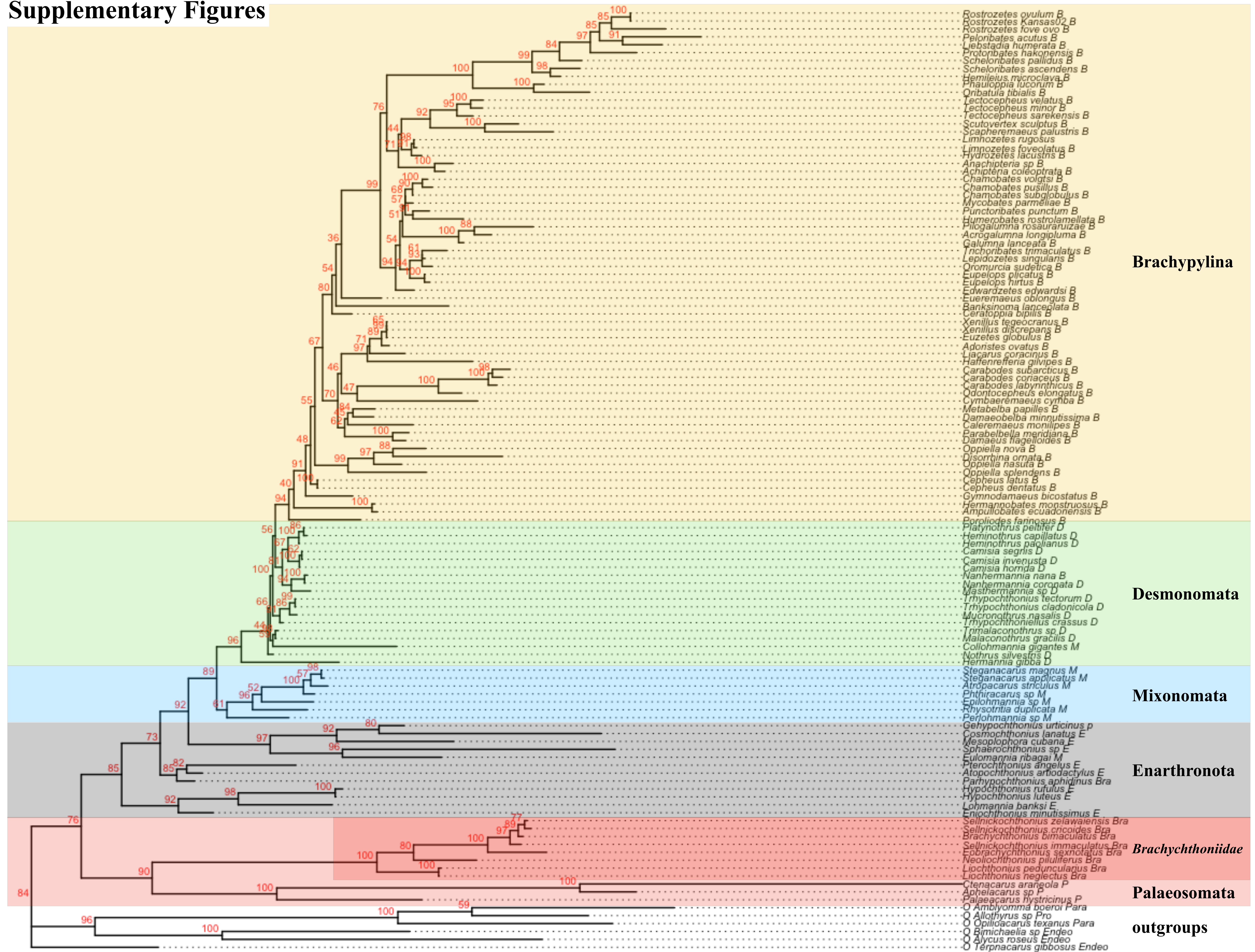
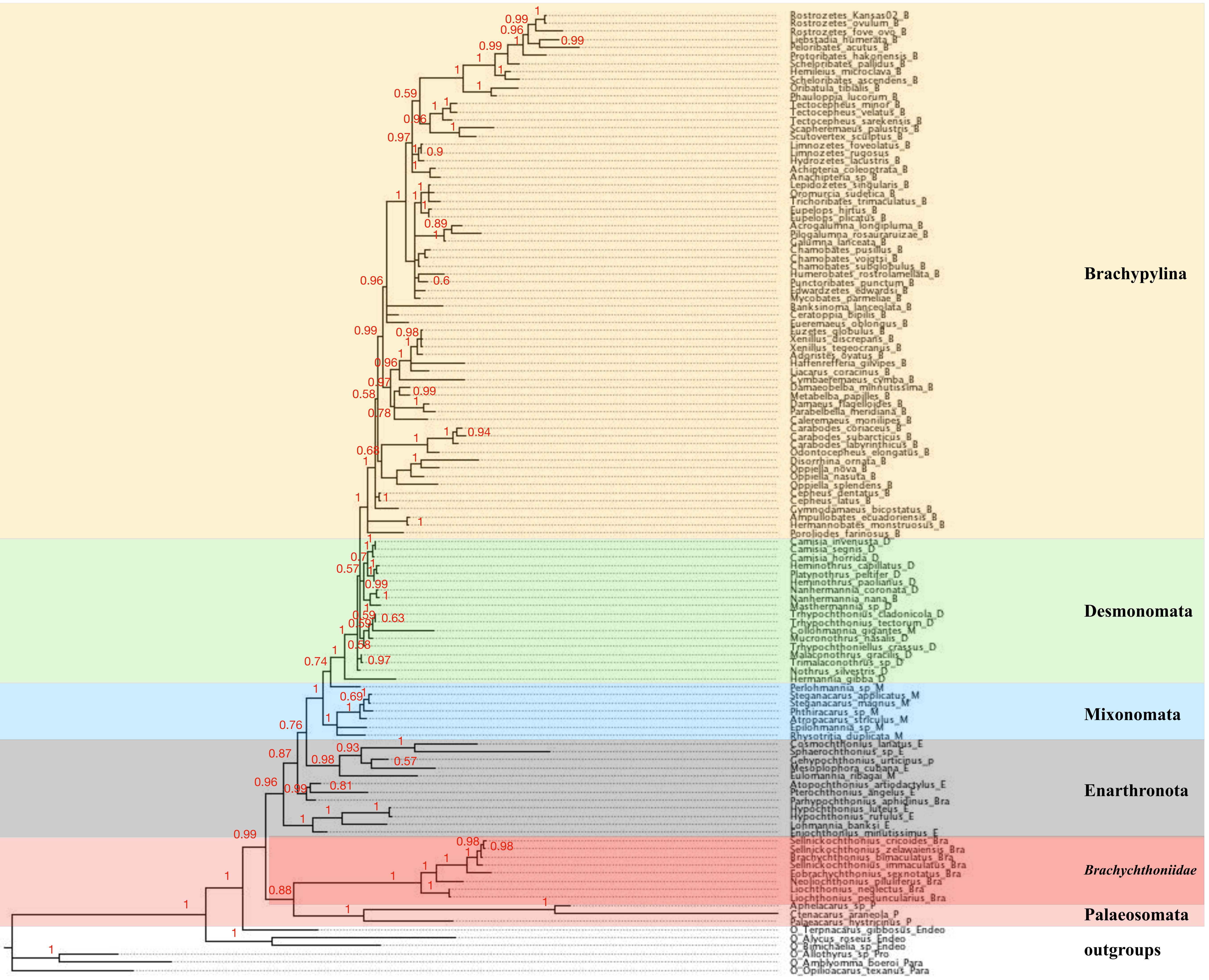


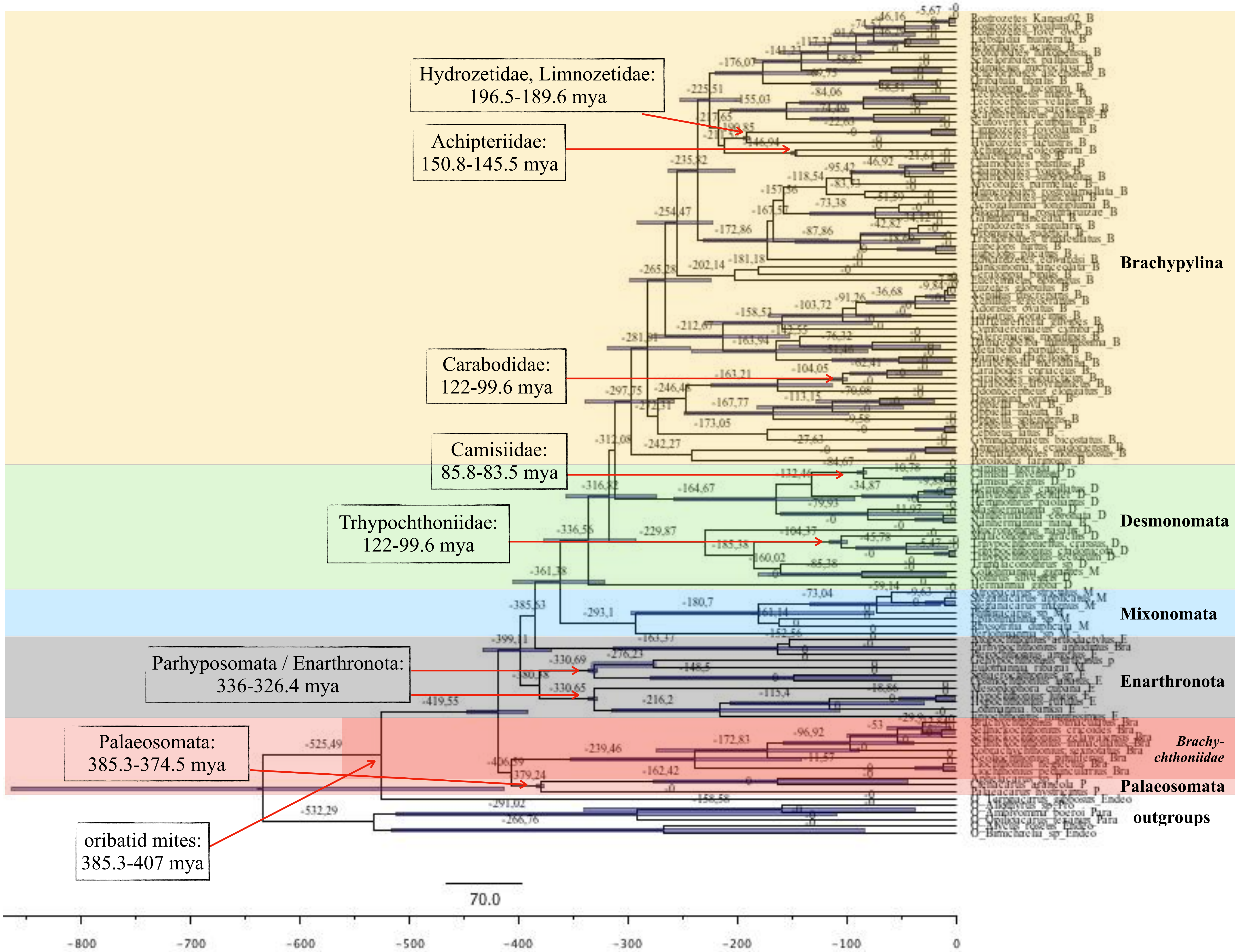
Supplementary Figures



Supplementary Figure 1. Maximum Likelihood tree of the major groups of oribatid mites with 1000 bootstrap replicates.



Supplementary Figure 2. Bayesian inference tree of the major groups of oribatid mites with posterior probabilities after 4 million generations.



Supplementary Figure 3. BEAST phylogeny of the major groups of oribatid mites with estimated divergence times based on 9 fossil constraints. Calibration points are shown in boxes, for details refer to Supplementary Table 2.

Phylogeny of oribatid mites based on 18S rDNA. Phylogenetic trees were reconstructed with a 2,096 bp alignment of 18S rDNA of 113 oribatid mite species and three non-acariformes outgroup species (3 Parasitiformes). Maximum Likelihood (ML) analysis was performed in R using the pml function of the phangorn package, using the GTR+I+G model and 1,000 bootstrap replicates (Supplementary Figure 1). Bayesian Inference (BI) was conducted in MrBayes v3.2.6 using lset parameters nst=6 rates=invgamma and 4 million generations, all other parameters were set as default (Supplementary Figure 2).

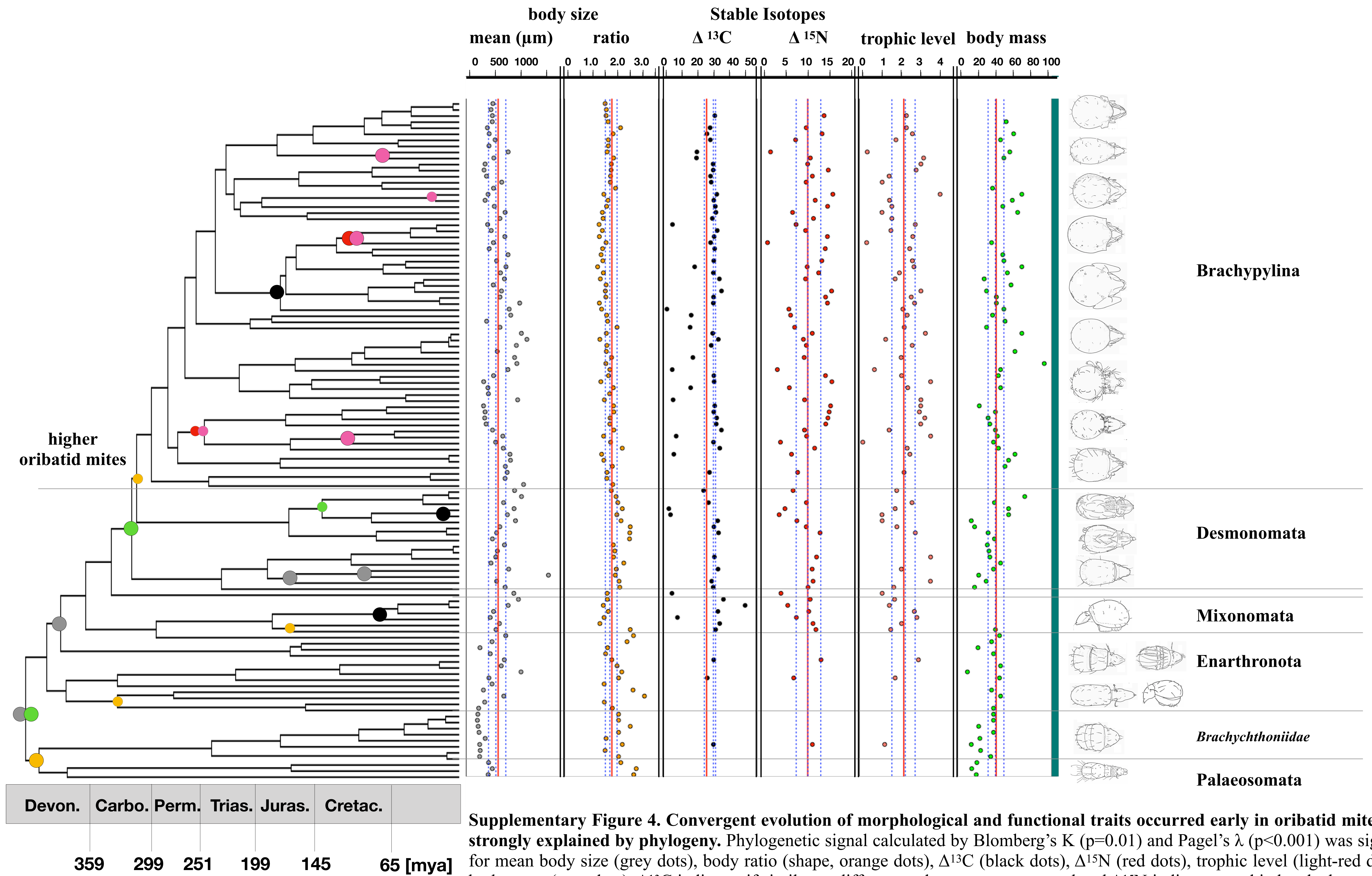
The supercohorts Brachypylina and Desmonomata (=Nothrina) included 64 and 17 species, respectively, and were recovered with 94 and 96 percent bootstrap support, respectively (posterior probability=1 for each). The tropical genus *Rostrozetes* represents Haplozetidae, because no sequences of temperate species were available, but body sizes and stable isotope data of three European species. The monotypic superfamily Collohmanniidae clustered within *Trimalacothonidae*. A similar relationship of *Collohmannia gigantea* was also recovered in Pachl et al. (2017)¹, also with equivocal node support.

The supercohort Mixonomata was recovered with 92 percent bootstrap (bs) support (posterior probability (pp)=1) and included three Phthiracaroida, one Euphthiracaroida and also included the superfamily Epilohmannioida which agrees with other studies¹⁻³. The position of Perlohmannioida differed in the ML and BI, being basal to Mixonomata in ML and ancestral to Desmonomata in the BI tree, the later is consistent with the BI analysis of a concatenated 18S+28S rDNA dataset¹ and the BEAST analysis in this study (Supplementary Figure 3).

The supercohort Endeostigmata was paraphyletic in both analyses (ML and BI), resolving into three monophyletic clades. First, the superfamily Hypochthonioida (bs=92, pp=1) including two species of the genus *Hypochthonius*, *Eniochthonius* and *Lohmannia* which is in agreement with Krantz and Proctor (2009)⁴ and with Dabert et al. (2010)⁵ but not with Pachl et al. (2017) which group Eniochthoniidae basal to Mesoplophoridae. Second, the superfamily Protoplophoridae (bs=97, pp=1), has not been recovered in this and previous molecular phylogenies, mainly due to strong differences in taxon sampling, e.g. of 14 enarthronote taxa in Pachl et al. (2017) of which only six overlap with this study. However, according to Krantz and Proctor (2009) Cosmochthoniidae and Sphaerochthoniidae are part of the superfamily Protoplophoroidea but, in contrast, here Mesoplophoridae are not part of Hypochthoniidae. The position of Eulohmanniidae is traditionally close to Mixonomata but has been recovered as sister taxon to *Gehypochthonius* in Pachl et al. (2017). Consistent with Schaefer et al. (2010) and Pachl et al. (2017), the supercohort Parhyposomata, here represented by the genera *Gehypochthonius* and *Parhypochthonius* were not recovered but clustered among other enarthronote mites. Third, the superfamily Atopochthonioida was recovered (bs=85, pp=0.99) and included the genera *Atopochthonius* and *Pterochthonius* and additionally the parhyposomatid genus *Parhypochthonius* (Parhyposomata).

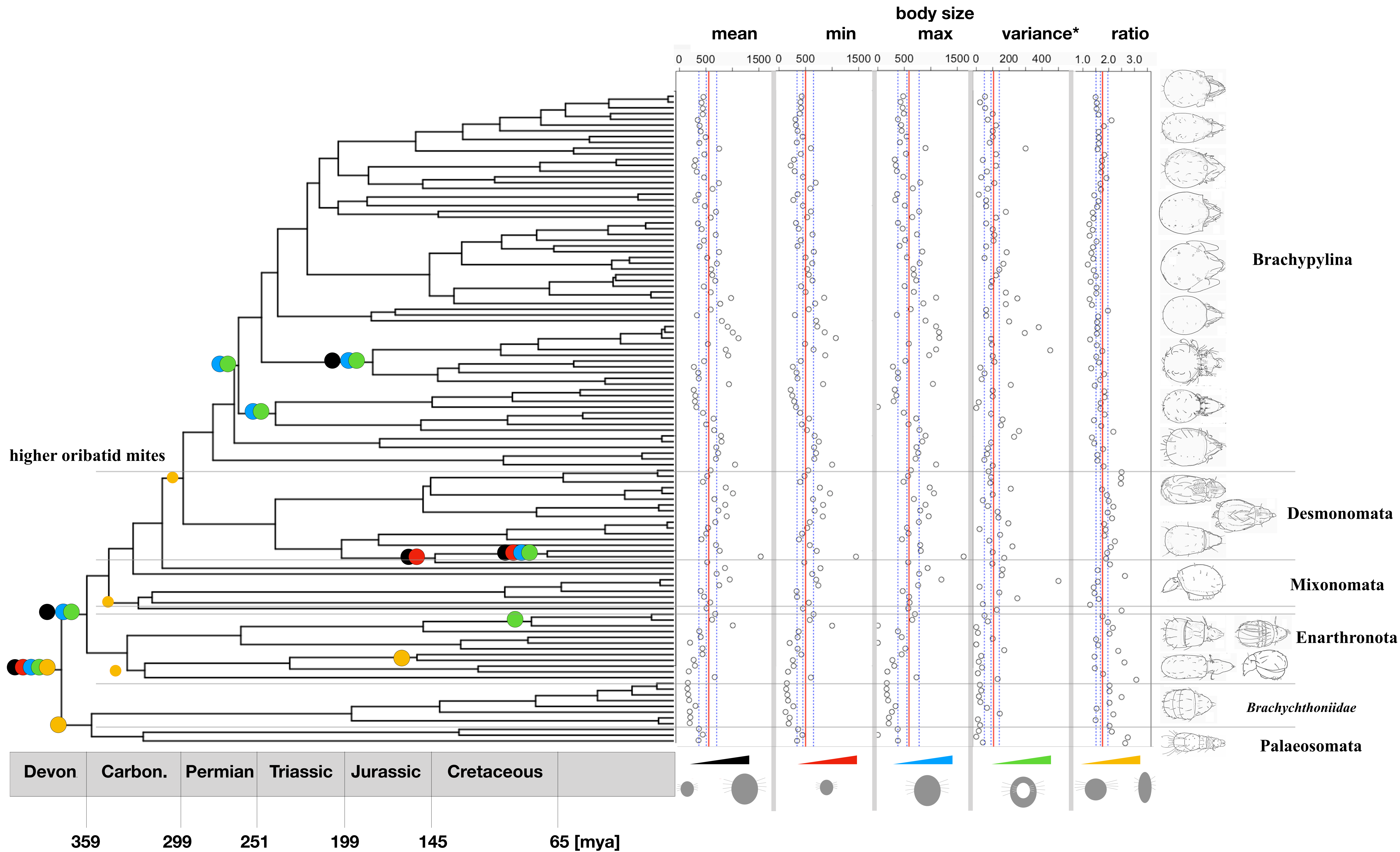
Notably, the enarthronote superfamily Brachychthonioida (bs=100, pp=1) clustered with medium support (bs=90, pp=0.88) as sister group to Palaeosomata (bs=96, pp=1) which has not been shown before. Klimov & OConnor (2013)⁶ included one Palaeosomata and two Brachychthoniidae in their study and recovered them as sister taxa as well, but not as monophyletic. This indicates that these taxa require more attention to resolve the early-derived relationships within oribatid mites and among acariformes.

The choice of outgroups and their effects on molecular clock estimates: The topology is robust and the mean molecular divergence time estimate of ~415 mya (ranging from 371-447 mya) between [Palaeosomata + Brachychthoniidae] and [Enarthronota + remaining oribatid mites] is reasonable and predating the oldest oribatid mite fossils (i.e., Enarthronota) only by about 25 my. The phylogeny among basal Acariformes and of Acari among Chelicerata is still not resolved with the molecular data currently available, irrespective of the number of genes and the methods used (e.g. secondary structures of rDNA to improve alignment quality) to improve the phylogeny on the base of Acariformes/Acari (e.g. Pepato & Klimov 2015). This study concentrated on nodes within the group of oribatid mites, and used only Acari as outgroups to infer the basal relationships and the origin of the stem group of oribatid mites. The position of Acariformes among Chelicerates is out of the scope of this study and adding more outgroups would not change the internal topology and age of internal nodes of the investigated ingroup (i.e. oribatid mites without Astigmata).



Supplementary Figure 4. Convergent evolution of morphological and functional traits occurred early in oribatid mites and is strongly explained by phylogeny. Phylogenetic signal calculated by Blomberg's K ($p=0.01$) and Pagel's λ ($p<0.001$) was significant for mean body size (grey dots), body ratio (shape, orange dots), $\Delta^{13}\text{C}$ (black dots), $\Delta^{15}\text{N}$ (red dots), trophic level (light-red dots) and body mass (green dots). $\Delta^{13}\text{C}$ indicates if similar or different carbon sources are used and $\Delta^{15}\text{N}$ indicates trophic levels due to nitrogen enrichment in higher levels.

Circles on nodes highlight nodes which traits significantly differ on decending clades (PIC), size of points indicate if the trait increased (large) or decreased (small). Trait distribution is mapped along the phylogeny next to species. Red lines in trait columns show the mean distribution, dashed blue lines show the 25% and 75% quantiles and median of the respective trait. Images of common body forms within each taxonomic group are provided on the right). Body mass was calculated after Caruso & Migliorini (2009)⁷ using body length and width of drawings in Weigmann 2006⁸.



Supplementary Figure 5. Convergent evolution of body size related traits occurred early in oribatid mites and is strongly explained by phylogeny. Phylogenetic signal calculated by Blomberg's K ($p=0.01$) and Pagel's λ ($p<0.001$) was significant for mean ($K=0.34$, $\lambda=0.78$), minimum (min: $K=0.35$, $\lambda=0.79$), maximum size (max: $K=0.037$, $\lambda=0.73$), variance ($\lambda=0.25$, $p<0.05$) and ratio (body length/width: $K=0.7$, $\lambda=0.93$). Circles on nodes indicate significant trait divergence calculated by phylogenetic independent contrast (PIC) (black=mean size, red=minimum size, blue=maximum size, green= variance, orange=ratio). Trait distribution is mapped along the phylogeny next to species. Red lines in trait columns show the mean distribution, dashed blue lines show the 25% and 75% quantiles and median of the respective trait. Images of common body forms within each taxonomic group are provided on the right.); * indicates significance only for Pagel's λ .

Supplementary Tables

Supplementary Table 1. Taxonomic overview of species included in the phylogeny and NCBI accession numbers. All six, commonly recognized, groups (infraorders) of oribatid mites were covered^{9,10}, and three acariform (Endeostigmata) and three parasitiform mites were included as outgroups for the phylogenetic tree. The majority of oribatid mite species belong to the infraorder Brachypylina (=higher oribatid mites) and represents a balanced taxon sampling across all infraorders, because of 10,000 described oribatid mites species, about ~8,000 belong to Brachypylina¹¹. When available on NCBI, more than one species per genus were included when available on NCBI (highlighted in grey). The dataset included 103 oribatid mite species of Brachypylina (n=63), Desmonomata (n=17), Mixonomata (n=9), Enarthronota (n=17, including 8 Brachychthoniidae), Parhyposomata (n=2) and Palaeosomata (n=3).

infraorder	superfamily	family	genus	species (available as 18S sequence)	Accno (NCBI)
<i>Brachypylina</i>	Achipteroidea	Achipteriidae	Achipteria	<i>Achipteria_coleoprata</i>	EF091418
		Achipteriidae	Anachipteria	<i>Anachipteria_sp</i>	JQ000048
			Lepidozetes	<i>Lepidozetes_singularis</i>	EU432193
	Ameroidea	Caleremaeidae	Caleremaeus	<i>Caleremaeus_monilipes</i>	MK630361
			Carabodidae	Carabodes	<i>Carabodes_coriaceus</i>
				<i>Carabodes_labyrinthicus</i>	KX397629
				<i>Carabodes_subarcticus</i>	EF091429
			Odontocephalus	<i>Odontocephalus_elongatus</i>	EU432200
		Tectocephidae	Tectocephus	<i>Tectocephus_minor</i>	EF093778
			<i>Tectocephus_sarekensis</i>	EF093777	
			<i>Tectocephus_velatus</i>	EF093780	
	Cepheoidea	Cepheidae	Cepheus	<i>Cepheus_dentatus</i>	MK630354
				<i>Cepheus_latus</i>	EU432206
	Ceratozetoidea	Ceratozetidae	Edwardzetes	<i>Edwardzetes_edwardsi</i>	MH198178
			Oromurcia	<i>Oromurcia_sudetica</i>	EU432194
			Trichoribates	<i>Trichoribates_trimaculatus</i>	EU432195
		Chamobatidae	Chamobates	<i>Chamobates_pusillus</i>	EU432188
				<i>Chamobates_subglobulus</i>	EU432190
				<i>Chamobates_voigtsi</i>	EU432189
		Euzetidae	Euzetes	<i>Euzetes_globulus</i>	AF022030
Humerobatidae		Humerobates	<i>Humerobates_rostromellata</i>	EU432196	

Supplementary Table 1 continued

	Mycobatidae	Mycobates	<i>Mycobates_parmeliae</i>	EU432191
	Punctoribatidae	Punctoribates	<i>Punctoribates_punctum</i>	XX123456
Cymbaeremaeoidea	Cymbaeremaeidae	Cymbaeremaeus	<i>Cymbaeremaeus_cymba</i>	EU432201
		Scapheremaeus	<i>Scapheremaeus_palustris</i>	EU433989
Damaeioidea	Belbodamaeidae	Damaeobelba	<i>Damaeobelba_minutissima</i>	MH198179
	Damaeidae	Damaeus	<i>Damaeus_flagelloides</i>	KR081608
		Metabelba	<i>Metabelba_papilles</i>	MH198180
		Parabelbella	<i>Parabelbella_meridiana</i>	KR081627
Eremaeioidea	Eremaeidae	Eueremaeus	<i>Eueremaeus_oblongus</i>	GQ864287
Galumnoidea	Galumnidae	Acrogalumna	<i>Acrogalumna_longipluma</i>	GQ864304
		Galumna	<i>Galumna_lanceata</i>	EU432197
		Pilogalumna	<i>Pilogalumna_rosauraruizae</i>	KJ423065
Gustavioidea	Liacaridae	Adoristes	<i>Adoristes_ovatus</i>	GQ864286
		Liacarus	<i>Liacarus_coracinus</i>	KR081619
	Tenuialidae	Haffenrefferia	<i>Haffenrefferia_gilvipes</i>	MK630363
Gustavioidea	Xenillidae	Xenillus	<i>Xenillus_discrepans</i>	EU432203
			<i>Xenillus_tegeocranus</i>	KR081637
	Ceratoppiidae	Ceratoppia	<i>Ceratoppia_bipilis</i>	EU432204
Gymnodamaeioidea	Gymnodaemaeidae	Gymnodamaeus	<i>Gymnodamaeus_bicostatus</i>	GQ864285
Hermannielloidea	Hermanniellidae	Ambullopates	<i>Ambullopates_ecuadoriensis</i>	KR081601
		Hermannobates	<i>Hermannobates_monstruosus</i>	KR081617
Hydrozetoidea	Hydrozetidae	Hydrozetes	<i>Hydrozetes_lacustris</i>	EU433987
	Limnozetididae	Limnozetes	<i>Limnozetes_foveolatus</i>	KX397634
			<i>Limnozetes_rugosus</i>	KX397636
Licneremaeoidea	Scutoverticidae	Scutovertex	<i>Scutovertex_sculptus</i>	GQ864305
Liodoidea	Liodidae	Poroliodes	<i>Poroliodes_farinosus</i>	EF203779
Oppioidea	Oppiidae	Disorrhina	<i>Disorrhina_ornata</i>	MH198181
		Oppiella	<i>Oppiella_nasuta</i>	MK630355
			<i>Oppiella_nova</i>	KR081626
			<i>Oppiella_splendens</i>	MK630356
	Thyrisomidae	Banksimona	<i>Banksimona_lanceolata</i>	MK630359

Supplementary Table 1 continued

	Oripodoidea	Haplozetidae	Peloribates	<i>Peloribates_acutus</i>	AB818529	
			Rostrozetes	<i>Rostrozetes_fove_ovo</i>	MK630358	
				<i>Rostrozetes_Kansas02</i>	MK630357	
				<i>Rostrozetes_ovulum</i>	HM070342	
		Hemileiidae	Hemileus	<i>Hemileius_microclava</i>	KR081616	
		Liebstadiidae	Liebstadia	<i>Liebstadia_humerata</i>	KR081620	
		Oribatulidae	Oribatula	<i>Oribatula_tibialis</i>	EU433990	
			Phauloppia	<i>Phauloppia_lucorum</i>	EU432198	
		Protoribatidae	Protoribates	<i>Protoribates_hakonensis</i>	AB818528	
		Scheloribatidae	Scheloribates	<i>Scheloribates_ascendens</i>	EU432199	
	Phenopeloidea	Phenopelopidae	Eupelops	<i>Eupelops_hirtus</i>	EF093782	
				<i>Eupelops_plicatus</i>	EF093782	
Desmonomata	Crotonioidea	Camisiidae	Camisia	<i>Camisia_horrida</i>	EU432207	
				<i>Camisia_invenusta</i>	EU432208	
				<i>Camisia_segnis</i>	EU432209	
				Heminothrus	<i>Heminothrus_capillatus</i>	GQ864288
					<i>Heminothrus_paolianus</i>	EF091423
					<i>Platynothrus_peltifer</i>	EF091422
		Nothridae	Nothrus	<i>Nothrus_silvestris</i>	EF091425	
	Hermannioidea	Hermannidae	Hermannia	<i>Hermannia_gibba</i>	EF091426	
	Nanhermannioidea	Nanhermanniidae	Masthermannia	<i>Masthermannia_sp</i>	KY922217	
				Nanhermannia	<i>Nanhermannia_coronata</i>	EF091421
					<i>Nanhermannia_nana</i>	KR081624
	Trhypochthonioidea	Malaconothridae	Malaconothrus	<i>Malaconothrus_gracilis</i>	EF091424	
Trimalaconothrus				<i>Trimalaconothrus_sp</i>	EU43221	
Trhypochthoniidae		Mucronothrus	<i>Mucronothrus_nasalis</i>	EF081299		
			Trhypochthoniellus	<i>Trhypochthoniellus_crassus</i>	EF081300	
			Trhypochthoniidae	Trhypochthonius	<i>Trhypochthonius_cladonicola</i>	JQ000047
				<i>Trhypochthonius_tectorum</i>	AF022041	
Mixonomata		Collohmannioidae	Collohmanniidae	Cohllohmanna	<i>Collohmanna_gigantes</i>	KR081604
	Epilohmannioidea	Epilohmanniidae	Epilohmannia	<i>Epilohmannia_sp</i>	EU432213	
	Eulohmannioidea	Eulohmanniidae	Eulohmannia	<i>Eulohmannia_ribagai</i>	EU432211	
	Euphtiracaroidae	Euphtiracaridae	Rhysotritia	<i>Rhysotritia_duplicata</i>	EF091417	
	Perlohmannioidea	Perlohmanniidae	Perlohmannia	<i>Perlohmannia_sp</i>	EU432212	
	Phthiracaroidae	Phthiracaridae	Phthiracarus	<i>Phthiracarus_sp</i>	KR081629	
				Atropacarus	<i>Atropacarus_striculus</i>	EF091416
				Steganacarus	<i>Steganacarus_applicatus</i>	GQ864301
			<i>Steganacarus_magnus</i>	AF022040		

Supplementary Table 1 continued

<i>Enarthronota</i>	Atopochthonoidea	Atopochthoniidae	Atopochthonius	<i>Atopochthonius_artiodactylus</i>	EU432216
		Pterochthoniidae	Pterochthonius	<i>Pterochthonius_angelus</i>	EU432214
	Brachychthonoidea	Brachychthoniidae	Brachychthonius	<i>Brachychthonius_bimaculatus</i>	MK630360
			Eobrachychthonius	<i>Eobrachychthonius_sexnotatus</i>	MK630362
		Liochthonius		<i>Liochthonius_neglectus</i>	MK630364
				<i>Liochthonius_peduncularius</i>	MK630365
		Neoliochthonius		<i>Neoliochthonius_piluliferus</i>	MK630366
		Sellnickochthonius		<i>Sellnickochthonius_cricoides</i>	MK630367
			<i>Sellnickochthonius_immaculatus</i>	MN065455	
			<i>Sellnickochthonius_zelawaiensis</i>	MH198174	
	Cosmochthonoidea	Cosmochthoniidae	Cosmochthonius	<i>Cosmochthonius_lanatus</i>	JN585919
		Sphaerochthoniidae	Sphaerochthonius	<i>Sphaerochthonius_sp</i>	JN585916
	Hypochthonoidea	Eniochtoniidae	Eniochthonius	<i>Eniochthonius_minutissimus</i>	EF091428
		Hypochthoniidae	Hypochthonius	<i>Hypochthonius_luteus</i>	EU152475
<i>Hypochthonius_rufulus</i>				EF091427	
Lohmannioidea	Lohmanniidae	Lohmannia	<i>Lohmannia_banksi_E</i>	AF022036	
Mesoplophoroidea	Mesoplophoridae	Mesoplophora	<i>Mesoplophora_cubana</i>	EU432217	
<i>Parhyposomata</i>	Parhypochthonoidea	Gehypochthoniidae	Gehypochthonius	<i>Gehypochthonius_urticinus</i>	EU433994
		Parhypochthoniidae	Parhypochthonius	<i>Parhypochthonius_aphidinus</i>	EU432215
<i>Palaeosomata</i>	Ctenacaroida	Aphelacaridae	Aphelacarus	<i>Aphelacarus_sp</i>	DQ648879
		Ctenacaridae	Ctenacarus	<i>Ctenacarus_araneola</i>	EU433991
	Palaeacaroida	Palaeacaridae	Palaeacarus	<i>Palaeacarus_hystricinus</i>	EF204472
<i>Acariformes</i> <i>outgroups</i>	Endeostigmata	Alycidae	Alycus	<i>Alycus_roseus</i>	GQ864294
			Bimichaelia	<i>Bimichaelia_sp</i>	GQ864295
		Terpnacaridae	Terpnacarus	<i>Terpnacarus_gibbosus</i>	AY620904
<i>Parasitiformes</i> <i>outgroups</i>	Holothyrida	Holothyroidea	Allothyrus	<i>Allothyrus_sp</i>	AF018655
	Ixodidae	Amblyomminae	Amblyomma	<i>Amblyomma_boeroi</i>	FJ464420
	Opilioacariformes	Opilioacarida	Opilioacarus	<i>Opilioacarus_texasus</i>	AF115375

Supplementary Table 2. Taxonomic assignment, geological age of fossil oribatid mites, and prior settings used in the molecular clock analysis.

	family	species (available as 18S sequence)	fossil specimen	geological epoch	time (mya)	reference	exponential calibration prior in BEAST
Brachypylina	Achipteriidae	<i>Achipteria_coleoprata</i> <i>Anachipteria_sp</i>	<i>Achipteria (?) obscura</i>	Tithonian	150.8-145.5	Krivolutsky & Krassilov 1977 ¹²	mean:2.0 offset:145.0
	Carabodidae	<i>Carabodes_coriaceus</i> <i>Carabodes_labyrinthicus</i> <i>Carabodes_subarcticus</i>	<i>Cretaceobodes martinezae</i>	Albian	122-99.6	Arillo et al. 2008 ¹³	mean:7.0 offset:99.0
	Hydrozetidae Limnozeteidae	<i>Hydrozetes_lacustris</i> <i>Limnozetes_foveolatus</i> <i>Limnozetes_rugosus</i>	<i>Hydrozetes sp.</i>	Sinemurian	196.5-189.6	Sivhead & Wallwork 1978 ¹⁴	mean:2.0 offset:189.0
Desmonomata	Camisiidae	<i>Camisia_horrida</i> <i>Camisia_invenusta</i> <i>Camisia_segnis</i>	<i>Eocamisia sukatshevae</i>	Santonian	85.8-83.5	Bulanova-Zachvatkina 1974 ¹⁵	mean:3.0 offset:82.0
	Trhypochthoniidae	<i>Mucronothrus_nasalis</i> <i>Trhypochthoniellus_crassus</i> <i>Trhypochthonius_cladonicola</i> <i>Trhypochthonius_tectorum</i>	<i>Trhypochthonius lopezvallei</i>	Albian	122-99.6	Arillo et al. 2012 ¹⁶	mean:7.0 offset:99.0
Parhyposomata / Enarthronota	Cosmochthoniidae	<i>Cosmochthonius_lanatus</i>	<i>Carbochthonius antrimensis</i>	Brigantian	336-326.4	Subías & Arillo 2002 ¹⁷	mean:3.0 offset:328.0
	Gehypochthoniidae	<i>Gehypochthonius_urticinus</i>	<i>Gehypochthonimimus hibernicus</i>				
	Sphaerochthoniidae Eulohmanniidae	<i>Sphaerochthonius_sp</i> <i>Eulohmannia_ribagai</i>					

Supplementary Table 2 continued

Enarthro-nota	Hypochthoniidae	<i>Hypochthonius_luteus</i> <i>Hypochthonius_rufulus</i>	<i>Paleohypochthonius</i> <i>jerami</i>				
	Mesoplophoridae	<i>Mesoplophora_cubana</i>	<i>Archaeoplophora_bella</i>	Brigantian	336-326.4	Subías & Arillo 2002 ¹⁷	mean:3.0 offset:328.0
	Eniochtoniidae	<i>Eniochthonius_minutissimus</i>					
	Lohmanniidae	<i>Lohmannia_banksi</i>					
Palaeoso-mata	Aphelacaridae	<i>Aphelacarus_sp</i>	<i>Monoaphelacarus</i> <i>carboniferus</i>				
	Ctenacaridae	<i>Ctenacarus_araneola</i>	<i>Ctenacaronychus</i> <i>nortoni</i>	lower Frasnian	385.3-374.5	Subías & Arillo 2002 ¹⁷ , Norton et al. 1988 ¹⁸	mean:3.0 offset:377.0
	Palaeacaridae	<i>Palaeacarus_hystricinus</i>	<i>Paleoctenacarus</i> <i>simmsi</i>				
oribatid mites				Emsian	407-397.5		
				Givetian	385.3-391.8	Shear et al. 1984 ¹⁹	mean:5.0 offset:384.0

Supplementary Table 3. Fossil data that decreased the likelihoods and posterior probabilities and were not used in the final molecular clock analyses. Fossil dates either strongly underestimated the age of species or could not be applied because too few specimens of the genus or family were present in the dataset.

	family	species (available as 18S sequence)	fossil specimen	geological epoch	time (mya)	reference	
Brachyphyllina	Cepheoidea	<i>Cepheus dentatus</i>	<i>Eupterotegaeus bitranslamellatus</i>	Albian	122-99.6	Arillo et al. 2012 ¹⁶	
		<i>Cepheus latus</i>	<i>Ommatocepheus nortoni</i>				
	Cymbaeremaeidae	<i>Cymbaeremaeus cymba</i>	<i>Ametroproctus valeiae</i>	Albian	122-99.6	Arillo et al. 2012 ¹⁶	
		<i>Scapheremaeus palustris</i>	<i>Scapheremaeus</i>	late Eocene	42-37	O'Dowd et al. 1991 ²⁰	
	Eremaeidae	<i>Eueremaeus oblongus</i>	<i>Eremaeus denaius</i>	Oligocene/Miocene	25	Woolley 1971 ²¹	
	Galumnidae	<i>Acrogalumna longipluma</i>		middle Eocene	42-49	O'Dowd et al. 1991 ²⁰	
		<i>Galumna lanceata</i>					
		<i>Pilogalumna rosauraruizae</i>					
	Oppiidae	<i>Dissorhina ornata</i>		<i>Dissorhina paleokrasica</i> n. sp., <i>D. nuda</i> n. sp.	late Pliocene	3.8, 2.7	Mico 2015 ²²
			<i>Oppiella nasuta</i> <i>Oppiella nova</i> <i>Oppiella splendens</i>	<i>Oppia hurdi</i>	Oligocene/Miocene	25	Woolley 1971 ²¹
Scutoverticidae		<i>Scutovertex sculptus</i>	<i>Hypovertex hispanicus</i> (<i>Scutoverticidae</i>)	Albian	122-99.6	Arillo et al. 2012 ¹⁶	
	<i>Phauloppia lucorum</i>	<i>Phauloppia</i>	middle Eocene	42-49	O'Dowd et al. 1991 ²⁰		
Des.	Hermanniiidae	<i>Hermannia gibba</i>	<i>Hermannia sellnicki</i> n. sp.	Lutetian	~44	Norton 2006 ²³	
Mixonomata	Collohmanniidae	<i>Collohmanna gigantes</i>	<i>Collohmanna schusteri</i> n. sp.	Lutetian	~44	Norton 2006 ²³	
	Phthiracaroida	<i>Phthiracarus</i> sp		Carboniferous trace fossils (coprolites)	359.2-299	Labandeira et al. 1997 ²⁴	
		<i>Atropacarus striculus</i>					
		<i>Steganacarus applicatus</i>					
		<i>Steganacarus magnus</i>					
Euphtiracarid.	<i>Rhysotritia duplicata</i>						

Supplementary Table 4. Summary of morphological traits (body size and body shape) used in this study. Minimum and maximum size values were collected from Weigman (2006)⁸ if not stated otherwise in the reference column. Body shape (ratio of length/width) was obtained by measuring drawings of species from literature. If sequences were not assigned to species (i.e., sp.) or the exact species was not available as drawing, measurements were taken from a congeneric species as listed in the column references. Sequences of the species of the genus *Haplozetes* were not available at GenBank and were replaced by species of the genus *Rostrozetes*.

infraorder	species (available 18S sequence)	body size [μm]				body form	reference
		mean	minimum	maximum	variance	(ratio= length/width)	
Brachypylina (higher oribatid mites)	<i>Achipteria_coleoptrata</i>	590	530	650	120	1.41	
	<i>Anachipteria_sp</i>	690	600	780	180	1.38	<i>Anachipteria nitens</i>
	<i>Lepidozetes_singularis</i>	460	415	505	90	1.53	
	<i>Caleremaeus_monilipes</i>	355	330	380	50	1.82	
	<i>Carabodes_coriaceus</i>	645	565	725	160	1.43	
	<i>Carabodes_labyrinthicus</i>	505	430	580	150	1.71	
	<i>Carabodes_subarcticus</i>	445	400	490	90	1.84	
	<i>Odontocephus_elongatus</i>	655	525	785	260	2.18	
	<i>Tectocephus_minor</i>	280	220	340	120	1.73	Dogan & Ayyildiz 2000 ²⁵
	<i>Tectocephus_sarekensis</i>	327.5	295	360	65	1.71	Knülle 1953 ²⁶
	<i>Tectocephus_velatus</i>	300	280	320	40	1.74	Dogan & Ayyildiz 2000 ²⁵
	<i>Cepheus_dentatus</i>	795	750	840	90	1.44	
	<i>Cepheus_latus</i>	785	670	900	230	1.35	
	<i>Edwardzetes_edwardsi</i>	770	680	860	180	1.35	
	<i>Oromurcia_sudetica</i>	677.5	630	725	95	1.3	
	<i>Trichoribates_trimaculatus</i>	620	560	680	120	1.5	<i>Trichoribates novus</i>
	<i>Chamobates_pusillus</i>	420	370	470	100	1.38	
	<i>Chamobates_subglobulus</i>	685	630	740	110	1.26	
	<i>Chamobates_voigtsi</i>	350	320	380	60	1.25	
	<i>Euzetes_globulus</i>	1115	1070	1160	90	1.28	
	<i>Humerobates_rostromellata</i>	747.5	655	840	185	1.33	
	<i>Mycobates_parmeliae</i>	462.5	410	515	105	1.53	
	<i>Punctoribates_punctum</i>	380	350	410	60	1.39	
	<i>Cymbaeremaeus_cymba</i>	745	690	800	110	1.68	
	<i>Scapheremaeus_palustris</i>	465	450	480	30	1.91	
	<i>Damaeobelba_minutissima</i>	272.5	260	285	25	1.32	
	<i>Damaeus_flagelloides</i>	935	830	1040	210	1.46	

Supplementary Table 4 continued

<i>Metabelba_papilles</i>	465	410	520	110	1.63	
<i>Parabelbella_meridiana</i>	364.5	348	381	33	1.67	
<i>Eueremaeus_oblongus</i>	590	560	620	60	1.97	<i>Eueremaeus silvestris</i>
<i>Acrogalumna_longipluma</i>	707.5	625	790	165	1.19	
<i>Galumna_lanceata</i>	600	530	670	140	1.42	
<i>Pilogalumna_rosauraruizae</i>	525	500	550	50	1.4	<i>Pilogalumna crassiclava</i>
<i>Adoristes_ovatus</i>	537.5	490	585	95	1.54	
<i>Liacarus_coracinus</i>	875	650	1100	450	1.75	
<i>Haffenrefferia_gilvipes</i>	920	870	970	100	1.52	
<i>Xenillus_discrepans</i>	1007.5	860	1155	295	1.54	
<i>Xenillus_tegeocranus</i>	910	720	1100	380	1.56	
<i>Ceratoppia_bipilis</i>	800	700	900	200	1.55	
<i>Gymnodamaeus_bicostatus</i>	695	660	730	70	1.78	
<i>Ampullobates_ecuadoriensis</i>	688.5	664	713	49	1.56	
<i>Hermannobates_monstruosus</i>	730	697	763	66	1.56	
<i>Hydrozetes_lacustris</i>	480	450	510	60	1.55	
<i>Limnozetes_foveolatus</i>	300	270	330	60	1.61	<i>Limnozetes ciliatus</i>
<i>Limnozetes_rugosus</i>	357.5	350	365	15	1.44	
<i>Scutovertex_sculptus</i>	625	590	660	70	1.7	
<i>Poroliodes_farinosus</i>	1050	1000	1100	100	1.8	
<i>Disorrhina_ornata</i>	300	250	350	100	1.84	
<i>Oppiella_nasuta</i>	292.5	285	300	15	1.69	
<i>Oppiella_nova</i>	270	220	320	100	1.83	
<i>Oppiella_splendens</i>	320	320			1.68	<i>Oppiella escotata</i>
<i>Banksinoma_lanceolata</i>	330	300	360	60	1.59	
<i>Peloribates_acutus</i>	440	390	490	100	1.61	<i>Peloribates europaeus</i>
<i>Rostrozetes_fove_ovo</i>	442	415	469	54	1.53	<i>Haplozetes vindobonensis</i>
<i>Rostrozetes_Kansas02</i>	416.5	405	428	23	1.54	<i>Haplozetes elegans</i>
<i>Rostrozetes_ovulum</i>	452.5	426	479	53	1.49	<i>Haplozetes tenuifusus</i>
<i>Hemileius_microclava</i>	376	335	417	82	1.62	
<i>Liebstadia_humerata</i>	345	310	380	70	2.11	
<i>Oribatula_tibialis</i>	470	410	530	120	1.83	
<i>Phauloppia_lucorum</i>	750	600	900	300	1.57	

Supplementary Table 4 continued

Brachy- pylina	<i>Protoribates_hakonensis</i>	380	320	440	120	1.81	
	<i>Scheloribates_ascendens</i>	495	445	545	100	1.62	
	<i>Eupelops_hirtus</i>	975	850	1100	250	1.26	
	<i>Eupelops_plicatus</i>	590	500	680	180	1.52	
Desmonomata	<i>Camisia_horrida</i>	892.5	825	960	135	2.13	
	<i>Camisia_invenusta</i>	735	670	800	130	1.96	
	<i>Camisia_segnis</i>	865	830	900	70	2.18	
	<i>Heminothrus_capillatus</i>	1010	960	1060	100	1.93	<i>Platynothrus capillatus</i>
	<i>Heminothrus_paolianus</i>	660	640	680	40	2	<i>Heminothrus paolianus longisetosus</i>
	<i>Platynothrus_peltifer</i>	875	770	980	210	1.74	
	<i>Nothrus_silvestris</i>	760	710	810	100	1.94	
	<i>Hermannia_gibba</i>	860	780	940	160	1.57	
	<i>Masthermannia_sp</i>	442.5	400	485	85	2.47	<i>Masthermannia mammillaris</i>
	<i>Nanhermannia_coronata</i>	525	480	570	90	2.48	
	<i>Nanhermannia_nana</i>	587.5	550	625	75	2.5	
	<i>Malacothonrus_gracilis</i>	415	375	455	80	2.24	<i>Malacothonrus monodactylus</i>
	<i>Trimalaconothrus_sp</i>	525	476	574	98	2.05	mean: <i>T. glaber</i> , <i>T. angulatus</i> , <i>T. maior</i> , <i>T. foveolatus</i> , <i>T. tardus</i> , <i>T. sculptus</i> , <i>T. vietsi</i>
	<i>Mucronothrus_nasalis</i>	690	580	800	220	2.08	
<i>Trhypochthoniellus_crassus</i>	507.5	435	580	145	1.83	mean: <i>T. longiseta</i> , <i>T. setosa</i>	
<i>Trhypochthonius_cladonicola</i>	540	530	550	20	1.88		
<i>Trhypochthonius_tectorum</i>	677.5	580	775	195	1.82		
Mixonomata	<i>Collohmanna_gigantes</i>	1535	1450	1620	170	1.9	
	<i>Epilohmannia_sp</i>	512.5	450	575	125	2.5	
	<i>Eulomannia_ribagai</i>	665	600	730	130	3.07	
	<i>Rhysotritia_duplicata</i>	580	560	600	40	1.28	
	<i>Perlohmanna_sp</i>	702.5	625	780	155	2.63	<i>Perlohmanna nasuta</i>
	<i>Phthiracarus_sp</i>	400	330	470	140	1.45	<i>Phthiracarus anonymys</i> (Grandjean 1934) ²⁷
	<i>Atropacarus_striculus</i>	465	340	590	250	1.62	Kamill & Baker 1980 ²⁸
	<i>Steganacarus_applicatus</i>	750	740	760	20	1.42	Bernini & Avanzati 1989 ²⁹
	<i>Steganacarus_magnus</i>	950	700	1200	500	1.58	<i>Steganacarus magna forma magnus</i> (Bernini & Avanzati 1988) ³⁰

Supplementary Table 4 continued

Enarthronota	<i>Atopochthonius_artiodactylus</i>	200	200			1.59	
	<i>Pterochthonius_angelus</i>	400	350	450	100	1.51	
	<i>Brachychthonius_bimaculatus</i>	165	157	173	16	2.5	
	<i>Eobrachychthonius_sexnotatus</i>	302.5	270	335	65	1.53	<i>Eobrachychthonius latior</i>
	<i>Liochthonius_neglectus</i>	193.5	182	205	23	2.03	
	<i>Liochthonius_peduncularius</i>	205	200	210	10	1.49	
	<i>Neoliochthonius_piluliferus</i>	194.5	123	266	143	2.18	
	<i>Sellnickochthonius_cricoides</i>	150	135	165	30	2.03	
	<i>Sellnickochthonius_immaculatus</i>	182.5	170	195	25	2.03	
	<i>Sellnickochthonius_zelawaiensis</i>	159	148	170	22	2.03	
	<i>Cosmochthonius_lanatus</i>	175	170	180	10	1.78	
	<i>Sphaerochthonius_sp</i>	292.5	275	310	35	1.46	<i>Sphaerochchthonius splendidus</i>
	<i>Eniochthonius_minutissimus</i>	375	370	380	10	2.03	
	<i>Hypochthonius_luteus</i>	615	580	650	70	1.97	
	<i>Hypochthonius_rufulus</i>	675	650	700	50	1.76	
	<i>Lohmannia_banksi_E</i>	1000	1000			2.16	<i>Lohmannia paradoxa</i>
<i>Mesoplophora_cubana</i>	435	420	450	30	1.45	<i>Mesoplophora aficana</i> (Els 1966) ³¹	
Parhypo- somata	<i>Gehypochthonius_urticinus</i>	267.5	260	275	15	2.61	<i>Gehypochthonius rhadamanthus</i>
	<i>Parhypochthonius_aphidinus</i>	435	350	520	170	2.37	
Palaeo- somata	<i>Aphelacarus_sp</i>	440	440			2.74	<i>Aphelacarus acarinus</i>
	<i>Ctenacarus_araneola</i>	367.5	360	375	15	2.12	
	<i>Palaeacarus_hystricinus</i>	360	340	380	40	2.64	Schatz 2004 ³²

Supplementary Table 5. Trophic traits used in this study and list of taxa for which stable isotope data were available. Data are from four 4 habitats³³⁻³⁸ to cover a wide taxon sampling (in total 70 species) which covers most species from the morphological trait dataset and phylogeny (in total 103 species). Trophic levels were assigned based on ¹⁵N values within communities (to account for site specific differences of the baseline values of ¹³C and ¹⁵N, which can geographically strongly differ) and categorized into four, very general functional groups. Within each trophic level, species were hierarchical assigned to their ¹⁵N position within their community. For better visualisation in the graph showing the distribution of trophic traits along the phylogeny, we added the factor 28 to ¹³C and the factor 8 to ¹⁵N values to obtain only positive values, which are also listed in this table. Species are listed from lowest to highest trophic levels and include Brachypylyna (n=49), Desmonomata (n=12), Mixonomata (n=6) and Enarthronota (n=3, including a single measurement for Brachychthoniidae).

infraorder	species	¹³ C	¹⁵ N	¹³ C + 28	¹⁵ N + 8	trophic level	functional group
Brachypylyna	<i>Carabodes_labyrinthicus</i>	1.78	-4.34	29.78	3.66	0	specialist
Brachypylyna	<i>Mycobates_parmeliae</i>	-0.03	-7.31	27.97	0.69	0.2	
Brachypylyna	<i>Phauloppia_lucorum</i>	-8.66	-6.62	19.34	1.38	0.23	
Brachypylyna	<i>Cymbaeremaeus_cymba</i>	-24.23	-5.06	3.77	2.94	0.6	
Brachypylyna	<i>Anachipteria_sp</i>	3.34	-1.57	31.34	6.43	0.99	
Desmonomata	<i>Camisia_invenusta</i>	-25.31	-4.64	2.69	3.36	0.99	
Desmonomata	<i>Camisia_horrida</i>	4.46	-0.57	32.46	7.43	1	primary decomposer
Desmonomata	<i>Hermannia_gibba</i>	-24.4	-4.23	3.6	3.77	1	
Brachypylyna	<i>Scutovertex_sculptus</i>	0.38	1.55	28.38	9.55	1	
Enarthronota	<i>Neoliochthonius_piluliferus</i>	1.75	2.98	29.75	10.98	1.13	
Brachypylyna	<i>Euzetes_globulus</i>	4.87	0.92	32.87	8.92	1.18	
Brachypylyna	<i>Carabodes_subarcticus</i>	6.8	1.17	34.8	9.17	1.36	
Brachypylyna	<i>Tectocephus_sarekensis</i>	-0.08	2.98	27.92	10.98	1.36	
Mixonomata	<i>Steganacarus_applicatus</i>	21.75	-2.67	49.75	5.33	1.37	
Brachypylyna	<i>Limnozetes_foveolatus</i>	1.98	3.63	29.98	11.63	1.38	
Mixonomata	<i>Epilohmannia_sp</i>	3.15	3.79	31.15	11.79	1.44	
Brachypylyna	<i>Chamobates_pusillus</i>	4.17	1.42	32.17	9.42	1.45	
Brachypylyna	<i>Achipteria_coleoprata</i>	1.07	3.23	29.07	11.23	1.5	
Brachypylyna	<i>Hydrozetes_lacustris</i>	2.95	6.47	30.95	14.47	1.5	

Supplementary Table 5 continued

Desmonomata	<i>Malaconothrus_gracilis</i>	1.62	1.98	29.62	9.98	1.6	primary decomposer	
Mixonomata	<i>Steganacarus_magnus</i>	8.06	2.47	36.06	10.47	1.64		
Desmonomata	<i>Camisia_segnis</i>	-26.37	-3.31	1.63	4.69	1.67		
Enarthronota	<i>Eniochthonius_minutissimus</i>	-2.09	-1.31	25.91	6.69	1.67		
Brachypylylina	<i>Oromurcia_sudetica</i>	5.52	1.4	33.52	9.4	1.67		
Brachypylylina	<i>Scheloribates_ascendens</i>	-0.18	-0.87	27.82	7.13	1.71		
Desmonomata	<i>Platynothrus_peltifer</i>	-4.46	-1.45	23.54	6.55	1.75		
Desmonomata	<i>Nanhermannia_nana</i>	2.08	1.57	30.08	9.57	1.76		
Brachypylylina	<i>Galumna_lanceata</i>	1.72	4.42	29.72	12.42	1.89		
Brachypylylina	<i>Liacarus_coracinus</i>	-11.19	1.09	16.81	9.09	1.98		
Desmonomata	<i>Nothrus_silvestris</i>	4.67	2.92	32.67	10.92	1.99		
Brachypylylina	<i>Metabelba_papilles</i>	2.05	5.98	30.05	13.98	2		secondary decomposer
Mixonomata	<i>Rhysotritia_duplicata</i>	5.71	3.12	33.71	11.12	2		
Brachypylylina	<i>Edwardzetes_edwardsi</i>	-27.56	-2.41	0.44	5.59	2.07		
Brachypylylina	<i>Hermannobates_monstruosus</i>	-0.72	-0.41	27.28	7.59	2.13		
Brachypylylina	<i>Eueremaeus_oblongus</i>	-12.93	-1.1	15.07	6.9	2.14		
Brachypylylina	<i>Liebstadia_humerata</i>	-0.35	1.57	27.65	9.57	2.25		
Brachypylylina	<i>Oribatula_tibialis</i>	-0.35	1.57	27.65	9.57	2.25		
Brachypylylina	<i>Rostrozetes_fove_ovo</i>	2.61	5.66	30.61	13.66	2.25		
Brachypylylina	<i>Ceratoppia_bipilis</i>	-12.25	-2.02	15.75	5.98	2.29		
Brachypylylina	<i>Odontocephelus_elongatus</i>	5.75	3.49	33.75	11.49	2.29		
Brachypylylina	<i>Caleremaeus_monilipes</i>	-12.6	-2.3	15.4	5.7	2.32		
Brachypylylina	<i>Punctoribates_punctum</i>	2.54	5.93	30.54	13.93	2.42		
Brachypylylina	<i>Cepheus_latus</i>	-23.22	-1.81	4.78	6.19	2.43		
Brachypylylina	<i>Eupelops_plicatus</i>	1.83	6	29.83	14	2.5		
Desmonomata	<i>Heminothrus_paolianus</i>	-1.35	1.61	26.65	9.61	2.54		
Brachypylylina	<i>Xenillus_tegeocranus</i>	0.38	1.6	28.38	9.6	2.55		
Brachypylylina	<i>Pilogalumna_rosauraruizae</i>	1.95	5.14	29.95	13.14	2.56		
Brachypylylina	<i>Protoribates_hakonensis</i>	-2.31	5.2	25.69	13.2	2.56		
Brachypylylina	<i>Chamobates_subglobulus</i>	2.2	6.43	30.2	14.43	2.58		
Brachypylylina	<i>Acrogalumna_longipluma</i>	-10.09	1.78	17.91	9.78	2.64		
Mixonomata	<i>Atropacarus_striculus</i>	4.6	2.15	32.6	10.15	2.66		
Brachypylylina	<i>Eupelops_hirtus</i>	1.68	6.44	29.68	14.44	2.67		

Supplementary Table 5 continued

Brachypylylina	<i>Chamobates_voigtsi</i>	-24.07	-0.76	3.93	7.24	2.71	secondary decomposer
Desmonomata	<i>Nanhermannia_coronata</i>	5	4.74	33	12.74	2.71	
Brachypylylina	<i>Tectocephus_minor</i>	1.62	6.62	29.62	14.62	2.75	
Mixonomata	<i>Phthiracarus_sp</i>	-20.93	-0.69	7.07	7.31	2.79	
Enarthronota	<i>Hypochthonius_rufulus</i>	1.92	4.96	29.92	12.96	2.87	
Brachypylylina	<i>Disorrhina_ornata</i>	1.93	6.83	29.93	14.83	2.92	
Brachypylylina	<i>Oppiella_nova</i>	2.57	7.15	30.57	15.15	2.99	
Brachypylylina	<i>Oppiella_splendens</i>	3.53	6.04	31.53	14.04	2.99	
Brachypylylina	<i>Damaeus_flagelloides</i>	-23.68	1.18	4.32	9.18	3	predator/scavenger
Brachypylylina	<i>Tectocephus_velatus</i>	1.55	1.9	29.55	9.9	3	
Brachypylylina	<i>Trichoribates_trimaculatus</i>	6.85	7.39	34.85	15.39	3	
Brachypylylina	<i>Oppiella_nasuta</i>	3.81	6.54	31.81	14.54	3.2	
Brachypylylina	<i>Xenillus_discrepans</i>	1.29	2.95	29.29	10.95	3.23	
Desmonomata	<i>Trimalaconothrus_sp</i>	0.67	3.14	28.67	11.14	3.49	
Brachypylylina	<i>Carabodes_coriaceus</i>	-21.74	1.65	6.26	9.65	3.5	
Brachypylylina	<i>Damaeobelba_minutissima</i>	2.2	7.45	30.2	15.45	3.5	
Desmonomata	<i>Trhypochthoniellus_crassus</i>	2.38	3.94	30.38	11.94	3.5	
Brachypylylina	<i>Limnozetes_rugosus</i>	3.89	7.67	31.89	15.67	3.99	

Supplementary References

1. Pachtl, P., Lindl, A. C., Krause, A., Scheu, S., Schaefer, I., Maraun, M. The tropics as ancient cradle of oribatid mite diversity. *Acarologia* 57, 309-322 (2017).
2. Schaefer, I., Norton, R. A., Scheu, S. & Maraun, M. Arthropod colonization of land – linking molecules and fossils in oribatid mites (Acari, Oribatida). *Mol. Phylogenet. Evol.* 57, 113-121 (2010).
3. Pepato, A. R. & Klimov, P. B. Origin and higher-level diversification of acariform mites – evidence from nuclear ribosomal genes, extensive taxon sampling, and secondary structure alignment. *BMC Evol. Biol.* 15, 178 (2015).
4. Krantz, G. W. & Proctor, D. E. in *A manual of Acarology*. eds Krantz GW and Walter DE (3rd edition, Texas Tech University Press 2009); chapter 15, pp. 430 ff.
5. Dabert, M., Witalinski, W., Kazmierski, A., Olszanowski, Z. & Dabert, J. Molecular phylogeny of acariform mites (Acari, Arachnida): strong conflict between phylogenetic signal and long-branch attraction artifacts. *Mol. Phylogenet. Evol.* 56, 222-241 (2010)
6. Klimov, P. B. & OConnor, B. Is permanent parasitism reversible? - critical evidence from early evolution of house dust mites. *Syst. Biol.* 62, 411-423 (2013).
7. Caruso, T. & Migliorini, M. Euclidean geometry explains why lengths allow precise body mass estimates in terrestrial invertebrates: the case of oribatid mites. *J Theor Biol*, **256**, 436-440 (2009).
8. Weigmann, G. Hornmilben (Oribatida) Die Tierwelt Deutschlands, 76. Teil. Goecke & Evers, Keltern (2006).
9. Grandjean, F. Essai de classification des Oribates (Acarieus). *Bull. Soc. Zool. Fr.* **78**, 421-446 (1954).
10. Grandjean F. Considération sur le classement des Oribates leur division en 6 groupes majeurs. *Acarologia* **11**, 127-153 (1969).
11. Schatz, H. Die Oribatidenliteratur und die beschriebenen Oribatidenarten (1758-2001): Eine Analyse. *Abh. Ber. Naturk. Görlitz* **74**, 37-45 (2002).
12. Krivolutsky, D. A. & Krassilov, V. A. *Oribatid mites from Upper Jurassic, USSR*. In: Skarlato OA, Balasho S (Eds.), Morphology and diagnostics of mites. *Academy of Sciences of the USSR, Leningrad*, pp. 16-24 (1977).
13. Arillo, A., Subías, L. S. & Shtanchaeva, U. A new fossil oribatid mite, *Ommatocephus nortoni* sp. nov. (Acariformes, Oribatida, Cepheidae) from a new outcrop of lower Cretaceous Álava amber (northern Spain). *System. Appl. Acarol.* **13**, 252-255 (2008).
14. Sivhead, U. & Wallwork, J. A. An early Jurassic oribatid mite from southern Sweden. *Geol. Foren. Stock. For.* **100**, 65-70 (1978).
15. Bulanova-Zachvatkina, Y. M. A new genus of mite (Acariformes, Oribatei) from the upper Cretaceous of Taymyr. *Paleont. Journ.* **8**, 247-250 (1974).

16. Arillo, A., Subías, L. S. & Shtanchaeva, U. A new species of fossil oribatid mite (Acariformes, Oribatida, Trhypochthoniida) from the lower Cretaceous amber of San Just (Teruel Province, Spain). *System. Appl. Acarol.* **17**, 106-112 (2012).
17. Subías, L. S. & Arillo, A. Oribatid fossil mites from the upper Devonian of South Mountain, New York and the lower Carboniferous of County Antrim, North Ireland (Acariformes, Oribatida). *Estud. Museo Cienc. Nat. Álava* **17**, 93-106 (2002).
18. Norton, R. A., Bonamo, P. M., Crierson, J. D. & Shear, W. A. Oribatid mite fossils from a terrestrial deposit near Gilboa, New York. *J. Paleontol.* **62**, 259-269 (1988).
19. Shear, W. A., Bonamo, P. M., Grierson, J. D., Rolfe, W. D. I., Laidlaw Smith, E. & Norton, R. A. Early land animals in North America: evidence from Devonian age arthropods from Gilboa, New York. *Science* **224**, 492-494 (1984).
20. O'Dowd, D. J., Brew, C. R., Christophel, D. C. & Norton, R. A. Mite-plant associations from the Eocene of Southern Australia. *Science* **252**, 99-101 (1991).
21. Woolley, T. A. Fossil oribatid mites in amber from Chiapas, Mexico (Acarina: Oribatei: Cryptostigmata). *Univ. Calif. Publ. Entomol.* **63**, 91-99 (1971).
22. Mico, L. Oribatid mite fossils from pre-Quaternary sediments in Slovenian caves III. Two new species of *Dissorhina* (Oppiidae) from the Pliocene. *Acarologia* **55**, 449-457 (2015).
23. Norton, R. A. First record of *Collohmanna* (*C. schusteri* n. sp.) and *Hermannia* (*H. sellnicki* n. sp.) from Baltic amber, with notes on Sellnick's genera of fossil oribatid mites (Acari: Oribatida). *Acarologia* **46**, 111-125 (2006).
24. Labandeira, C. C., Phillips, T. L. & Norton, R. A. Oribatid mites and the decomposition of plant tissues in Paleozoic coal-swamp forests. *Palaios* **12**, 319-353 (1997).
25. Doğan, S. & Ayyıldız, N. Systematic investigation on *Tectocephus* Berlese, 1895 (Acari: Oribatida) species in Erzincan and Erzurum plains. *Türk. entomol. Der.* **24**, 69-80 (2000).
26. Knülle, W. Die Arten der Gattung *Tectocephus* Berlese (Acarina: Oribatei). *Zool. Anz.* **152**, 280-305 (1953).
27. Grandjean, F. *Phthiracarus anonymum*, n. sp. *Rev. Fr. Entomol.* **1**, 51-59 (1934).
28. Kamill, B. W. & Baker, A. The genus *Atropacarus* Ewing (Acari: Cryptostigmata). *Bull. Br. Mus. Nat. Hist. (Zool.)* **39**, 189-204 (1980).
29. Bernini, F. & Avanzati, A.M. Notulae oribatologicae xlix. taxonomic revision of *Steganacarus* (*Steganacarus*) *aplicatus* (Sellnick, 1920) and the description of a new west Mediterranean *Steganacarus* species (Acarida, Oribatida). *Int. J. Acarol.* **15**, 65-74 (1989).
30. Bernini, F. & Avanzati, A. M. Taxonomic revision of *Steganacarus* (*Steganacarus*) *magnus* (Nicolet, 1855) (Acarida, Oribatida). *J. Nat. Hist.* **22**, 435-464 (1988).
31. Els, A. J. Studies on the South African Oribatei Inferiores (Acarina) part 1: description of *Mesoplophora africana* (Balogh 1958). *S. Afr. J. Sci.* **62**, 76-79 (1966).

32. Schatz, H. *Palaeacarus hystricinus* Trägårdh, 1932 (Acari: Oribatida: Palaeacaridae), a remarkable oribatid mite from Tyrol. *Ber. nat.-med. Verein Innsbruck*, **91**, 339-340 (2004).
33. Erdmann, G., Otte, V., Langel, R., Scheu, S. & Maraun, M. The trophic structure of bark-living oribatid mite communities analysed with stable isotopes (^{15}N , ^{13}C) indicates strong niche differentiation. *Exp. Appl. Acarol.* **41**, 1-10 (2007).
34. Lehmitz, R. & Maraun M. Small-scale spatial heterogeneity of stable isotopes signatures ($\delta^{15}\text{N}$, $\delta^{13}\text{C}$) in *Sphagnum* sp. Transfers to all trophic levels in oribatid mites. *Soil Biol. Biochem.* **100**, 242-251 (2016).
35. Maaß, S., Maraun, M., Scheu, D., Rillig, M. C. & Caruso, T. Environmental filtering vs. resource-based niche partitioning in diverse soil animal assemblages. *Soil Biol. Biochem.* **85**, 145-152 (2015).
36. Maraun, M., Erdmann, G., Fischer, B. M., Pollierer, M. M., Norton, R. A., Schneider, K. & Scheu, S. Stable isotopes revisited: Their use and limits for oribatid mite trophic ecology. *Soil Biol. Biochem.* **43**, 87-882 (2011).
37. Ilardi, M., Dick, J. T. A., Maraun, M. & Caruso, T. Response of native forest floor microarthropods to the invasive macroinvertebrate *Arcitalitrus dorrieni* (Crustacea; Amphipoda) (in preparation, 2019).
38. Magilton, M., Maraun, M., Emmerson, M. & Caruso, T. Oribatid mites reveal that competition for resources and trophic structure combine to regulate the assembly of diverse soil animal communities. (in preparation, 2019).
39. Original stable isotope data were obtained from the authors on request.