

## Supplementary Materials for

### Structure and genome ejection mechanism of *Staphylococcus aureus* phage P68

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#### This PDF file includes:

Fig. S1. Purified sample of P68 contains native virions, particles in process of genome release, and empty particles; attachment of P68 virions to *S. aureus* cell walls; and interactions of P68 virions with liposomes.

Fig. S2. Resolution and interpretability of cryo-EM reconstructions.

Fig. S3. Details of P68 head.

Fig. S4. Incorporation of P68 portal complex into capsid and changes in the structure of P68 portal complex upon genome release.

Fig. S5. Structures of P68 tail fiber and tail spike.

Fig. S6. Schemes of cryo-EM reconstruction strategies.

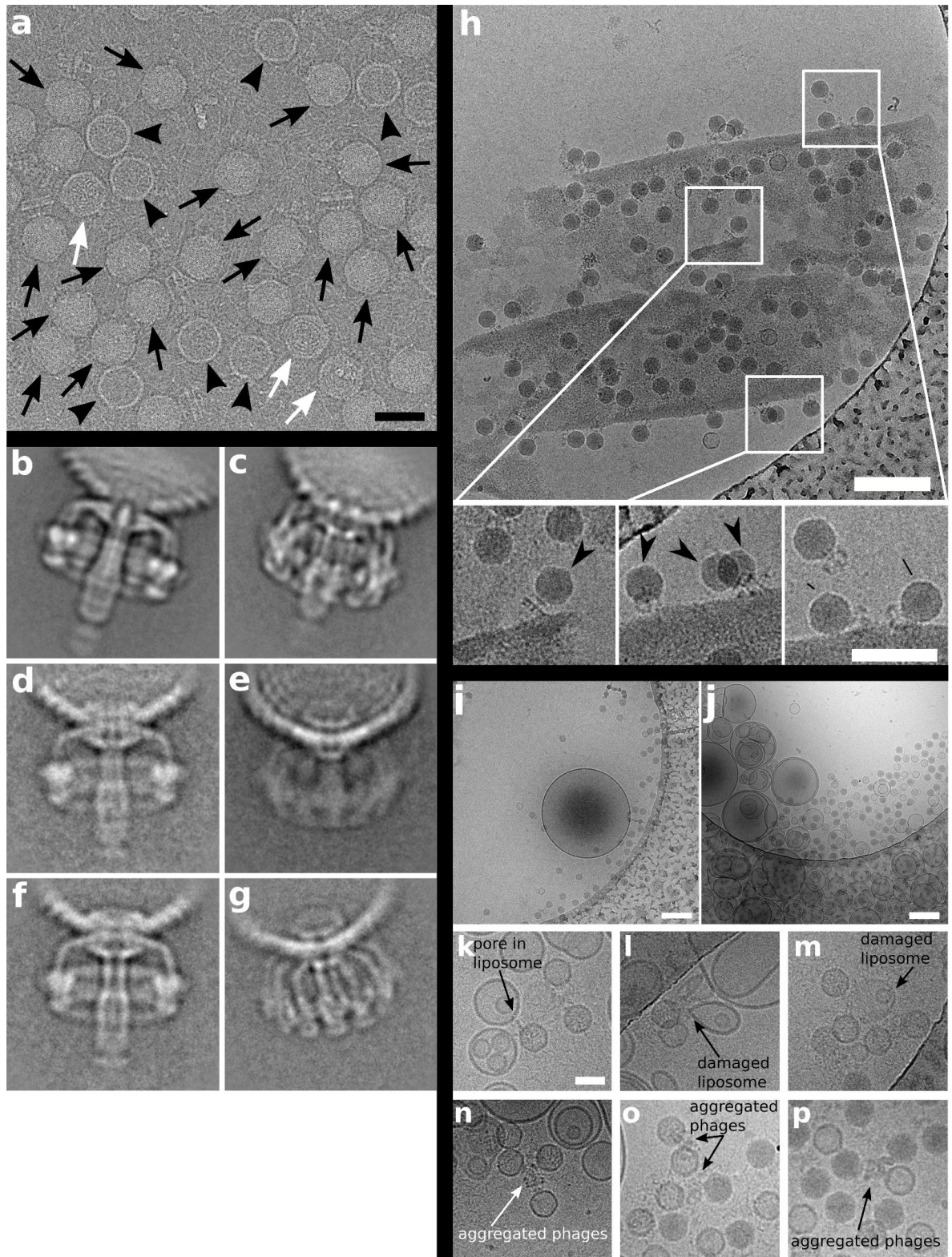
Table S1. Cryo-EM structure quality indicators.

Table S2. List of P68 proteins.

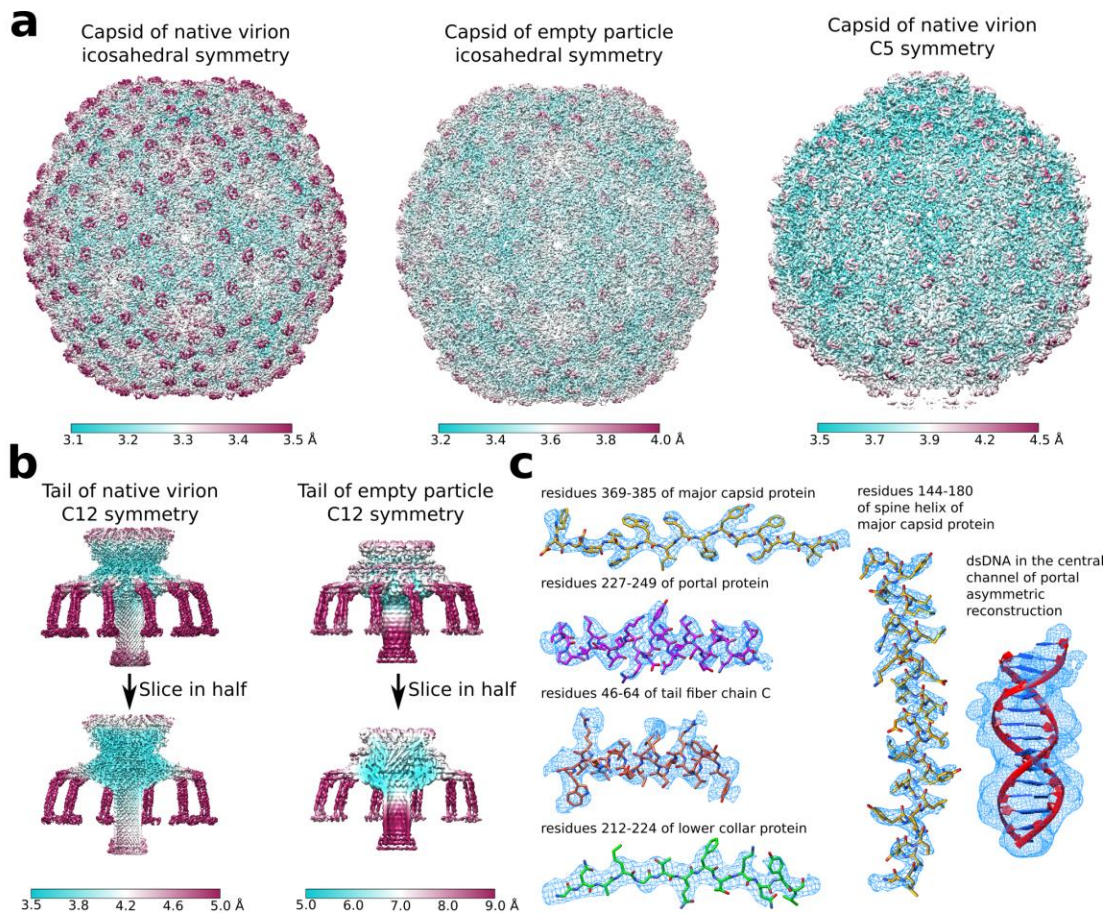
Table S3. HHpred searches for homologs of P68 tail fiber, head fiber, tail knob, and tail spike.

Table S4. X-ray structure quality indicators.

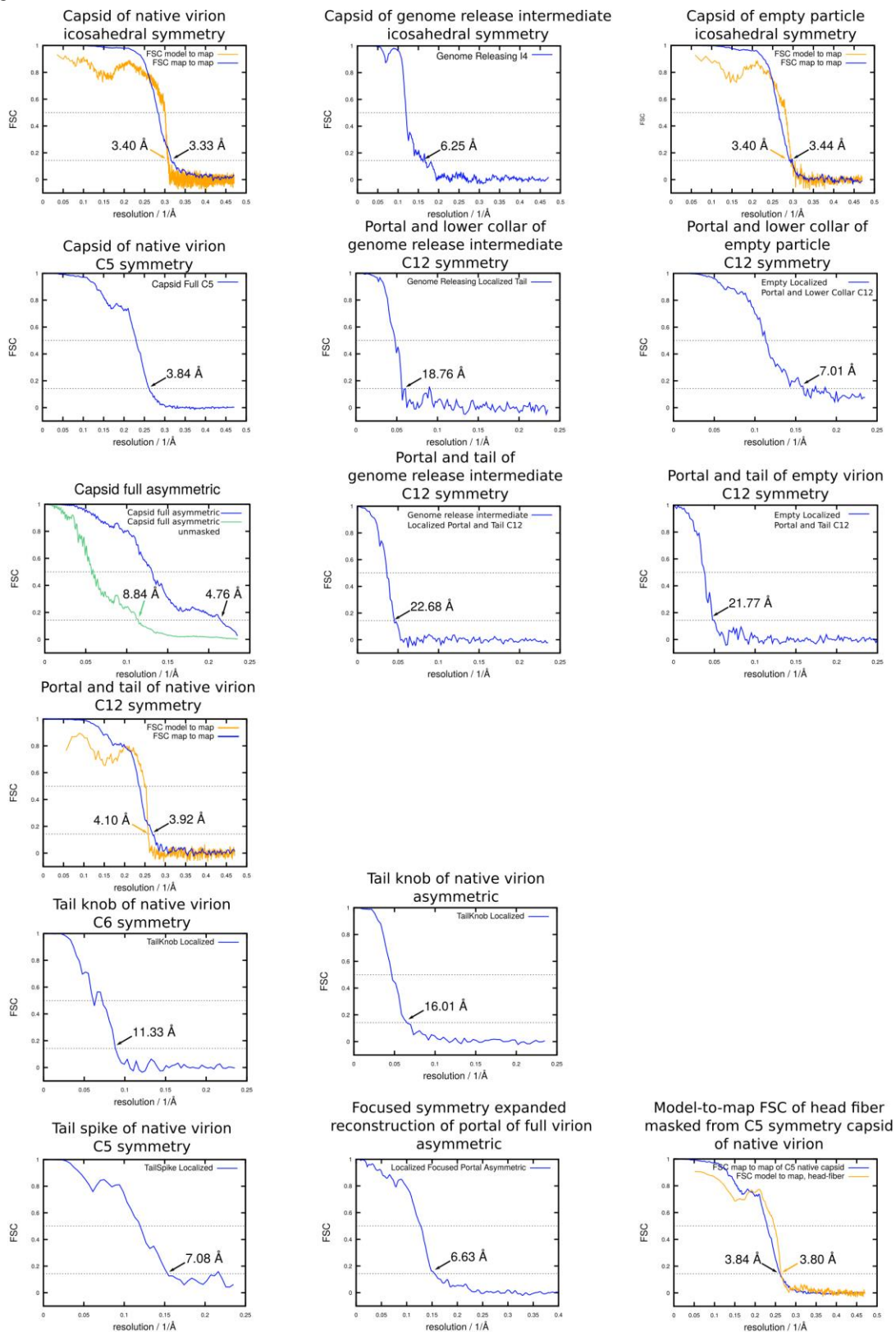
Supplementary figures:



**Fig. S1. Purified sample of P68 contains native virions, particles in process of genome release, and empty particles; attachment of P68 virions to *S. aureus* cell walls; and interactions of P68 virions with liposomes.** (a) Electron micrograph of P68 showing native virions (black arrows), particles in process of genome release (white arrows), and empty particles (black arrowheads). Scale bar 50 nm. (b to g) Reference-free two-dimensional class averages of native virions with complete tails (b); genome release intermediates with complete (c) and shortened tails (d); and empty particles with complete (e) and shortened tails (fg). (h) Attachment of P68 to fragments of *S. aureus* cell wall. At the stage of initial attachment the tail axis of P68 is not perpendicular to the surface of the cell wall. Scale bar 200 nm, scale bar in inset 100 nm. (i to p) P68 particles do not bind to liposomes. P68 virions were mixed with liposomes with lipid composition that mimics that of *S. aureus* membrane at neutral (i) and acidic (j) pH. (k to m) Liposomes in mixture with P68 at acidic pH are distorted. (n-p) P68 particles aggregate with each other by interactions of their tails at acidic pH. Scale bars 200 nm in (ij) and 50 nm in (k-p).



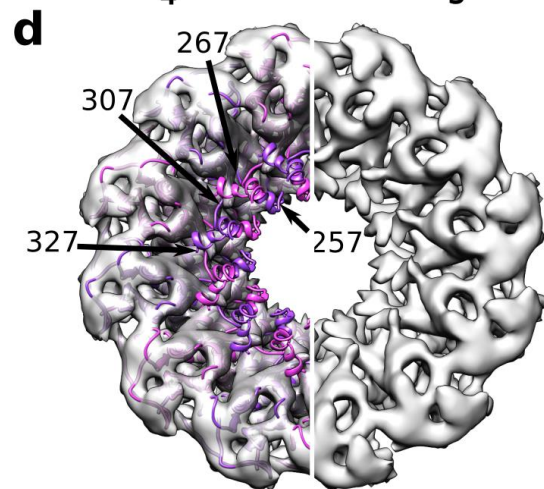
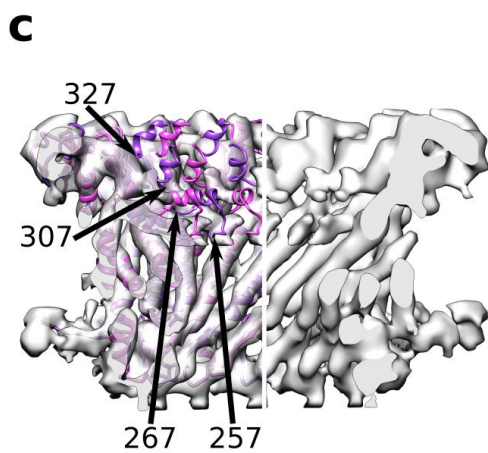
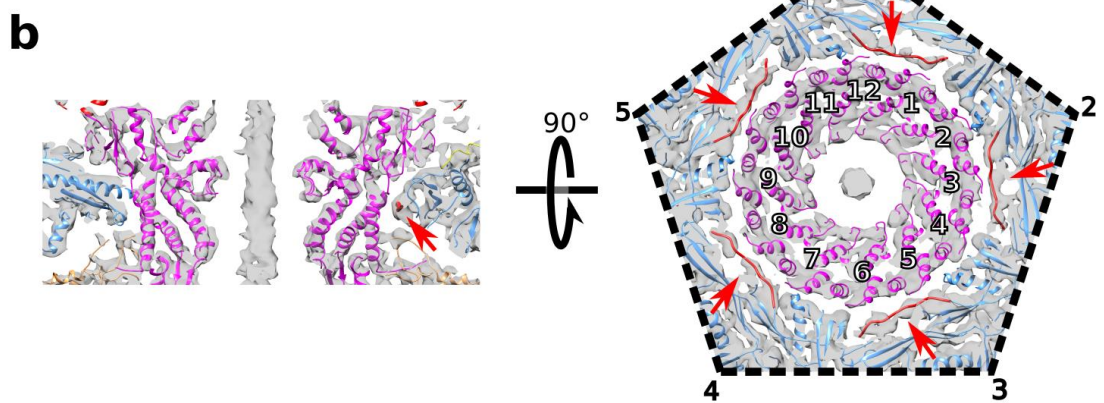
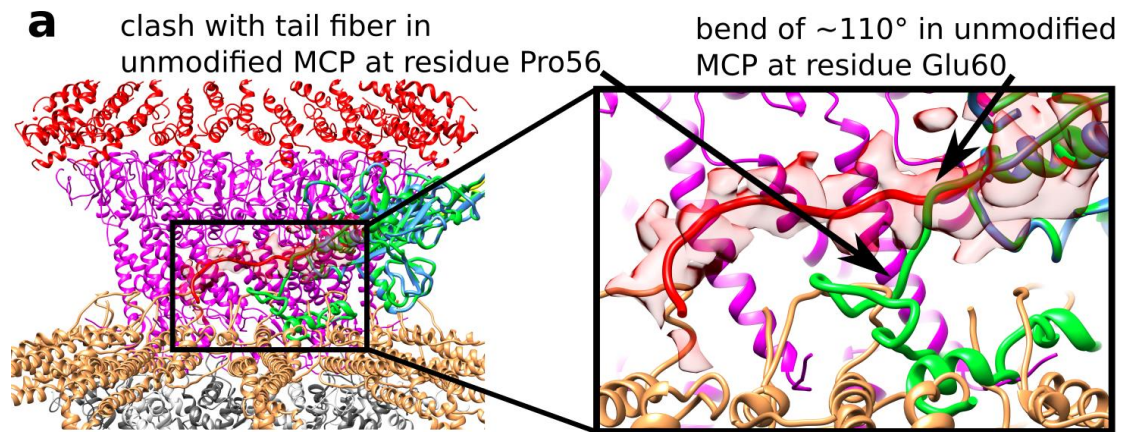
**Fig. S2. – continues on next page.**

**d**

**Fig. S2. Resolution and interpretability of cryo-EM reconstructions.** (ab) Local resolution maps of cryo-EM reconstructions described in manuscript. (c) Representative cryo-EM density maps with the P68 virion proteins shown in stick representations. (d) FSC curves of cryo-EM reconstructions.



**Fig. S3. Details of P68 head.** (a to e) Insertion and peripheral domains of major capsid protein form cleft that binds extended loop of neighboring capsid protein within pentamer or hexamer. (a) Overview of capsid protein interactions in P68 head. The major capsid proteins are shown in cartoon representation in grey, with insertion domains highlighted in blue, peripheral domains in green, and extended loops in yellow. Interactions of major capsid proteins in pentamers (b) and hexamers (c). Details of binding of extended loops by insertion and peripheral domains in pentamers (d) and hexamers (e). (f) Spacing of DNA layers in native P68 virions and genome release intermediate. The DNA of native virions is shown in red and that of genome release intermediates in blue. Phage proteins are shown in yellow. (gh) Comparison of structures of side chains of phe 259 of major capsid proteins in hexamers that form attachment sites for head fibers (a) and in hexamers that do not bind head fibers (b). Cryo-EM maps are shown as blue mesh. (ij) Inner core protein of P68 contains predicted pore lining helix. Sequence analysis of inner core protein of P68 (gp22). The program PSIPRED predicts propensity of several of its sequences to form  $\alpha$ -helices (a). Longest  $\alpha$ -helix of inner core protein has properties of pore-lining helix (b), indicating that it may participate in forming the pore in the cytoplasmic membrane for genome delivery.

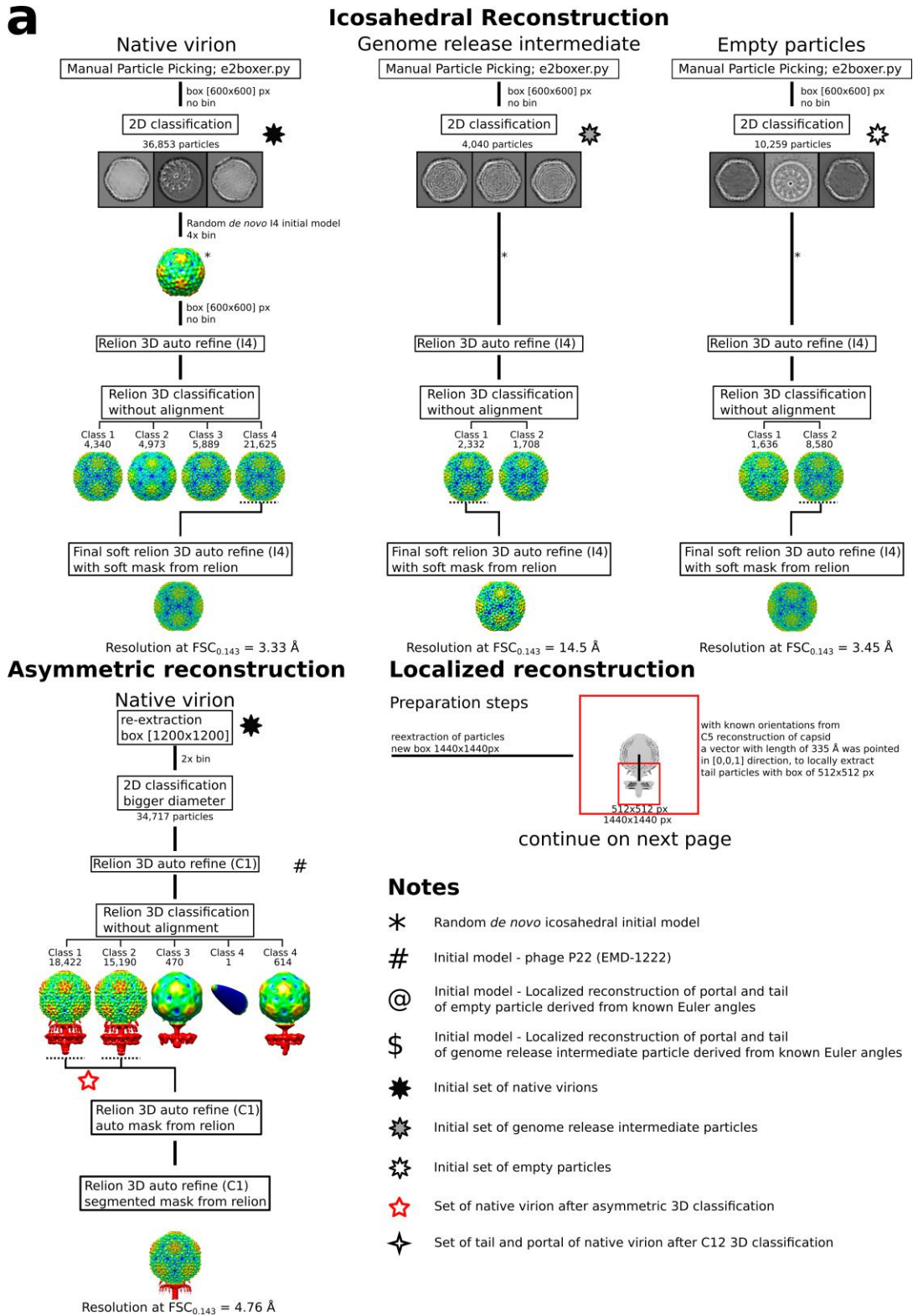


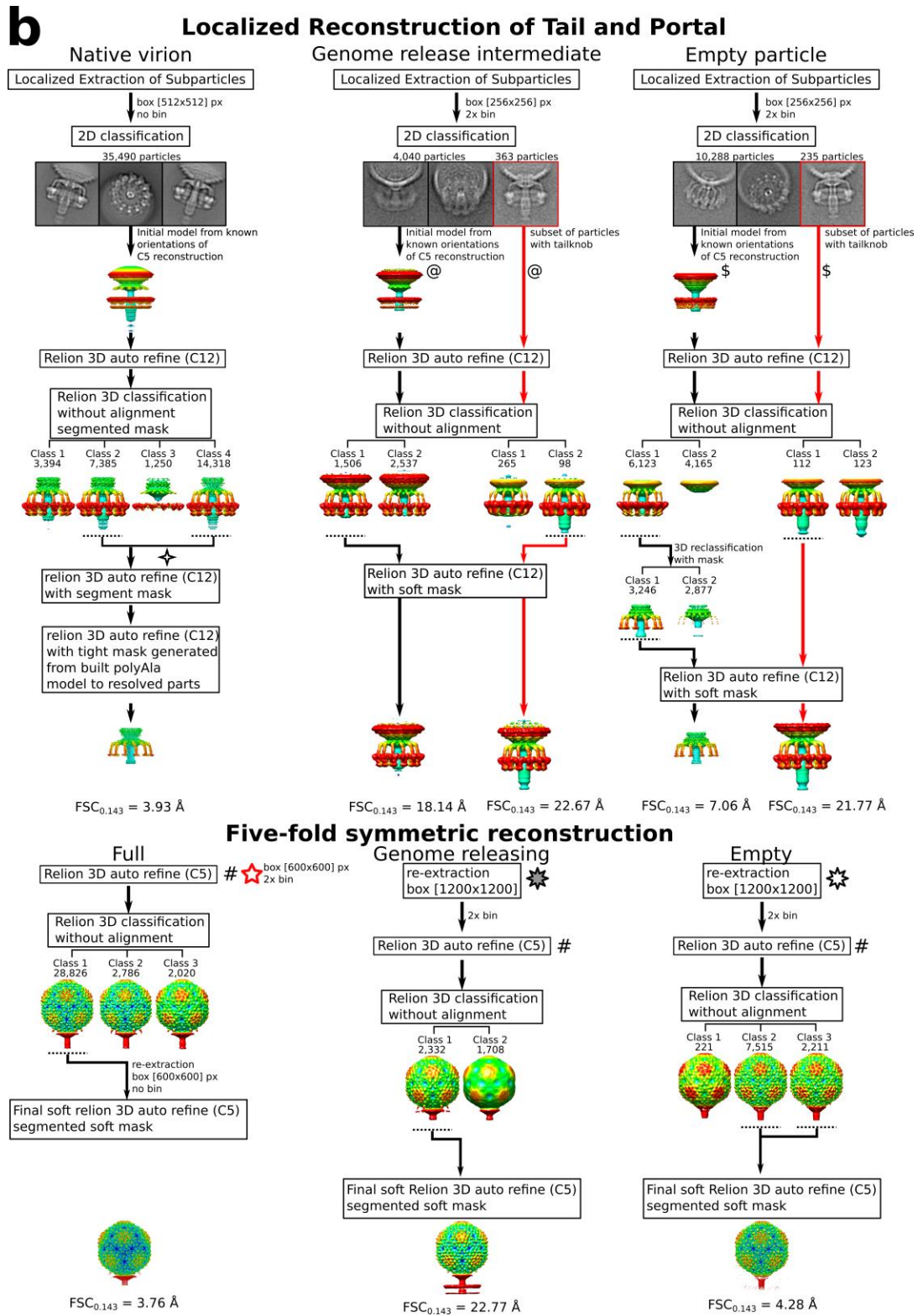


**Fig. S4. Incorporation of P68 portal complex into capsid and changes in the structure of P68 portal complex upon genome release.** (ab) Comparison of structures of N-terminal arms of major capsid proteins that interact with pentamers of capsid proteins and portal complex. Portal proteins are shown in magenta, major capsid proteins in blue, N-terminal arms of capsid proteins interacting with pentamers of capsid proteins in green, and N-terminal arms of capsid proteins interacting with portal complex in red. (a) If the N-terminal arm of major capsid protein binding to the portal complex had the same structure as in the vertices occupied by capsid proteins, the N-terminal arm would clash with the portal and tail fibers. (b) Side and top views of capsid-portal interactions. (cd) Differences between structures of portal complexes of native P68 virion (left halves of images and cartoon structures) and empty particle (right halves of images). Side-view of portal complex cut in half (a) and top view of portal complex (b). Cryo-EM densities of the portal complexes are shown as grey semi-transparent surfaces.



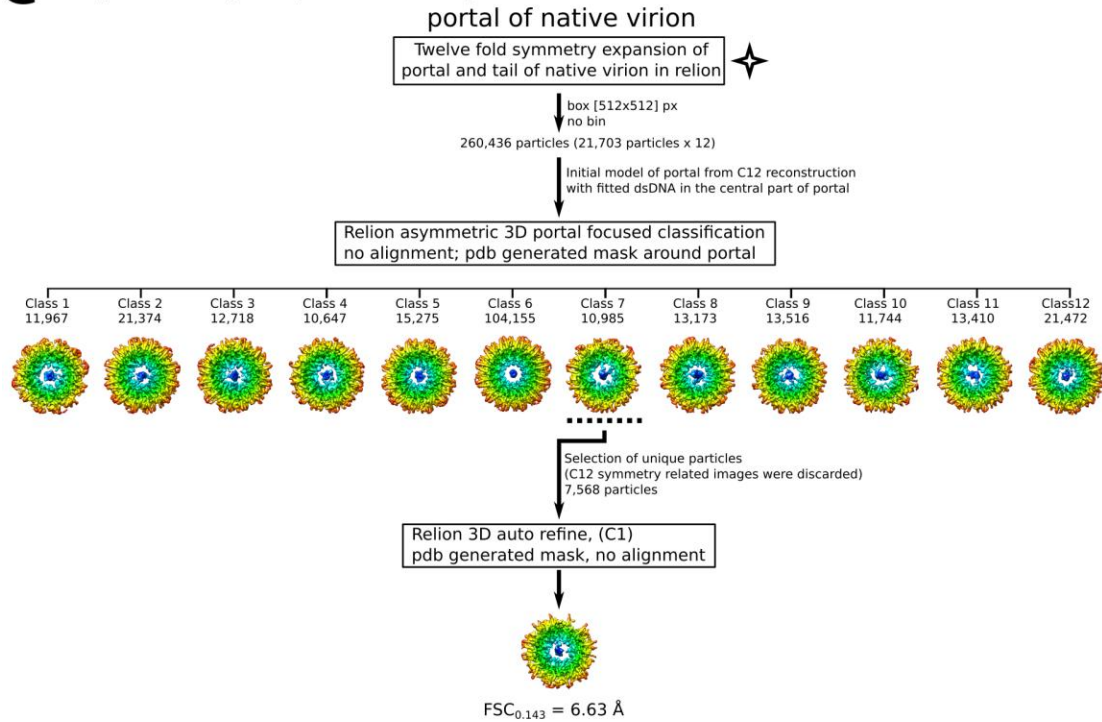
**Fig. S5. Structures of P68 tail fiber and tail spike.** (a to d) Comparison of structures of platform domains of P68 solved by X-ray crystallography at 2.0 Å in blue (a) and phi11 in green (b). Residues conserved among the two viruses are shown in red in both of the structures. Residues that are involved in the receptor binding of phi11 and the corresponding residues of P68 are shown as sticks and indicated with labels. (c) Superposition of the two structures. (d) Structure-based sequence alignment of platform domains of P68 and phi11. Relative distances of the corresponding C $\alpha$  atoms are indicated. Residues of phi11 involved in receptor binding are highlighted with a green background. Residues conserved among the two viruses are shown in red. (e) X-ray crystallography structure solved at 2.0 Å resolution of the tail-fiber shows swapping between platform and tower domains of tail fiber proteins of P68 and phi11. (f to h) Asymmetric reconstruction of tail knob and tail spike complexes. (fg) The surface of the map is rainbow colored based on its distance from the tail axis. (h) Correlation coefficients of tail knob and tail spike parts of the reconstructions with rotated copies of themselves. The results indicate that the tail knob has sixfold symmetry and the tail spike has fivefold symmetry.

**a****Fig. S6. Schemes of cryo-EM reconstruction strategies.**



**Fig. S6. Schemes of cryo-EM reconstruction strategies - continued.**

## C Symmetry Expanded Asymmetric Reconstruction of Portal



## Localized reconstruction of Tail knob and Tail spike

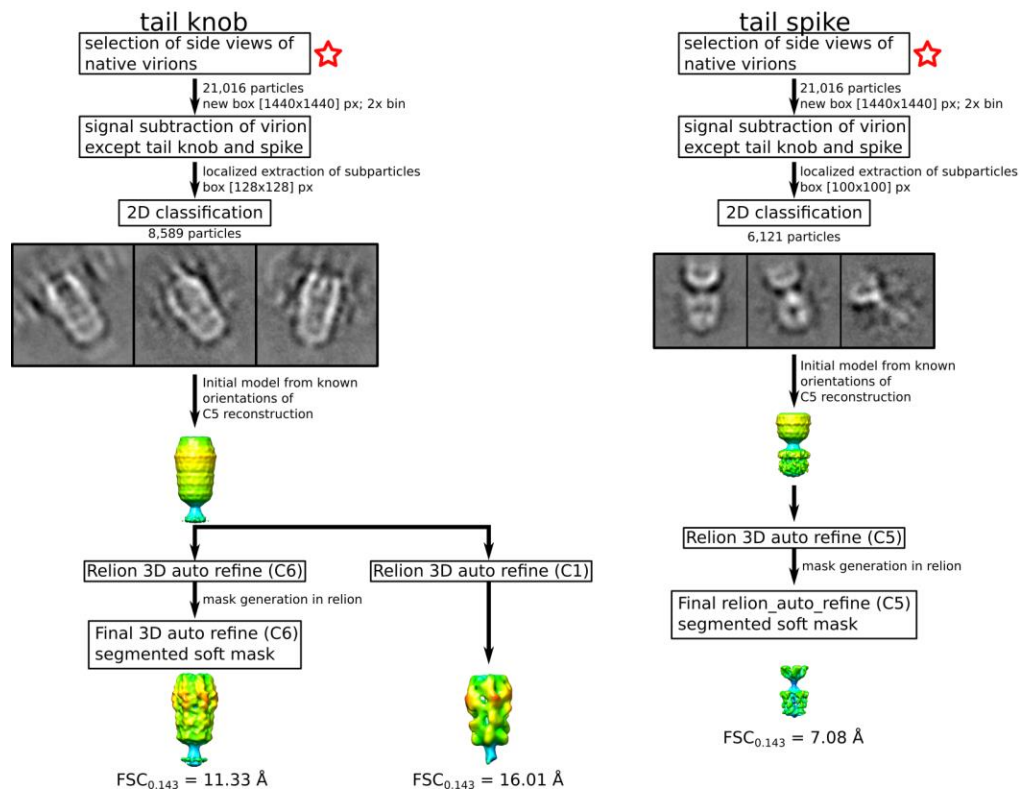


Fig. S6. Schemes of cryo-EM reconstruction strategies - continued.



**Table S2. List of P68 proteins.**

ORF No.	Protein name/ Function	Structural?/ Found in MS	Amino acid sequence length	Symmetry	No. of chains in ASU	No. of modelled residues	Not modelled residues	Model Completeness	Homologous proteins PDB codes
1	hypothetical protein/-	No/No	65	-	-	-	-	-	-
2	hypothetical protein/-	No/No	100	-	-	-	-	-	-
3	hypothetical protein/-	No/No	78	-	-	-	-	-	-
4	hypothetical protein/ Putative ssDNA binding protein	No/Yes	122	-	-	-	-	-	-
5	hypothetical protein/-	No/No	58	-	-	-	-	-	-
6	hypothetical protein/-	No/No	136	-	-	-	-	-	-
7	hypothetical protein/-	No/Yes	55	-	-	-	-	-	-
8	hypothetical protein/-	No/No	160	-	-	-	-	-	-
9	Putative Encapsidation Protein	No/No	378	-	-	-	-	-	-
10	DNA polymerase	No/No	755	-	-	-	-	-	-
11	Tailspike/ Cell Wall Degrading Enzyme	Yes/Yes	479	C5	-	-	-	-	4F88, 5T1Q, 2K3A, 2LRJ, 4CT3, 5D74, 5UDN, 4CGK, 4H29
12	Holin	No/No	140	-	-	-	-	-	-
13	Tailknob	Yes/Yes	587	C6	-	-	-	-	5F85, 4EP0
14	Head Fiber/ Receptor Binding	Yes/Yes	481	C3 (trimer) C5 (in whole phage, pentamer of trimers)	1	A: 54	A: 1-56-481	A: 11.4%	(C-terminal part) 4HEM, 2F0C, 3DA0, 3D8M, 2DSD, 1ZRJ
15	Holin	No/No	92	-	-	-	-	-	-
16	Endolysin	No/No	250	-	-	-	-	-	5UDN, 5D74, 4CT3, 2K3A, 5O1Q, 2MK5
17	Tail Fiber/ Receptor Binding	Yes/Yes	647	C3 (trimer) C12 (dodecamer of trimers, in native phage structure)	3	A: 633 B: 636 C: 617	A: 1-7; 322-327; 646-647 B: 1-4; 322-327; 646-647 C: 1-23; 322-327; 646-647	A: 97.8% B: 96.2% C: 95.3%	5EFV
18	Lower Collar	Yes/Yes	251	C12	1	A: 221	A: 1-2; 155-183	A: 88.0%	-
19	Portal	Yes/Yes	327	C12	1	A: 301	A: 1-5; 82-103	A: 92.0%	1FOU, 2JES, 3JL4, 3JAY
20	Major Capsid Protein	Yes/Yes	408	Quasi-icosahedral	4	A: 393 B: 386 C: 386 D: 393	A: 1-3; 396-408 B: 1-10; 396-408 C: 1-10; 396-408 D: 1-3; 396-408	A: 96.3% B: 94.6% C: 94.6% D: 96.3%	all HK97 folds
21	Astrotzka Protein	Yes/Yes	60	Quasi-icosahedral	4	A: 54 B: 55 C: 27 D: 95	A: 1-4; 60 B: 1-4; 60 C: 1-4; 32-60 D: 1-4; 60	A: 91.6% B: 91.6% C: 46.6% D: 91.6%	-
22	Inner Core Protein	Yes/Yes	133	C3, C2 C12 (dodecamer of hexamers in native phage)	6	A: 17 B: 17 C: 23 D: 22 E: 21 F: 21	A: 1-75; 94-133 B: 1-75; 94-133 C: 1-73; 98-133 D: 1-73; 97-133 E: 1-73; 96-133 F: 1-73; 96-133	A: 12.7% B: 12.7% C: 17.2% D: 16.5% E: 15.7% F: 15.7%	-



**Table S3. HHpred searches for homologs of P68 tail fiber, head fiber, tail knob, and tail spike.**

**Tower domain of tail fiber protein HHblits homolog search**

Protein HHblits hit	UNIPROT code	Probability / % <sup>#</sup>	E-value <sup>%</sup>	P-value <sup>*</sup>	Score <sup>5</sup>	Aligned Cols <sup>α</sup>	Query HMM <sup>β</sup>	Template HMM <sup>γ</sup>	Sequence identity / %	Similarity <sup>κ</sup>
major teichoic acid biosynthesis protein C, <i>S. aureus</i>	A0A1D7UFH3	97.0	$3.3 \times 10^{-3}$	$1.1 \times 10^{-10}$	75.3	166	3-174	263-429	31	0.501
muramidase, <i>B. campestris</i>	W7D9P4	96.1	$9.5 \times 10^{-4}$	$3.1 \times 10^{-9}$	61.4	83	97-179	7-89	34	0.547
muramidase, <i>B. thermosphacta</i>	W7CFP7	96.0	$1.5 \times 10^{-3}$	$4.9 \times 10^{-9}$	64.9	161	4-174	290-452	29	0.492
bacteriophage related putative muramidase, <i>B. subtilis</i>	E0TX48	94.0	$4.3 \times 10^{-3}$	$1.4 \times 10^{-7}$	59.5	127	1-137	1007-1140	29	0.563
major teichoic acid biosynthesis protein C, <i>S. aureus</i> phage P240	A0A1X9IGV7	92.8	$1.2 \times 10^{-1}$	$4.1 \times 10^{-7}$	51.5	150	6-171	173-335	31	0.588

**C-terminal domain of head fiber HHpred homolog search**

Protein HHpred hit	pdb code	Probability / % <sup>#</sup>	E-value <sup>%</sup>	P-value <sup>*</sup>	Score <sup>5</sup>	Aligned Cols <sup>α</sup>	Query HMM <sup>β</sup>	Template HMM <sup>γ</sup>	Sequence identity / %	Similarity <sup>κ</sup>
receptor binding protein; phage TP901-1	4IOS	94.5	0.45	$4.6 \times 10^{-6}$	33.4	77	23-101	7-87	3	-0.123
receptor binding protein; lactococcal phage p2	3DA0	92.4	2.2	$2.2 \times 10^{-5}$	32.0	71	29-101	44-118	11	0.059
receptor binding domain; phage bLL170	2FSD	90.5	3.4	3.5	33.2	87	3-99	32-124	15	0.211

**Tail knob protein HHpred homolog search**

Protein HHpred hit	pdb code	Probability / % <sup>#</sup>	E-value <sup>%</sup>	P-value <sup>*</sup>	Score <sup>5</sup>	Aligned Cols <sup>α</sup>	Query HMM <sup>β</sup>	Template HMM <sup>γ</sup>	Sequence identity / %	Similarity <sup>κ</sup>
bacteriophage C1 tail knob	4EO2	100.0	$2 \times 10^{-117}$	$2 \times 10^{-112}$	981.4	563	2-587	1-576	30	0.558
bacteriophage phi29 tail knob	5FB4	100.0	$2 \times 10^{-113}$	$2 \times 10^{-118}$	951.2	544	1-579	5-598	19	0.272

**Tail spike protein HHpred homolog search**

Protein HHpred hit	pdb code	Probability / % <sup>#</sup>	E-value <sup>%</sup>	P-value <sup>*</sup>	Score <sup>5</sup>	Aligned Cols <sup>α</sup>	Query HMM <sup>β</sup>	Template HMM <sup>γ</sup>	Sequence identity / %	Similarity <sup>κ</sup>
PlyCA, PlyCB; Lysin	4F88	100.0	$2.0 \times 10^{-75}$	$2.0 \times 10^{-80}$	609.36	418	4-476	7-463	19	0.215
N-acetylmuramoyl-L-alanine Amidase	5T1Q	99.3	$1.3 \times 10^{-14}$	$1.0 \times 10^{-75}$	118.45	6	350-477	243-358	17	0.201
CHAP domain protein	2K3A	99.3	$8.1 \times 10^{-15}$	$4.8 \times 10^{-18}$	106.56	93	342-476	39-153	20	0.149
staphyloxanthin biosynthesis protein	2LRJ	99.2	$3.8 \times 10^{-14}$	$2.3 \times 10^{-17}$	97.77	104	352-475	8-111	23	0.234
peptidoglycan protease, phage K	4CT3	98.9	$1.5 \times 10^{-10}$	$1.5 \times 10^{-15}$	104.46	106	350-463	44-154	19	0.143
putative phage lysin, phage phi7917	5D74	98.5	$7.6 \times 10^{-9}$	$7.8 \times 10^{-14}$	103.0	96	352-456	46-159	18	0.108
CHAP domain, <i>S. pyogenes</i> MGAS5005	5UDN	98.4	$5.6 \times 10^{-8}$	$5.7 \times 10^{-13}$	95.4	113	350-476	20-150	17	0.115
CHAP domain, <i>S. pneumoniae</i>	4CGK	98.0	$9.0 \times 10^{-7}$	$9.2 \times 10^{-12}$	87.6	107	352-476	284-390	17	0.046
endopeptidase	4HZ9	97.4	$5.6 \times 10^{-5}$	$5.7 \times 10^{-10}$	65.0	78	354-445	17-95	19	0.240
CHAP domain	4Q4G	92.5	0.82	$8.4 \times 10^{-6}$	49.9	90	351-458	374-472	14	-0.030

<sup>#</sup> - estimated probability of the template to be (at least partly) homologous to the query sequence  
<sup>%</sup> - the expected number of false positives per database search with a score at least as good as the score of this sequence match  
<sup>\*</sup> - equal to the E-value divided by the number of hidden markov models in the searched database  
<sup>5</sup> - gives the total score that includes the score from the secondary structure comparison  
<sup>α</sup> - the total number of matched columns in the query/template alignment  
<sup>β</sup> - range of amino acid sequence from the query hidden markov model used for comparison  
<sup>γ</sup> - range of amino acid sequence from the template hidden markov model used for comparison  
<sup>κ</sup> - secondary structure similarity

**Table S4. X-ray structure quality indicators.**

<b>X-ray Data Collection</b>		Tail Fiber
Space group		R3
Cell dimensions		
	<i>a, b, c</i> (Å)	100.73, 100.73, 125.46
	$\alpha, \beta, \gamma$ (°)	90.0, 90.0, 120.0
Wavelength (Å)		0.97857
Resolution (Å)		2.0 (2.0502 - 2.0002)
R <sub>merge</sub>		0.216
Completeness (%)		99.9 (100.0)
<b>Refinement</b>		
Resolution (Å)		2.0
No. reflections		32065 (2146)
R <sub>work</sub> /R <sub>free</sub> (%)		16.4/20.2 (21.45/22.89)
No. of atoms (non-hydrogen)		
	Protein	4021
	Water	436
B-factors		
	Protein	27.1
	Water	30.5
R.m.s. deviations		
	Bond lengths (Å)	0.003
	Bond angles (°)	0.57
Ramachandran outliers (%)		0.0
Rotamer outliers (%)		0.2