Supplementary Data



Figure S1. Inhibition HMGB1 impairs total cell expansion and colony-forming capacity of MDS-L. (A) RTPCR analysis of HMGB1 mRNA expression at 72 h in MDS-L cells with either non-targeting siRNA control (non-target) or HMGB1-specific siRNA (siHMGB1). *P <0.0001 for siHMGB1 compared to non-target. n= 3/group. (B) Left, representative flow cytometric analysis of HMGB1 expression of HMGB1 at 72 h for specified treatments. SSC, side-scattered light. Right, bar graphs indicate percentage of HMGB1+ population and average expression of HMGB1 in mean fluorescence intensity (MFI) value. *P <0.0001. n= 5-6/group. (C) Total cell expansion and (D) Number of colony-forming cells (CFCs) per 1000 cells per dish containing the indicated siRNA. *P <0.05. n= 4-6/group. (E) Apoptotic (Annexin V+ 7AAD-) and necrotic (Annexin V+ 7AAD+) populations in MDS-L containing either non-target siRNA or HMGB1-

specific siRNA. *P= 0.001 and 0.02 for Annexin V+ 7AAD- and Annexin V+ 7AAD+,

respectively. *n*= 4/group. Student's 2-tailed unpaired *t* tests were used in these analyses.

Supplementary Figure 2



Figure S2. Neutrophil Elastase (NE) is differentially expressed in MDS cells compared to CD34+ Cord blood. mRNA expression of NE in primary MDS cells compared to CD34+ cord blood. NE was not detected (ND) in MDS-L cells. *P= 0.02 for MDS compared to cord blood by Mann-Whitney analysis. n= 3-6/group. Student's 2-tailed unpaired t test was used in this analysis.



Figure S3. Dual therapy with sivelestat and chemotherapy is additive to impair MDS function compared to chemotherapy alone. (A) Left, total cell expansion of primary marrow cells from MDS patient 0449 after 72 h incubation with 300 µg/ml sivelestat (Sive) or DMSO. n=4/treatment. Right, number of CFCs following the treatment with sivelestat or DMSO. 5,500 cells for 0449 were plated per dish. n= 3/group. (B) Flow cytometric analysis of HMGB1 expression in CD34⁺ cells from MDS patient 0449. (C) Total cell expansion of MDS-L in cultures at 72 h and 7d with DMSO, 300 µg/ml sivelestat, 1 µM azacitidine, or both. * $P \le$ 0.001 for sivelestat, Aza and

Aza+Sive compared to DMSO. P = 0.005 and <0.0001 for Aza compared to Aza + Sive in 72h and 7d, respectively. n= 5-6/group. (**D**) CFCs from 72 h cultures with treatments indicated in (**C**). Number of cells from culture per dish: 1,000 cells. * $P \le 0.001$ for sivelestat, Aza and Aza+Sive compared to DMSO. P < 0.001 for Aza compared to Aza + Sive. n= 6/group. (**E**) Percentage annexin V+ cells by flow cytometric analysis for 72 h cultures at conditions indicated in (**C**). *P<0.0001 for sivelestat, Aza and Aza+Sive compared to DMSO. P <0.0001 for Aza compared to DMSO. P <0.0001 for Sivelestat, Aza and Aza+Sive compared to DMSO. P <0.0001 for Aza compared to Aza + Sive. n= 5/group. (**F**) Percentage Annexin V+ cells of primary CD34+ MDS cells in culture with DMSO, 75 nM decitabine (Dec) or 75 nM decitabine and 300 µg/ml sivelestat (Dec+Sive) for 7 days. DMSO is shown in gray, Dec in blue and Dec+Sive in red. *P= 0.002 and <0.0001 for Dec and Dec+Sive compared to DMSO. P = 0.03 for Dec+Sive compared to Dec, respectively. n= 4/group. Student's 2-tailed unpaired t tests were used in these analyses.



Figure S4. Sivelestat preserved splenic architecture in mice transplanted with MDS-L cells. The efficacy of sivelestat in an MDS xenograft model was tested in vitro and in vivo. NSG mice were transplanted with MDSL cells that were pretreated with 300 μ g/ml sivelestat for 72h in vitro (a-d). Spleens were harvested for analysis at 17 weeks post-transplantation. (**A**) Wright-Giemsa staining of spleen aspirates showing a lower level of MDS-L engraftment in sivelestat-treated animals. (**B**) Percentages of human CD45 cell engraftment in the spleen aspirates by

flow cytometric analysis. (**C**) Spleen weights are shown. n= 4-5/group. (**D**) Hematoxylin and eosin stain of section of spleen. Scale bar is 20 μ M. For testing drug in vivo, NSG mice were given sivelestat via IP injection (5 mg/kg for 7 days) 24 h after MDSL transplantation (E-F). (**E**) Spleen weights are shown. n= 5-7/group. (**F**) Hematoxylin and eosin stain of section of spleen. Scale bar 20 μ M. Student's 2-tailed unpaired *t* tests were used in these analyses.



Figure S5. Sivelestat inhibits the release of HMGB1 in primary MDS, and modulates TLRrelated signaling pathways. (A) HMGB1 protein expression by western analysis of conditioned

media from CD34+ MDS cells treated with 300 μg/ml sivelestat or DMSO for 12h. Right, quantification of HMGB1 level in conditioned media was normalized to REVERT Total Protein Stain. *n*= 1 for biologic sample; 2 technical replicates/group. (**B**) Quantitative RTPCR measurement of expression of TLR6 and TLR9 in MDS-L cells following culture with DMSO or 300 µg/ml sivelestat for 4-8 h. **P* = 0.006 and <0.0001for TLR6 and TLR9, respectively. *n*= 3/group. (**C**) Left, representative flow cytometry plots of isotype (grey) and anti-phospho-ERK1/2 (blue) in MDS-L cells after incubation with sivelestat for 12 h. Positive gating was defined by isotype control in the individual treatment. Right, quantification of phospho-ERK1/2. **P* = 0.0004 and <0.0001 for 300 µg/ml and 600 µg/ml sivelestat vs. DMSO, respectively. *n*= 4-8/group. (**D**) Levels of phospho-ERK1/2 and total ERK1/2 in MDS-L cells following 12h culture with sivelestat or DMSO. Right, quantification of phospho-ERK1/2 was normalized to total ERK for each sample. **P* =0.02 for sivelestat compared to DMSO, *n*=2-3/group. (**E**) Protein expression of IKBα in MDS-L cells treated with sivelestat or DMSO for 12h. **P* = 0.01 for sivelestat vs DMSO, *n*=4-5/group. Student's 2-tailed unpaired *t* tests were used in these analyses.

Supplementary Fig. 6



Figure S6. Sivelestat modulates the innate immune response in MDS-L cells via the NF_KB pathway. Proteome Profiler Arrays for the NF_KB pathway of protein lysates from MDS-L cells cultured with DMSO or sivelestat (300 or 600 µg/ml) for 24 h are shown. (**A**) Images of membranes from Proteome Profiler Arrays are shown following culture sivelestat compared to control DMSO. Four pairs of reference spots are shown at the following locations: A1, A12, F1 are positive references and F12 is negative control. Proteins were assayed in duplicates. Protein targets selected in Fig. 6 are highlighted with red boxes as follows: CARD6 (B3), FADD (B7), IRAK1 (C5), TNFRSF3 (C10), ReIA pS529 (D7), c-ReI (D8), TNFRII (E6), and TNFRSF10A (E8). (**B**) Levels of proteins are quantified with ImageJ software following sivelestat treatment compared to control DMSO cultures.





Figure S7. Interleukin Receptor Associated Kinase-1 (IRAK1) is differentially expressed in MDS cells compared to CD34+ Cord blood. Real-time PCR analysis of IRAK1 expression in primary MDS cells and MDS-L cells compared to CD34+ Cord blood. Data are normalized to GAPDH internal control and shown relative to CD34+ cord blood expression. *P= 0.005 and 0.0001 for MDS and MDS-L compared to cord blood, respectively. n= 6/group. Student's 2-tailed unpaired *t* tests were used in these analyses.