## SUPPLEMENTAL DATA

Supplemental Table 1. The MRM Settings and Retention Time for Metabolites Analyzed in This Study.

Metabolite	RT (min) <sup>a</sup>	Prec Ion	Prod Ion	Dwell	Frag (V)	CE (V)	Cell Acc (V)	Polarity
NaAD	8.53	665	136	30	380	35	3	Positive
NAD	8.61	664	136	30	380	37	3	Positive
NaMN	8.64	336	124	30	380	13	3	Positive
NMN	8.81	335	123	30	380	8	3	Positive
NA <i>O</i> G	3.78	286	124	30	380	30	3	Positive
NA <i>N</i> G	8.57	286	124	30	380	11	3	Positive
NaR	8.15	256	124	30	380	9	3	Positive
NR	8.21	255	123	30	380	9	3	Positive
NA	5.79	124	80	30	380	20	3	Positive
NAM	1.21	123	80	30	380	13	3	Positive
MeNA	1.43 <sup>b</sup>	138	52	30	380	50	3	Positive
Тд	7.48	138	92	30	380	22	3	Positive
Na <i>N</i> P	8.42	256	124	30	380	9	3	Positive
Na <i>N</i> Rha	8.25	270	124	30	380	12	3	positive

<sup>a</sup> Prec Ion: Precursor Ion, Prod Ion: Product Ion, Frag: Fragmentor Voltage, CE: Collision Energy, Cell

Acc: Cell Accelerator Voltage, RT: Retention Time.

<sup>b</sup> The retention time of MeNA was obtained with different LC-MS program (see Wu et al., 2018).

## Supplemental Table 2. Primers Used in This Study.

Primer Name	Sequence information (5' to 3') <sup>1</sup>	Function	
76C5FOR	GGAATTCATGGAGAAGAGTAATGGCCTTCGAGTGA (EcoR I site was underlined)	Protein expression by pMal-c2x	
76C5REV	GC <u>TCTAGA</u> CTAAAAAGATGATATAATCAATCAAATTTTGTAACGATTG (Xba I site was underlined)		
76C5F1_site mut	TGGGCTCCGCAACAA <mark>GAG</mark> GTTCTAAAGCATCGAGCCATTGG	Site-directed mutagenesis of 76C5 Site-directed mutagenesis of 76C5	
76C5R1_site mut	TCGATGCTTTAGAAC <mark>CTC</mark> TTGTTGCGGAGCCCATTTCAC		
76C5F2_site mut	CACATAATGGTTGG <mark>AAC</mark> TCGACTGTTGAGAGTGTTTGTGAAGGC		
76C5R2_site mut	CTCTCAACAGTCGA <mark>GTT</mark> CCAACCATTATGTGTCAGGAATCCC		
76C5F3_site mut	AGAGTGTTTGTGAA <mark>GGC</mark> GTCCCTATGATCTGTTTGCCTTTTC	Site-directed	
76C5R3_site mut		mutagenesis of 76C5	
76C4FOR	GGAATTCATGGAGAAGAGTAATGGCCTGCGAGT (EcoR I site was underlined)	te was underlined) FAGAGATTGATATG (Xba I site was by pMal-c2x	
76C4REV	GC <u>TCTAGA</u> CTAGAAAGATGATATAATTAATCAAATTTTGTAGAGATTGATATG (Xba I site was underlined)		
76C4F1_site mut	GGGCTCCACAACAA <mark>GAC</mark> GTTCTAAAGCATCGAGCTATTGGAGG	Site-directed mutagenesis of 76C4	
76C4R1_site mut	TCGATGCTTTAGAAC <mark>GTC</mark> TTGTTGTGGAGCCCATTTCACTATCT		
76C4F2_site mut	CACATAATGGTTGG <mark>AGC</mark> TCGACGGTTGAGAGTGTTTGTGA	Site-directed	
76C4R2_site mut		76C4	
76C4F3_site mut	AGAGTGTTTGTGAA <mark>G<b>CA</b>GTCCCTATGATCTGTTTGCCTTTTCG</mark>	Site-directed mutagenesis of 76C4	
76C4R3_site mut	ACAGATCATAGGGACTGCTTCACAAACACTCTCAACCGTCG		

<sup>1</sup> mutated nucleotide and corresponding amino acid codon were highlighted.

Sample name	Tissue	Age	Growth Condition
Sample_1	Young Seedlings	8 days	1/2 MS, long day (16/8)
Sample_2	Young seedling leaf	8 days	1/2 MS, long day (16/8)
Sample_3	Young seedling roots	8 days	½ MS, long day (16/8)
Sample_4	Young Seedlings	15 days	1/2 MS, long day (16/8)
Sample_5	Young seedling leaf	15 days	½ MS, long day (16/8)
Sample_6	Young seedling roots	15 days	1/2 MS, long day (16/8)
Sample_7	Young Seedlings	21 days	½ MS, long day (16/8)
Sample_8	Young seedling leaf	21 days	1/2 MS, long day (16/8)
Sample_9	Young seedling roots	21 days	½ MS, long day (16/8)
Sample_10	Rosette leaf	30 days	Soil, long day (16/8)
Sample_11	Cauline leaf	30 days	Soil, long day (16/8)
Sample_12	Senescent leaves	35 days	Soil, long day (16/8)
Sample_13	1st node	35 days	Soil, long day (16/8)
Sample_14	Stem, 2nd internode	35 days	Soil, long day (16/8)
Sample_15	Inflorescence	35 days	Soil, long day (16/8)
Sample_16	Flowers stage 12	30 days	Soil, long day (16/8)
Sample_17	Flowers stage 13	30 days	Soil, long day (16/8)
Sample_18	Flowers stage 14	30 days	Soil, long day (16/8)
Sample_19	Flowers stage 15	30 days	Soil, long day (16/8)
Sample_20	Siliques, with seeds stage 3 and 4	30 days	Soil, long day (16/8)
Sample_21	Siliques, with seeds stage 5	30 days	Soil, long day (16/8)
Sample_22	Adult roots	35 days	Soil, long day (16/8)
Sample_23	Mature seeds	42 days	Soil, long day (16/8)
Sample_24	Fresh harvested pollen	30 days	Soil, long day (16/8)

Supplemental Table 3. List of 24 tissues used for NAD metabolomics analysis in this study.



**Supplemental Figure 1.** LC-QQQ-MS analysis (MRM, m/z 256 > 124) of nicotinate *N*-arabinoside (black line), nicotinate *N*-xyloside (red line) and the extract from leaf tissue of 8-day-old seedlings (blue line). Both nicotinate *N*-arabinoside and nicotinate *N*-xyloside are prepared by enzymatic reactions catalyzed by UGT76C5.



**Supplemental Figure 2.** Expression pattern of *At3g13050* in Arabidopsis, which was extracted from TAIR website (https://www.arabidopsis.org/; Winter et al., 2007).



Supplemental Figure 3. Quantification of nicotinate conjugates secreted in the liquid medium. In these experiments, around 3 mg of seeds, after sterilized with 75% alcohol and washed with ddH<sub>2</sub>O five times, were grown in 20 mL ½ MS liquid medium in a growth chamber for 15 days. The plant materials were weighed and 500 µL of liquid medium was mixed with equal volume of acetonitrile. The liquid samples were injected on UPLC-MS/MS for analysis after 10 minutes of maximum speed centrifugation with a microcentrifuge and filtered through a 0.22-µm syringe filter. Bars shown mean  $\pm$  SD (n = 3). Asterisks indicate significant differences from the Col-0 plants; \*\*\* P < 0.001, \*\* P < 0.01 (two-tailed Student's *t* test). F.W., fresh weight.

## REFERENCES

- Winter D, Vinegar B, Nahal H, Ammar R, Wilson GV, Provart NJ (2007) An "electronic fluorescent pictograph" browser for exploring and analyzing large-scale biological data sets. *PLoS One* 2, e718. doi: 10.1371/journal.pone.0000718
- Wu, R.R., Zhang, F.X., Liu, L.Y., Li, W., Pichersky, E., and Wang, G.D. (2018). MeNA, controlled by reversible methylation of nicotinate, is an NAD precursor that undergoes long-distance transport in Arabidopsis. *Mol. Plant* 11, 1264-1277. doi: 10.1016/j.molp.2018.07.003