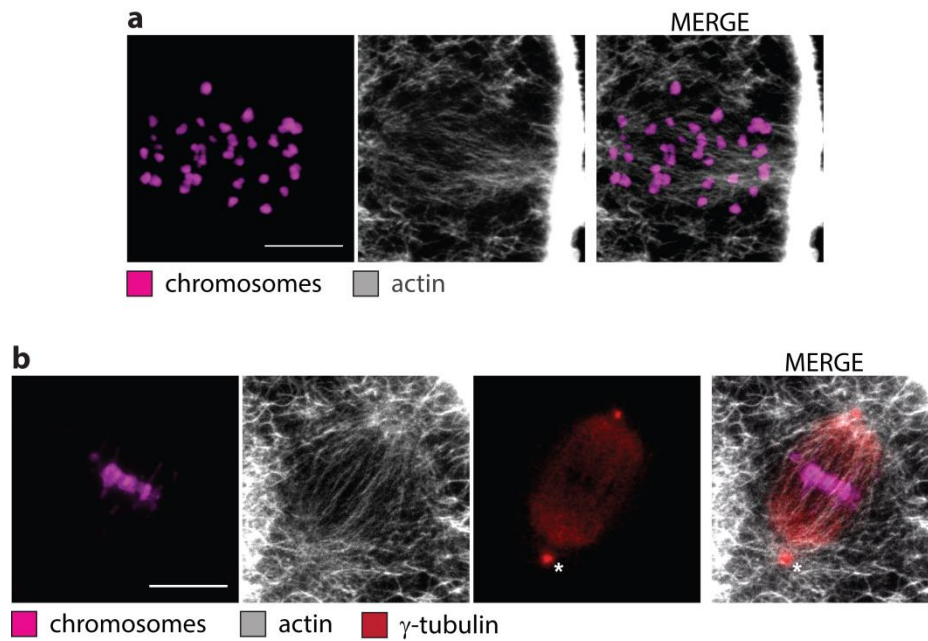


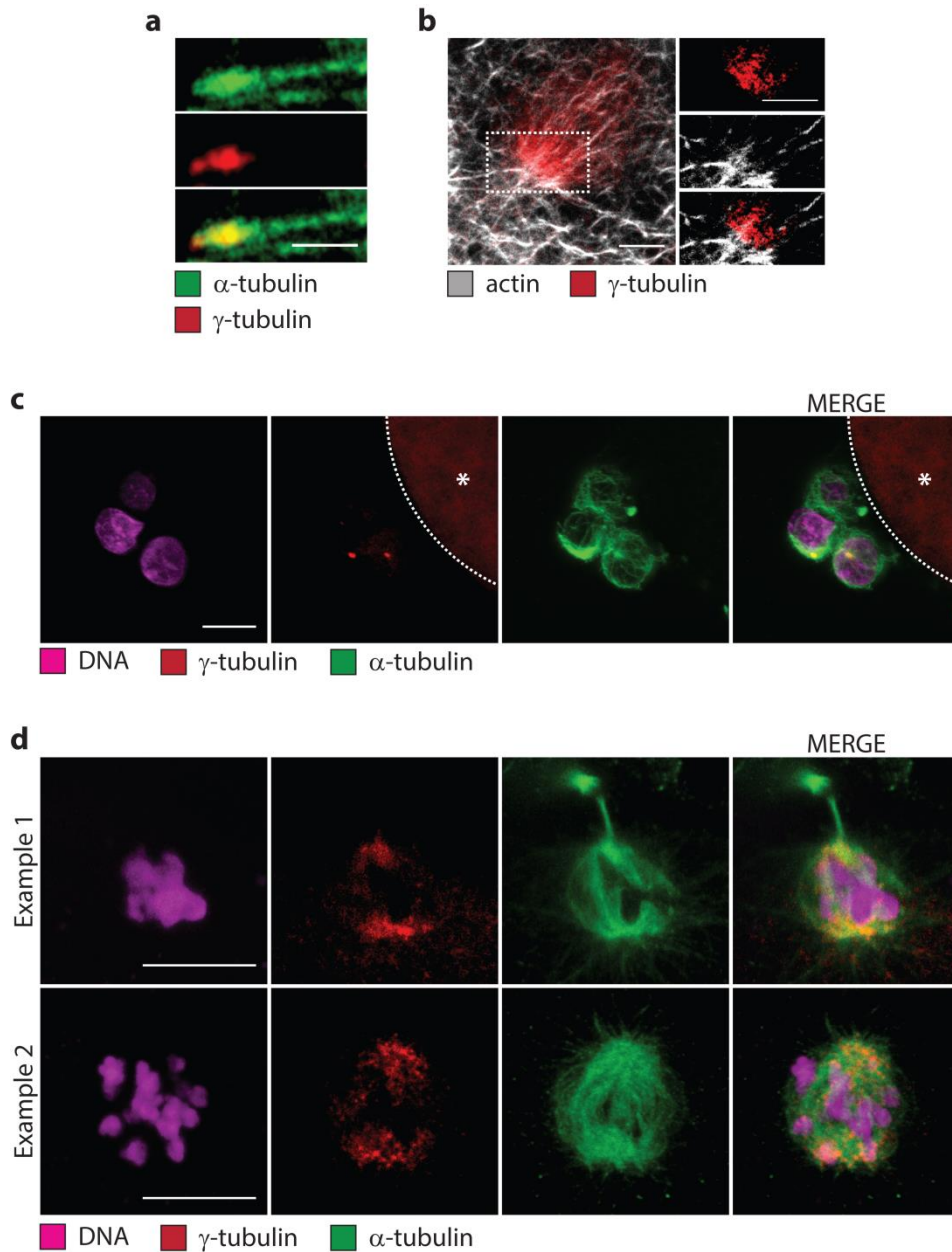
Supplementary Information

**Actin-microtubule interplay coordinates spindle assembly in
human oocytes**

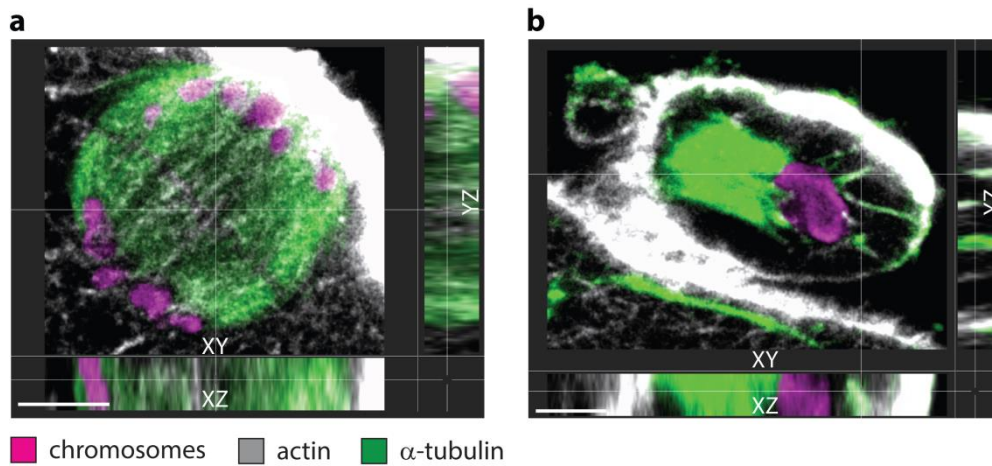
Roeles and Tsiavaliaris



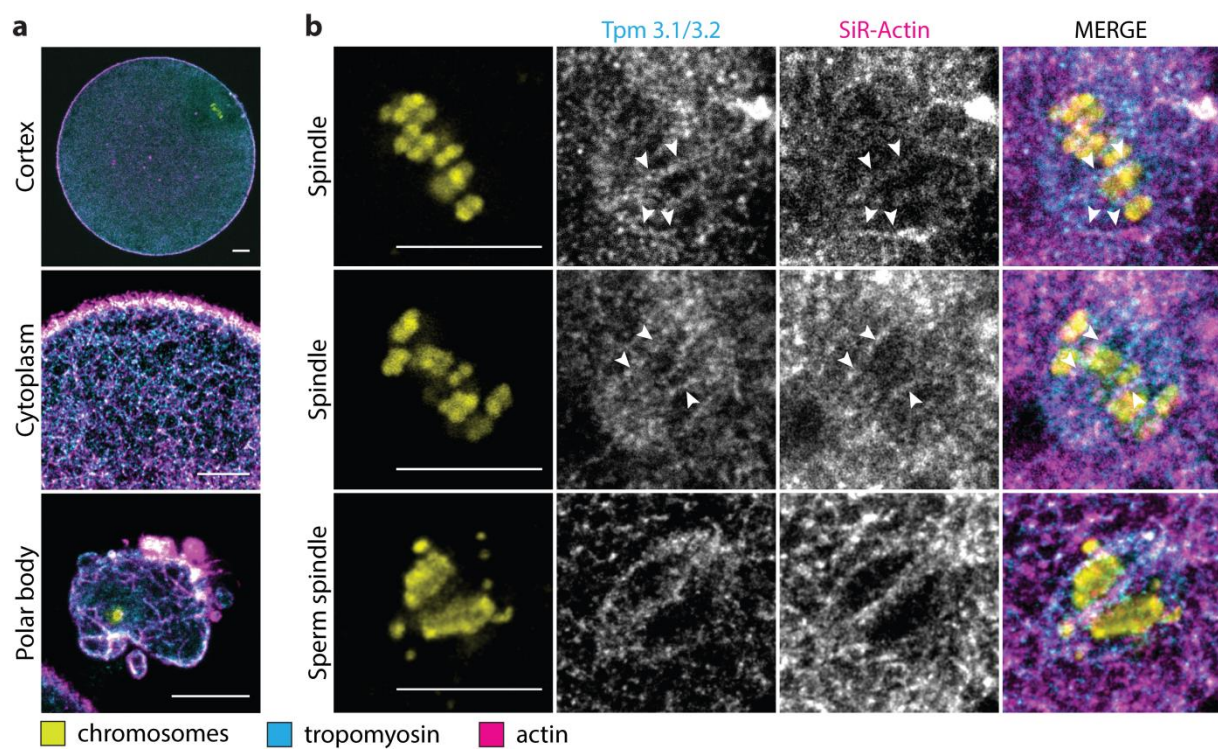
Supplementary Fig. 1 Actin pervades the spindle of oocytes with abnormal chromosome number and associates with microtubules surrounding sperm DNA. **a**, Representative z-projection of a diploid metaphase II oocyte stained for chromosomes (Hoechst) and actin (phalloidin); z = 25 sections. **b**, Immunofluorescence staining of γ -tubulin together with chromosomes (Hoechst) and actin (phalloidin) at a spindle structure surrounding the sperm DNA in an ICSI treated oocyte. Asterisk marks sperm centrosome; z = 13 sections. Scale bars, 10 μ m.



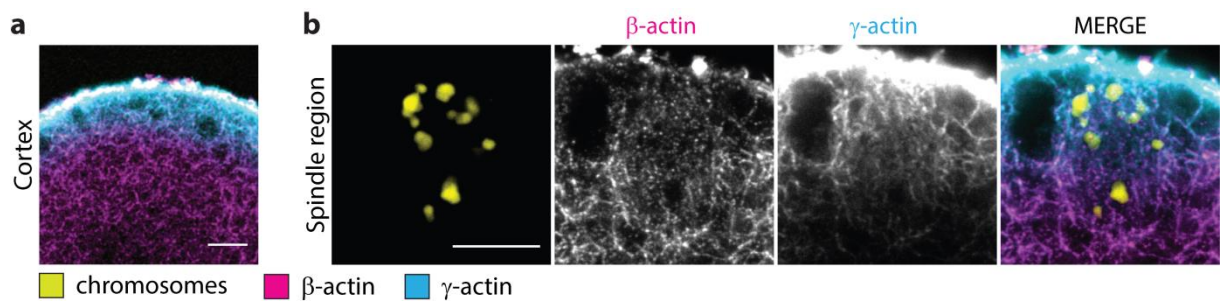
Supplementary Fig. 2 Characterization of γ -tubulin distribution in human oocytes. **a**, Immunofluorescence image showing microtubules (α -tubulin) and γ -tubulin. Scale bar, 2 μ m. **b**, Pole region of a metaphase II oocyte showing γ -tubulin and actin (phalloidin); $z = 7$ sections. Scale bar, 5 μ m. **c**, Distribution of γ -tubulin in oocyte cytoplasm and cumulus cells. Immunofluorescence staining shows DNA (Hoechst), microtubules (α -tubulin) and γ -tubulin. Asterisk marks oocyte; $z = 15$ sections. Scale bar, 10 μ m. **d**, Immunofluorescence projections showing chromosomes (Hoechst), microtubules (α -tubulin) and γ -tubulin in oocytes at early meiosis II; $z = 11$ sections. Scale bars, 10 μ m.



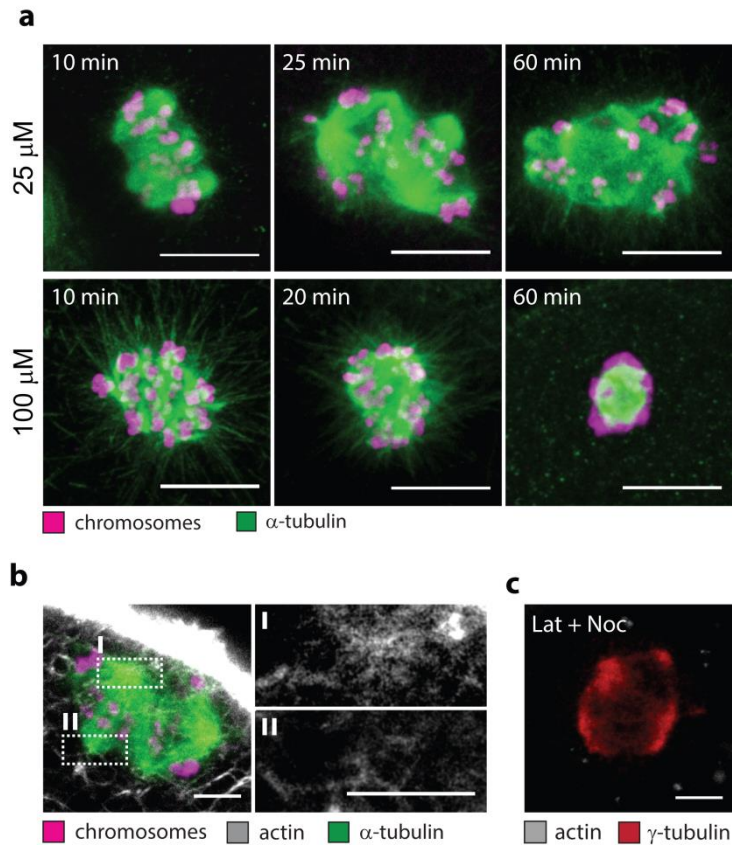
Supplementary Fig. 3 Actin and microtubules adopt distinct and overlapping distribution patterns. **a** and **b**, Orthogonal sections showing chromosomes (Hoechst), actin (phalloidin) and microtubules (α -tubulin) inside the centre volume of anaphase I (**a**) and telophase I (**b**) spindles. Depth (z) = 3 μm . Scale bars, 10 μm .



Supplementary Fig. 4 Spindle actin is associated with tropomyosins in human oocytes. **a**, Representative images showing tropomyosin Tpm3.1/3.2 and actin (SiR-actin) together with chromosomes (Hoechst) at the cortex, cytoplasm and polar body of metaphase II oocytes; $z = 1, 2$ and 1 sections. **b**, Close-up view of Tpm3.1/3.2 and actin (SiR-actin) at spindles surrounding maternal and paternal (sperm) chromatin in ICSI treated oocytes; $z = 3, 4$ and 3 sections. Arrowheads mark signal overlap at the spindle. Scale bars, 10 μm .

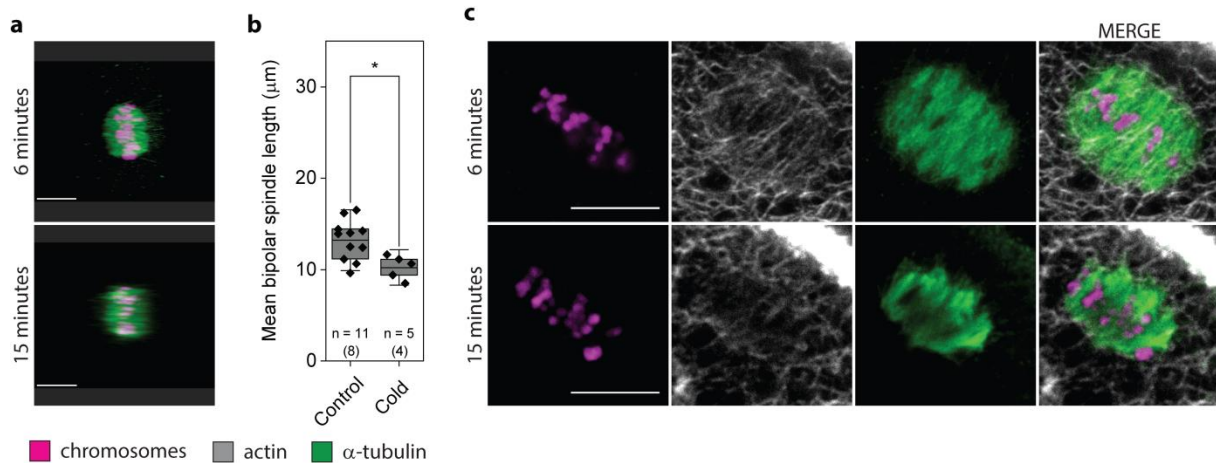


Supplementary Fig. 5 Different specific actin isoforms meet in the vicinity of the meiotic spindle. **a** and **b**, Total view and spindle close-up of metaphase II oocytes stained for chromosomes (Hoechst), β -actin and γ -actin; $z = 1$ and 9 sections. Scale bars, $10 \mu\text{m}$.

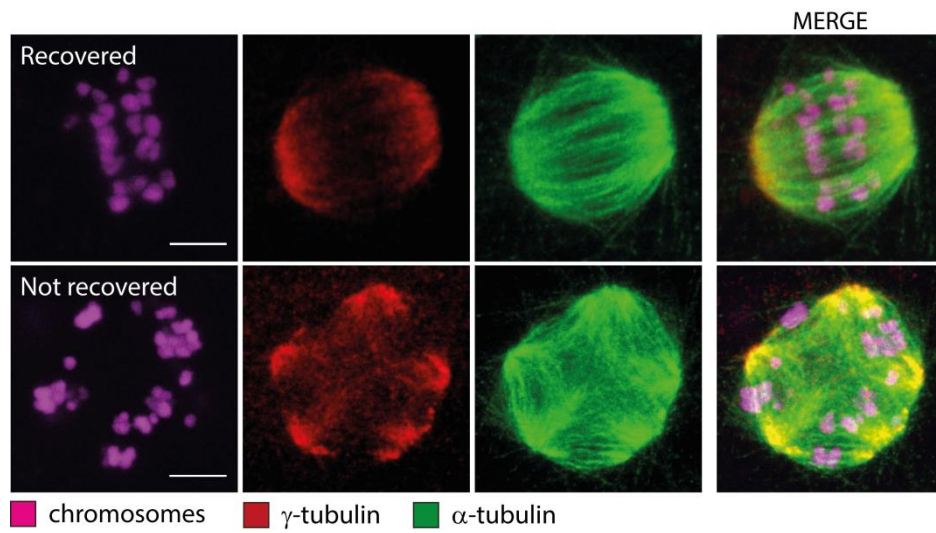


Supplementary Fig. 6 Microtubule decomposition by nocodazole affects spindle actin organization.

a, Immunofluorescence projections exemplifying chromosome and microtubule arrangement in nocodazole-treated oocytes. $z = 11$ sections. Scale bars, 10 μ m. **b**, Close-up view of actin accumulations (phalloidin) at different multipoles of a nocodazole-treated oocyte stained for chromosomes (Hoechst), actin (phalloidin) and α -tubulin; $z = 4$ sections. Scale bars, 5 μ m. **c**, Representative image of an oocyte treated with latrunculin-B and subsequently incubated in both latrunculin-B and nocodazole showing actin (phalloidin) and γ -tubulin; $z = 5$ sections. Scale bar, 5 μ m.



Supplementary Fig. 7 Microtubule decomposition by cold affects spindle actin organization. **a**, 3D snapshots of chromosomes (Hoechst) and microtubules (α -tubulin) exemplifying short spindles with broad poles in cold-treated oocytes. Scale bars, 10 μm . **b**, Mean length of bipolar spindles in untreated MII oocytes and oocytes exposed to cold for 6 to 15 minutes. Control measurements are equal to those shown in Fig. 3b. **c**, Example z-projections of cold-treated oocytes after 6 and 10 minutes showing chromosomes (Hoechst), actin (phalloidin) and α -tubulin; z = 3 and 4 sections. Scale bars, 10 μm . n = total number of oocytes. The number of donors is specified within parentheses. Centre lines are mean values; boxes illustrate 25th and 75th percentiles. Error bars represent standard deviation. Significance was tested using the two-tailed two sample t-test. Asterisk denotes p-value: * < 0.05. Source data are provided as a Source Data file.



Supplementary Fig. 8 γ -tubulin shows bipolar distribution in recovered oocytes. Representative z-projections of recovered and not recovered MII oocytes after nocodazole treatment and 60 to 90 minutes regeneration time showing chromosomes (Hoechst), α -tubulin and γ -tubulin; z = 4 sections. Scale bars, 5 μ m.