

Variation in the coding region of the heavy metal ATPase *BrHMA3* controls natural variation in cadmium accumulation in widely eaten *Brassica rapa* vegetables

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Supplementary Figures S1–S10 and Tables S1-S4

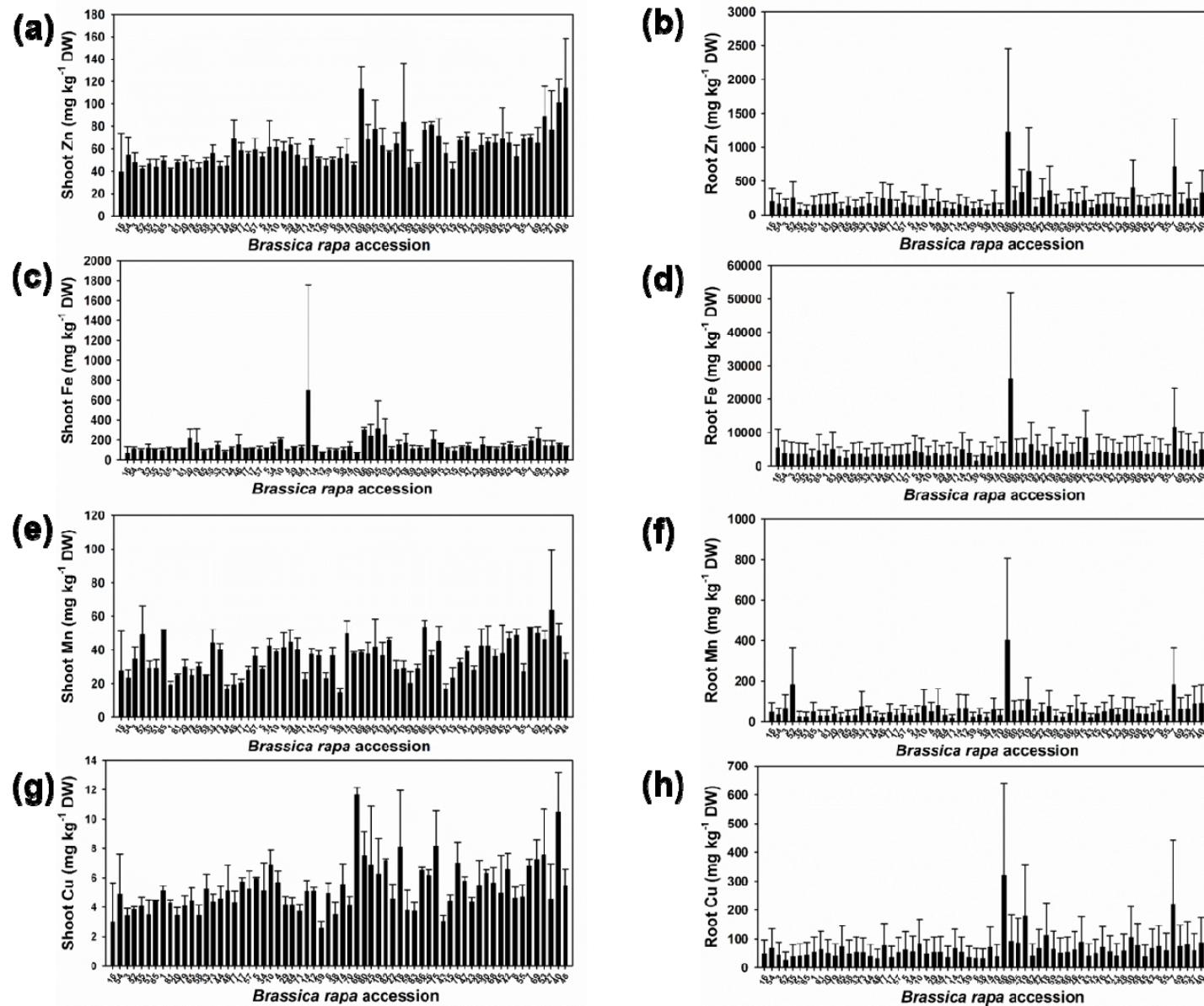


Fig. S1. Concentrations of Zn (a, b), Fe (c, d), Mn (e, f) and Cu (g, h) in the shoots (a, c, e, g) and roots (b, d, f, h) of 64 *Brassica rapa* accessions grown in hydroponic culture. Data are the means \pm SD ($n = 3$).

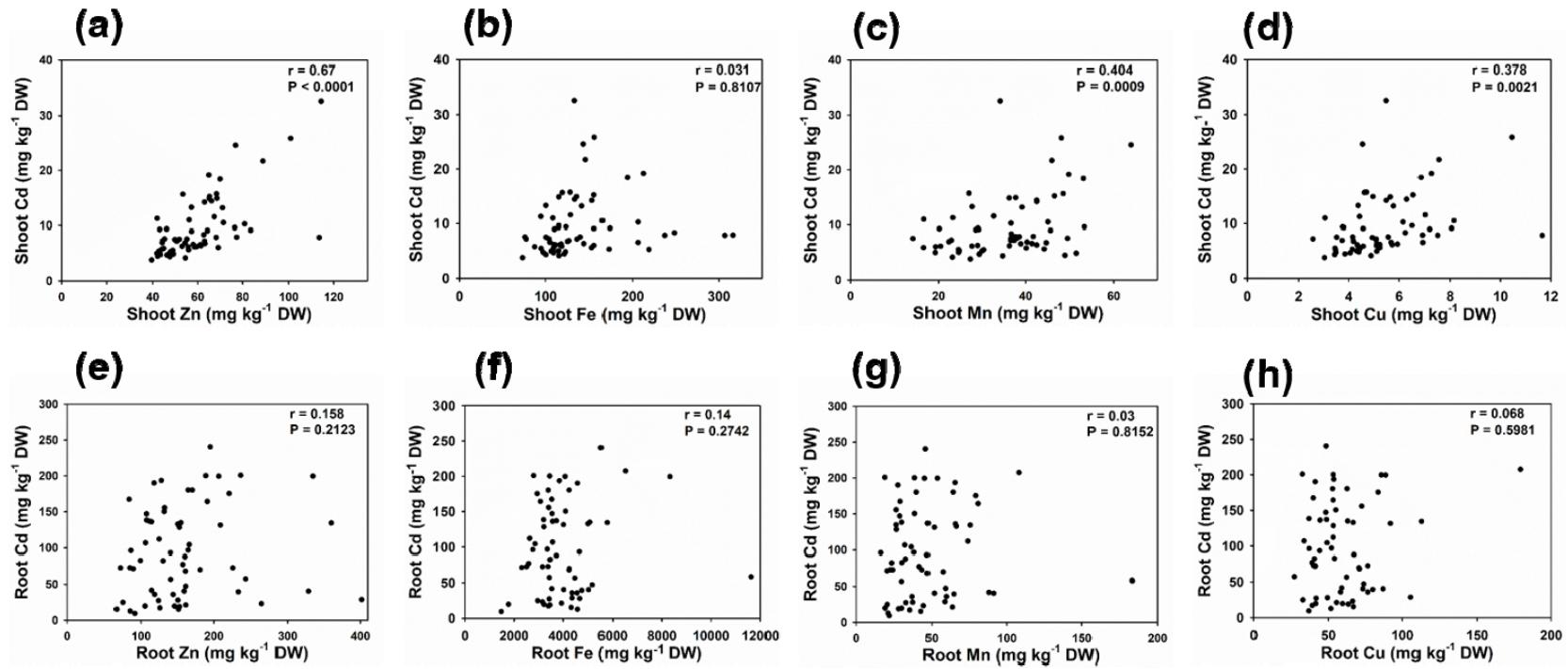


Fig. S2. Relationships between the concentration of Cd with those of Zn (a, e), Fe (b, f), Mn (c, g) and Cu (d, h) in the shoots (a – d) and roots (e – h) of 64 *Brassica rapa* accessions grown in hydroponic culture.

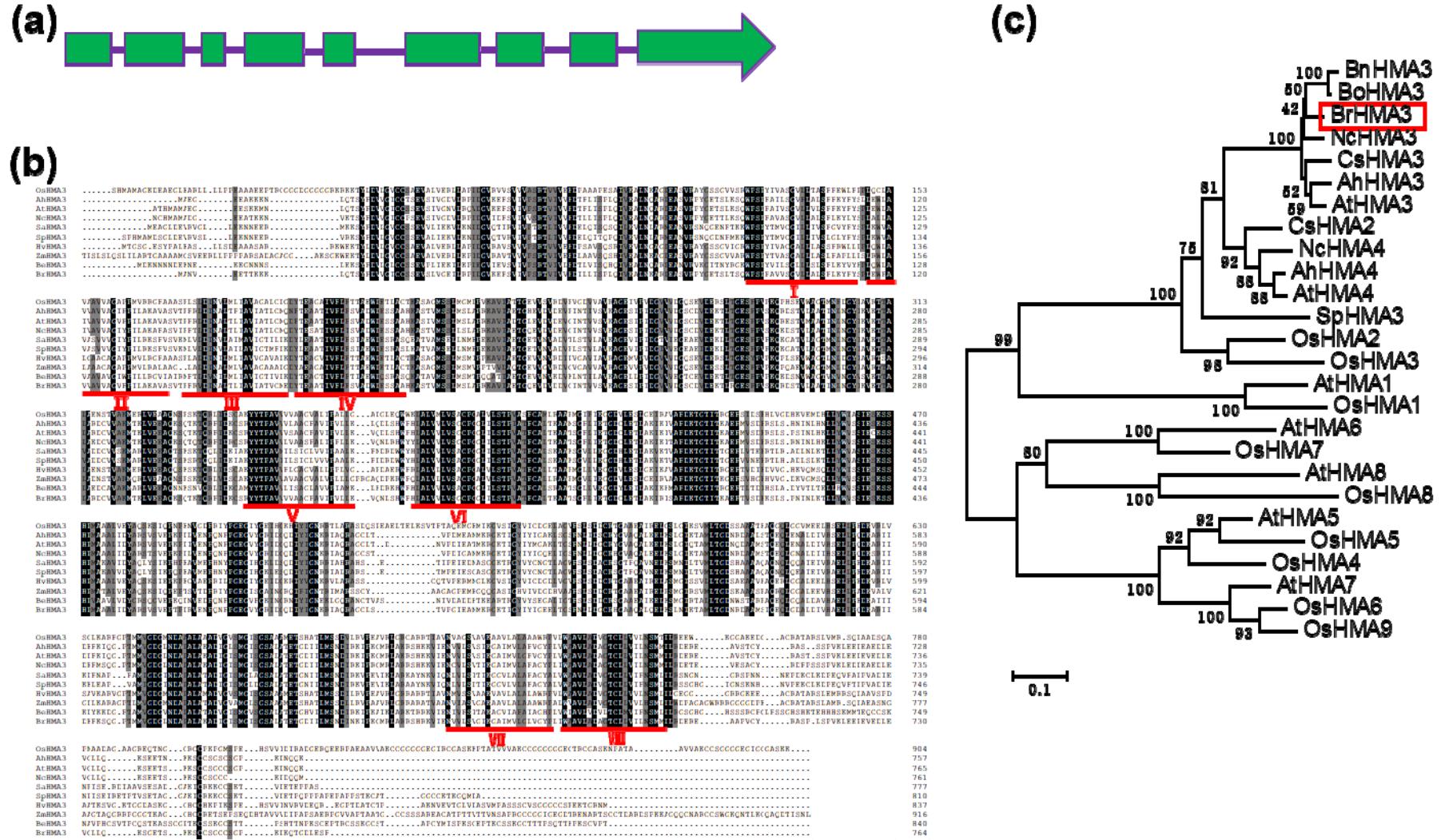


Fig. S3. *BrHMA3* gene in *Brassica rapa*. Schematic gene structure (a), sequence alignment of OsHMA3, AtHMA3, AhHMA3, NcHMA3, SaHMA3, SpHMA3, HvHMA3, ZmHMA3, BcHMA3, BrHMA3, (b), and phylogenetic analysis of P_{1B}-ATPases (c). Transmembrane domains (underlined in b) were predicted by TOPCONS.

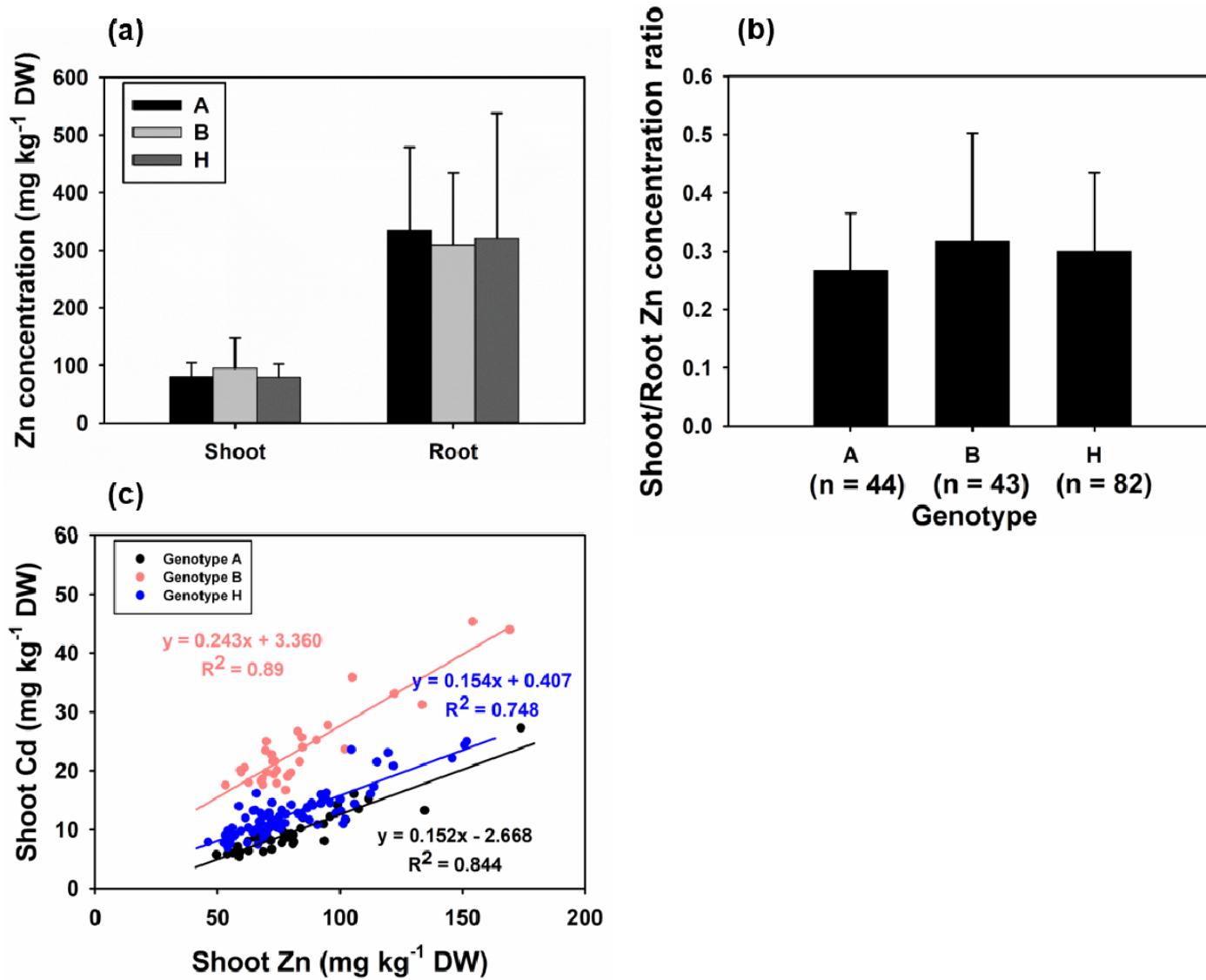
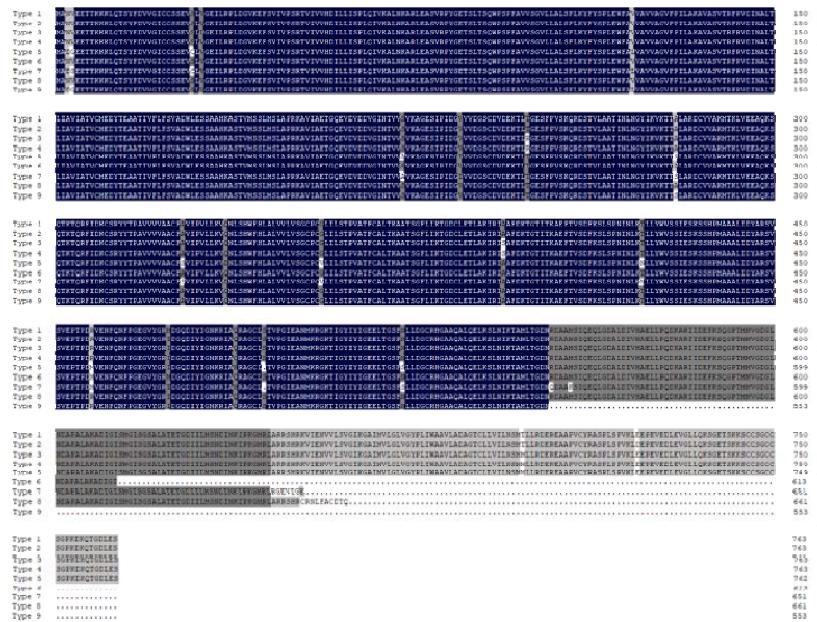


Fig. S4. The concentrations of Zn in the shoots and roots (a), the ratio of shoot/root Zn concentrations (b), and the relationship between the concentrations of Cd and Zn in the shoots (c) of 151 F₂ plants grouped according to *BrHMA3* genotypes. A, B, and H represent *BrHMA3* genotypes of GJCGB, DQMY939 and heterozygous. Data are the means ± SD ($n = 43 - 82$). DW, dry weight.

(a)**(b)**

Haplotypes	Frequency
Low-Cd	
Type1	3
Type2	2
Type3	8
Type4	2
Type5	27
High-Cd	
Type6	4
Type7	1
Type8	11
Type9	6

Fig S5. Amino acid sequence alignment of nine protein haplotypes of BrHMA3 among the 64 *B. rapa* accessions (a) and the frequency distribution of each haplotype (b). Sequences were aligned using the DNAMAN programme.

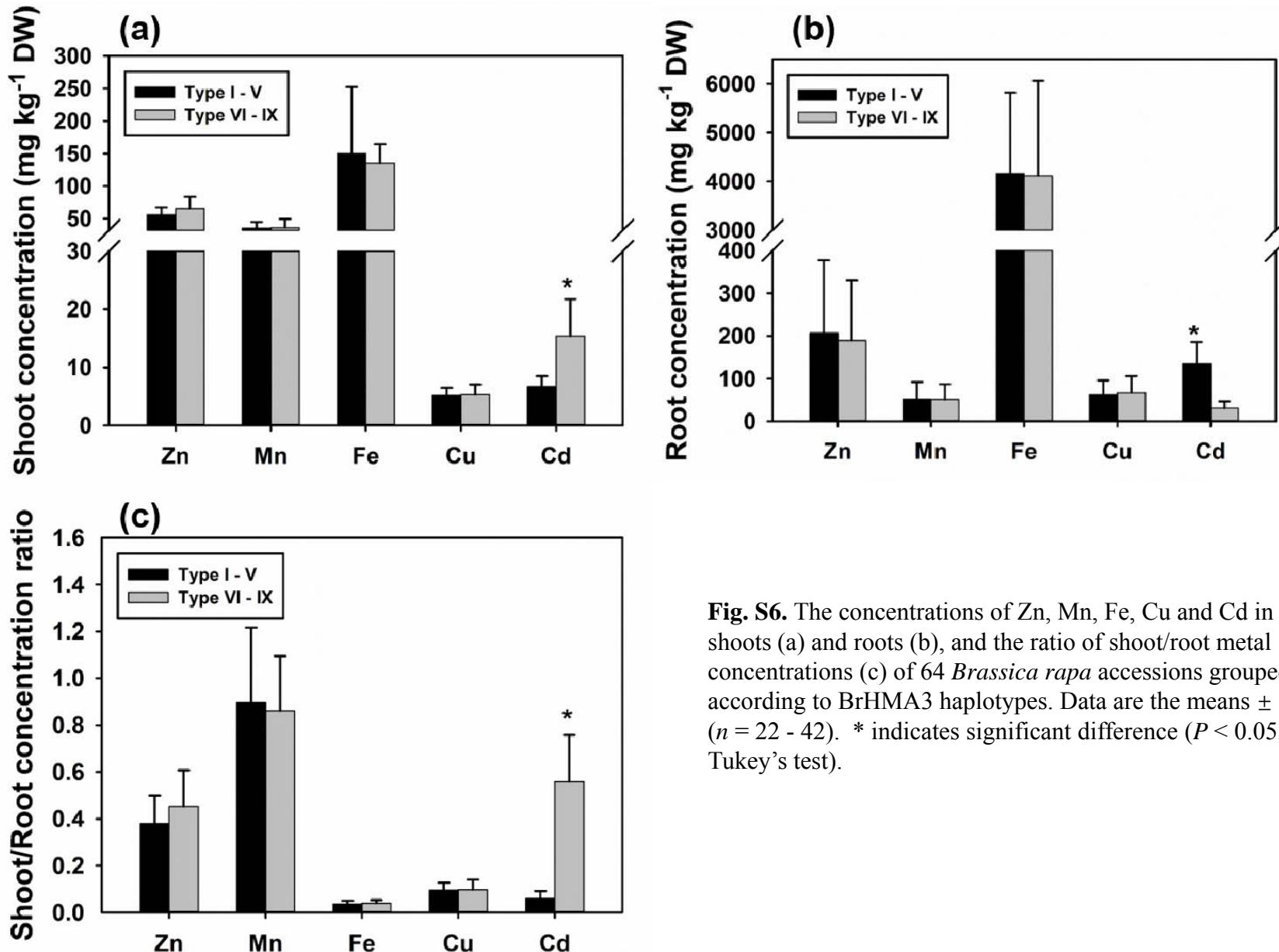


Fig. S6. The concentrations of Zn, Mn, Fe, Cu and Cd in the shoots (a) and roots (b), and the ratio of shoot/root metal concentrations (c) of 64 *Brassica rapa* accessions grouped according to BrHMA3 haplotypes. Data are the means \pm SD ($n = 22 - 42$). * indicates significant difference ($P < 0.05$, Tukey's test).

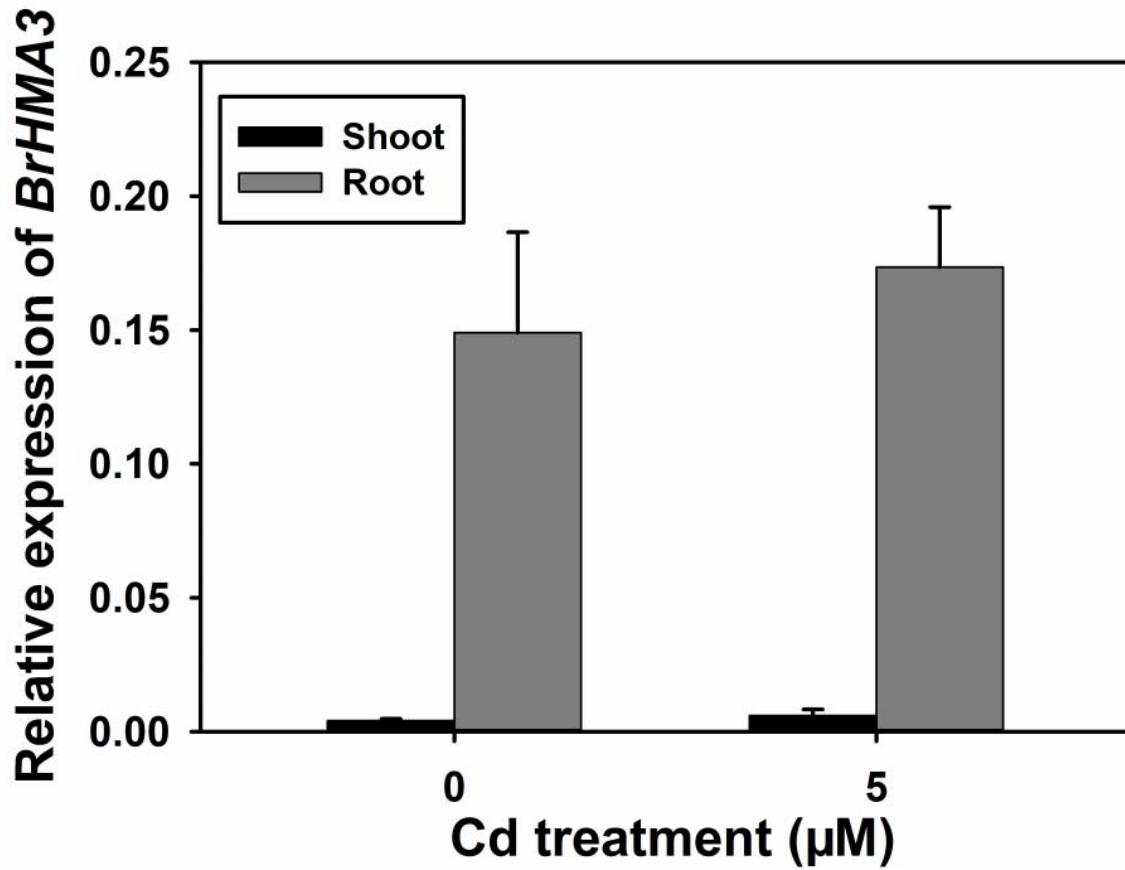


Fig. S7. The expression pattern of *BrHMA3* in the roots and shoots of *Brassica rapa* (cv. Chiifu) grown in hydroponic culture with or without 5 μM Cd for 3 d. Expression of *BrHMA3* was calculated as $2^{-\Delta CT}$ relative to *Bra005178*. Data are the means \pm SD ($n = 3$).

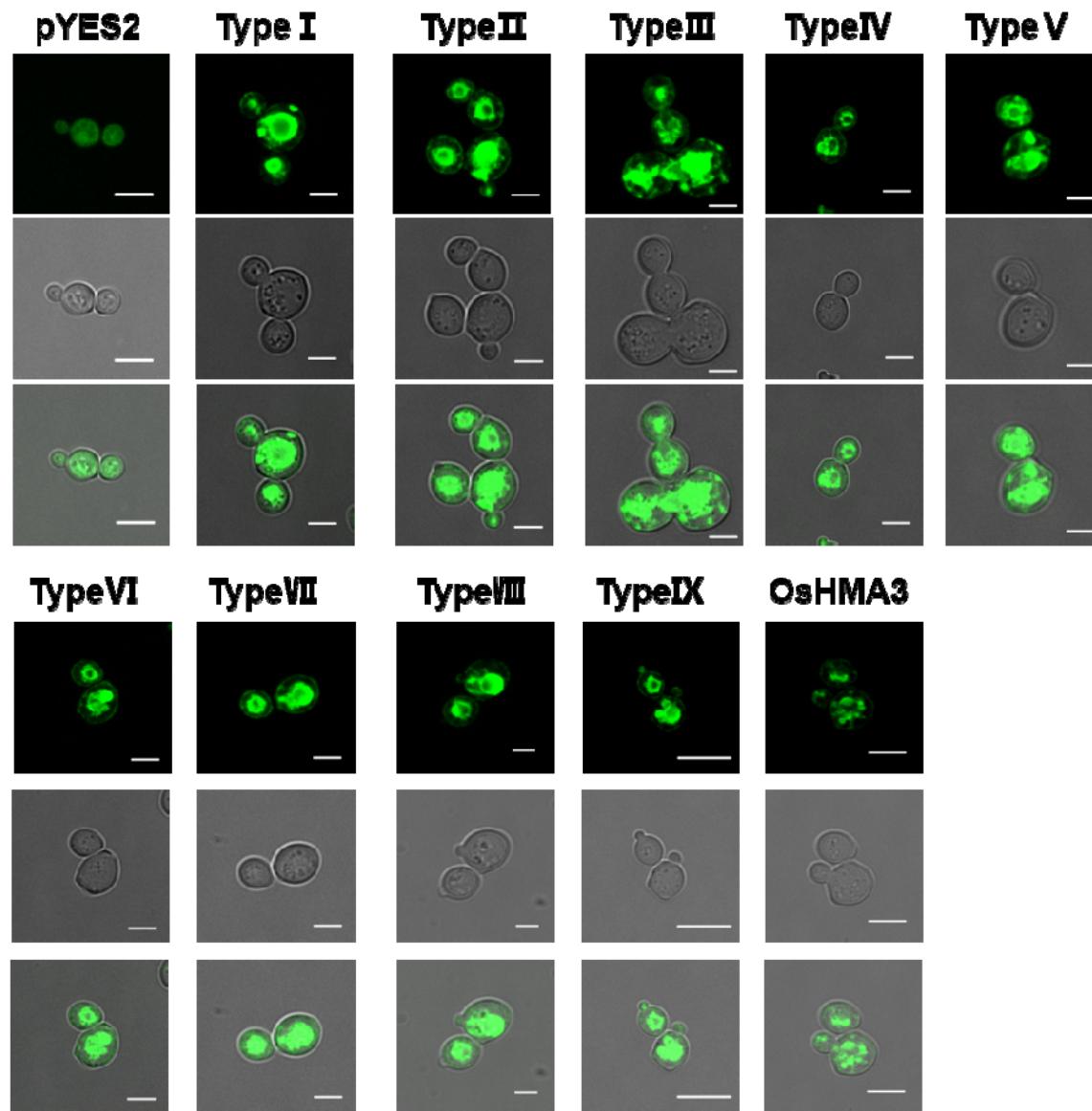


Fig. S8. Intracellular localization of GFP-BrHMA3 and GFP-OsHMA3 expressed in yeast strain INVSc1.
Bar = 2 μ m.

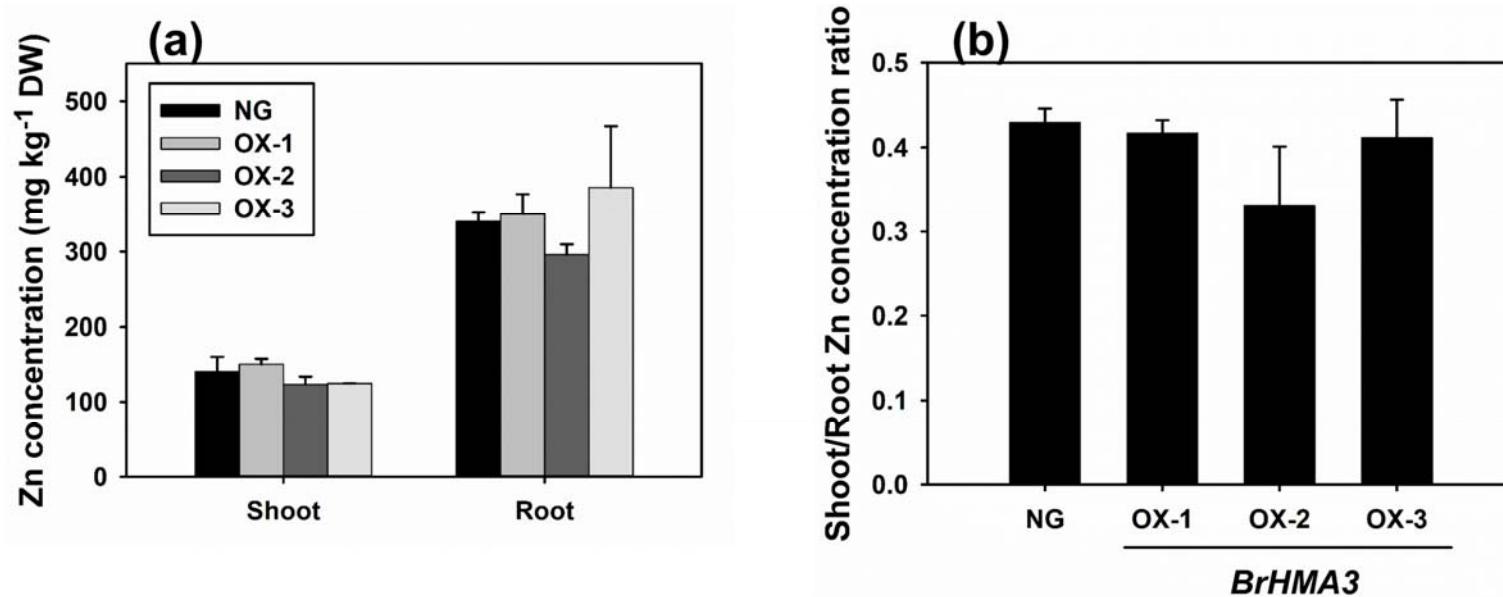


Fig. S9. The concentrations of Zn in the roots and shoots (a) and the shoot/root Zn concentration ratio of transgenic rice plants expressing *BrHMA3* (b). NG, non-transgenic segregation line control; OX, overexpressing lines. Data are the means \pm SD ($n = 3$).

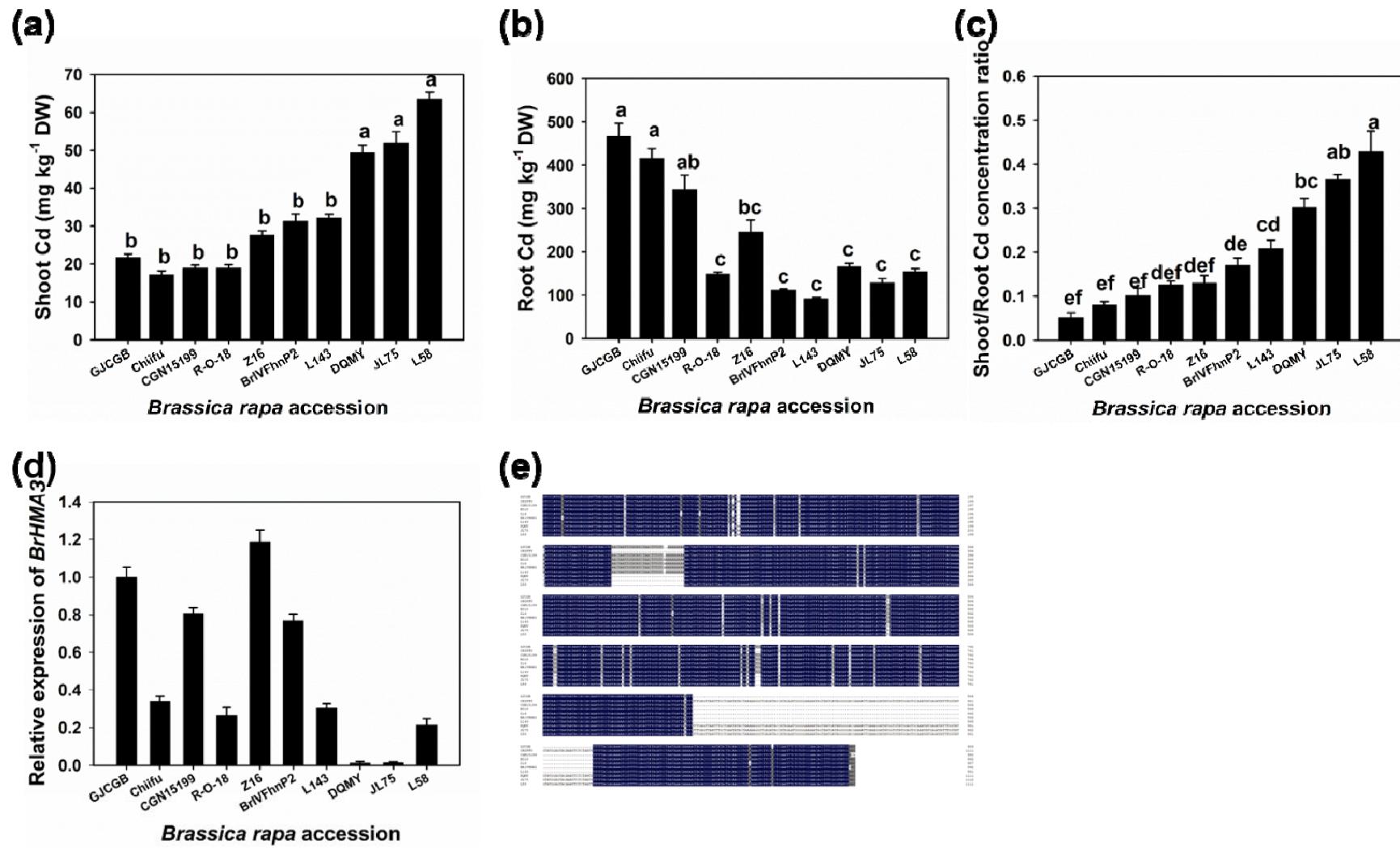


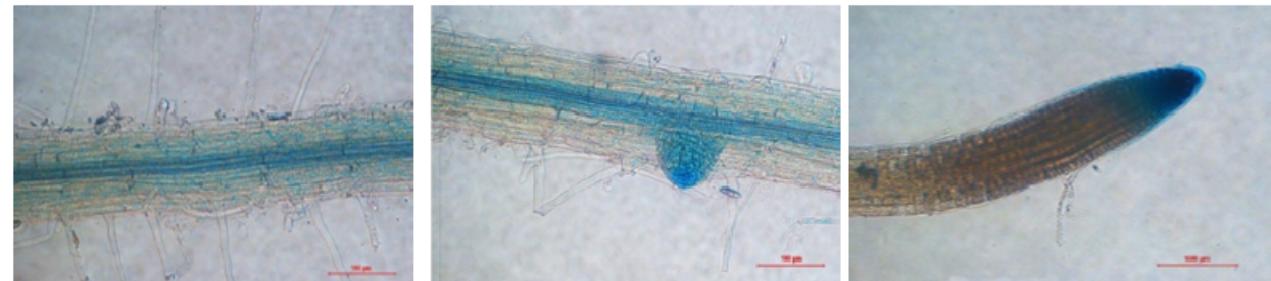
Fig. S10. Cadmium concentrations in the shoots (a) and roots (b), and the ratio of shoot/root Cd concentrations of ten accessions of *Brassica rapa* grown in hydroponic culture with 0.5 μ M Cd for 10 d; relative expression of *BrHMA3* in the roots (d) and the alignment of *BrHMA3* promoter sequences in the ten accession of *B. rapa* (e). Data are the means \pm SD ($n = 3$). Different letters above the bars indicate significant difference at $P < 0.05$ (Tukey's test).

Promoter Sequence

Type A



Low Cd In shoot



Type B



High Cd In shoot

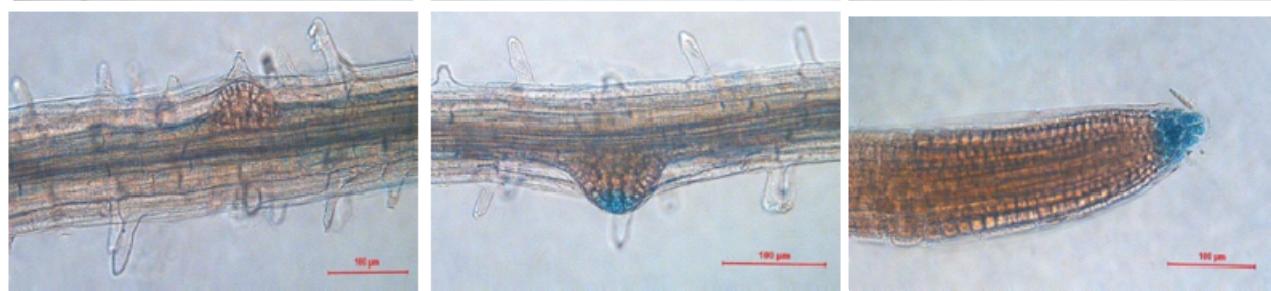


Fig. S11. *BrHMA3* promoter activity. GUS staining of transgenic *Arabidopsis thaliana* containing two types of *Pro^{BrHMA3}::GUS* constructs. The blue and yellow parts indicate 35 bp deletion and 152 bp insertion. Bar = 100 µm.

Table S1. The code, name, subspecies, and BrHMA3 haplotype of 64 *Brassica rapa* accessions used in the present study

BrHMA3 Haplotype	Subspecies	Cultivar name	Code
Type I	Brassica chinensis L.	Gaojiaochigengbai(GJCGB)	3
Type I	Brassica pekinensis	DatouqingV02A155(DTQV02A155)	34
Type I	Brassica pekinensis	Chiifu	52
Type II	Brassica chinensis L.	Chiqingcai(CQC)	4
Type II	Brassica pekinensis	V02A0555xiaoshizitou(V02A0555XSZT)	57
Type III	Brassica chinensis L.	Sijixiangexiaobaicai(SJXCB)	10
Type III	Brassica chinensis L.	Shanghaiyueman(SHSYM)	14
Type III	Brassica chinensis L.	Zhongbayewutaihai(ZBYWTC)	19
Type III	Brassica narinosa L. H. Bailey	Shangdong4(SD4)	26
Type III	Brassica pekinensis	Fushan212(FS212)	54
Type III	Brassica pekinensis	Zicaitai(ZCT)	65
Type III	Brassica parachinensis L. H. Bailey	Gailiangshiyuehong(GLSYH)	66
Type III	Brassica parachinensis L. H. Bailey	SynBr05	86
Type III	Brassica parachinensis L. H. Bailey	Huangqinqinggengbaicai(HGQGBC)	17
Type IV	Brassica chinensis L.	Lvlingwutaihai(LLWTC)	18
Type IV	Brassica narinosa L. H. Bailey	V02B0645Huomojiuquanxinwu(V02B0645HQQXH)	1
Type V	Brassica chinensis L.	Shianyoucui(SAYC)	5
Type V	Brassica chinensis L.	Zhongheqingcui(ZHQC)	6
Type V	Brassica chinensis L.	Material from Holland by Zhaojianjun	12
Type V	Brassica chinensis L.	V02B0591Takucuai(V02B0591TKC)	16
Type V	Brassica chinensis L.	Z16	20
Type V	Brassica chinensis L.	Jianchun(JC)	25
Type V	Brassica chinensis L.	424-2-3	29
Type V	Brassica chinensis L.	14Hetaowenxiaoqingkou(14HTWXQK)	33
Type V	Brassica chinensis L.	Beijingxinsanhaob(BJXSH)	35
Type V	Brassica chinensis L.	V02A1351Shimabaicai(V02A1351SMBC)	44
Type V	Brassica chinensis L.	CGN07201	46
Type V	Brassica chinensis L.	Kenshin	51
Type V	Brassica chinensis L.	Yunhongzhongjiang(YHZJ)	58
Type V	Brassica campestris L.	Liuye70tianyouqingcaixin(LY70TYQCX)	70
Type V	uncultivated	Chamagudaoyebaicai(CMGDYBC)	71
Type V	Yellow Sarson	R-o-18	73
Type V	Yellow Sarson	L143	74
Type V	Komatsuna	L41	75
Type V	Komatsuna	CGN17281	76
Type V	Brassica juncea var.multisecta L.H.Bailey	Xizhangyeshengbaicai8#(XZYSBC8#)	77
Type V	Brassica rapa L.	CGN15199	79
Type V	Brassica rapa L.	CGN15220	80
Type V	Brassica rapa L.	CGN15201	81
Type V	Brassica rapa L.	CGN06721	82
Type V	Brassica pekinensis	Sijichun (SJC)	84
Type V	Rapid cycling	SynBr02	85
Type VI	Brassica chinensis L.	Shanghaiqing(SHQ)	7
Type VI	Brassica chinensis L.	Suzhouqing(SZQ)	8
Type VI	Brassica pekinensis	Xianfengxiayang(XFXY)	22
Type VI	Brassica pekinensis	Beijing75(BJ75)	30
Type VII	Brassica campestris L.	L58DH	68
Type VIII	Brassica chinensis L.	Jingyanwuyueman(JYWYM)	15
Type VIII	Brassica pekinensis	BrVfVfh P1	23
Type VIII	Brassica pekinensis	Xinfeng90(XF90)	38
Type VIII	Brassica pekinensis	33--3	40
Type VIII	Brassica pekinensis	Huabai2(HB2)	42
Type VIII	Brassica pekinensis	V02A1499Zhulongcui(V02A1499ZLC)	43
Type VIII	Brassica pekinensis	Xinlv85(XL85)	47
Type VIII	Brassica pekinensis	Xiakang40(XK40)	48
Type VIII	Brassica pekinensis	Zhulongcui(ZLC)	53
Type VIII	Brassica pekinensis	Qingbai'er5-4-2(QBE5-4-2)	55
Type VIII	Brassica campestris L.	Dongguanbaishayouqing45tiantiancaixin(DGBSYQ45TTCX)	69
Type IX	Brassica pekinensis	Ji'nanagenV02A0806(JNDGV02A0806)	27
Type IX	Brassica pekinensis	Jnlv75(JL75)	28
Type IX	Brassica pekinensis	L488-3	39
Type IX	Brassica pekinensis	V02A1396Xingchengxiaocuocai(V02A1396XCXCC)	45
Type IX	Brassica pekinensis	Daqingmaye939(DQMY939)	59
Type IX	Brassica rapa L.	CGN06688	83

Table S2. Nine *Brassica rapa* accessions selected for the soil pot experiment.

BrHMA3 Haplotype	Subspecies	Name	Code
Type I	<i>Brassica pekinensis</i>	Chiifu	52
Type II	<i>Brassica pekinensis</i>	V02A0555xiaooshizitou(V02A0555XSZT)	57
Type III	<i>Brassica pekinensis</i>	Shangdong4(SD4)	26
Type IV	<i>Brassica chinensis</i> L.	Huaguanqinggengbaicai(HGQGBC)	17
Type V	<i>Brassica pekinensis</i>	Z16	20
Type VI	<i>Brassica chinensis</i> L.	Shanghaiqing(SHQ)	7
Type VII	<i>Brassica campestris</i> L.	L58DH	68
Type VIII	<i>Brassica pekinensis</i>	BrIVFhn P1	23
Type IX	<i>Brassica pekinensis</i>	Daqingmaye939(DQMY939)	59

Table S3. Primers used in the present study

Purpose	Forward(5' - 3')	Reverse (5' - 3')
BrHMA3 ORF (Genomic DNA)	TCCGAACACCTTCGCGTAT	GTACTTACCAATTCTACAGATT
PYES2-BrHMA3	AGGGAAATTAAAGCTTGGTACCATGGCCAACGTTGAAGAGAGACAAC	GGCGGCCGTTACTAGTGGATCCTCAGAAACTCTAAGTCCCTGTTT
BrHMA3-GFP	CTTCGAATTCTGCAGTCGACGGTACCATGGCCAACGTTGAAGAGAGACA	CTCGCCCTGCTCACCATCAGGATCCAGAAACTCTAAGTCCCTGTT
Genetic marker (Promoter)	ATAACCTAATAATACCACGAC	CGTTGGATACCGCAAAGGTGTT
Genetic marker (ORF)	GCAATGCTCACAGGAGATAGC	GCTGTAAAAATGATAACGTTTGGTTATT
Pro^{BrHMA3}-GUS	TTAATTAAATGGCCAACGTTGAAGAGACAAC	GGCGCGCCTCAGAAACTCTAAGTCCCTGTT
Arabidopsis Actin	TCGTTGCCCTCCAGAGA	GGTACTGAGGGAGGCCAAGAT
AthMA3 promoter	TAAAACGACGGCCAGTGCCAAGCTTACGCTTGAAACAGTTGAGAAAAA	GTTGTCCTTCAACGTTGGCCATTGAGATCTGGACTAAATTG
Rice actin	GGTCAACTTGTGATTCCCTCT	AACCGCAAATCAAAGAACG
OsHMA3 qRT-PCR	TCCATCCAACCAAACCCGGAAA	TGCCAATGTCTCTGTTCCC
Brassica rapa Actin	CTCACCGAAGCACCTCTCAA	GACCACTGGCGTAAAGGGAG
BrHMA3 qRT-PCR	TTCTTGAGATGCAGGGACTT	AGTCCCTGTTGCTTGCC

Table S4. Ten *Brassica rapa* accessions selected for promoter analysis

BrHMA3 Haplotype	Subspecies	Name
Type I	Brassica chinensis L.	Gaojiaochigengbai(GJCGB)
Type V	Brassica pekinensis	Z16
Type V	Yellow Sarson	R-o-18
Type I	Brassica pekinensis	Chiifu
Type V	Yellow Sarson	L143
Type IX	Brassica pekinensis	Jinlv75(JL75)
Type VII	Brassica campestris L.	L58DH
Type IX	Brassica pekinensis	Daqingmaye939(DQMY939)
Type V	Brassica rapa L.	CGN15199
Type I	Brassica pekinensis	BrIVFhn P1