

Comparison of three seemingly similar lytic polysaccharide monoxygenases from *Neurospora crassa* suggests different roles in plant biomass degradation

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## **SUPPORTING INFORMATION**

**Supplementary Tables (S1) and Figures (S1-S10)**

TABLE S1

**Table S1.** X-ray data collection and refinement statistics for the structure of *Nc*LPMO9A (catalytic domain)

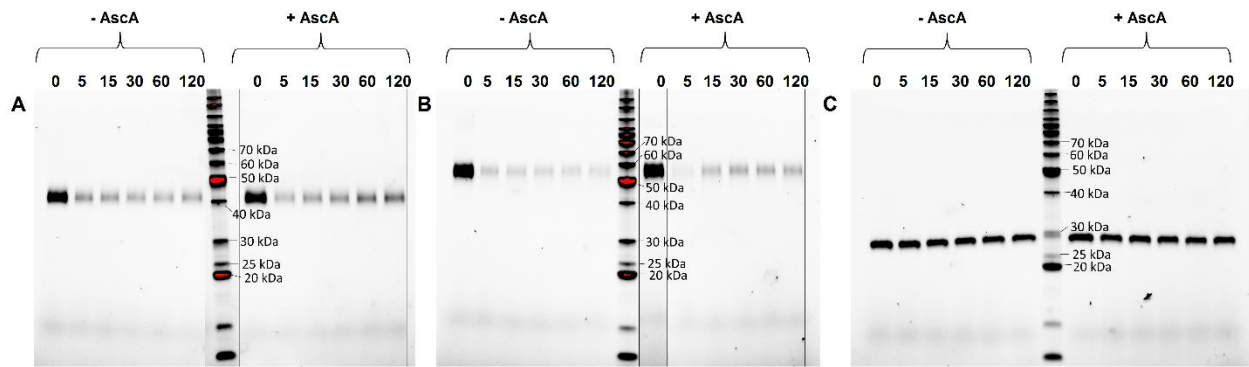
<b>Data collection</b>	
Space group	<i>P</i> 3 <sub>2</sub> 21
Cell dimensions (a, b, c) (Å)	a=80.4, b=80.4, c=57.9
$\alpha$ , $\beta$ , $\gamma$ (°)	90, 90, 120
No. of molecules per asymmetric unit	1
Station (synchrotron)	ID23-2 (ESRF, Grenoble)
Wavelength (Å)	0.8726
Resolution* (Å)	44.51-1.6 (1.63-1.6)
No. of observations	269267
No. of unique reflections	28860
Completeness* (%)	100 (100)
<i>R</i> <sub>merge</sub> <sup>1</sup> (%)	9.8 (100)
Mean( <i>I</i> / <i>sd</i> ( <i>I</i> ))*	14 (2.3)
CC1/2*	99.9 (69.8)
Multiplicity*	9.3 (8.9)
<b>Refinement</b>	
<i>R</i> <sub>work</sub> / <i>R</i> <sub>free</sub> (%)	0.154/0.178
R.m.s.d., bond lengths (Å)	0.009
R.m.s.d., bond angles (°)	1.35
No. of reflections	27351
No. of protein atoms	1632
No. of solvent molecules	243
No. of sulfate ions	3
No. of mannose residues	2
No. of metal atoms	2 (1 Cu, 1 Li)
Average B factor (Å <sup>2</sup> ) for protein residues	
Overall	17.73
Main chain atoms	17.15
Side chain atoms	18.4
Average B factor (Å <sup>2</sup> ) for heteroatoms	
Water molecules	30.06
Metal atoms	14.57 (Cu) 24.27 (Li)
Mannose residues	50.93
Sulfate ions	63.97
Ramachandran plot <sup>2</sup> (%)	
Favoured region	98.14
Outliers	0
PDB entry	5FOH

\*Highest resolution shell is shown in parentheses

$$^1R_{\text{merge}} = \frac{\sum_{hkl} \sum_i |I_i(hkl) - \langle I(hkl) \rangle|}{\sum_{hkl} \sum_i I_i(hkl)}$$

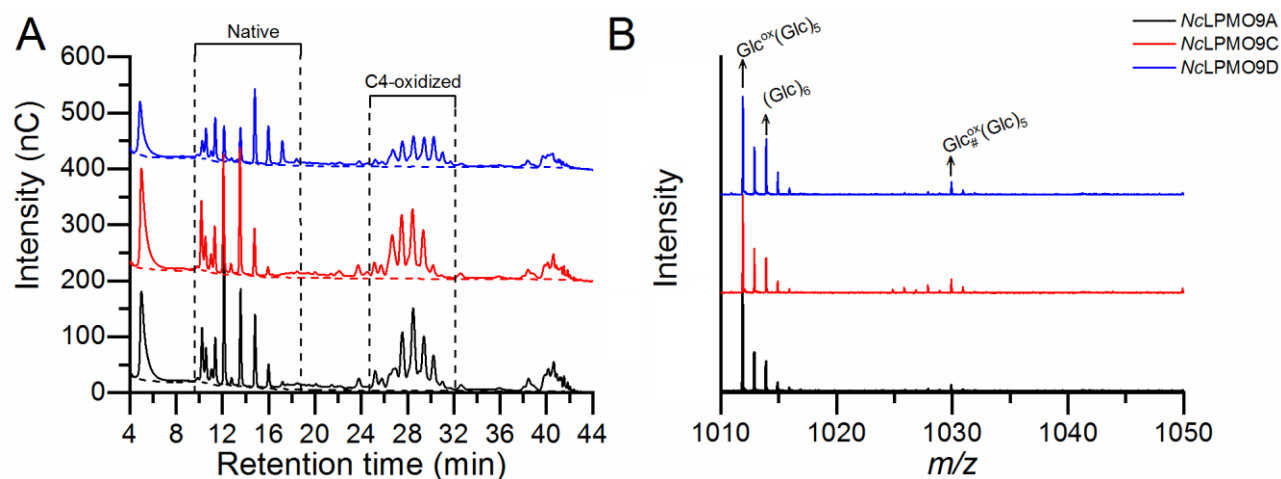
<sup>2</sup>Calculated using a strict boundary Ramachandran plot

FIGURE S1



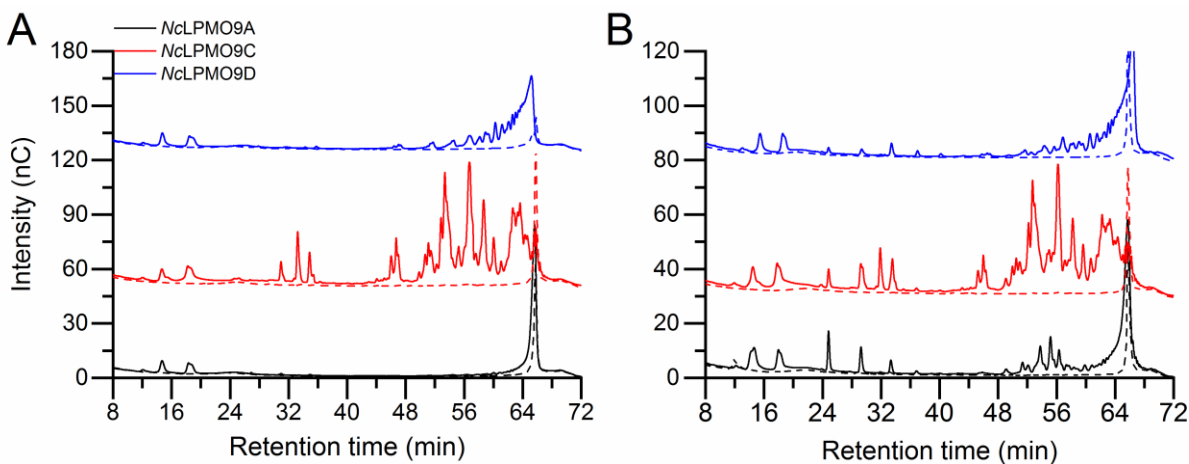
**Figure S1. SDS-PAGE of unbound C4 oxidizing *NcLPMOs* during incubation with 2 mg·ml<sup>-1</sup> PASC, in the absence (-AscA) or presence (+AscA) of 1 mM ascorbic acid.** The amounts of unbound (A) *NcLPMO9A*, (B) *NcLPMO9C*, and (C) *NcLPMO9D* were determined by SDS-PAGE at different time points after starting the incubation of enzyme with substrate. The unbound fractions were separated by filtration through 0.22 μM filter and 2.5 μL of the filtrates were incubated with 2.5 μL of SDS sample buffer and loaded onto the gels.

FIGURE S2



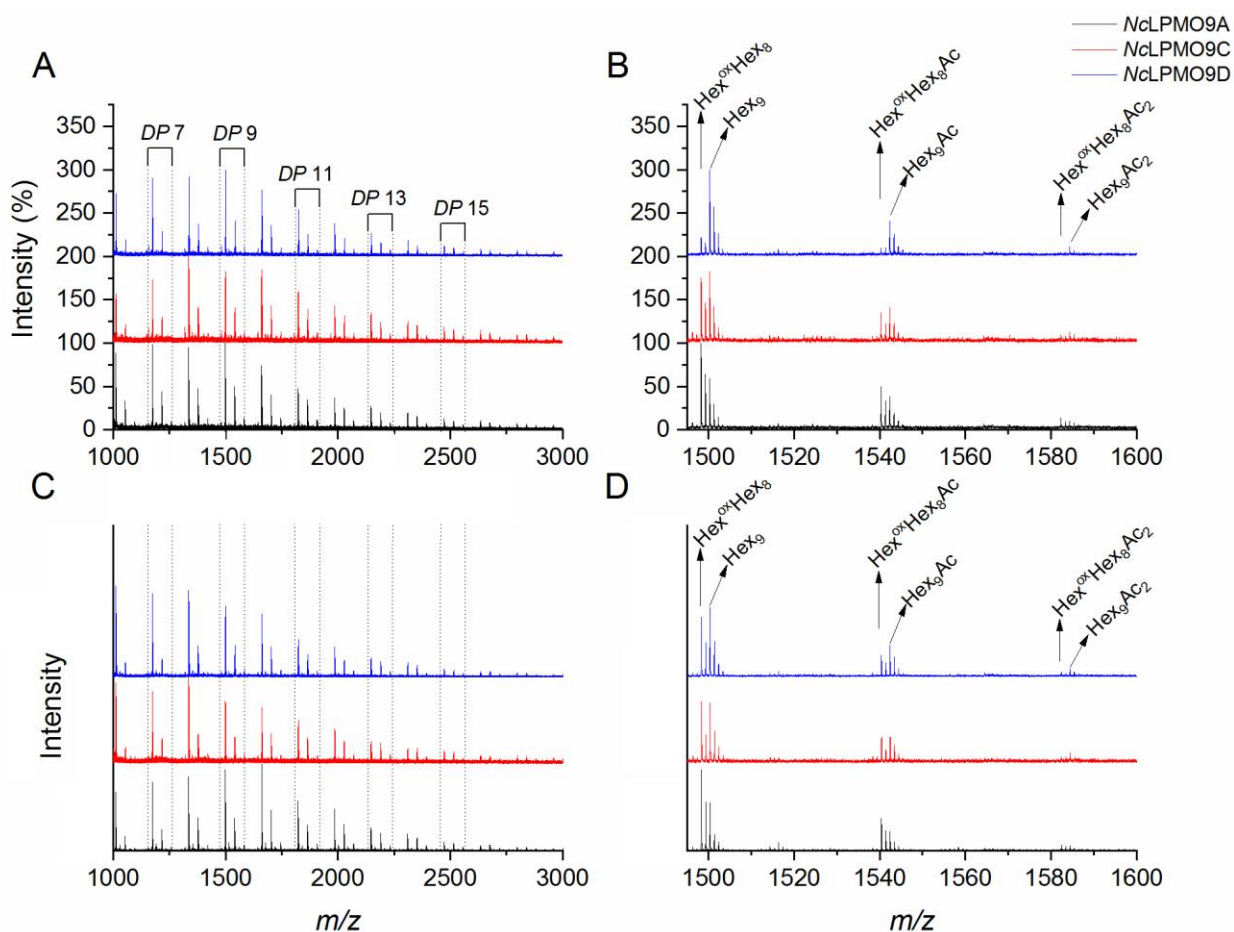
**Figure S2. Soluble products generated from PASC by C4 oxidizing *NcLPMOs*.** (A) HPAEC-PAD profiles of reaction mixtures containing *NcLPMO9A* (black), *NcLPMO9C* (red) or *NcLPMO9D* (blue), and PASC, with (solid lines) and without (dashed lines) ascorbic acid. The produced oxidized cello-oligosaccharides are labeled in the figure and annotations are based on previous work (Isaksen, Westereng et al. 2014). (B) Close-up of the DP 6 cluster in a MALDI-ToF MS spectrum of the products, showing the sodium adduct of the native,  $(\text{Glc})_6$ , the oxidized keto-form,  $\text{Glc}^{\text{ox}}(\text{Glc})_5$ , and the oxidized gemdiol form,  $\text{Glc}_{\#}^{\text{ox}}(\text{Glc})_5$ .

FIGURE S3



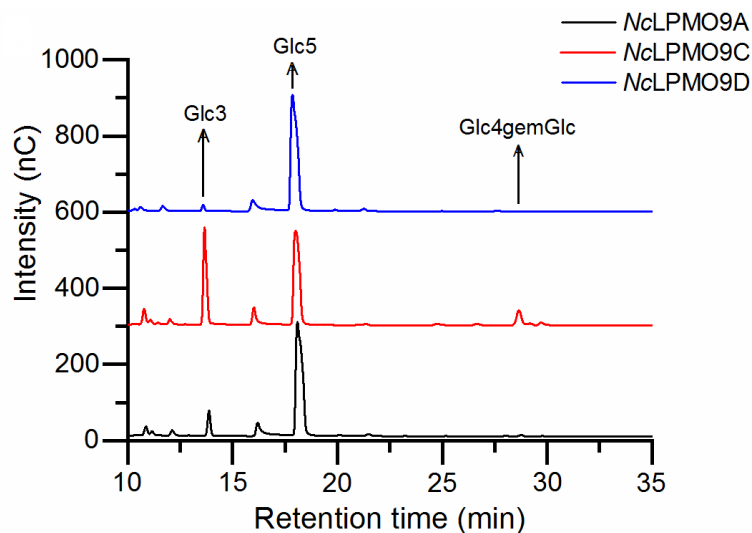
**Figure S3. HPAEC-PAD profiles of soluble reaction products generated by C4 oxidizing NcLPMOs from (A) TXG and (B) TXG coated on PASC.** Reaction mixtures contained 1  $\mu\text{M}$  NcLPMO9A (black), 1  $\mu\text{M}$  NcLPMO9C (red) or 1  $\mu\text{M}$  NcLPMO9D (blue), and (A) 2  $\text{mg}\cdot\text{mL}^{-1}$  TXG, or (B) 2  $\text{mg}\cdot\text{mL}^{-1}$  TXG and 2  $\text{mg}\cdot\text{mL}^{-1}$  PASC, with (solid lines) and without (dashed lines) ascorbic acid.

FIGURE S4



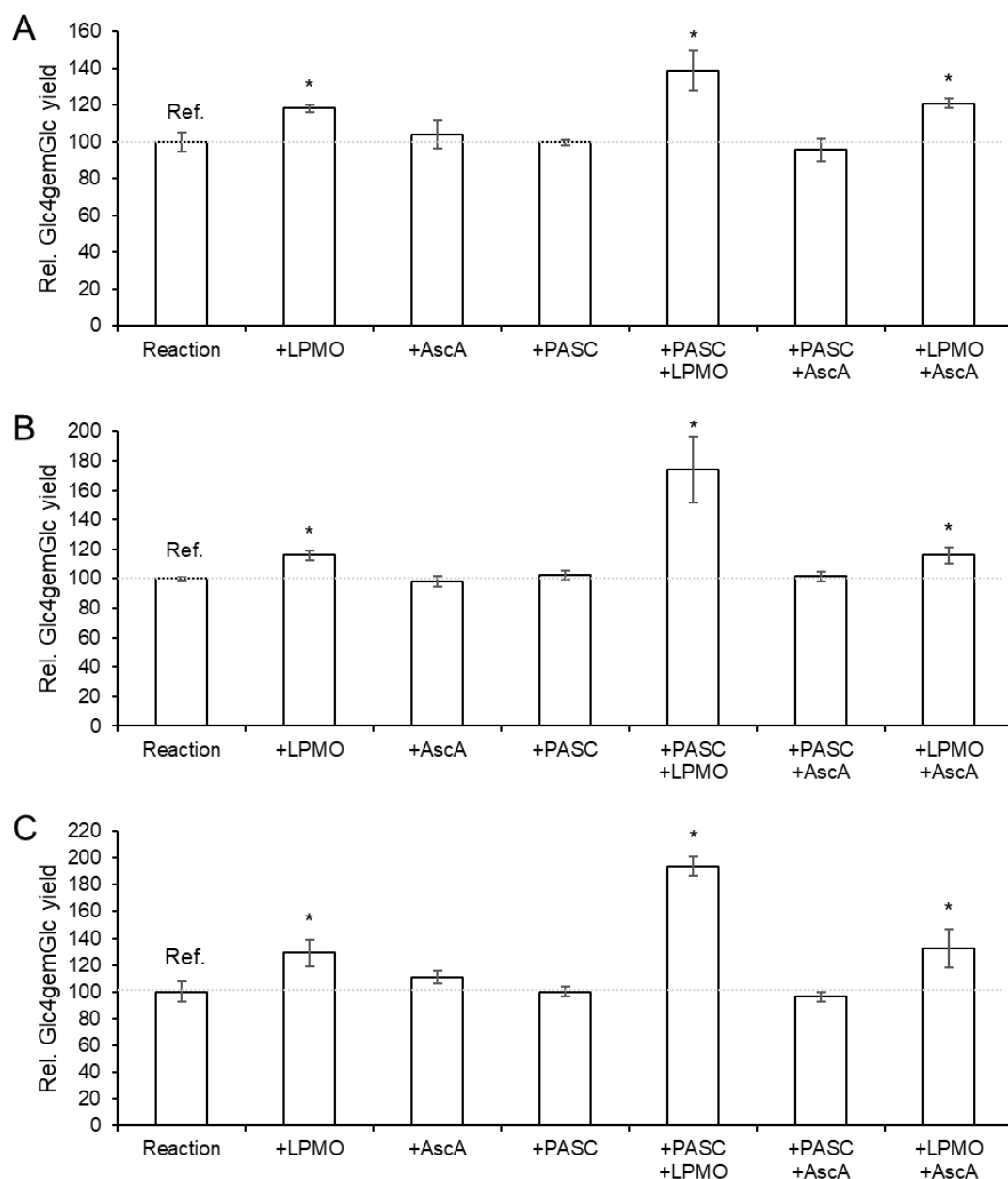
**Figure S4. Reaction products generated from konjac glucomannan (KGM), or from KGM coated on PASC.** The figures show MALDI-ToF MS spectra of products generated from KGM (A), with a close-up of the DP 9 cluster (B), or KGM coated on PASC (C), with a close-up of the DP 9 cluster (D), by *NcLPMO9A* (black), *NcLPMO9C* (red) and *NcLPMO9D* (blue). Brackets indicate product clusters of the same DP. Due to the identical  $m/z$  values of KGM and PASC products, it is not possible to distinguish them by MALDI-ToF MS, however, the appearance of acetylated and double acetylated products (typical for KGM), as well as their oxidized forms, confirms activity on KGM (acetylated products were not observed in reactions with only PASC; Fig. S2B). Abbreviations: Hex, hexose (+162 Da); Ac, acetyl group (+42 Da); ox, oxidized (-2 Da for keto form). The species in panels B and D include: native DP9, ( $m/z$  1500), oxidized DP9 (-2 Da;  $m/z$  1498) for DP9), and their acetylated (+42 Da;  $m/z$  1542 and 1540, respectively) and double acetylated (+84 Da;  $m/z$  1584 and 1582, respectively) forms.

FIGURE S5



**Figure S5. Reaction products generated by C4 oxidizing *NcLPMOs* from cellopentaose.** HPAEC-PAD chromatograms of products generated in reactions containing 1  $\mu$ M *NcLPMO9A* (black line), 1  $\mu$ M *NcLPMO9C* (red line) or 1  $\mu$ M *NcLPMO9D* (blue line), and 2.4 mM cellopentaose in the presence of 1 mM ascorbic acid (the oxidized product, Glc<sub>4</sub>gemGlc, is labeled). In control reactions without ascorbic acid, the cellopentaose was not cleaved.

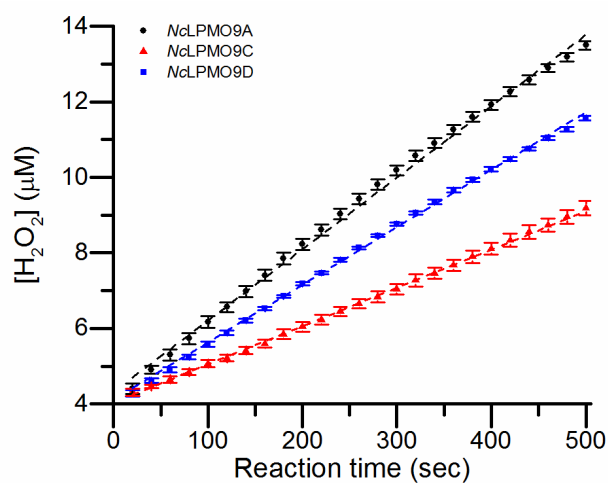
FIGURE S6



**Figure S6. Control experiments to assess the cause of the arrest in product formation by LPMOs.** Reactions were set up as in Figure 6C:  $2 \text{ mg}\cdot\text{ml}^{-1}$  PASC was incubated with  $1 \mu\text{M}$  (A) *NcLPMO9A*, (B) *NcLPMO9C* or (C) *NcLPMO9D* and  $3.3 \text{ mM}$  AscA in  $50 \text{ mM}$  Bis-Tris, pH 6.5, at  $45 \text{ }^\circ\text{C}$  and  $1000 \text{ rpm}$ . After  $240 \text{ min}$  incubation (i.e., the end point in Fig. 6C),  $100 \mu\text{l}$  of the reaction mixtures were supplemented with  $50 \mu\text{l}$  buffer or buffer containing various combinations of PASC, LPMO and AscA, followed by incubation for another  $120 \text{ min}$ . Solubilized oxidized products were enzymatically converted to Glc4gemGlc using  $1 \mu\text{M}$  *TrCel7A*, and the concentrations of Glc4gemGlc were determined by HPAEC-PAD, as in Figure 6. Samples were run in triplicates; error bars represent standard deviations. Product levels with statistically significant (one-tailed Student's t-test at  $\alpha=0.05$  significance level, probability  $p<0.05$ ) differences from the reference ("Buffer") are marked with an asterisk. Note that additional Glc4gemGlc was only produced when the reaction mixture was supplemented with LPMO; addition of PASC, AscA or both, in the absence of added LPMO, did not have a significant effect on the Glc4gemGlc yield.

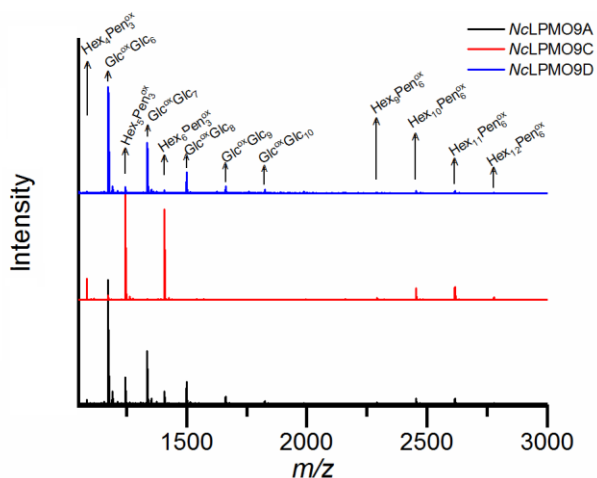


FIGURE S7



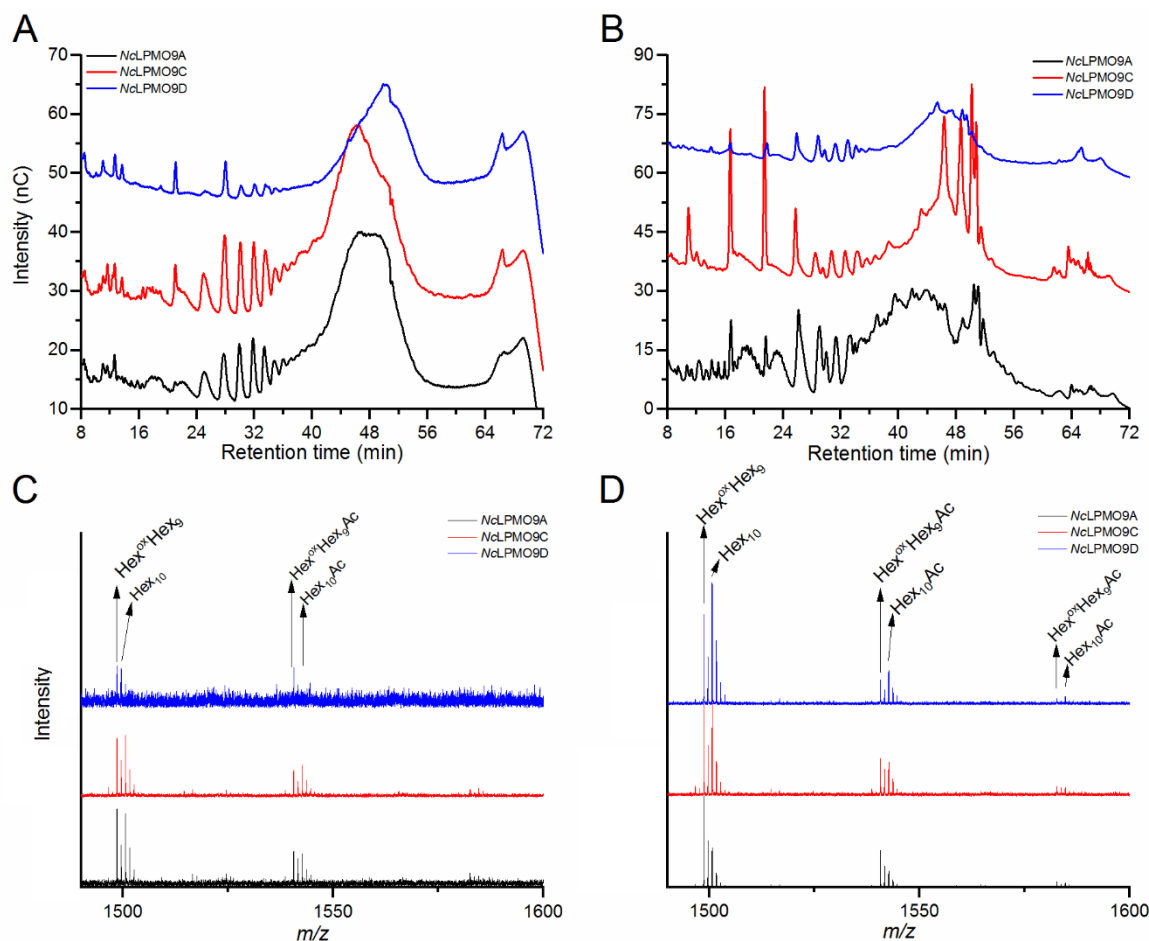
**Figure S7. Generation of H<sub>2</sub>O<sub>2</sub> by C4 oxidizing NcLPMOs.** 1 μM of NcLPMO9A (black), NcLPMO9C (red) or NcLPMO9D (blue) were incubated with 50 μM ascorbic acid in the absence of substrate, and the apparent production of H<sub>2</sub>O<sub>2</sub> was measured as described in “Materials and methods”. The figure shows data points in the linear region of the progress curves and error bars indicate standard deviations calculated from three experiments.

FIGURE S8



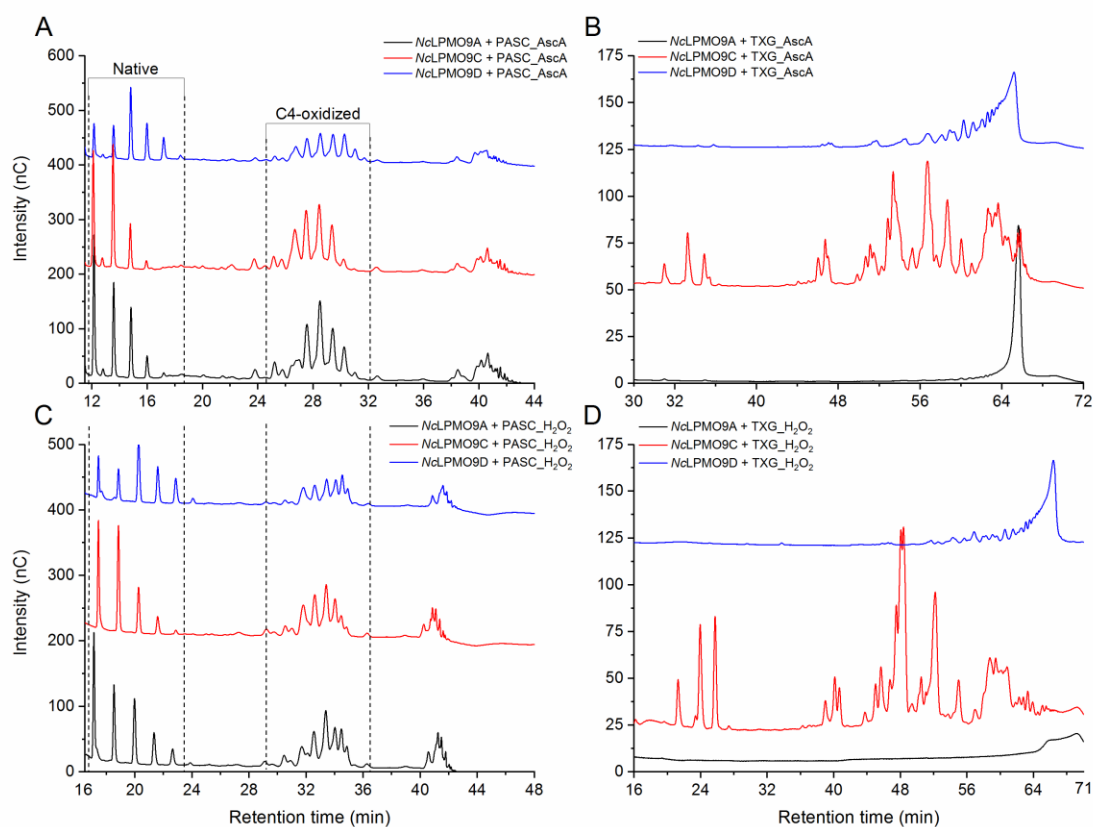
**Figure S8. Reaction products generated by C4 oxidizing *NcLPMOs* from TXG coated on PASC with  $\text{H}_2\text{O}_2$  as a co-substrate.** MALDI-ToF MS spectra showing product profiles generated from a mixture of  $2 \text{ mg}\cdot\text{mL}^{-1}$  TXG and  $2 \text{ mg}\cdot\text{mL}^{-1}$  PASC by  $1 \mu\text{M}$  *NcLPMO9A* (black), *NcLPMO9C* (red) or *NcLPMO9D* (blue) in  $50 \text{ mM}$  Bis-Tris, pH 6.5, at  $45^\circ\text{C}$  and  $1000 \text{ rpm}$ , with addition of  $\sim 45 \mu\text{M}$   $\text{H}_2\text{O}_2$  to the reactions every 15 min for 4 h. Prior to every addition of  $\text{H}_2\text{O}_2$ ,  $\sim 12 \mu\text{M}$  of ascorbic acid was added to ensure reduction of the LPMO. Control reactions were done in the absence of  $\text{H}_2\text{O}_2$  meaning that only  $\sim 12 \mu\text{M}$  ascorbic acid was added every 15 min for 4 h. In the control reactions only minute amounts of the oxidized products were detected. Oxidized products characteristic for xyloglucan ( $\text{Hex}_4\text{Pen}_3^{\text{ox}}$ ;  $\text{Hex}_5\text{Pen}_3^{\text{ox}}$ ;  $\text{Hex}_6\text{Pen}_3^{\text{ox}}$ ;  $\text{Hex}_9\text{Pen}_6^{\text{ox}}$ ;  $\text{Hex}_{10}\text{Pen}_6^{\text{ox}}$ ;  $\text{Hex}_{11}\text{Pen}_6^{\text{ox}}$ ; and  $\text{Hex}_{12}\text{Pen}_6^{\text{ox}}$ ) and for cellulose ( $\text{Glc}^{\text{ox}}(\text{Glc})_6$ ;  $\text{Glc}^{\text{ox}}(\text{Glc})_7$ ;  $\text{Glc}^{\text{ox}}(\text{Glc})_8$ ;  $\text{Glc}^{\text{ox}}(\text{Glc})_9$ ; and  $\text{Glc}^{\text{ox}}(\text{Glc})_{10}$ ), were identified. Abbreviations: Hex, hexose (+162 Da); Pen, pentose (+132 Da); Glc, glucose; ox, oxidized. For more discussion of the structure of the products, see the legend of Fig. 4 in the main manuscript.

FIGURE S9



**Figure S9. Reaction products generated by C4 oxidizing NcLPMOs from KGM or KGM coated on PASC with H<sub>2</sub>O<sub>2</sub> as a co-substrate.** Reaction mixtures contained 1 μM NcLPMO9A (black), 1 μM NcLPMO9C (red) or 1 μM NcLPMO9D (blue), and 2 mg·mL<sup>-1</sup> KGM, or 2 mg·mL<sup>-1</sup> KGM and 2 mg·mL<sup>-1</sup> PASC. (A) HPAEC-PAD profiles of soluble reaction products generated from 2 mg·mL<sup>-1</sup> KGM by 1 μM NcLPMO9A (black), 1 μM NcLPMO9C (red) or 1 μM NcLPMO9D (blue), in 50 mM Bis-Tris, pH 6.5, at 45 °C and 1000 rpm, with addition of ~45 μM H<sub>2</sub>O<sub>2</sub> to the reactions every 15 min for 4 h. Prior to every addition of H<sub>2</sub>O<sub>2</sub>, ~12 μM of ascorbic acid was added to ensure reduction of the LPMO. Products appear in the HPAEC-PAD chromatograms in the range between 24 and 40 min. (B) HPAEC-PAD profiles of soluble reaction products generated from a mixture of 2 mg·mL<sup>-1</sup> KGM and 2 mg·mL<sup>-1</sup> PASC by 1 μM NcLPMO9A (black), 1 μM NcLPMO9C (red) or 1 μM NcLPMO9D (blue), in 50 mM Bis-Tris, pH 6.5, at 45 °C and 1000 rpm, using the same feeding of H<sub>2</sub>O<sub>2</sub> and ascorbic acid as in panel A. Sharp peaks appearing at 11, 16, 21 and 25 min (absent in the reactions with only KGM) are cello-oligomers; peaks reflecting C4-oxidized cello-oligomers are not clearly visible because they overlap with peaks of KGM-derived products. (C) MALDI-ToF MS of soluble reaction products generated from KGM, showing a close up of the DP9 cluster. (D) MALDI-ToF MS of soluble reaction products generated from KGM coated on PASC, showing a close up of the DP9 cluster. Abbreviations: Hex, hexose (+162 Da); Ac, acetyl group (+42 Da); ox, oxidized (-2 Da for keto form). Control reactions without addition of H<sub>2</sub>O<sub>2</sub>, i.e. addition of ascorbic acid only, showed minute amounts of products, due to LPMO reactions fueled by ascorbic acid, at very low concentration, and O<sub>2</sub> (not shown).

FIGURE S10



**Figure S10. Comparison of the soluble products generated by C4 oxidizing *NcLPMOs* from PASC or TXG in reactions with only  $O_2$  or with  $H_2O_2$ .** This figure is a compilation of Figures 7A, 7B, S2A and S3A showing similarities between the product profiles generated in reactions with  $O_2$  and mM amounts of ascorbic acid (A, B) or with  $H_2O_2$  and priming amounts of ascorbic acid (C, D). Due to considerable drift in the chromatographic system, retention times vary, but the similarities in the peak profiles are nevertheless clear. (A and C) HPAEC-PAD profiles of soluble products generated by 1  $\mu M$  *NcLPMO9A* (black line), 1  $\mu M$  *NcLPMO9C* (red line) or 1  $\mu M$  *NcLPMO9D* (blue line), in reactions with 2  $mg \cdot mL^{-1}$  PASC and (A) 1 mM ascorbic acid or (C) addition of  $\sim 45 \mu M$   $H_2O_2$  to the reactions every 15 min for 4 h (prior to every addition of  $H_2O_2$ ,  $\sim 12 \mu M$  of ascorbic acid was added to ensure reduction of the LPMO), in 50 mM Bis-Tris, pH 6.5, at 45  $^\circ C$  and 1000 rpm. Dashed lines indicate the elution regions of native and oxidized cello-oligomers. (B and D) HPAEC-PAD profiles of soluble products generated by 1  $\mu M$  *NcLPMO9A* (black line), 1  $\mu M$  *NcLPMO9C* (red line) or 1  $\mu M$  *NcLPMO9D* (blue line), in reactions with 2  $mg \cdot mL^{-1}$  TXG and (B) 1 mM ascorbic acid or (D) addition of  $\sim 45 \mu M$   $H_2O_2$  to the reactions every 15 min for 4 h (prior to every addition of  $H_2O_2$ ,  $\sim 12 \mu M$  of ascorbic acid was added to ensure reduction of the LPMO), in 50 mM Bis-Tris, pH 6.5, at 45  $^\circ C$  and 1000 rpm.