

OXIDATIVE POTENTIAL VERSUS BIOLOGICAL EFFECTS:

**A review on the relevance of cell-free/abiotic assays as predictors of toxicity from
airborne particulate matter.**

SUPPLEMENTARY MATERIALS

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References to publications cited in the tables are given at the end of the document.

Table S1. Overview of cell culture and animal studies where biological effects of PM and combustion particles have been examined and compared with acellular oxidative capacity

Particles	Ox-capacity (Cell-free)	Conc. (OP)	Biological endpoints	Conc. (Bio-endpoints)	Association (Ox-capacity vs Bio-endpoints)	Reference
Utah Valley PM ₁₀ (3 samples)	Deoxyribose assay	500 µg/mL (?) (not clearly specified)	<i>In vitro</i> cytotoxicity and induction of IL-6 and CXCL8 in BEAS-2B cells	125-500 µg/mL	No apparent association	Frampton et al. 1999
Water soluble and insoluble fractions of Utah Valley PM ₁₀ (2 samples)	Deoxyribose assay	500 µg/mL (?) (not clearly specified)	<i>In vitro</i> CXCL8 release in BEAS-2B cells, <i>in vivo</i> inflammation in rat lungs (PMN and total protein in BALF)	<i>In vitro</i> : 500 µg/well in 12 well plate <i>In vivo</i> : 100-1000 µg/rat	Possible association with both endpoints	Ghio et al. 1999
8 coal fly ashes, crystalline silica, TiO ₂ and coal dust (11 samples)	ESR with and without H ₂ O ₂ and DMPO	30 mg/mL	<i>In vitro</i> cytotoxicity (MTT) and 8-oxo-dG in RLE cells.	Cytotox: not specified LC50 = 0.5-5.1 mg/mL 8-oxo-dG: LC50 conc. 40 cm ² /mL and 5x10 ⁷ particles	Statistical significant correlation with 8-oxo-dG, but not with cytotoxicity <i>Note: Correlation apparently obtained between OH[•]-formation at equal mass and 8-oxo-dG at equal surface area. No correlation with 8-oxo-dG at equal particle number or LC50 conc. TiO₂ (no OH[•] but 2nd most potent inducer of 8-oxo-dG) was excluded from the correlation analysis.</i>	van Maanen et al. 1999
PM _{2.5} from different locations (5 samples)	ESR	-	<i>In vitro</i> DNA single strand breaks (comet assay) in IB3-1 and K652 cells	33 µg/mL	No apparent association <i>Note: Antioxidant treatment abolished the DNA damage, hence the authors suggested that PM-radicals could be responsible for the effects</i>	Dellinger et al. 2001
Utah Valley PM (TSP) treated with metal chelator ± replacement of metals (3 samples)	Deoxyribose assay	1 mg/mL	<i>In vitro</i> CXCL8 release in BEAS-2B cells, <i>in vivo</i> cytotoxicity and inflammation in rat lungs (LDH and total protein in BALF, neutrophilia, fluid infiltrates, and epithelial thickening in lung sections)	<i>In vitro</i> : 62.5-1000 µg/mL <i>In vivo</i> : 1000 µg/rat	Possible association with <i>in vitro</i> and <i>in vivo</i> cytotoxicity. No apparent association with inflammatory reactions <i>in vitro</i> and <i>in vivo</i>	Molinelli et al. 2002
PM ₁₀ , PM _{2.5} and UFP, from different sites and seasons (15 samples)	DTT	5-50 µg/mL	<i>In vitro</i> induction of HO-1 expression, glutathione depletion and mitochondrial damage in RAW264.7 cells, and induction of HO-1 in BEAS-2B cells	RAW264.7: 12-100 µg/mL BEAS-2B: 50 µg/mL	Strong correlation with HO-1 expression in RAW264.7 cells shown by linear regression ($r^2 = 0.97$). Apparent association with other endpoints. <i>Note: OP^{DTT} highly correlated with PAH content ($r^2 = 0.98$)</i>	Li et al. 2003
PM ₁₀ and PM _{2.5}	ESR with H ₂ O ₂ and DMPO	~0.1-2.5 mg/mL (not clearly specified)	<i>In vitro</i> induction of 8-oxo-dG in A549 cells	50 µg/mL	No difference in ability to induce 8-oxo-dG despite considerable variation in acellular oxidative capacity	Shi et al. 2003
PM ₁₀ (3 samples)	ESR with H ₂ O ₂ and DMPO	200 µg/mL	<i>In vitro</i> cytotoxicity and induction of TNF-α, IL-6 and NO in RAW264.7 cells	15-1000 µg/mL	No apparent association	Salonen et al. 2004
Coarse and fine PM from 2 locations (4 samples)	ESR with H ₂ O ₂ and DMPO	~0.18 mg/mL	<i>In vitro</i> CXCL8 and TNF-α in human whole blood, <i>in vivo</i> inflammation (PMN and TNF-α, MIP-2 and LDH) and glutathione depletion in rat lungs (BALF)	<i>In vitro</i> : ~11-355 µg/mL (?) (not clearly specified) <i>In vivo</i> : ~32 mg/rat	No apparent association	Schins et al. 2004

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Table S1 (continued). Overview of cell culture and animal studies where biological effects of PM and combustion particles have been examined and compared with acellular oxidative capacity

Particles	Ox-capacity (Cell-free)	Conc. (OP)	Biological endpoints	Conc. (Bio-endpoints)	Association (Ox-capacity vs Bio-endpoints)	Reference
EC, DEP and CB of different sizes (4 samples)	ESR Oxidation of methionine	100 µg/mL	<i>In vitro</i> oxidative stress (8-isoprostane), intracellular ROS, induction of LTB ₄ , PGE ₂ , and AA, and activation of cPLA ₂ in canine AM	1-32 µg/mL (3-240 cm ² /mL depending on particle type)	Possible association with LTB ₄ and 8-isoprostane Note: No apparent association with AA, PGE ₂ , cPLA ₂ , or ROS	Beck-Speier et al. 2005
PM ₁₀ and PM _{2.5} sampled at different locations and seasons (10 samples)	DTT	Coarse: 40 µg/mL Fine: 10 µg/mL	<i>In vitro</i> cell death (apoptosis and necrosis) in A549 cells and DNA single strand breaks (comet assay) in THP-1 cells	A549: 80 µg/cm ² THP-1: 10 µg/mL	No apparent association	De Vizcaya-Ruiz et al. 2006
PM ₁₀ and PM _{2.5} sampled at different locations and time points (15 samples)	ESR with H ₂ O ₂ and DMPO	Fine: 95 µg/mL Coarse: 80 µg/mL	<i>In vitro</i> DNA single strand breaks (comet assay) in A549 cells	20 µg/cm ²	No overall/general association when considering all PM sampled together <i>Note: Correlation obtained when urban and rural PM were analyzed separately implying involvement of other factors</i>	Shi et al. 2006
PM ₁₀ , PM _{2.5} and UFP from wildfire smoke	ESR with and without H ₂ O ₂ and DMPO	- -	<i>In vitro</i> intracellular H ₂ O ₂ production, lipid peroxidation and DNA strand breaks in RAW264.7 cells	100 µg/mL	Claimed association with intracellular H ₂ O ₂ and lipid peroxidation, but no statistical analysis presented	Leonard et al. 2007
Combustion particles (2) with high and low spin densities	ESR	-	<i>In vitro</i> particle uptake, LTB ₄ release and mitochondrial damage in NR8383 cells, <i>in vivo</i> mitochondrial damage in AM, nitrotyrosine staining and NOx in lungs of mice	<i>In vitro</i> : 10 µg/mL <i>In vivo</i> : 10 µg/m ³ (6h/day, 4 days)	The high-free radical particle induced more effects <i>in vitro</i> and <i>in vivo</i> than the low-free radical particle <i>Note: Effects on LTB₄ directly proportional with increased uptake of high-free radical particles.</i>	Repine et al. 2008
PM _{2.5} sampled during/after a wildfire (5 samples)	DTT	-	<i>In vitro</i> intracellular ROS in primary rat AM	-	No apparent association	Verma et al. 2009
NIST SRM 1648 and SRM 2975, and Toronto PM _{2.5}	DTT	100 or 60 µg/mL	<i>In vitro</i> cytotoxicity and CXCL8 release in A549 cells	50-1000 µg/mL	No apparent association	Akhtar et al. 2010
Size-fractionated PM from 4 locations (20 samples)	ESR with H ₂ O ₂ and DMPO	-	<i>In vitro</i> cytotoxicity, CXCL8 release and oxidative DNA damage (comet assay with FPG) in A549 cells	100 µg/mL	Weak, but statistically significant association with cytotoxicity ($r = 0.366$, $P < 0.001$), CXCL8 ($r = 0.336$, $P < 0.01$) and oxidative DNA damage ($r = 0.559$, $P < 0.001$) <i>Note: OP of smallest size-fraction not associated with biological effects</i>	Wessels et al. 2010

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Table S1 (continued). Overview of cell culture and animal studies where biological effects of PM and combustion particles have been examined and compared with acellular oxidative capacity

Particles	Ox-capacity (Cell-free)	Conc. (OP)	Biological endpoints	Conc. (Bio-endpoints)	Association (Ox-capacity vs Bio-endpoints)	Reference
PM ₁₀ , PM _{2.5} and UFP sampled at 8 locations (24 samples)	DTT	-	<i>In vitro</i> cytotoxicity (MTT test) and induction of TNF- α , IL-6 and MIP-2 in RAW264.7 cells	6.25-100 $\mu\text{g}/\text{mL}$	Statistical significant correlation with cytotoxicity ($\beta = -145$, $P < 0,0001$) <i>Note: Significant correlation with cytokines obtained after excluding the sample with the highest oxidative capacity ($\beta = 45/80/647$, $P < 0,0001$)</i>	Steenhof et al. 2011
Wood smoke particles and fine PM (4 samples)	DCFH ESR with DMPO	1.56-50 $\mu\text{g}/\text{mL}$ 1 mg/mL	<i>In vitro</i> intracellular ROS, DNA damage (comet assay with and without FPG, 8-oxodG, ϵdA , ϵdG , and DNA adducts), and induction of MCP-1, TNF- α , LFA-1, CXCL8, HO-1, OGG1 in THP-1 and A549 cells, <i>in vivo</i> inflammation, oxidative stress and DNA damage in rat lung and liver	<i>In vitro</i> : 1.56-100 $\mu\text{g}/\text{mL}$ <i>In vivo</i> : 0.64 mg/kg (~128 $\mu\text{g}/\text{rat}$)	No apparent association <i>Note: the particle with the highest oxidative capacity also induced the highest level of HO-1 expression in THP-1 cells</i>	Danielsen et al. 2010, and 2011*
PM from 2 locations sampled at two different time points (4 samples)	ESR with H ₂ O ₂ and DMPO	-	<i>In vitro</i> RBC-hemolysis	80 $\mu\text{g}/\text{mL}$	No apparent association	Quintana et al. 2011
Diesel and biodiesel exhaust particles (8 samples)	DTT Depletion of AA	10 $\mu\text{g}/\text{mL}$ 12 $\mu\text{g}/\text{mL}$	<i>In vitro</i> cytotoxicity and IL-6 release in BEAS-2B cells	3.125-200 $\mu\text{g}/\text{mL}$	No apparent association	Gerlofs-Nijland et al. 2013
Day-to-day variations in PM _{2.5} (10 days)	DTT	-	<i>In vitro</i> ROS formation in primary rat AM	PM _{2.5} : 3-87 $\mu\text{g}/\text{m}^3$	Statistical significant correlation ($r_s = 0.86$) <i>Note: OP^{DTT} also strongly correlated with organic carbon ($r_s = 0.80$) and water-soluble organic carbon ($r_s = 0.90$)</i>	Delfino et al. 2013
PM ₁₀ (6 samples)	DTT	-	<i>In vitro</i> intracellular ROS and TNF- α and CXCL8 release in J774A.1 and A549 cells	ROS: 100-400 $\mu\text{g}/\text{mL}$ Cytokines: 25-100 $\mu\text{g}/\text{mL}$	No statistical significant correlation	Lu et al. 2014
Welding fumes (3 samples)	ESR with H ₂ O ₂ and DMPO		Cytotoxicity (mitochondrial dysfunction) intracellular ROS (ESR w/ DMPO), DNA damage (Comet assay), phagocytosis, cytokine release (TNF- α , IL-6, IL-1 β) in RAW264.7 cells	50 and 250 $\mu\text{g}/\text{mL}$	No apparent association	Badding et al. 2014
PM _{2.5} sampled over 2 months at 2 sites (54 samples)	DTT	- Section of PM-filter (different conc. for each sample)	<i>In vitro</i> induction of TNF- α , and IL-6 release in A549 cells (24 and 48 h)	- Aqueous extracts of PM-filter sections (corresponding to 0.5-2.5 mg/mL ?)	Statistically significant correlation (TNF- α : $R^2 = 0.65-0.74$, $p < 0.001$; IL-6: $R^2 = 0.80-0.91$, $p < 0.001$) <i>Note: Exposure concentrations not clearly specified but appears extremely high</i>	Liu et al. 2014

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Table S1 (continued). Overview of cell culture and animal studies where biological effects of PM and combustion particles have been examined and compared with acellular oxidative capacity

Particles	Ox-capacity (Cell-free)	Conc. (OP)	Biological endpoints	Conc. (Bio-endpoints)	Association (Ox-capacity vs Bio-endpoints)	Reference
Water-soluble fractions of PM of different size, from separate sites and seasons (20 samples)	DDT	- Not clearly stated (different conc. for each sample)	<i>In vitro</i> Cytotoxicity (MTT and LDH) and DNA damage (Comet assay)	- Not clearly stated (different conc. for each sample)	No apparent overall association when considering all PM samples together. However, moderate but statistically significant correlation between OP ^{DDT} and MTT when PM sampled at different sites and seasons were analyzed separately ($R^2 = 0.30-0.46$), and strong correlation between OP ^{DDT} and LDH in the winter samples ($R^2 = 0.78-0.79$). Implies that other factors contribute.	Velali et al. 2015
8 diesel exhaust particles	DDT	-	<i>In vitro</i> cytotoxicity in SVEC4-10 cells: cell proliferation (colony formation), MTT test and wound healing (scratch test)	5-100 µg/mL	No apparent association (2-fold variation in OP, but no variation in cytotoxicity)	Fox et al. 2015
108 PM samples	ESR with H ₂ O ₂ and DMPO	-	<i>In vitro</i> cytotoxicity (Neutral red uptake), cytokines (IL-8) and DNA damage (Comet assay with FPG) in BEAS-2B cells	12.5-100 µg/mL	No statistical significant correlation	Van Den Heuvel et al 2016
4 diesel exhaust particles	DDT assay	-	<i>In vitro</i> gene expression of TNF-α and HO-1 in Raw 264.7 cells	-	Statistically significant correlation with both endpoints	Karavalakis et al. 2017
9 PM _{2.5} and 1 diesel exhaust particle	DDT assay, depletion of GSH and AA, plasmid scission assay	10-100 µg/mL	<i>In vitro</i> intracellular ROS (DCFH), gene expression of antioxidants (SOD and HO-1), cytokines (IL-6) and metabolizing enzymes (CYP1A1) in NCH-H292 cells	1-10 µg/mL	Statistically significant correlation <i>Note: OP^{AA} and OP^{GSH} strongest correlated with all endpoints. OP^{GSH} and OP^{DDT} not correlated with IL-6.</i>	Crobeddu et al. 2017
95 PM ₁₀ samples	ESR with H ₂ O ₂ and DMPO	-	<i>In vitro</i> cytotoxicity (Neutral red uptake), and cytokines (IL-8) in BEAS-2B cells. Bacterial mutagenicity by Ames test.	12.5-100 µg/mL	No statistical significant correlation with IL-8 or cytotoxicity Statistically significant correlation between OP and bacterial mutagenicity.	Van Den Heuvel et al. 2018
PM _{0.25} from two airports	ESR with H ₂ O ₂ and DMPO, depletion of GSH and AA	50 µg/mL 12.5 µg/mL	<i>In vitro</i> cytotoxicity (MTT), intracellular ROS (H ₂ DCFDA), and cytokine release (TNF-α, IL-6 and IL-8) in 16HBE cells.	10 and 100 µg/mL	Statistical significant correlation between OP and intracellular ROS No apparent association between OP and cytotoxicity or proinflammatory cytokines	He et al. 20018
3 diesel particles	DDT (PM ethanol extracts)	-	<i>In vitro</i> gene expression of metabolizing enzymes (CYP1A1 and -1B1), cytokine release (TNF-α and IL-8), intracellular Ca ²⁺ .	CYP1/cytokines: 200 µg/mL (50 µg/cm ²) Ca ²⁺ : 2.3 mg/ml	Statistical significant correlation between OP and CYP1 expression ($R^2 > 0.9$), but low correlation with cytokines ($R^2 = 0.2$ and 0.26)	Jaramillo et al. 2018

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Table S2. Overview of epidemiological and human exposure studies where health effects of PM and combustion particles have been examined and compared with acellular oxidative capacity

Particles	OP (Cell-free)	Number of samples	Biological endpoints	Conc. (Bio-endpoints)	Association (Ox-capacity vs Bio-endpoints)	Reference
Utah valley PM ₁₀ collected different times (3 samples)	Deoxyribose assay	3	Pulmonary inflammation in human volunteers (total cells, PMNs, total protein, albumin, fibronectin, α 1-antitrypsin, tissue factor, fibrinogen, IL-1 β , TNF- α , and CXCL8 in BALF)	100 and 500 μ g/person	Possible association with all endpoints <i>Note: effects may be due to high zinc levels which co-variated with redox active metals</i>	Ghio and Devlin 2001
Metal-rich and metal-poor PM _{2.5} (2 samples)	ESR with H ₂ O ₂ and DMPO	2	Pulmonary inflammation in human volunteers (total cells, PMNs, monocytes, IL-6, IL-8, TNF- α in BALF and ROS generation by BAL-cells)	100 μ g/person	Possible association with all endpoints <i>Note: Effects were also associated with the most abundant metal, zinc</i>	Schaumann et al. 2004
TSP, PM ₁₀ and PM _{2.5} sampled at 6 schools sampled at 4 days	ESR with ascorbate and DMPO	-	Lung function (FEV ₁ , FVC, FEF _{50%}) in 651 children (8-13 years) attending the same 6 schools	Outdoor air PM conc.: TSP: 59-101 μ g/m ³ PM ₁₀ : 28-59 μ g/m ³ PM _{2.5} : 15-24 μ g/m ³	Statistically significant negative association with lung function (P < 0.05)	Hogervorst et al. 2006
Predicted weekly PM ₁₀ mass and oxidative potential, 2002-2006	Depletion of GSH in synthetic lung lining fluid	Modeled based on 34 monitoring sites and 841 measures	Carotide intima-media thickness in cohort of 2348 (mean age: 61 yr)	25.0 \pm 0.6 μ g/m ³ (mean \pm SD)	PM ₁₀ mass more strongly correlated than PM ₁₀ OP	Tonne et al. 2012
Day-to-day variations in PM _{2.5} (10 days)	DTT	10	Airway inflammation (exhaled NO) in 45 children with asthma (9-18 yr)	3-87 μ g/m ³	Positive association with exhaled NO in children	Delfino et al. 2013
PM ₁₀ levels measured on day of admission and 14 days before/after	Antioxidant depletion (GSH, AA, uric acid) in synthetic lung lining fluid	-	160 asthma/COPD exacerbations in 151 patients (bi-directional case-crossover study)	-	No statistical significant correlation with asthma/COPD admissions <i>Note: The same authors had earlier reported an association with PM₁₀ mass</i>	Canova et al. 2014
PM _{2.5} sampled from June 2012 to April 2018	DTT assay on water-soluble PM _{2.5} extracts	227 samples. DTT activity modelled for 1998-2009	Emergency department visits for asthma/wheezing and congestive heart failure	-	OP more strongly associated than PM _{2.5} mass	Bates et al. 2015
The RAPTES-project Real life PM-exposure at 5 locations	Depletion of GSH and AA	-	Airway inflammation (exhaled NO), lung function (FEV ₁ , FVC), nasal inflammation (IL-6, IL-8, and lactoferrin), and various vascular inflammatory and coagulative markers in 31 human volunteers	5 h exposure PM ₁₀ : 26-394 μ g/m ³ PM _{2.5} : 16-140 μ g/m ³ PM _{10-2.5} : 9-252 μ g/m ³ PNC: 9-67 x10 ³ /cm ³	No apparent association between acellular oxidative capacity of the different PM-fractions sampled at each location and any of the assessed endpoints	Janssen et al. 2015; Steenhof et al. 2013 and 2014; Strak et al. 2012, 2013a, and 2013b*

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Table S2 (continued). Overview of epidemiological and human exposure studies where health effects of PM and combustion particles have been examined and compared with acellular oxidative capacity

Particles	OP (Cell-free)	Number of samples	Biological endpoints	Conc. (Bio-endpoints)	Association (Ox-capacity vs Bio-endpoints)	Reference
PM ₁₀ and PM _{2.5} , central London, UK	Depletion of GSH and AA in synthetic lung lining fluid	Daily OP levels for 685/703 days in 2011-2012	All-cause non-accidental mortality, death from cardiovascular and respiratory disease, stratified by age (0-14, 15-64 and 65+ years)	PM10: 9-32 µg/m ³ PM2.5: 5-25 µg/m ³ (10 th -90 th percentile)	No statistical significant association with OP in complete population. PM ₁₀ OP ^{AA} showed stronger negatively association with respiratory mortality in elderly (65+), than PM ₁₀ and PM _{2.5} mass.	Atkinson et al. 2016
PM _{2.5} , Netherlands and Belgium	DTT assay, ESR with H ₂ O ₂ and DMPO	Measures of PM ^{2.5} OP from 40 sites, applied to land-use regression (LUR) model	Doctor diagnosed asthma, prevalence of asthma symptoms, hay fever, and rhinitis by age 14. Allergic sensitization, lung function (FEV ₁ , FEVC, FEF ₂₅₋₇₅) and FeNO at age 12, in a birth cohort of 3701 children	-	OP ^{DTT} , but not OP ^{ESR} , statistically significantly associated with asthma incident and symptoms, rhinitis, and lung function. <i>Note: Most associations with lung function, but not symptoms, were lost when adjusting for NO₂.</i>	Yang et al. 2016
PM _{2.5} , Atlanta, USA	DTT assay	OP of water-soluble fraction of 196 daily samples (2011-2012), used to model OP for 1998-2009	Hospital visits for respiratory (pneumonia, COPD, asthma/wheeze) and cardiovascular disease (ischemic heart disease, congestive heart failure) in Atlanta 1998-2009	-	Modelled OP ^{DTT} were statistically significantly associated with asthma,/wheeze and IHD <i>Note: association with PM_{2.5} not assessed, but OP^{DTT} co-varied strongly with PM_{2.5} mass (r = 0.49-0.86)</i>	Abrams et al. 2017
PM _{2.5} , southeastern USA	DTT assay, depletion of AA	OP of water-soluble fraction of 500 samples, used to model OP for 1998-2009	Hospital visits for respiratory (pneumonia, COPD, asthma/wheeze) and cardiovascular disease (ischemic heart disease, congestive heart failure) in Atlanta 1998-2009	-	Modelled OP ^{DTT} , but not OP ^{AA} , were statistically significantly associated with asthma,/wheeze and CHF <i>Note: association with PM_{2.5} not assessed, but OP^{DTT} co-varied strongly with PM_{2.5} mass (r = 0.49-0.86)</i>	Fang et al. 2016
PM _{10-2.5} , PM _{2.5-0.18} , PM _{0.18} and black carbon, Los Angeles, USA	DTT assay	Five days of PM sampling (coarse, fine, ultrafine) prior to each clinical visit in study.	Microvascular function (reactive hyperemia index: RHI) in a cohort panel study of 93 non-smoking adults (65-96 years old)	-	PM _{0.18} OP ^{DTT} , but not PM _{2.5-0.18} OP ^{DTT} or PM _{10-2.5} OP ^{DTT} , was statistically significantly associated with reduced RHI.	Zhang et al. 2016
PM _{2.5} , Montreal, Canada	Depletion of GSH and AA in synthetic lung lining fluid	Personal exposure of 62 asthmatic children of 10 days	Airway inflammation (FeNO)	-	OP ^{GSH} , but not OP ^{AA} or PM _{2.5} mass, was statistically significantly associated with FeNO	Maikawa et al. 2016
PM _{2.5} , Ontario, Canada	Depletion of GSH and AA in synthetic lung lining fluid	PM _{2.5} sampled in 30 cities in Ontario	Cause-specific mortality (all non-accidental, lung cancer, cardio-metabolic, ischemic heart disease, respiratory disease)	-	OP ^{GSH} more strongly associated with lung cancer deaths than PM _{2.5} mass, while PM _{2.5} mass showed stronger association with all non-specific mortality. <i>No associations were observed with OP^{AA}.</i>	Weichenthal et al. 2016A

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Particles	OP (Cell-free)	Number of samples	Biological endpoints	Conc. (Bio-endpoints)	Association (Ox-capacity vs Bio-endpoints)	Reference
PM _{2.5} , Ontario, Canada	Depletion of GSH and AA in synthetic lung lining fluid	PM _{2.5} sampled in 16-31 cities in Ontario	Emergency room visits for respiratory illness and myocardial infarctions, and birth outcomes (preterm birth and low birth weight).	-	OP ^{GSH} modified association between PM _{2.5} respiratory illness, myocardial infarctions and birth outcomes (stronger increases in PM _{2.5} -associated risks were observed in regions with high OP ^{GSH}). <i>No effect-modifications were observed with OP^{AA}.</i>	Weichenthal et al. 2016B, 2016C, Lavigne et al. 2018**
PM ₁₀ , PM _{2.5} and PM _{10-2.5} , Netherlands	DTT assay, ESR with H ₂ O ₂ and DMPO	Measures of PM _{2.5} OP from 40 sites, applied to land-use regression (LUR) model	Diabetes prevalence among 289.703 adults	-	OP ^{DTT} , more strongly associated with diabetes than PM-mass. NO association with OP ^{ESR} . <i>Note: strong associations also observed for NO₂</i>	Strak et al. 2017
PM _{10-2.5} , PM _{2.5-0.15} and PM _{0.3} Ontario, Canada	Depletion of GSH and AA in synthetic lung lining fluid	Controlled exposures to CAPs (3x53)	Clinical trial with 53 healthy volunteers (mean age 28) in chamber Blood biomarkers: ET1, IL-6, CRP, VEGF, MDA, S100, NSE, UCHL1, cortisol, BDNF Urinary biomarkers: VEGF, 8-OHdG, MDA, VMA, HVA, cortisol	PM _{10-2.5} (µg/m ³): 212.6 ± 51.8 PM _{2.5-0.15} (µg/m ³): 238.4 ± 62.0 PM _{0.3} (µg/m ³): 120.0 ± 72.0	OP ^{GSH} was statistically significantly associated with blood IL-6, VEGF and S100, and urinary 8-OHdG. OP ^{AA} was associated with blood UCHL1, and urinary MDA. PM _{2.5} mass was associated with blood MDA and urinary 8-OHdG. <i>Note: as several GSH measurements were below detection levels, OP^{GSH} associations assessed based on a binary model (above or below detection limit)</i>	Liu et al. 2018

Studies marked in green show significant correlation/association between at least one OP-measure and all biological effects assessed, confirmed by statistical analysis. Studies marked in blue show significant correlation/association between at least one OP-measure and one biological effect confirmed by statistical analysis. **The three publications from Ontario, by Weichenthal and colleagues were counted as one, as these were based on the same exposures (PM mass and OP metrics from same sites and sample periods).

Fig S1

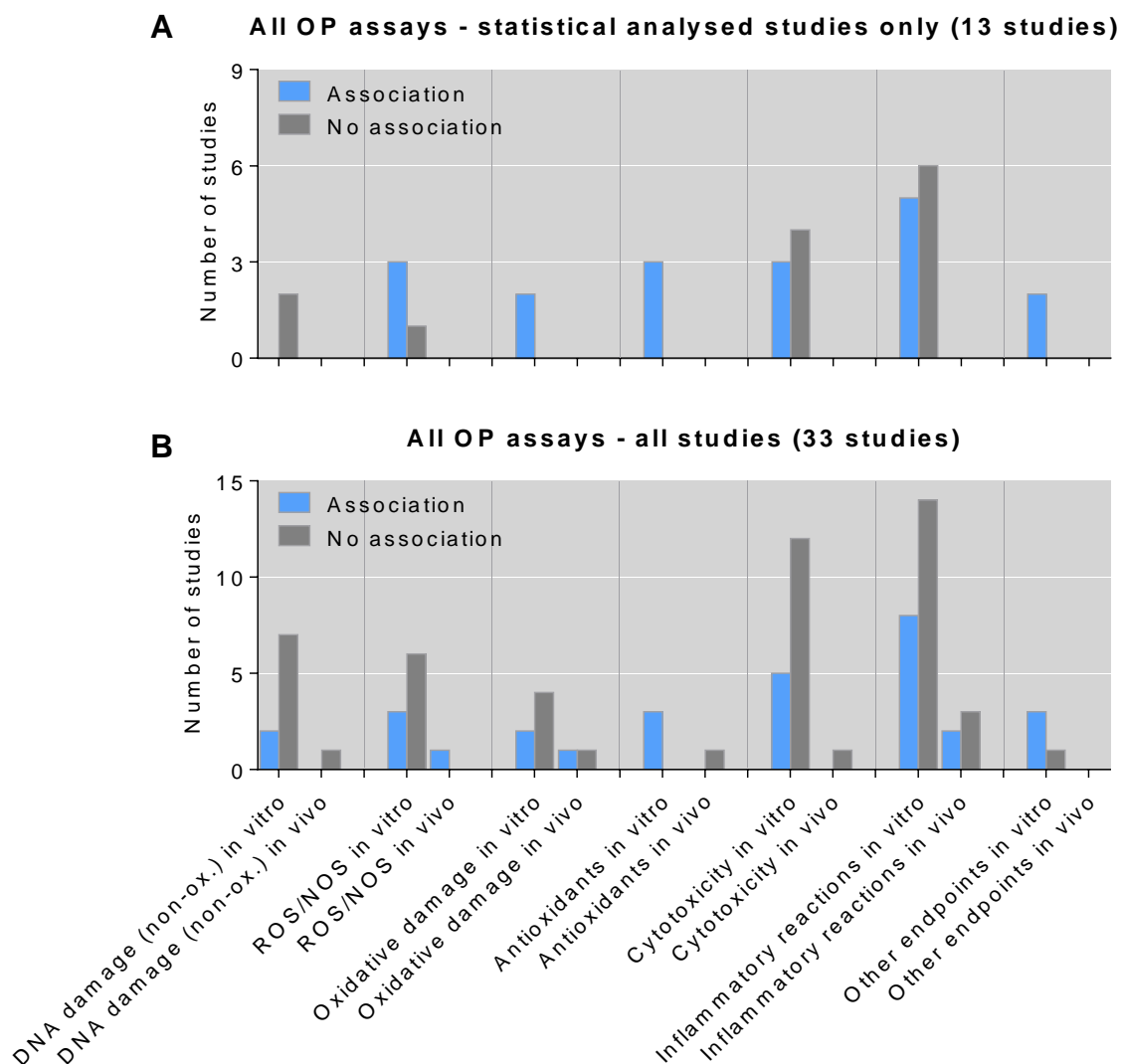


Figure S1. Possible association between all OP assays (pooled) and biological effects of PM in experimental studies *in vitro* and *in vivo*. The figure displays the number of studies showing an association or no association between OP measured by any assay specific biological effects *in vitro* (cell cultures), by statistical analysis only (A) or also including studies where statistics were not applied (B). Data in B was based on comparing rank order of OP vs rank order of biological effects of different PM samples. “Oxidative damage” include lipid peroxidation and oxidative DNA damage. “Other endpoints” include cellular signaling, proliferation. As some studies have explored association between OP and several different biological effects, the sum of the individual columns exceeds the total number of studies given in the figure title.

Fig S2

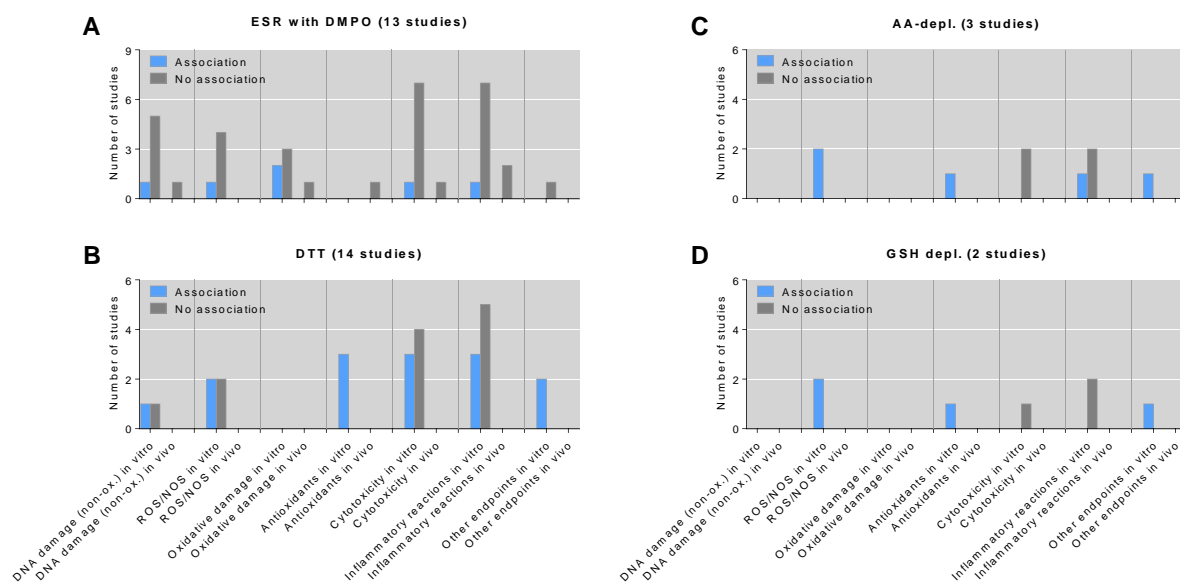


Figure S2. Possible association between specific OP assays and biological effects of PM in experimental studies *in vitro*. The figure displays the number of studies showing an apparent association or no apparent association between OP measured by ESR with DMPO as spin trap (A), DTT-assay (B), AA-depletion (C) or GSH-depletion (D) and specific biological effects *in vitro* (cell cultures). The data includes not only results from studies applying statistical analysis of the association between OP and effects, but also studies where statistical analysis was not applied. When statistical analysis was not available, the rank order of OP vs rank order of biological effects of different PM samples were compared. “Oxidative damage” include lipid peroxidation and oxidative DNA damage. “Other endpoints” include cellular signaling, proliferation. As some studies have explored association between OP and several different biological effects, the sum of the individual columns exceeds the total number of studies given in the figure title. AA – ascorbic acid; DMPO - 5,5-dimethyl-pyrroline N-oxide; DTT – dithiothreitol; ESR – electron spin resonance; GSH – reduced glutathione.

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