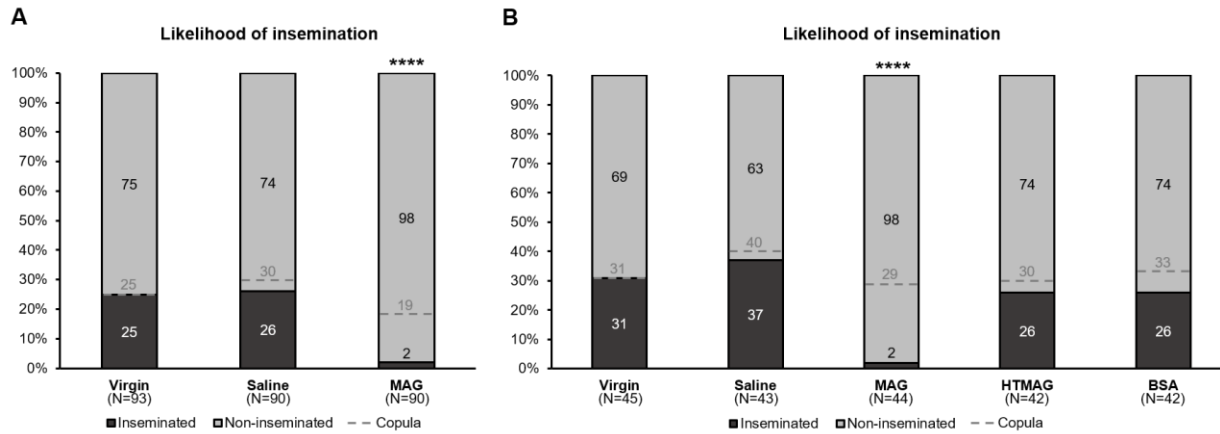


1 **Supplemental Information: Male accessory gland molecules inhibit harmonic**
 2 **convergence in the mosquito *Aedes aegypti***

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4



5

6 **Figure S1. MAG-induced inhibition of harmonic convergence correlates with**

7 **lower insemination rates.** Treatment had a significant effect on the proportion of

8 apparent copulas that resulted in female insemination (χ^2 : treatment, $P < 0.05$ for both

9 sets of replicates). (A) MAG-injected females (2%; $N = 90$) were successfully inseminated

10 less often compared to virgin (25%; $N = 93$) and saline-injected (26%; $N = 90$) control

11 females. (B) Heat-treated MAG (HTMAG; 26%; $N = 42$) and BSA-injected (26%; $N = 42$)

12 females did not display lower insemination rates compared to virgin (31%; $N = 45$) and

13 saline-injected (37%; $N = 43$) control females, but were more likely to be inseminated

14 than MAG-injected (2%; $N = 44$) females. Treatment did not have a significant effect on

15 apparent copula formation ($N = 89-93$ and $N = 43-45$ pairs per group for replicates 1-7

16 and 4-7, respectively; χ^2 : treatment, $P \geq 0.124$ for both sets of replicates) and copula

17 formation did not correlate with HC (χ^2 : $P \geq 0.268$ for both sets of replicates). Mated

18 groups were excluded from the mating outcome analyses because they were already

19 100% inseminated prior to testing. Dashed gray lines represent the percent of apparent

20 copulas formed in each group based on our visual analysis. Asterisks denote

21 significance for insemination data compared to the saline control group. **** $P \leq 0.0001$.

22

Treatment	Converging sex								Converging harmonics							
	M		F		B		U		2:1		3:2		4:3		5:3	
	Ratio	%	Ratio	%	Ratio	%	Ratio	%	Ratio	%	Ratio	%	Ratio	%	Ratio	%
Virgin	15/42	36	1/42	2	2/42	5	24/42	57	18/42	43	9/42	21	0/42	0	15/42	36
Saline	12/40	30	-	-	4/40	10	24/40	60	19/40	47.5	6/40	15	2/40	5	13/40	32.5
Mated	5/20	25	-	-	-	-	15/20	75	10/20	50	2/20	10	0/20	0	8/20	40
MAG	5/18	28	-	-	-	-	13/18	72	9/18	50	4/18	22	0/18	0	5/18	28
HTMAG	2/19	11	-	-	-	-	17/19	89	10/19	53	2/19	10	1/19	5	6/19	32
BSA	5/20	25	-	-	-	-	15/20	75	14/20	70	2/20	10	1/20	5	3/20	15
Total	44/159	28	1/159	<1	6/159	4	108/159	68	80/159	50	25/159	16	4/159	3	50/159	31

23

24 **Table S1. Harmonic convergence is most often achieved by males at the female**

25 **second and male first harmonics.** Converging sex and converging harmonics data

26 presented for each treatment group. Most often the converging sex shifted its flight

27 tones too early in an interaction to determine which sex's flight tone modulations

28 achieved HC (U, unknown: 68% of converging pairs). However, when either sex's flight

29 tone modulations resulted in HC in a detectable manner, HC was almost always
30 achieved by the male alone (M, male: 28%) rather than the female alone (F, female:
31 <1%) or both sexes (B, both: 4%). The sex responsible for convergence did not vary
32 between treatment groups (N=18–42 and N=9–21 per group for trials 1–7 and 4–7,
33 respectively; χ^2 : $P \geq 0.601$ for both sets of replicates). Convergence occurred most often
34 by the merging of the female second and male first harmonics (2:1; 50%) and at similar
35 harmonic combinations across all replicates (N=18–42 and N=9–21 per group for
36 replicates 1–7 and 4–7, respectively; χ^2 : $P \geq 0.731$ for both sets of replicates). Data for
37 each treatment group were pooled across replicates 1–7 for trials including virgin,
38 saline, mated, and MAG groups or across replicates 4–7 for trials containing additional
39 HTMAG and BSA groups. M, male; F, female; B, both; U, unknown. 2:1: Female second
40 and male first harmonics; 3:2: Female third and male second harmonics; 4:3: Female
41 fourth and male third harmonics; 5:3: Female fifth and male third harmonics.

42

43 **Supplemental Experimental Procedures**

44

45 *Mosquito rearing and maintenance*

46 *Aedes aegypti* mosquitoes (Thai strain) originated from collections in Bangkok,
47 Thailand (15°72'N, 101°75'E) and have been maintained in colony with annual
48 supplementation of F₁ eggs since 2009. Mosquitoes were reared in an environmental
49 chamber as described previously [1] to obtain uniform, medium-body-sized adults (male
50 wing lengths: 2.21 ± 0.08 mm; females: 2.87 ± 0.12 mm) [2]. Briefly, eggs were vacuum
51 hatched and 200 larvae were placed in plastic trays containing 1 L of distilled water and
52 4 fish food pellets the following day (Hikari Cichlid Gold, Hayward, CA, USA). Male and
53 female pupae were visually separated by size and females were isolated in test tubes to
54 confirm their sex and ensure their virgin status prior to transfer into 8 L plastic buckets,
55 where they fed on a 10% sucrose solution *ad libitum*.

56

57 *MAG extract preparation, mosquito injections, and antennae removal*

58 A total of fifty MAGs were dissected from males at two-to-three days post-
59 eclosion and pooled at a 1:1 ratio in a 50 μ l solution (one-quarter MAG-equivalent
60 injections) of modified phosphate-buffered saline (PBS; pH 6.9) or at a 2:1 ratio in 25 μ l
61 PBS (for one-half MAG-equivalent injections). Standard injections were performed at
62 the one-quarter MAG-equivalent dose as this concentration is effective for inducing a
63 complete refractory mating response in *Ae. aegypti* [3] and corresponds approximately
64 to a single male ejaculate [4]. The buffer was comprised of 133.58 mM sodium chloride
65 (Fisher Scientific, Pittsburgh, PA, USA), 2.63 mM potassium chloride (Fisher), 9.75 mM
66 sodium phosphate dibasic (Fisher), 3 mM potassium phosphate monobasic (Fisher),
67 and 2 mM calcium chloride dihydrate (MilliporeSigma, St. Louis, Missouri, USA) in Milli-
68 Q water (MilliporeSigma). Buffer was filtered through a Durapore Membrane Filter (0.22
69 μ m pore size; MilliporeSigma). Dissected MAGs were then homogenized, sonicated for
70 30 s, and centrifuged at 4 °C and 14,500 rcf for 15 min, and the resulting supernatant
71 was used for MAG injections.

72 Injections were performed on female mosquitoes at two-to-three days post-
73 eclosion using a Nanoject III Programmable Nanoliter Injector (Drummond Scientific
74 Company, Broomall, PA, USA) and finely pulled glass capillary needles to inject liquids

75 into the lateral thorax at the anepisternal cleft. MAG contents were injected at a one-
76 quarter MAG-equivalent concentration, or 250 nl injections of the 1 MAG/ μ l PBS
77 solution, and in the case of the MAG dosage experiment, at one-half MAG-equivalent
78 concentration, or 250 nl of 2 MAG/ μ l PBS solution. For injection of heat-treated MAG
79 contents, MAG homogenate was heated for 5 min at 95 °C in a Bio-Rad C1000 Thermal
80 Cycler (Bio-Rad Laboratories, Hercules, CA, USA) [1]. For experiments comparing BSA
81 and MAG injections, protein concentrations were verified for equivalency using a Pierce
82 BCA Protein Assay Kit (Thermo Fisher Scientific, Waltham, MA, USA). For BSA
83 injections, we injected BSA protein powder dissolved in PBS at approximately 1 μ g/ μ l
84 concentration, as this is close to the average one-quarter MAG-equivalent homogenate
85 concentration of 1.03 ± 0.27 ug/ μ l (N=8 samples). To control for potential needle injury
86 effects, we included a cohort of females injected with 250 nl of PBS alone (saline
87 treatment).

88 All injected females as well as non-injected controls, including mated females
89 (males added at a 1:1 male to female ratio), were placed in 2 L wax-lined cardboard
90 recovery cages and transferred to an environmental chamber under the conditions
91 referenced above. Mosquitoes were held for two days prior to audio recordings and
92 were provided with 10% sugar-soaked cotton pads. Males were removed from the
93 mated group cage 24 h prior to recordings.

94

95 *Audio recording and analysis of pre-copulatory flight interactions*

96 Females were tethered using nail glue (L.A. Colors, Ontario, CA, USA or similar
97 product) at the dorsal mesothorax by a human hair attached to an insect pin [5]. Similar
98 to previously described recording methods [7], tethered females were placed
99 approximately 3 cm away from either a particle velocity microphone or an
100 omnidirectional microphone (NR-21358 and FG-23329-C05, respectively; Knowles,
101 Itasca, IL, USA) attached to a custom amplifier [6], in a 20 x 14 x 10 cm plastic
102 recording arena. Upon initiation of female flight, audio recordings using Audacity 2.1.3
103 and 2.2.2 software (<https://www.audacityteam.org/>, accessed May 23, 2017 and
104 updated April 13, 2018) were initiated and three virgin males were added into the arena.
105 The recording was stopped after the first male-female pre-copulatory flight interaction
106 lasting > 1 s was recorded and the female either persistently rejected or formed an
107 apparent mating copula with the male. A flight interaction was defined as a male
108 approaching a female in flight and rapidly modulating his flight tones in an attempt to
109 secure, harmonically converge, and mate with the female. To be considered a copula,
110 the couple had to be connected for > 8 s, as this is the minimum amount of time
111 necessary for insemination to occur [8,9]. After audio recordings, female spermathecae
112 were dissected to verify insemination status. If an apparent copula led to successful
113 insemination, the copula was considered a mating copula; if not, it was considered a
114 pseudocopula [10,11]. All recordings were performed at 28.04 ± 1.75 °C and $49.63 \pm$
115 6.96% RH.

116 To determine whether the male and female wing beat harmonics converged
117 during flight interactions, audio files were analyzed as spectrograms using Raven Pro
118 1.5 software (Bioacoustics Research Program, Cornell Laboratory of Ornithology,
119 Ithaca, NY, USA). Potential instances of convergence were systematically tested at all
120 harmonic combinations below 3,000 Hz, including the female second and male first

121 harmonics (2:1), the female third and male second harmonics (3:2), the female fourth
122 and male third harmonics (4:3), the female fifth and male third harmonics (5:3), and the
123 female fifth and male fourth harmonics (5:4). Because convergence was never detected
124 at 5:4, this harmonic combination was excluded in all analyses. To be considered a true
125 instance of convergence, male and female harmonics had to be within 5 Hz of each
126 other for at least 1 s [12]. The sex responsible for HC was defined as the individual,
127 male or female (or both), who shifted his or her flight tone during a courtship interaction
128 to achieve convergence. If HC occurred too early in an interaction to determine which
129 sex's flight tone shift achieved convergence, the sex responsible for convergence was
130 deemed unknown. Average flight tone frequencies during male-female interactions
131 across treatment groups were similar for most replicates (N=90–93 and N=43–45 per
132 group for replicates 1–7 and 4–7, respectively; two-way ANOVA: treatment, $P > 0.05$ for
133 nine out of eleven replicates), with males flying at a fundamental frequency of $841.82 \pm$
134 56.59 Hz and females flying at 476.81 ± 36.11 Hz across all treatment groups and
135 replicates. This, together with the converging harmonics data (Figure S1), suggests that
136 the differences in ability to induce male convergence between mated or MAG-injected
137 females and virgin females are caused by subtle (that is, ≥ 5 Hz tone shifts), rather than
138 drastic changes in courtship flight tone frequencies.

139

140 *Statistical analysis*

141 All data were analyzed using IBM SPSS Statistics software (SPSS version 24,
142 IBM Corp., Armonk, NY). For all *post hoc* pairwise comparisons, significant differences
143 were identified only for those comparisons with P values that were less than 0.05 and
144 were less than their Benjamini-Hochberg critical values with a false discovery rate of
145 0.25 [13–15]. All means are presented \pm standard deviation (SD). HC, copulation, and
146 insemination data were analyzed using a binary logistic generalized linear model, with
147 treatment and replicate effects tested using Wald Chi-Square (χ^2) test. When comparing
148 proportions of insemination events, mated groups (which had 100% inseminated
149 outcomes) were excluded from the analyses to run the binary logistic model, as the 95%
150 confidence intervals as calculated by $3/N$ (where N = the total number of outcomes
151 across all replicates) for these groups were < 0.1 [16]. Fundamental flight tone data were
152 analyzed with a two-way ANOVA test and copulation-convergence correlation,
153 converging harmonics, and converging sex data were analyzed using a χ^2 test.

154 For HC data, after testing for replicate effects and finding none ($P = 0.810$), we
155 combined the seven replicate experiments conducted with virgin, saline, mated, and
156 MAG groups. Four of the seven experiments also contained additional heat-treated
157 MAG and BSA controls and were tested for replicate effects ($P = 0.362$), combined into a
158 single analysis, and analyzed separately. Data from all experiments on copulation and
159 insemination rates ($P > 0.05$ for three out of four sets of replicates), fundamental flight
160 tone frequency ($P > 0.05$ for nine out of eleven replicates), as well as converging
161 harmonics and converging sex data ($P > 0.05$ for all twenty-two replicates), were likewise
162 combined into the aforementioned replicate groupings to ensure robust sample sizes in
163 all analyses.

164

165

166

167 **Supplemental References**

- 168 1. Villarreal, S.M., Pitcher, S., Helinski, M.E.H., Johnson, L., Wolfner, M.F., and
169 Harrington, L.C. (2018). Male contributions during mating increase female survival
170 in the disease vector mosquito *Aedes aegypti*. *J. Insect Physiol.* 108, 1–9.
- 171 2. Helinski, M.E.H., and Harrington, L.C. (2011). Male Mating History and Body Size
172 Influence Female Fecundity and Longevity of the Dengue Vector *Aedes aegypti*.
173 *J. Med. Entomol.* 48, 202–211.
- 174 3. Helinski, M.E.H., Deewatthanawong, P., Sirot, L.K., Wolfner, M.F., and
175 Harrington, L.C. (2012). Duration and dose-dependency of female sexual
176 receptivity responses to seminal fluid proteins in *Aedes albopictus* and *Ae.*
177 *aegypti* mosquitoes. *J. Insect Physiol.* 58, 1307–1313.
- 178 4. Alfonso-Parra, C., Avila, F.W., Deewatthanawong, P., Sirot, L.K., Wolfner, M.F.,
179 and Harrington, L.C. (2014). Synthesis, depletion and cell-type expression of a
180 protein from the male accessory glands of the dengue vector mosquito *Aedes*
181 *aegypti*. *J. Insect Physiol.* 70, 117–124.
- 182 5. Cator, L.J., Ng'Habi, K.R., Hoy, R.R., and Harrington, L.C. (2010). Sizing up a
183 mate: variation in production and response to acoustic signals in *Anopheles*
184 *gambiae*. *Behav. Ecol.* 21, 1033–1039.
- 185 6. Arthur, B.J., Sunayama-Morita, T., Coen, P., Murthy, M., and Stern, D.L. (2013).
186 Multi-channel acoustic recording and automated analysis of *Drosophila* courtship
187 songs. *BMC Biol.* 11, 11.
- 188 7. Villarreal, S.M., Winokur, O., and Harrington, L. (2017). The Impact of
189 Temperature and Body Size on Fundamental Flight Tone Variation in the
190 Mosquito Vector *Aedes aegypti* (Diptera: Culicidae): Implications for Acoustic
191 Lures. *J. Med. Entomol.* 54, 1116–1121.
- 192 8. Ponlawat, A., and Harrington, L.C. (2009). Factors Associated with Male Mating
193 Success of the Dengue Vector Mosquito, *Aedes aegypti*. *Am. J. Trop. Med. Hyg.*
194 80, 395–400.
- 195 9. Degner, E.C., and Harrington, L.C. (2016). Polyandry Depends on Postmating
196 Time Interval in the Dengue Vector *Aedes aegypti*. *Am. J. Trop. Med. Hyg.* 94,
197 780–785.
- 198 10. Craig, G.B. (1967). Mosquitoes: Female monogamy induced by male accessory
199 gland substance. *Science.* 156, 1499–1501.
- 200 11. Gwadz, R.W., Craig, G.B., and Hickey, W.A. (1971). Female sexual behavior as
201 the mechanism rendering *Aedes aegypti* refractory to insemination. *Biol. Bull.*
202 140, 201–214.
- 203 12. Cator, L.J., and Harrington, L.C. (2011). The harmonic convergence of fathers
204 predicts the mating success of sons in *Aedes aegypti*. *Anim. Behav.* 84, 627–633.

- 205 13. Benjamini, Y., and Hochberg, Y. (1995). Controlling the False Discovery Rate: A
206 Practical and Powerful Approach to Multiple Testing. *J. R. Stat. Soc. B.* 57, 289–
207 300.
- 208 14. Benjamini, Y., and Yekutieli, D. (2001). The Control of the False Discovery Rate in
209 Multiple Testing under Dependency. *Ann. Stat.* 29, 1165–1188.
- 210 15. McDonald, J.H. (2014). *Handbook of Biological Statistics, Third Edition*
211 (Baltimore, MD: Sparky House Publishing).
- 212 16. van Belle, G. (2008). *Statistical Rules of Thumb, Second Edition* (Hoboken, NJ:
213 John Wiley & Sons, Inc.).
- 214