

SMCHD1 mutation spectrum for facioscapulohumeral muscular dystrophy type 2 (FSHD2) and *Bosma arhinia* microphthalmia syndrome (BAMS) reveals disease-specific localization of variants in the ATPase domain

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## **Supplementary Materials and Methods**

### Genetic analysis of D4Z4 repeats

Determination of the D4Z4 repeat size and allelic background was performed by Southern blotting and hybridization with probes p13E-11, 4qA and 4qB as described previously. Hybridization conditions slightly vary between the different laboratories. In our laboratory hybridization with probe p13E-11 was performed in a buffer with 10% Dextran sulphate, 1M NaCl, 50 mM Tris-HCl, pH 7.5, 1% SDS and 250ug/ml Salmon sperm DNA at 65°C. 4qA and 4qB hybridizations were done in a phosphate buffer with 10% polyethylene glycol 6000.<sup>1</sup>

Methylation at D4Z4 was determined either by Southern blotting at the FseI site in D4Z4 or by bisulphite conversion and PCR at the DR1 site.<sup>2</sup> For Southern blotting based methylation analysis with probe p13E-11 we used the hybridization buffer with 10% Dextran sulphate (see before). Calculations of repeat size corrected methylation compared to controls (delta1) and SMCHD1 pathogenic variant carriers (delta2) were done as described previously.<sup>3</sup>

### SMCHD1 sequencing and variant prediction

SMCHD1 variants were identified by Sanger sequencing or by whole exome or whole genome sequencing (WES/WGS) followed by confirmation using Sanger sequencing. All variants identified in FSHD2, BAMS and controls have been submitted to the Leiden Open Variation Database (LOVD, [www.lovd.nl](http://www.lovd.nl)). The putative effects of the SMCHD1 variants were investigated through prediction algorithms using Alamut Visual v.2.4.2 (Interactive Biosoftware, <https://www.interactive-biosoftware.com/alamut-visual/>) or Variant Effect Predictor (VEP) in Ensemble (<https://www.ensembl.org/info/docs/tools/vep/index.html>). This includes SIFT (Sorting Intolerant from Tolerant, <http://sift.jcvi.org>), Polyphen2 (<http://genetics.bwh.harvard.edu/pph2/>) and Align GVGD (Grantham Variation and Grantham Deviation, <http://agvgd.hci.utah.edu/about.php>). Splicing predictions were done in Alamut.

### Statistical analysis

The FseI methylation level, delta1 and delta2 methylation score (in Supplementary figure 2) were compared using the unpaired t-test in Graphpad Prism 7. For the comparisons shown in Supplementary table 4, we used a Pearson chi-square test with Yates' continuity correction in R 3.3.2. For the visualization, the R packages ggplot2 and trackViewer from R/Bioconductor were used.<sup>4</sup>

## Supplementary Tables

nr	publication	identified variants	new variants
1	Lemmers et al., 2012 (PMID: 23143600) <sup>8</sup>	15	15
2	Sacconi et al., 2013 (PMID: 24075187) <sup>13</sup>	3	3
3	Mitsuhashi et al., 2013 (PMID: 24128691) <sup>10</sup>	1	1
4	Winston et al., 2015 (PMID: 24755953) <sup>15</sup>	1	1
5	Lemmers et al., 2015 (PMID: 25256356) <sup>3</sup>	51	36
6	Larsen et al., 2015 (PMID: 25370034) <sup>7</sup>	11	11
7	Smith et al., 2015 (poster ASHG 2015)	8	8
8	Lemmers et al., 2015 (PMID: 25820463) <sup>9</sup>	2	2
9	Boogaard et al., 2016 (PMID: 25782668) <sup>14</sup>	5	5
10	Hamanaka et al., 2016 (PMID: 27061275) <sup>6</sup>	11	11
11	Gaillard et al., 2016 (PMID: 27634379) <sup>5</sup>	1	1
12	Nguyen et al., 2017 (PMID: 28744936) <sup>12</sup>	1	1
13	Mul et al., 2018 (PMID: 29980640) <sup>11</sup>	23	6
		Total	101

### Supplementary table 1

Chronological overview of 101 published SMCHD1 variants involved in FSHD2.<sup>3, 5-15</sup> Third column shows all variants described in the publication. Some variants were described in consecutive publications and overlap. In the last column we only mention the non-overlapping variants.

1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
ID	Family	Nr	M/F	cDNA (NM 015295.2)	gene (hg19 (GRCh37.p5))	protein (NP_056110.2)	type	ORF	position	Fsel	delta1	delta2	dbSNP	AFR	EUR	Allele Frequency	SAS	Ensemble	Alamut	VG1	GV	GD	Prediction	Publication
F1	R1871	1	M	c.101G>C	g.2656175G>C	R34P	M	P-ORF	exon 1	11	-37	-3						probably_damaging(0.969)	Deleterious(0)	C0	241	40	2	
F2	LOVD12	1	M	c.311A>G	g.2666917A>G	N104S	M	P-ORF	exon 3	NA	NA	NA	rs375795924	1.13E-04	7.99E-05	8.38E-04	0	benign(0)	Tolerated(0.13)	C0	223	0	0	ASHG conference 2015 (Smith, 2015)
F3	Rf6	30	F	c.320T>C	g.2666926T>C	L107P	M	P-ORF	exon 3	3	-36	-6	rs1135402737					probably_damaging(0.998)	Deleterious(0)	C65	0	98	3	PMID: 29980640 (Mul, 2018)
F4	LOVD11	1	M	c.328G>A	g.2666934G>A	A110T	M	P-ORF	exon 3	NA	NA	NA						possibly_damaging(0.865)	Deleterious(0.01)	C0	27	37	2	PMID:25370034 (Larsen, 2014)
F5	Rf866	1	M	c.410G>A	g.2667016G>A	G137E	M	P-ORF	exon 3	22	-21	10	rs1057519644					probably_damaging(0.989)	Deleterious(0.01)	C0	79	30	2	PMID: 25256356 (Lemmers, HMG 2015)
F6	LOVD21	1	M	c.448G>C	g.2673303G>C	D150H	M	P-ORF	exon 4	NA	NA	NA						probably_damaging(1)	Deleterious(0)	C65	0	81	3	ASHG conference 2015 (Smith, 2015)
F7	Rf1685	1	M	c.562G>A	g.2674068G>A	G188R	M	P-ORF	exon 5	6	NA	NA						probably_damaging(1)	Deleterious(0)	C65	0	125	3	
F8	LOVD23	1	M	c.565A>G	g.2674071A>G	M189V	M	P-ORF	exon 5	NA	NA	NA						probably_damaging(0.977)	Deleterious(0)	C15	0	21	2	ASHG conference 2015 (Smith, 2015)
F9	Rf1142	1	M	c.580C>T	g.2674086C>T	L194F	M	P-ORF	exon 5	12	-28	2						probably_damaging(0.999)	Deleterious(0)	C15	0	22	2	PMID: 25256356 (Lemmers, HMG 2015)
F10	Rf677	1	M	c.610A>G	g.2674116A>G	K204E	M	P-ORF	exon 5	6	-32	-4	rs1184311800					possibly_damaging(0.851)	Deleterious(0)	C55	0	57	3	PMID: 29980640 (Mul, 2018)
F11	IW1	2	M	c.724G>G	g.2688477G>A	A242T	M	P-ORF	exon 6	14	NA	NA						probably_damaging(0.959)	Deleterious(0)	C55	0	58	3	PMID: 29980640 (Mul, 2018)
F12	Rf1247	1	F	c.787C>G	g.2688659C>G	H263D	M	P-ORF	exon 7	4	-34	-5						probably_damaging(0.996)	Deleterious(0)	C55	0	81	3	PMID: 25256356 (Lemmers, HMG 2015)
F13	Rf2542	1	M	c.790G>A	g.2688962G>A	E254K	M	P-ORF	exon 7	6	-33	-4						probably_damaging(0.996)	Deleterious(0)	C55	22	192	3	PMID: 27061275 (Hamanaka, 2016)
F14	LOVD36	1	F	c.848A>G	g.2688720A>G	Y283C	M	P-ORF	exon 7	4	NA	NA	rs886041921					probably_damaging(0.996)	Deleterious(0)	C55	22	192	3	
F15	Rf294	2	F	c.848A>G	g.2688720A>G	Y283C	M	P-ORF	exon 7	5	-37	-6	rs886041921					probably_damaging(0.996)	Deleterious(0)	C55	22	192	3	
F16	IW20	1	M	c.1031G>A	g.2694682G>A	R344Q	M	P-ORF	exon 9	24	NA	NA	rs370983669	4.11E-04	3.02E-05	0	0	benign(0.031)	Deleterious(0)	C35	0	43	3	
F17	Rf742	1	M	c.1058A>G	g.2697047A>G	Y353C	M	P-ORF	exon 9	9	-41	-6						probably_damaging(0.997)	Deleterious(0)	C65	0	194	3	PMID: 23143600 (Lemmers, 2010)
F18	Rf959	3	M	c.1058A>G	g.2697047A>G	Y353C	M	P-ORF	exon 9	12	NA	NA						probably_damaging(0.997)	Deleterious(0)	C65	0	194	3	PMID: 25256356 (Lemmers, HMG 2015)
F19	Rf1196	1	F	c.1273G>G	g.2697970G>A	G425R	M	P-ORF	exon 10	1	-37	-8						probably_damaging(0.925)	Deleterious(0)	C65	0	125	3	PMID: 25256356 (Lemmers, HMG 2015)
F20	IW35	1	M	c.1273G>A	g.2697970G>A	G425R	M	P-ORF	exon 10	17	NA	NA						probably_damaging(0.955)	Deleterious(0)	C65	0	125	3	
F21	IW39	1	M	c.1282C>T	g.2697979C>T	R428C	M	P-ORF	exon 10	14	NA	NA						probably_damaging(0.996)	Deleterious(0)	C65	0	179	3	
F22	LOVD1	1	M	c.1433G>A	g.2700627G>A	G478E	M	P-ORF	exon 11	12	NA	NA						benign(0.142)	Deleterious(0)	C65	0	98	2	PMID:25370034 (Larsen, 2014)
F23	Rf739	1	M	c.1436G>C	g.2700630G>C	R479P	M	P-ORF	exon 11	10	-32	-1						probably_damaging(0.997)	Deleterious(0)	C65	0	103	3	PMID: 23143600 (Lemmers, 2010)
F24	Rf2377	1	M	c.1436G>G	g.2700630G>T	R479L	M	P-ORF	exon 11	13	-31	1						probably_damaging(0.995)	Deleterious(0)	C65	0	102	3	PMID: 23143600 (Lemmers, 2010)
F25	Rf2456	1	M	c.1436G>G	g.2700630G>A	R479Q	M	P-ORF	exon 11	2	-48	-13						probably_damaging(0.992)	Deleterious(0)	C35	0	43	3	
F26	Rf300	1	M	c.1474T>C	g.2700743T>C	C492R	M	P-ORF	exon 12	7	-29	-2						probably_damaging(0.996)	Deleterious(0)	C65	0	180	3	PMID: 23143600 (Lemmers, 2010)
F27	Rf1729	1	F	c.1556T>C	g.2700825T>C	F519S	M	P-ORF	exon 12	11	-31	-1						probably_damaging(0.994)	Deleterious(0)	C65	0	155	3	PMID: 29980640 (Mul, 2018)
F28	Rf1021	1	M	c.1580C>T	g.2700849C>T	T527M	M	P-ORF	exon 12	18	-25	6	rs397518422					probably_damaging(0.992)	Deleterious(0)	C0	78	52	2	PMID: 24075187 (Sacconi, 2013)
F29	IW31	1	M	c.1652A>G	g.2703694A>G	Q551R	M	P-ORF	exon 13	15	NA	NA						probably_damaging(0.968)	Deleterious(0.03)	C0	53	5	2	
F30	Rf2572	1	M	c.1786T>G	g.2703828T>G	W596G	M	P-ORF	exon 13	10	-21	3						probably_damaging(0.994)	Deleterious(0)	C65	0	184	3	
F31	LOVD5	1	F	c.1844T>A	g.2705693T>A	V815D	M	P-ORF	exon 14	17	NA	NA						probably_damaging(0.996)	Deleterious(0)	C65	0	152	3	PMID:25370034 (Larsen, 2014)
F32	IW8	1	F	c.1865C>T	g.2705714C>T	P622L	M	P-ORF	exon 14	8	NA	NA						probably_damaging(0.997)	Deleterious(0)	C65	0	98	3	
F33	IW11	1	F	c.192T>G	g.2705770G>T	V641L	M	P-ORF	exon 14	16	NA	NA	rs377559548	0	1.51E-05	0	0	benign(0.079)	Deleterious(0.03)	C0	29	5	1	
F34	Rf999	3	F	c.206T>C	g.2707585C>T	P890S	M	P-ORF	exon 16	7	-28	-1	rs397514623					probably_damaging(0.996)	Deleterious(0)	C65	0	73	3	PMID: 23143600 (Lemmers, 2010)
F35	Rf917	1	M	c.224T>C	g.2707911T>C	L749P	M	P-ORF	exon 17	11	-36	-3						probably_damaging(0.905)	Deleterious(0)	C45	14	87	3	PMID: 25256356 (Lemmers, HMG 2015)
F36	Rf1857	1	M	c.2321A>G	g.2718216A>G	Y774C	M	P-ORF	exon 18	6	NA	NA						probably_damaging(0.921)	Deleterious(0.01)	C25	102	155	3	
F37	Rf676	1	F	c.2545G>A	g.2722693G>A	D849N	M	P-ORF	exon 20	5	-37	-6						probably_damaging(0.996)	Deleterious(0)	C15	0	23	2	PMID: 23143600 (Lemmers, 2010)
F38	Rf1432	1	F	c.2768T>C	g.2728517T>C	L923P	M	P-ORF	exon 22	5	NA	NA						probably_damaging(0.942)	Deleterious(0)	C0	112	53	2	
F39	Rf1492	1	M	c.2932T>A	g.2729292T>A	L978H	M	P-ORF	exon 24	14	-28	3						probably_damaging(0.998)	Deleterious(0)	C0	235	89	2	
F40	Rf385	101	M	c.2941T>G	g.2729300T>G	Y981D	M	P-ORF	exon 24	21	-24	8						benign(0.158)	Deleterious(0)	C35	56	140	2	PMID: 27153398 (Boogaard, 2016)
F41	LOVD17	1	M	c.318T>G	g.2732401G>C	G1063R	M	P-ORF	exon 25	NA	NA	NA						probably_damaging(0.999)	Deleterious(0)	C65	0	125	3	ASHG conference 2015 (Smith, 2015)
F42	LOVD20	1	M	c.332T>C	g.2738441T>C	L1108P	M	P-ORF	exon 26	NA	NA	NA						possibly_damaging(0.888)	Deleterious(0.05)	C15	106	80	2	ASHG conference 2015 (Smith, 2015)
F43	IW3	1	M	c.3340G>A	g.2738458G>A	V1114I	M	P-ORF	exon 26	10	NA	NA	rs778206654	0	1.13E-04	0	5.80E-04	probably_damaging(0.956)	Deleterious(0)	C25	0	29	3	
F44	Rf1659	1	F	c.3811G>T	g.2747529G>T	I1271L	M	P-ORF	exon 30	18	NA	NA						benign(0.087)	Deleterious(0)	C25	0	31	2	
F45	Rf1430	1	F	c.3899T>A	g.2747617T>A	I1300K	M	P-ORF	exon 30	13	NA	NA	rs764012354	0	2.43E-05	0	0	probably_damaging(0.986)	Deleterious(0)	C35	29	91	3	
F46	LOVD7	1	F	c.4388A>C	g.2760691A>C	Q1463P	M	P-ORF	exon 35	13	NA	NA						probably_damaging(0.969)	Deleterious(0)	C65	0	75	3	PMID:25370034 (Larsen, 2014)
F47	Rf1126	204	F	c.4404G>A	g.2760707G>A	M1468I	M	P-ORF	exon 35	13	-29	1						benign(0)	Tolerated(0.64)	C0	163	0	0	PMID: 25256356 (Lemmers, HMG 2015)
F48	LOVD8	1	M	c.4454C>T	g.2762122C>T	P1485L	M	P-ORF	exon 36	18	NA	NA						probably_damaging(0.996)	Deleterious(0)	C15	99	94	2	PMID:25370034 (Larsen, 2014)
F49	Rf683	1	F	c.4661T>C	g.2763297T>C	F1554S	M	P-ORF	exon 37	11	-27	1						possibly_damaging(0.477)	Deleterious(0)	C65	0	155	3	PMID: 23143600 (Lemmers, 2010)
F50	Rf1853	1	M	c.5249A>G	g.2775805A>G	D1750G	M	P-ORF	exon 42	5	-27	-2						probably_damaging(0.999)	Deleterious(0)	C65	0	94	3	
F51	IW23	1	M	c.5249A>T	g.2775805A>T	D1750V	M	P-ORF	exon 42	12	NA	NA						probably_damaging(0.999)	Deleterious(0)	C65	0	152	3	
F52	IW6	1	M	c.5337A>G	g.2778227A>G	Y1846C	M	P-ORF	exon 44	11	NA	NA						probably_damaging(0.998)	Deleterious(0)	C65	0	194	3	
F53	Rf385	102	F	c.5598C>G	g.2784498C>G	R1866G	M	P-ORF	exon 45	10	-31	-1						probably_damaging(0.995)	Deleterious(0)	C65	0	125	3	PMID: 27153398 (Boogaard, 2016)
F54	Rf1807	1	M	c.5597G>A	g.2784497G>A	R1866Q	M	P-ORF	exon 45	14	-28	3	rs886044586					probably_damaging(0.992)	Deleterious(0)	C35	0	43	3	
F55	Rf975	201	F	c.244del	g.2656098del		D	D-ORF	exon 1	15	-38	-1												PMID: 25256356 (Lemmers, HMG 2015)
F56	IW10	1	F	c.173 174insCT	g.2656247_2656248insCT		D	D-ORF	exon 1	19	NA	NA												
F57	Rf743	1	M	c.182 183dup	g.2656256_2656257dup		D	D-ORF	exon 1	14	-30	2												
F58	Rf1472	1	M	c.182 183dup	g.2656256_2																			



ID	Family	Nr	SMCHD1	SMCHD1	SMCHD1	type	ORF	position	Fsel	delta1	delta2	dbSNP	AFR	EUR	EAS	SAS	Ensemble	Alamut	AGVGD (3)	GV	GD	Total (1+2+3)	Publication	
			cDNA (NM 015295.2)	gene (hg19 (GRCh37.p5)	protein (NP 056110.2)												PolyPhen (1)	SIFT(2)				Prediction		
F3	R16	265	F c.320T>C	q.266926T>C	L107P	M	P-ORF	exon 3	13	-38	-2	rs1135402737					probably_damaging(0.998)	Deleterious(0)	C65	0	98	3	PMID: 29980640 (Mul, 2018)	
F3	R16	52	F c.320T>C	q.266926T>C	L107P	M	P-ORF	exon 3	11	-34	-2	rs1135402737					probably_damaging(0.998)	Deleterious(0)	C65	0	98	3	PMID: 29980640 (Mul, 2018)	
F3	R16	262	M c.320T>C	q.266926T>C	L107P	M	P-ORF	exon 3	7	-36	-5	rs1135402737					probably_damaging(0.998)	Deleterious(0)	C65	0	98	3	PMID: 29980640 (Mul, 2018)	
F3	R16	37	F c.320T>C	q.266926T>C	L107P	M	P-ORF	exon 3	6	-35	-5	rs1135402737					probably_damaging(0.998)	Deleterious(0)	C65	0	98	3	PMID: 29980640 (Mul, 2018)	
F3	R16	53	M c.320T>C	q.266926T>C	L107P	M	P-ORF	exon 3	7	-39	-6	rs1135402737					probably_damaging(0.998)	Deleterious(0)	C65	0	98	3	PMID: 29980640 (Mul, 2018)	
F3	R16	51	F c.320T>C	q.266926T>C	L107P	M	P-ORF	exon 3	3	-42	-10	rs1135402737					probably_damaging(0.998)	Deleterious(0)	C65	0	98	3	PMID: 29980640 (Mul, 2018)	
F12	IW1	1	F c.724G>A	q.2688477G>A	A242T	M	P-ORF	exon 6	19	NA	NA						probably_damaging(0.999)	Deleterious(0)	C55	0	58	3	PMID: 29980640 (Mul, 2018)	
F18	Rf742	4	M c.1058A>G	q.2697047A>G	Y353C	M	P-ORF	exon 9	9	-42	-6						probably_damaging(0.997)	Deleterious(0)	C65	0	194	3	PMID: 23143600 (Lemmers, 2010)	
F28	Rf1021	3	M c.1580C>T	q.2700849C>T	T527M	M	P-ORF	exon 12	11	-27	1	rs397518422					probably_damaging(0.992)	Deleterious(0)	C0	78	52	2	PMID: 24075187 (Sacconi, 2013)	
F28	Rf1021	2	F c.1580C>T	q.2700849C>T	T527M	M	P-ORF	exon 12	24	-25	10	rs397518422					probably_damaging(0.992)	Deleterious(0)	C0	78	52	2	PMID: 24075187 (Sacconi, 2013)	
F34	Rf399	2	F c.2068C>T	q.2707565C>T	P690S	M	P-ORF	exon 16	10	-30	-1	rs397514623					probably_damaging(0.996)	Deleterious(0)	C65	0	73	3	PMID: 23143600 (Lemmers, 2010)	
F40	Rf385	203	M c.2941T>G and c.5596C>G	q.272930T>G and q.2784496C>G	Y981D	M	P-ORF	exon 24	5	-38	-7						benign(0.158)	Deleterious(0)	C35	56	140	2	PMID: 27153398 (Boogaard, 2016)	
F40	Rf385	206	F c.2941T>G and c.5596C>G	q.272930T>G and q.2784496C>G	Y981D	M	P-ORF	exon 24	1	-37	-9						benign(0.158)	Deleterious(0)	C35	56	140	2	PMID: 27153398 (Boogaard, 2016)	
F47	Rf1126	205	M c.4404G>A	q.2760707G>A	M1468I	M	P-ORF	exon 35	12	-35	-1						benign(0)	Tolerated(0.64)	C0	163	0	0	PMID: 25256356 (Lemmers, HMG 2015)	
F47	Rf1126	208	F c.4404G>A	q.2760707G>A	M1468I	M	P-ORF	exon 35	22	-34	5						benign(0)	Tolerated(0.64)	C0	163	0	0	PMID: 25256356 (Lemmers, HMG 2015)	
F55	Rf975	101	M c.244del	q.2656098del		D	D-ORF	exon 1	27	-26	11													PMID: 25256356 (Lemmers, HMG 2015)
F55	Rf975	301	F c.244del	q.2656098del		D	D-ORF	exon 1	13	-38	-3													PMID: 25256356 (Lemmers, HMG 2015)
F55	Rf975	302	F c.244del	q.2656098del		D	D-ORF	exon 1	13	-36	-1													PMID: 25256356 (Lemmers, HMG 2015)
F57	Rf743	2	F c.182_183dup	q.2656256_2656257dup		D	D-ORF	exon 1	12	-34	-1													
F57	Rf743	5	F c.182_183dup	q.2656256_2656257dup		D	D-ORF	exon 1	9	-31	-1													
F57	Rf743	4	M c.182_183dup	q.2656256_2656257dup		D	D-ORF	exon 1	17	-26	5													
F60	Rf1101	1	M c.582dupT	q.2674088dup		N	D-ORF	exon 5	11	-28	1													
F67	Rf1033	2	F c.1302_1306delTGA	q.2697999_2698003del		D	D-ORF	exon 10	10	-22	3	rs387907319												PMID: 25256356 (Lemmers, HMG 2015)
F69	Rf393	101	M c.1608del	q.2700875del		D	D-ORF	exon 12	12	-38	-3													PMID: 23143600 (Lemmers, 2010)
F69	Rf393	302	F c.1608del	q.2700875del		D	D-ORF	exon 12	19	-35	2													PMID: 23143600 (Lemmers, 2010)
F69	Rf393	303	M c.1608del	q.2700875del		D	D-ORF	exon 12	19	-26	6													PMID: 23143600 (Lemmers, 2010)
F69	Rf393	305	M c.1608del	q.2700875del		D	D-ORF	exon 12	21	-33	4													PMID: 23143600 (Lemmers, 2010)
F69	Rf393	206	F c.1608del	q.2700875del		D	D-ORF	exon 12	11	-36	-3													PMID: 23143600 (Lemmers, 2010)
F71	Rf1637	4	F c.2088_2138del	q.2707585_2707635del		D	D-ORF	exon 16	8	-30	-2													
F71	Rf1637	5	F c.2088_2138del	q.2707585_2707635del		D	D-ORF	exon 16	10	-31	-1													
F77	Rf909	202	F c.2665dupA	q.2724958dup		D	D-ORF	exon 21	7	-22	1													PMID: 25256356 (Lemmers, HMG 2015)
F77	Rf909	102	F c.2665dupA	q.2724958dup		D	D-ORF	exon 21	5	-28	-3													PMID: 25256356 (Lemmers, HMG 2015)
F88	Rf988	3	M 1.2 Mb deletion SMCHD1	1.2 Mb deletion SMCHD1		D	D-ORF	exon 12	26	-2	2													PMID: 25820463 (Lemmers, Hum Mut, 2015)
F88	Rf988	4	F 1.2 Mb deletion SMCHD1	1.2 Mb deletion SMCHD1		D	D-ORF	exon 12	31	-21	0													PMID: 25820463 (Lemmers, Hum Mut, 2015)
F88	Rf988	5	M 1.2 Mb deletion SMCHD1	1.2 Mb deletion SMCHD1		D	D-ORF	exon 12	16	-25	5													PMID: 25820463 (Lemmers, Hum Mut, 2015)
F89	Rf929	3	F 1.2 Mb deletion SMCHD1	1.2 Mb deletion SMCHD1		D	D-ORF	exon 12	20	-25	7													PMID: 25820463 (Lemmers, Hum Mut, 2015)
F90	Rf922	2	F c.412C>T	q.2667018C>T		N	D-ORF	exon 3	12	-42	-5													PMID: 25256356 (Lemmers, HMG 2015)
F99	LOVD28	2	F c.1819A>T	q.2703861A>T		N	D-ORF	exon 13	10	NA	NA													PMID: 27061275 (Hamanaoka, 2016)
F102	Rf947	3	M c.2656C>T	q.2724949C>T		N	D-ORF	exon 21	8	-33	-3	rs201632358												
F102	Rf947	2	F c.2656C>T	q.2724949C>T		N	D-ORF	exon 21	13	-26	3	rs201632358												
F107	Rf1727	1	F c.3631C>T	q.2740817C>T		N	D-ORF	exon 28	13	-31	1													
F108	Rf1552	201	M c.4267C>T	q.2751377C>T		N	D-ORF	exon 33	17	-27	5													
F108	Rf1552	102	F c.4267C>T	q.2751377C>T		N	D-ORF	exon 33	17	-25	6													
F113	Rf400	4	M c.5383C>T	q.2777820C>T		N	D-ORF	exon 43	18	-33	3	rs867694014												
F115	Rf629	2	F c.5602C>T	q.2784502C>T		N	D-ORF	exon 45	8	-24	1	rs1229050345												PMID: 25256356 (Lemmers, HMG 2015)
F115	Rf629	3	M c.5602C>T	q.2784502C>T		N	D-ORF	exon 45	3	-26	-3	rs1229050345												PMID: 25256356 (Lemmers, HMG 2015)
F115	Rf629	4	M c.5602C>T	q.2784502C>T		N	D-ORF	exon 45	8	-21	2	rs1229050345												PMID: 25256356 (Lemmers, HMG 2015)
F115	Rf629	5	M c.5602C>T	q.2784502C>T		N	D-ORF	exon 45	8	NA	NA	rs1229050345												
F115	Rf629	10	F c.5602C>T	q.2784502C>T		N	D-ORF	exon 45	14	-26	3	rs1229050345												
F115	Rf629	11	F c.5602C>T	q.2784502C>T		N	D-ORF	exon 45	12	-28	1	rs1229050345												
F115	Rf629	12	F c.5602C>T	q.2784502C>T		N	D-ORF	exon 45	6	-29	-2	rs1229050345												PMID: 25256356 (Lemmers, HMG 2015)
F118	LOVD32	2	M c.187_403>A	q.26861503>A		S3	D-ORF	intron 1	10	NA	NA													PMID: 27061275 (Hamanaoka, 2016)
F121	Rf744	4	F c.1843_15A>G	q.2705677A>G		S3	P-ORF	intron 13	10	-34	-3													
F124	Rf549	1	M c.2147_1G>C	q.2707804G>C		S3	P-ORF	intron 16	7	NA	NA													
F129	Rf1352	2	M c.3634_19A>G	q.2743740A>G		S3	D-ORF	intron 29	4	-28	-3													
F131	Rf1034	4	F c.4347_238A>G	q.2760414A>G		S3	D-ORF	intron 34	9	-30	-1													PMID: 25256356 (Lemmers, HMG 2015)
F131	Rf1034	5	F c.4347_238A>G	q.2760414A>G		S3	D-ORF	intron 34	9	-30	-1													
F131	Rf1034	3	F c.4347_238A>G	q.2760414A>G		S3	D-ORF	intron 34	29	-26	12													
F136	Rf691	1	M c.186+1G>A	q.2656261G>A		S5	P-ORF	intron 1	14	-31	1													
F136	Rf691	5	M c.186+1G>A	q.2656261G>A		S5	P-ORF	intron 1	9	-34	-3													
F139	Rf394	301	F c.873+1G>A	q.2688746G>A		S5	P-ORF	intron 7	10	-37	-4													
F154	Rf947	3	M c.2700+1G>T	q.2724994G>T		S5	D-ORF	intron 21	8	-33	-3													
F154	Rf947	2	F c.2700+1G>T	q.2724994G>T		S5	D-ORF	intron 21	13	-26	3													
F159	Rf1110	3	F c.3048+2T>C	q.2729409T>C		S5	P-ORF	intron 24	10	-41	-6													PMID: 24075187 (Sacconi, 2013)
F165	Rf874	3	F c.3274_3276+1del	q.2732488_2732491del		S5	P-ORF	intron 25	12	-35	-2													PMID: 23143600 (Lemmers, 2010)
F171	Rf1414	4	F c.3276_3276+4del	q.2732490_2732494del		S5	P-ORF	in																



## Supplementary table 2

Table summarizing the variant analysis in 187 unrelated FSHD2 families (F1-F187), 41 BAMS (B1-B41) families, 58 non-pathogenic variants (C1-C58) and 2 for which the pathogenicity is unclear (X1-X2), with proband in white and family members carrying the variant in grey (column 1). For each individual the family, personal number (nr) and gender (if known) is shown (columns 2, 3 and 4). For each variant, we provide cDNA (based on accession number NM\_015295.2), genomic (based on hg19, GRCh37.p5) and protein (NP\_056110.2) information (columns 5, 6 and 7). Column 8 describes the variant type (M=missense, D=insertion/deletion, N=nonsense, S3/S5 splice site variant at the 3' or 5' and SYN=synonymous). The variants are sorted by type and by position. ORF-disrupting (D-ORF) and ORF-preserving (P-ORF) consequence of the variant and the SMCHD1 exon number is shown in columns 9 and 10. The D4Z4 methylations values; FseI, delta 1 and delta2 for the individual are shown in columns 11,12 and 13. Values that were not determined or could not be calculated were indicated with NA. And for control individuals, variants that were analyzed in multiple individuals showing normal methylation values were marked normal. The dbSNP number and the frequency of the variant in the EXAC database (AFR=African, EUR=European; EAS=East Asian and SAS=South Asian populations) in columns 14-18). For missense variants we evaluated the pathogenic effect by the following prediction algorithms: PolyPhen (from Variant Effect Predictor in Ensemble; column 19), SIFT (from Alamut, column 20) and Align GVGD Class (columns 21, 22 and 23). Each program has a different pathogenicity score. A pathogenic prediction within one of the programs received a score of 1 (marked grey) and a benign prediction a score of 0 (marked white). The total prediction score for all 3 algorithms is shown in column 24, where we highlighted the variants in grey that obtained a false negative prediction (D4Z4 hypomethylation, but total prediction score <2), or a false positive prediction (normal D4Z4 methylation, but total prediction score >1). The last column shows the publication, in which the variant was first described.



	Mutation spectrum 2015 <sup>3</sup>		other published variants		new variants current study		total variants	
	n	%	n	%	n	%	n	%
Indel	8	15,7%	11	20,8%	16	19,3%	35	18,7%
Missense	13	25,5%	20	37,7%	21	25,3%	54	28,9%
Nonsense	5	9,8%	5	9,4%	18	21,7%	28	15,0%
Splice site	25	49,0%	17	32,1%	28	33,7%	70	37,4%
Total	51		53		83		187	

Supplementary table 3

SMCHD1 variant type for all variants published in our previous study (n=51)<sup>3</sup>, in 12 other publications (n=53, Supplementary table 1)<sup>5-15</sup>, and new variants (n=83) in the current study.

Distribution	ATPase domain	%	Remaining protein	%	P value	Total
Exons	3-12		1-2 / 13-48			
Amino acids	445	22,2%	1560	77,8%	NA	2005
Indels <sup>1</sup>	10	28,6%	25	71,4%	0,49	35
Missense	27	50,0%	27	50,0%	3,61E-06	54
Nonsense	7	25,0%	21	75,0%	0,90	28
Splicing (S3)	1	5,9%	17	94,4%	0.16 <sup>2</sup>	182
Splicing (S5)	8	15,4%	44	84,6%	0,32	52
						351

<sup>1</sup> Complete gene deletions were not included

<sup>2</sup> Chi-squared approximation may be incorrect

#### Supplementary table 4

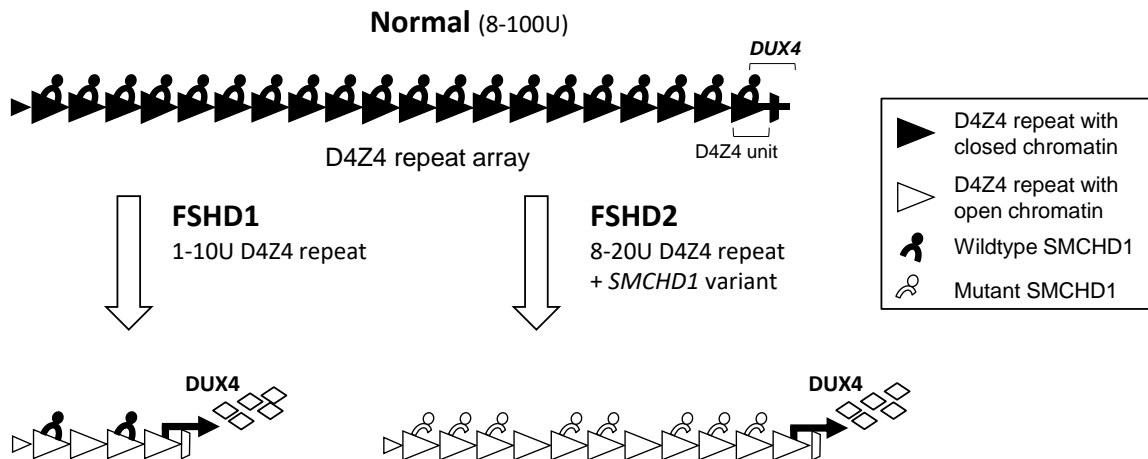
Distribution of the 187 FSHD2-related SMCHD1 variants in the C-terminal extended ATPase domain and in the rest of the protein. C-terminal extended ATPase domain is based on Gordon et al. 2017.<sup>16</sup> In contrast to the other variant types, we observe a significant enrichment (P value 1.59E-05) of missense variants in the extended ATPase domain compared with the size (445/2005 amino acids) of this region.

Proband ID	Sex	cDNA	Protein	Exon	Inheritance	PMID publication
K1	F	c.320T>C	L107P	3	NA	28067909 <sup>17</sup>
D1	M	c.386T>A	M129K	3	NA	28067909 <sup>17</sup>
New	F	c.386T>G	M129R	3	NA	new
11	F	c.400G>T	A134S	3	de novo	28067911 <sup>16</sup>
12	F	c.400G>T	A134S	3	de novo	28067911 <sup>16</sup>
M1 and 2*	F	c.403A>T	S135C	3	de novo	28067909 <sup>17</sup> /28067911 <sup>16</sup>
AF1 and 4*	F	c.403A>T	S135C	3	de novo	28067909 <sup>17</sup> /28067911 <sup>16</sup>
I1	M	c.404 G>A	S135N	3	de novo	28067909 <sup>17</sup>
R1	F	c.404G>A	S135N	3	Probably familial	28067909 <sup>17</sup>
3	M	c.404G>A	S135N	3	de novo	28067911 <sup>16</sup>
AK1	M	c.404G>T	S135I	3	de novo	28067909 <sup>17</sup>
1	M	c.407A>G	E136G	3	de novo	28067911 <sup>16</sup>
T1	M	c.408A>C	E136D	3	Paternal	28067909 <sup>17</sup>
AG1	F	c.410 G>A	G137E	3	NA	28067909 <sup>17</sup>
A1	F	c.415A>C	N139H	3	de novo	28067909 <sup>17</sup>
Y1	F	c.415A>C	N139H	3	NA	28067909 <sup>17</sup>
C1	M	c.423G>C	L141F	3	NA	28067909 <sup>17</sup>
E1	M	c.423G>C	L141F	3	NA	28067909 <sup>17</sup>
S1	F	c.423G>C	L141F	3	NA	28067909 <sup>17</sup>
V1	M	c.423G>T	L141F	3	de novo	28067909 <sup>17</sup>
AB1	M	c.511T>G	F171V	5	Probably familial	28067909 <sup>17</sup>
AA1	M	c.725C>G	A242G	6	de novo	28067909 <sup>17</sup>
10	F	c.1025G>C	W342S	8	de novo	28067911 <sup>16</sup>
O1	F	c.1034A>G	Q345R	8	Maternal	28067909 <sup>17</sup>
F1	M	c.1043A>G	H348R	9	NA	28067909 <sup>17</sup>
L1 and 13*	F	c.1043A>G	H348R	9	NA	28067909 <sup>17</sup> /28067911 <sup>16</sup>
N1 and 5*	M	c.1043A>G	H348R	9	de novo	28067909 <sup>17</sup> /28067911 <sup>16</sup>
X1	F	c.1043A>G	H348R	9	de novo	28067909 <sup>17</sup>
Z1	M	c.1043A>G	H348R	9	NA	28067909 <sup>17</sup>
AC1	M	c.1043A>G	H348R	9	de novo	28067909 <sup>17</sup>
AE1	M	c.1043A>G	H348R	9	de novo	28067909 <sup>17</sup>
14	F	c.1043A>G	H348R	9	NA	28067911 <sup>16</sup>
AH1	F	c.1199A>T	Q400L	10	Paternal	28067909 <sup>17</sup>
P1 and 6*	M	c.1259A>T	D420V	10	de novo	28067909 <sup>17</sup> /28067911 <sup>16</sup>
9	M	c.1259A>T	D420V	10	de novo	28067911 <sup>16</sup>
W1	M	c.1417G>C	E473Q	11	NA	28067909 <sup>17</sup>
8	F	c.1552A>G	K518E	12	NA	28067911 <sup>16</sup>
J1	M	c.1568C>A	T523K	12	NA	28067909 <sup>17</sup>
U1	F	c.1568C>A	T523K	12	NA	28067909 <sup>17</sup>
B1	M	c.1571A>G	N524S	12	NA	28067909 <sup>17</sup>
AJ1 and 7*	M	c.1655G>A	R552Q	13	de novo	28067909 <sup>17</sup> /28067911 <sup>16</sup>

Supplementary table 5

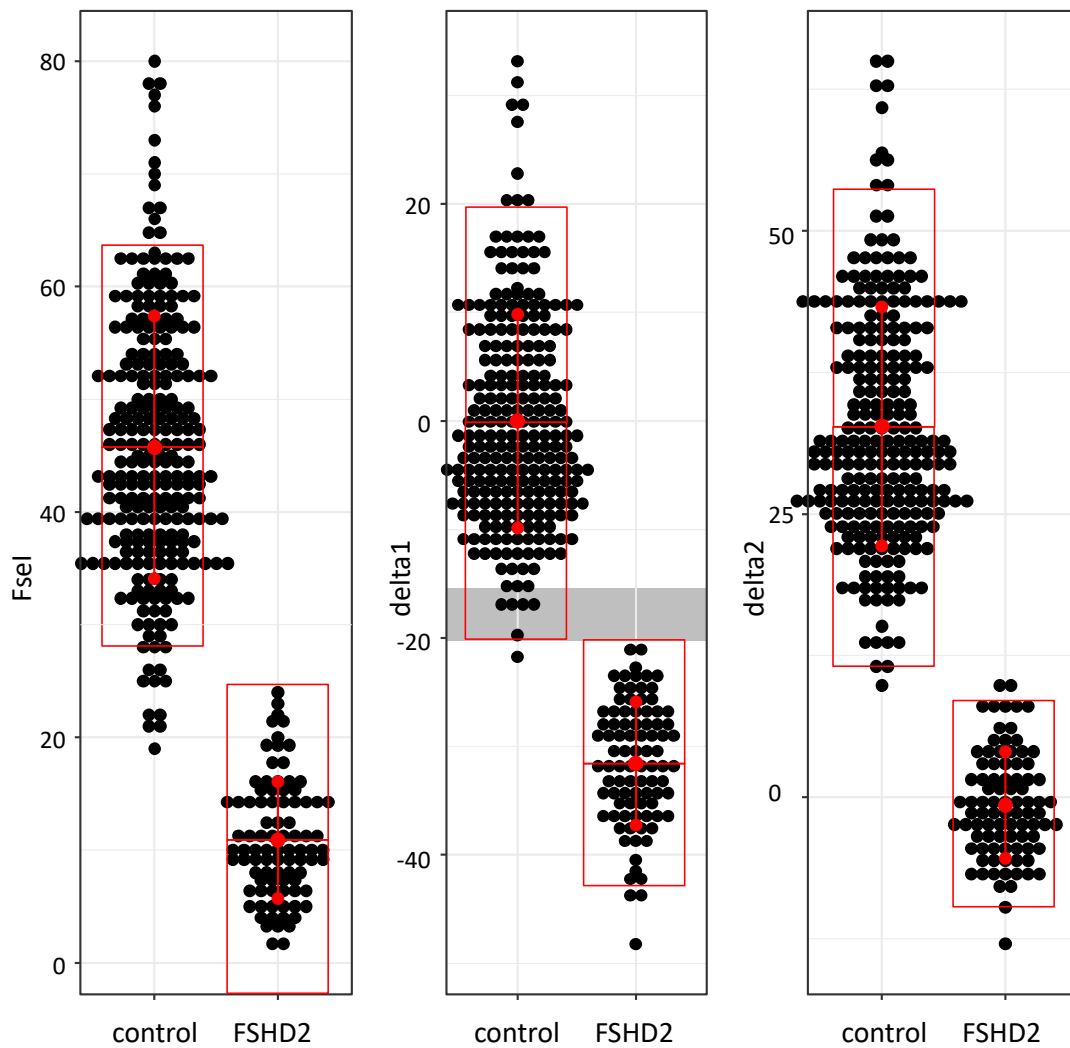
Overview of SMCHD1 variants identified in 41 BAMS families from previous publications.<sup>16, 17</sup> The 1<sup>st</sup> and 2<sup>nd</sup> columns show the reference ID and gender of the proband. The 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> columns shows the position of the variant in the cDNA, the protein and the exon and whether the variants occurred de novo, or not. The PMID of the publication describing the family is shown in the last column.

## Supplementary Figures



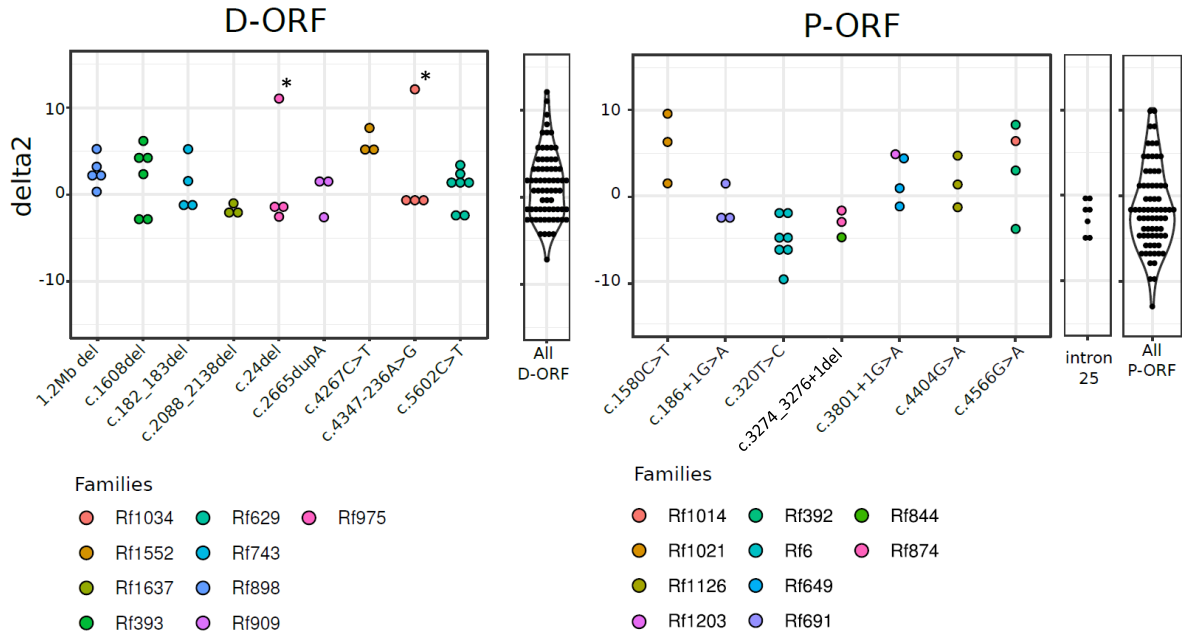
Supplementary figure 1

FSHD is caused by mis-expression of the transcription factor DUX4 in skeletal muscle, where it is normally repressed. A complete copy of the DUX4 retrogene is embedded in the most distal unit of the D4Z4 macrosatellite repeat on chromosome 4 and the region immediately distal to the repeat. In control individuals the repeat varies between 8-100 units, In most FSHD cases, the disease is caused by a D4Z4 repeat contraction to a size of 1-10 units (FSHD1). The less common form FSHD2 is caused by heterozygous variants in the chromatin modifier SMCHD1 in combination with a D4Z4 repeat of 8-20 units, also resulting in DUX4 expression in skeletal muscle. Individual D4Z4 units are depicted as open and filled (representing open and closed chromatin structure) triangles, DUX4 protein expression is indicated with diamonds. Wildtype and mutant SMCHD1 protein are indicated with a closed and open symbol, respectively.



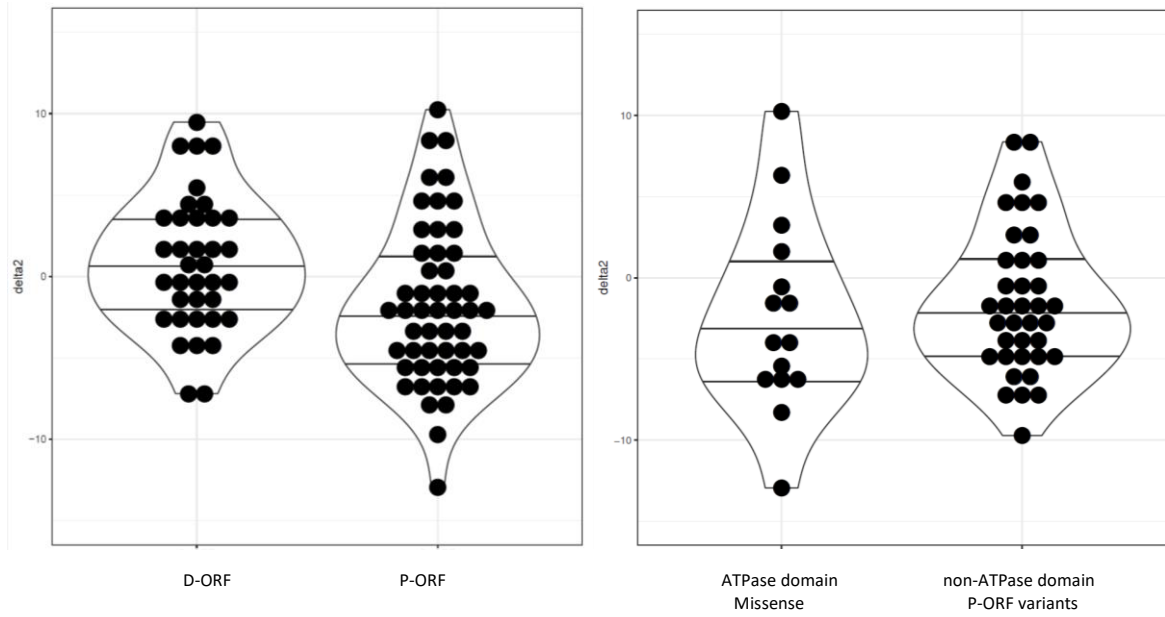
Supplementary figure 2

Threshold methylation values for control individuals (n=249) and for unrelated FSHD2 patients in this study (n=89). The red vertical line with dots indicates the average methylation and 1 SD, while the red box indicates the control and FSHD2 threshold for the different methylation values (1.5 or 2 SD). The FseI methylation value shows the highest variability due to the contribution of the D4Z4 repeat size. The average methylation in controls is 46.8% with a SD of 14.1% and the threshold for FSHD2 is defined at 25% (1.5 SD below the average). The delta1 value has an average value in controls of 0% with an SD of 10.0% and the threshold for FSHD2 is defined at -20.0% (2 SD below the average). We also define a delta1 grey zone between -15.0% to -20.0% where the milder methylation defect might occur due to variants in unknown epigenetic modifiers. The delta2 value is only valid for SMCHD1 mutation carriers. The average is -0.7% and the standard deviation is 4.7%.



Supplementary figure 3

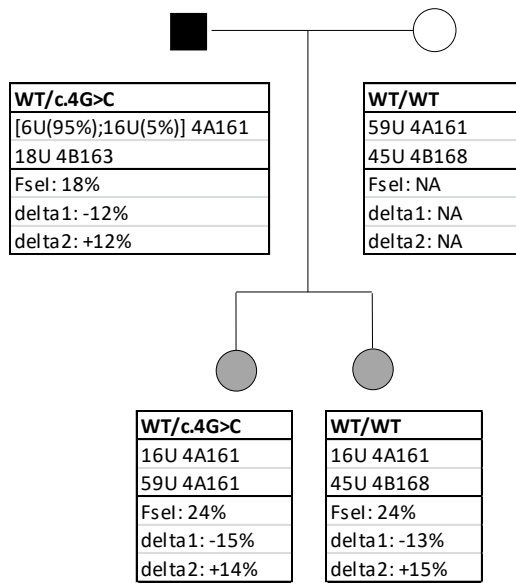
Delta2 methylation values for carriers of the same SMCHD1 variant in comparison to the distribution of all carriers of a disrupting ORF (D-ORF, left) or preserving ORF (P-ORF, right) variant. For most variants the methylation level of different carriers is comparable. Also different intron 25 variants which probably all result in the skip of exon 25 have a comparable delta2 methylation value (P-ORF, right). Individuals with exceptionally high D4Z4 methylation levels discussed in the text are marked with an asterisk.



Supplementary figure 4

Violin plot with delta2 methylation values for unrelated carriers of an D-ORF or P-ORF SMCHD1 variant (left) and for ATPase domain P-ORF missense variants and the other P-ORF variants (right). The lines indicate the average, the 25th and 75th percentiles.

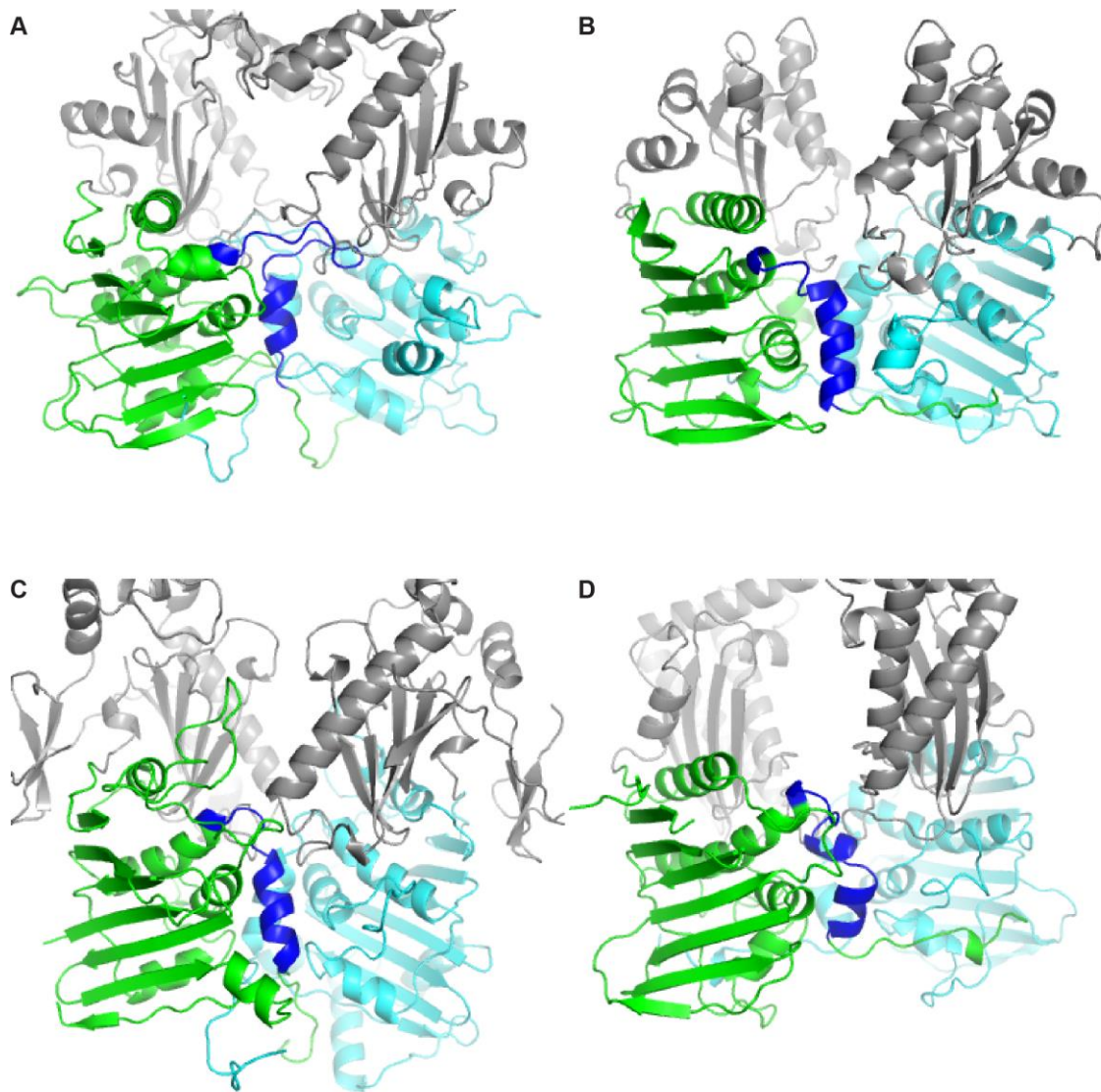
## Rf668



### Supplementary figure 5

Pedigree of family Rf668. Father and both daughters have an Fsel methylation below the FSHD2 threshold and low, but normal, delta1 scores. The father and oldest daughter are heterozygous for the SMCHD1 variant A2P, while the youngest daughter does not carry the variant. The first column shows information on the SMCHD1 variant, the 2<sup>nd</sup> and 3<sup>rd</sup> columns show repeat size and haplotype of both D4Z4 alleles at chromosome 4. The other columns show information on the Fsel, delta1 and delta2 methylation values.





Supplementary figure 6

Comparison of four GHKL ATPase dimers and their downstream ,or C-terminal extended, domains. (A) Computational model of SMCHD1, (B) *E. coli* MutL, (C) MORC2 and (D) *Mycobacterium tuberculosis* GyrB. In each panel, the two ATPase domains are colored in green and cyan, with a helix and loop at the dimer interface marked in dark blue. In SMCHD1, several BAMS mutations localize to this loop. Downstream domains (M-domain, transducer domain) are colored in grey. The ATPase domains show strong structural conservation, while the downstream domains show a much greater structural diversity.

## References

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