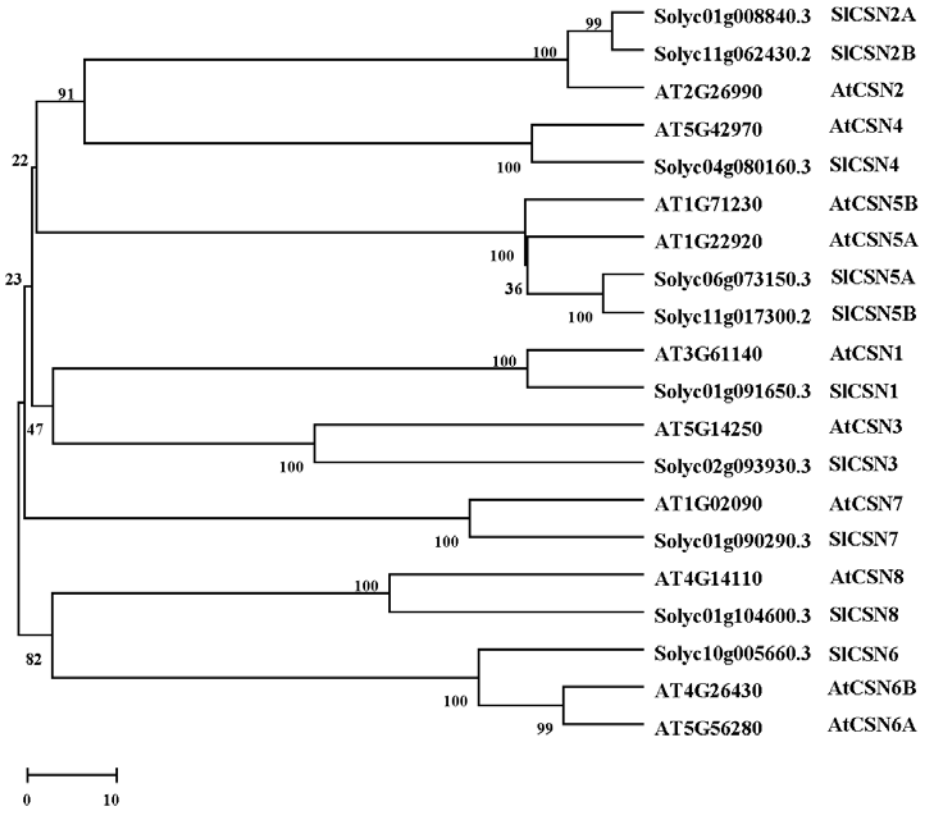
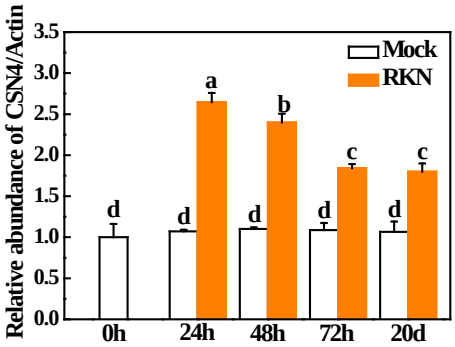
Supplemental Figure S1



Supplemental Figure S2

A



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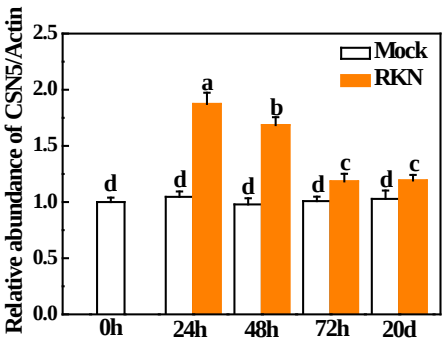
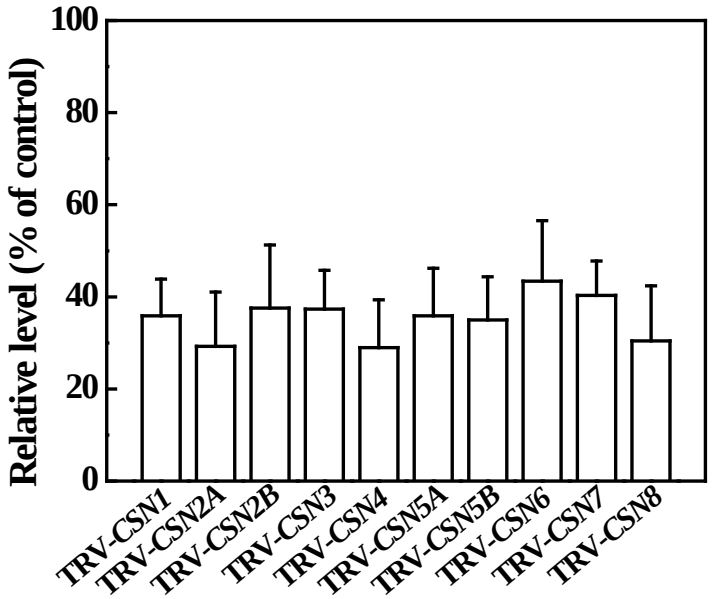
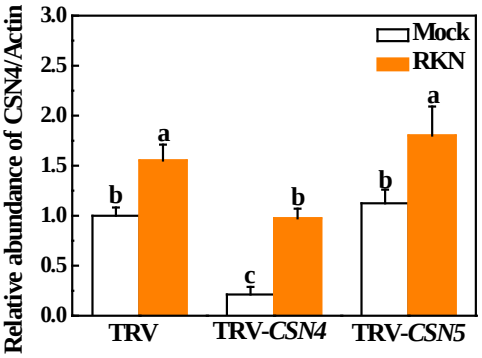


Supplemental Figure S3



Supplemental Figure S4

A



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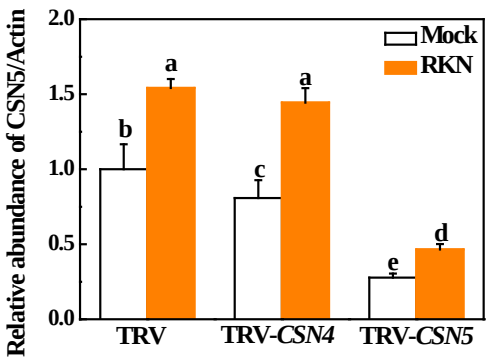

Supplemental Table S1 Primers used for VIGS vectors construction.

Gene name	Accession number	Restriction sites	PCR length	Forward primer	Reverse primer
CSN1	Solyc01g091650	BamHI/XhoI	299 bp (from +2 bp to +300 bp)	5′-agcggatccTGGAGCCTGACGAGGATT-3′	5′-agcctcgagATAATTAGTGCCCAAACGTC-3′
CSN2A	Solyc01g008840	BamHI/XhoI	300 bp (from +7 bp to +306 bp)	5′-agcggatccTCTGATGCGGATATGGA-3′	5′-agcctcgagACTATAGTTCCGTGTCACTG-3′
CSN2B	Solyc11g062430				
CSN3	Solyc02g093930	BamHI/XhoI	299 bp (from +2 bp to +300 bp)	5′-agcggatccTGAATCTGAACATGAATTCA-3′	5′-agcctcgagCAACTGGATTTGCTCTGTA-3′
CSN4	Solyc04g080160	EcoRI/BamHI	276 bp (from +713 bp to +988 bp)	5′-agcgaattcGTCCTCAACGATCTCGT-3′	5′-agcggatccGAATGCCCAACAAAGTGC-3′
CSN5A	Solyc06g073150	EcoRI/BamHI	367 bp (from +622 bp to +988 bp)	5′-agcgaattcTATAAGCCTCCAGATGACCC-3′	5′-agcggatccCGACAGTAATCTTAGCACTATCAC-3′
CSN5B	Solyc11g017300				
CSN6	Solyc10g005660	BamHI/XhoI	290 bp (from +11 bp to +300 bp)	5′-agcggatccCATCCAGCAGCGGCTTAACC-3′	5′-agcctcgagACACCTTCTTATAGAGCTC-3′
CSN7	Solyc01g090290	BamHI/XhoI	298 bp (from +325 bp to +533 bp)	5′-agcggatccAGTATTGCTGGCTGTCTTCC-3′	5′-agcctcgagACCAGTCAGTTAATGTCTG-3′
CSN8	Solyc01g104600	BamHI/XhoI	299 bp (from +2 bp to +300 bp)	5′-agcggatccTGGATACTTCTCAACTCAAC-3′	5′-agcctcgagTCCACGAATAGCCTCATGTA-3′

Supplemental Table S2 Primers used for qRT-PCR assays.

Gene name	Accession number	Forward primer	Reverse primer
ACTIN	Solyc11g005330	5′-TGTCCCTATTTACGAGGGTTATGC-3′	5′-CAGTTAAATCACGACCAGCAAGAT-3′
CSN1	Solyc01g091650	5′-GTCACTATGCTTCTTGTCTG-3′	5′-CGAGACAAATGGGTGGGTAT-3′
CSN2A	Solyc01g008840	5′-TATGGCAGAGAGAAAGTGGG-3′	5′-GTCATCCATTATTGTCCGTC-3′
CSN2B	Solyc11g062430	5′-GGGAGTATGGACGAATGAAT-3′	5′-AACCTCAGACTCCATCAGCA-3′
CSN3	Solyc02g093930	5′-TCACTGATTCATCTTGGGCA-3′	5′-CAACTTTTTCGACACTGCCA-3′
CSN4	Solyc04g080160	5′-GAAACCACATCAGCAAGCAC-3′	5′-ATCCCCGCATTCTATCTTCG-3′
CSN5A	Solyc06g073150	5′-CACGTTATGCATCCTTAATGG-3′	5′-ACCTGAGGATTCCGTCTGT-3′
CSN5B	Solyc11g017300	5′-TGGGCACTTAGTGGCAGC-3′	5′-TGGATCAGAGGGTTCAGTCT-3′
CSN6	Solyc10g005660	5′-TCCATCTACTCACTCACTTG-3′	5′-ACTCGCTTTCATAGATACTG-3′
CSN7	Solyc01g090290	5′-GGGCATAGTTAGAGGAAAGC-3′	5′-CTTCCTGTCCGATTCACTCA-3′
CSN8	Solyc01g104600	5′-TGGTGGCTGTTTGGAGGATT-3′	5′-AATAAGCGGAAAGAAGGAGC-3′
PI-2	Solyc03g020080	5′-GGATGCTTCTGGGATACGTT-3′	5′-TGGATGCTCCGAGACTACAG-3′
COI1	Solyc05g052620	5′-GGATGCTTCTGGGATACGTT-3′	5′-TGGATGCTCCGAGACTACAG-3′
LOXD	Solyc03g122340	5′-CCGTGGTTGACACATTATCG-3′	5′-ACAGCAGTCCGCCCTATTTA-3′
AOC	Solyc02g085730	5′-GCGCGAATTCACCTCAACAGATTCAACTAACACTG-3′	5′-GGGGCTCGAGTTAATTAGTGTAATTTTTCAGTGCGGC-3′
JAZ1	Solyc07g042170	5′-TTCCCTCAAGGTGGAATGAAGGCT-3′	5′-TCCGAAACTCGGAACCACCAAATC-3′
JAZ2	Solyc12g009220	5′-ACCTGATCAACCAGAGAAGGCA-3′	5′-AAACTCACACCAGATTGATCAGCTGT-3′
JAZ3	Solyc03g122190	5′-TTCCCTGCTGACAAAGCTAGAGCA-3′	5′-AGGGTGCAGATGAAACTGATCCGA-3′
JAZ4	Solyc12g049400	5′-GCCAAAGCCTCAGCAACAAAGGAT-3′	5′-ATCACTGCTCTGGCTTTCTCTGCT-3′
JAZ5	Solyc03g118540	5′-TCAGCTGTTCCGTCTAGCAGCATT-3′	5′-TGCATTTGGTGTAACAGGTGGTGC-3′

JAZ6	Solyc01g005440	5′-AGTCGATGCTGGTCTCAAACGTCA-3′	5′-TCGAAGACATTGACCATCCCACCA-3′
JAZ7	Solyc11g011030	5′-TTGCTATGGCTCGTAGAGCAACTC-3′	5′-TTTCCCAATGAACGCTTGACGACG-3′
JAZ8	Solyc06g068930	5′-TCGTCAACCTCCCAATCATAAC-3′	5′-GGAAAGGGTAGTGAGTGCATC-3′
JAZ9	Solyc08g036640	5′-TTTGGAGCTCACTCTTATGCCTCC-3′	5′-AGCTCAGTAGCATCGGAAACCACA-3′
JAZ10	Solyc08g036620	5′-GGAACTCACTCTTTCTCCTAGCAAC-3′	5′-TGGTGATGAAGGCTCAGACAGCTT-3′
JAZ11	Solyc08g036660	5′-GGAGTTTAGGCTTATGCCACCTTC-3′	5′-GGCTCAGATATTGGTGACAGACTC-3′
JAZ12	Solyc01g009740	5′-TGCGCATTCCGAGGCATGATGATA-3′	5′-CCTTCTTGCAATTGGCAACTCTGCT-3′

Supplemental Table S3 Primers used for yeast two-hybrid assays.

Gene name	Accession number	Restriction sites	Forward primer	Reverse primer
CSN4	Solyc04g080160	SmaI/XhoI	5′-agccccgggtATGGAGAGTGCGTTCGCTAGTG-3′	5′-agcctcgagCTAGACAGGAATAGGGAGCCCCT-3′
CSN5A	Solyc06g073150	SmaI/BamHI	5′-agccccgggtATGGACTCTCTGAATTCTTACGCA-3′	5′-agcggatccTCAGCTTTCGATCATGGGCTC-3′
CSN5B	Solyc11g017300	SmaI/BamHI	5′-agccccgggtATGGACGCTCTGAATTCTTACG-3′	5′-agcggatccTCAGGTTTCGACCATCGGCTCT-3′
JAZ1	Solyc07g042170	EcoRI/BamHI	5′-agcgaattcATGGCTTCATCGGAGATTGTGGATTCC-3′	5′-agcggatccCTAGTATTGCTCAGTTTTCACTGCAAATTGAC-3′
JAZ2	Solyc12g009220	SmaI/BamHI	5′-agccccgggtATGGGGTCATCGGAAAATATGGATTCCG-3′	5′-agcggatccCTAGAAATATTGCTCAGTTTTAACAAATTGAGCACCTAATCC-3′
JAZ3	Solyc03g122190	EcoRI/BamHI	5′-agcgaattcATGTCGAATTTATGTGACGCTCGCCG-3′	5′-agcggatccCTATAACTTGAAATTGAGATCGAGCTGATCTTCGCTA-3′
JAZ4	Solyc12g049400	SmaI/XhoI	5′-agccccgggtATGTCAAATAGGCAACTTTGTTCATTAGATAGTGAGAAATCAC-3′	5′-agcctcgagCTAGAAATTGAGATCAAAATGATCTCCACGAGTCCTT-3′
JAZ5	Solyc03g118540	SmaI/BamHI	5′-agccccgggtATGGAGAGAGATTTCATGGGGTTGACTGT-3′	5′-agcggatccCTACTTGACCAAACTGATTATGTGGGGAGG-3′
JAZ6	Solyc01g005440	SmaI/BamHI	5′-agccccgggtATGGAGAGGGACTTTATGGGATTGAATATCAAAGATT-3′	5′-agcggatccCTAGGTCTCCTTACCGGCTAACAGAG-3′
JAZ7	Solyc11g011030	EcoRI/BamHI	5′-agcgaattcATGGATTCAAGAATGGAGATAGATTTTATGGACCT-3′	5′-agcggatccTTAGTTTTCCCAATGAACGCTTGACGAC-3′
JAZ8	Solyc06g068930	EcoRI/BamHI	5′-agcgaattcATGCATTGGTCATATTCTAACAAGGCTCA-3′	5′-agcggatccCTAATTTGCAGCAGGAAGTGAAGTACTT-3′
JAZ9	Solyc08g036640	EcoRI/BamHI	5′-agcgaattcATGAGAAGAAATTGTAATTTGGAGCTCACTCTTATGCC-3′	5′-agcggatccCTATTTGTGATATGGCGAAGTTGCTTGAACTCT-3′
JAZ10	Solyc08g036620	SmaI/BamHI	5′-agccccgggtATGAGAAGAAAGTGTAATTTGGAACTCAC-3′	5′-agcggatccCTAGTGATGATATGGAGAAGTTATTTGAAT-3′
JAZ11	Solyc08g036660	EcoRI/BamHI	5′-agcgaattcATGAGAAGAAATTGTAATTTGGAGTTTAGGCTTATGCC-3′	5′-agcggatccCTAGTGATGATATGGCGAAGTTGTTTGAGTTCTACT-3′

JAZ12	Solyc01g009740	EcoRI/BamHI	5′-agcgaattcATGTCTTCAGGTACTGATAATGTGATTGG-3′	5′-agcggatccCTATTTCTTATTTTCAGGGGCAGCATGG-3′

Supplemental Table S4 Primers used for BiFC assays.

Gene name	Accession number	Restriction sites	Forward primer	Reverse primer
CSN4	Solyc04g080160	PacI/SpeI	5′-cccttaattaacATGGAGAGTGCGTTCGCTAGTG-3′	5′-gggactagtGACAGGAATAGGGAGCCCCTTCT-3′
CSN5A	Solyc06g073150	PacI/SpeI	5′-cccttaattaacATGGACTCTCTGAATTCTTACGCAT-3′	5′-gggactagtGCTTTCGATCATGGGCTC-3′
CSN5B	Solyc11g017300	PacI/SpeI	5′-cccttaattaacATGGACGCTCTGAATTCTTACG-3′	5′-gggactagtGGTTTCGACCATCGGCTCT-3′
JAZ2	Solyc12g009220	PacI/SpeI	5′-cccttaattaacATGGGGTCATCGGAAAATATGGATTCCG-3′	5′-gggactagtCTAGAAATATTGCTCAGTTTTAACAAATTGAGCACCTAATCC-3′
SIZF3	Solyc06g075780	PacI/SpeI	5′-cccttaattaacATGATAAAAATTAGAGAAG-3′	5′-gggactagtAGAGGATAAGAATGGAGGTAC-3′