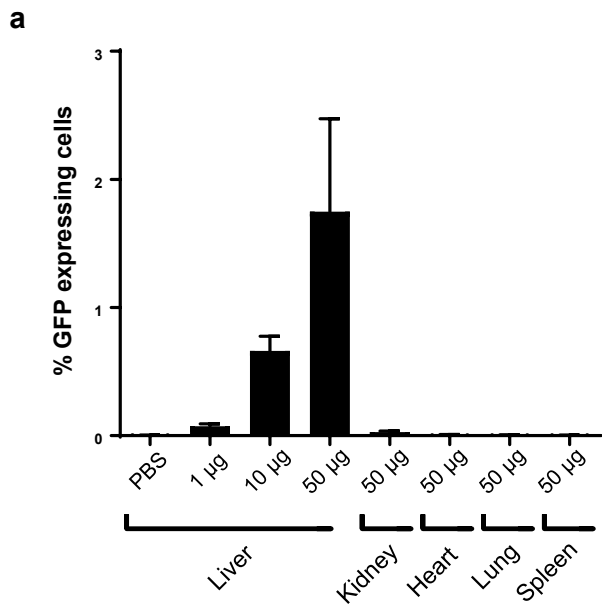


Supplementary Figure S4

Specific killing by cytotoxic CD4⁺ T cells in BALB/c mice

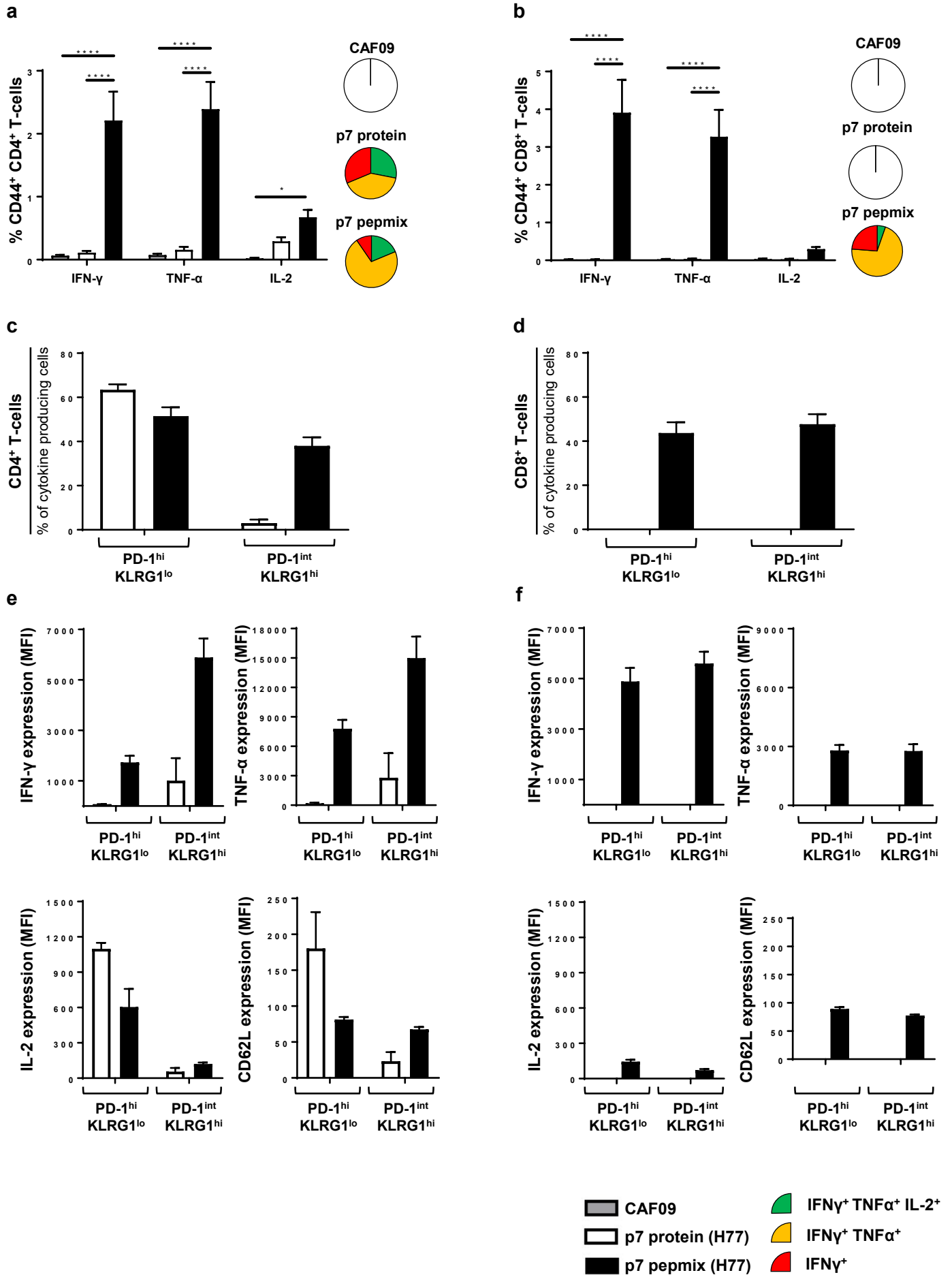
BALB/c mice were vaccinated three times at 2-week intervals with p7 pepmix (J4). (a) Splenocytes from individual mice were isolated and restimulated with a pool of overlapping p7 (J4) peptides and frequencies of antigen-specific, cytokine producing CD4⁺ CD4⁺ T-cells (upper panel) and CD4⁺ CD8⁺ T-cells (lower panel) is shown in the bar charts as means and standard errors of the mean (SEM) ($n = 5$ to 6 mice in each vaccine group). $**P < 0.01$. (b) Splenocytes from p7 pepmix (J4) vaccinated mice ($n = 5$ mice) were pooled restimulated with individual p7 peptides to map epitope-specific CD4⁺ CD4⁺ T-cells (upper panel) and CD4⁺ CD8⁺ T-cells (lower panel). (c) CFSE-labelled target cells from naïve mice were pulsed with the single p7 peptides #3 or #4 or left untouched and subsequently injected i.v. into CAF09 and p7 pepmix (J4) vaccinated mice 12 days after the final immunization. Eighteen hours after i.v. injection, splenocytes were isolated from individual mice and the three target cell populations were identified based on their CFSE- and MHC-II profile. The plots are representative for 5 mice in each vaccine group.

Supplementary Figure S5



Supplementary Figure S5. Surrogate challenge model based on liver specific expression of HCV p7 and GFP in naïve mice. (a) GFP-expressing liver cells from mice challenged with 1, 10 or 50 µg of p7(J4)-GFP plasmid, three days after hydrodynamic injection, were identified by FACS-analysis. Mice injected with 50 µg plasmid were also assessed for GFP-expressing cells in the kidney, heart, lung and spleen. Means and SEM are indicated on the graph (n = 5 to 6 mice per group). (b) DAPI-stained liver cells isolated from mice 4 days after hydrodynamic injection (H.I.) with PBS or p7(J4)-GFP plasmid were visualized by fluorescence microscopy to assess GFP-expression. (c) Representative FACS-plots showing gating of FITC-positives to identify GFP-expressing liver cells 4 days after injection with PBS (upper panel) or 100 µg p7(J4)-GFP plasmid (lower panel). (d) Similarly, GFP-expressing liver cells could be identified after hydrodynamic injection of plasmid DNA expressing strain H77 (genotype 1a) p7 and GFP, p7(H77)-GFP.

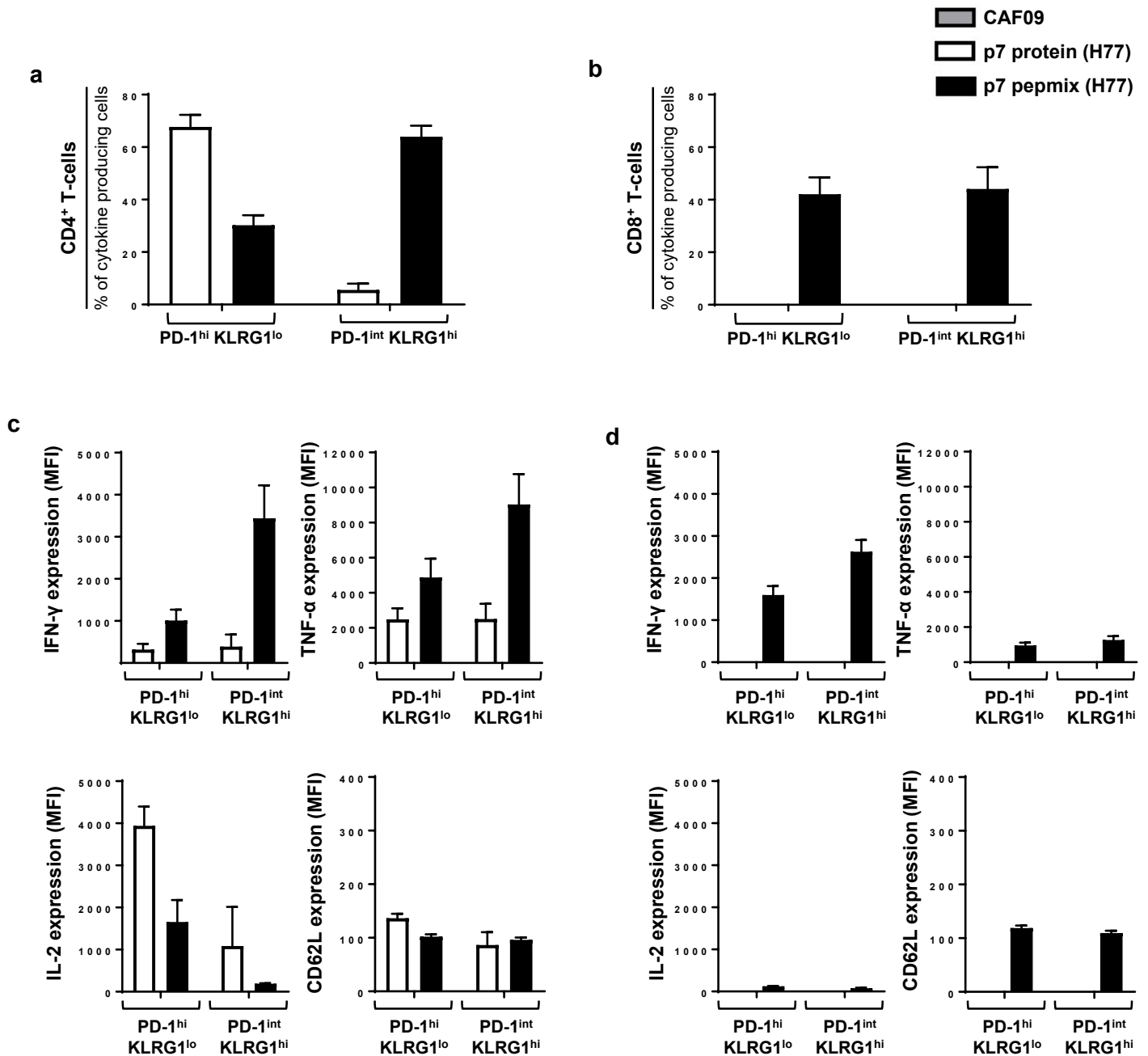
Supplementary Figure S6



Supplementary Figure S6

Pre-challenge cytokine- and phenotypic characterization of T-cell subsets in vaccinated mice. PBMCs purified from mice ten days after the final vaccination and immune responses were evaluated similar to post-challenge responses. Bars indicate frequencies of IFN- γ , TNF- α and IL-2 producing CD44⁺ CD4⁺ T-cells (**a**) or CD44⁺ CD8⁺ T-cells (**b**) and pie charts show the distribution of IFN- γ positives co-producing TNF- α alone or in combination with IL-2. The frequencies of p7-specific PD-1^{hi}/KLRG1^{lo} and PD-1^{int}/KLRG1^{hi} CD44⁺ CD4⁺ T-cells (**c**) and CD44⁺ CD8⁺ T-cells (**d**) is shown on graphs. Further analysis shows the relative expression (MFI) of IFN- γ , TNF- α , IL-2 and CD62L by these CD44⁺ CD4⁺ T-cells (**e**) and CD44⁺ CD8⁺ T-cells (**f**). Bars represent means and standard errors of the mean (SEM) (n = 7 to 12 mice in each vaccine group). *, $P < 0.05$; **** $P < 0.0001$.

Supplementary Figure S7



Supplementary Figure S7

Post-challenge characterization of T-cell subsets in vaccinated mice. Cytokine producing PBMCs purified from mice five days after challenge with p7(H77)-GFP plasmid (see Fig. 5a and 5b) were gated into PD-1^{hi}/KLRG1^{lo} and PD-1^{int}/KLRG1^{hi} T-cell subpopulations and assessed for their relative expression (MFI) of IFN-γ, TNF-α, IL-2 and CD62L (a and c, CD4⁺ T-cells; b and d, CD8⁺ T-cells) ten days after the final vaccination and immune responses were evaluated similar to post-challenge responses. Bars represent means and standard errors of the mean (SEM) (n = 7 to 12 mice in each vaccine KLRG group).

Supplementary Table S1.

In silico prediction of CD8⁺ T cell epitopes within p7 peptide #4 in CB6F1 mice

HCV strain	Epitope sequence	Allele	MHC-I IC50 (nM)
J4	AAYAFYGVWPL	H-2-K ^b	11.4
J4	YAFYGVWPL	H-2-D ^b	42.8
H77	AVYAFYGM	H-2-K ^b	5.6
H77	YAFYGMWPL	H-2-D ^b	38.6

Supplementary Table S1. Prediction of strong CD8⁺ T cell epitopes defined by the highest total score for proteasomal processing, TAP transport and MHC class I binding with IC50 values ≤ 500 nm were computed for H-2-K^b, H-2-D^b, H-2-K^d, H-2-D^d, H-2-L^d alleles.