

**Supplemental Information:**

**Figures S1-S5 and Table S1.**

Buechler...Turley, Figure S1

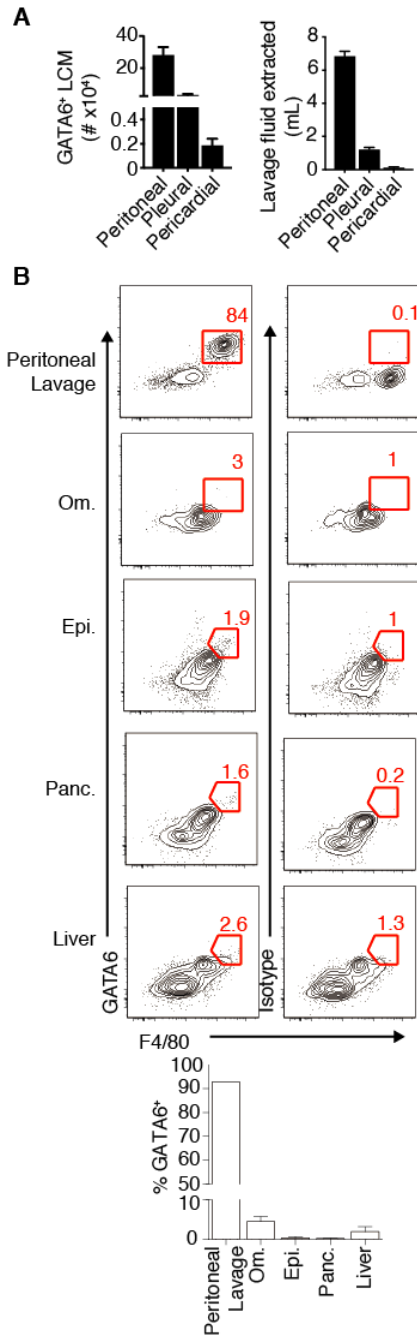


Figure S1. LCM numbers across cavities and mesothelial cells and fibroblasts at cavity surfaces exhibit *Wt1-Raldh1/2* axis.

*Related to Figure 1*

(A) GATA6<sup>+</sup> LCM number across cavities (left) and lavage fluid extracted (right). (B) Frequency of GATA6<sup>+</sup> F4/80<sup>+</sup> cells in peritoneal lavage and select cavity-associated tissues (top) and quantification (bottom). (A) Data are from 12-13 mice across >3 experiments. (B) Data are from 3 experiments. Peritoneal lavage quantification is from one experiment, representative of ≥3. (A-B) Mean values + SEM are shown.

Buechler...Turley, Figure S2

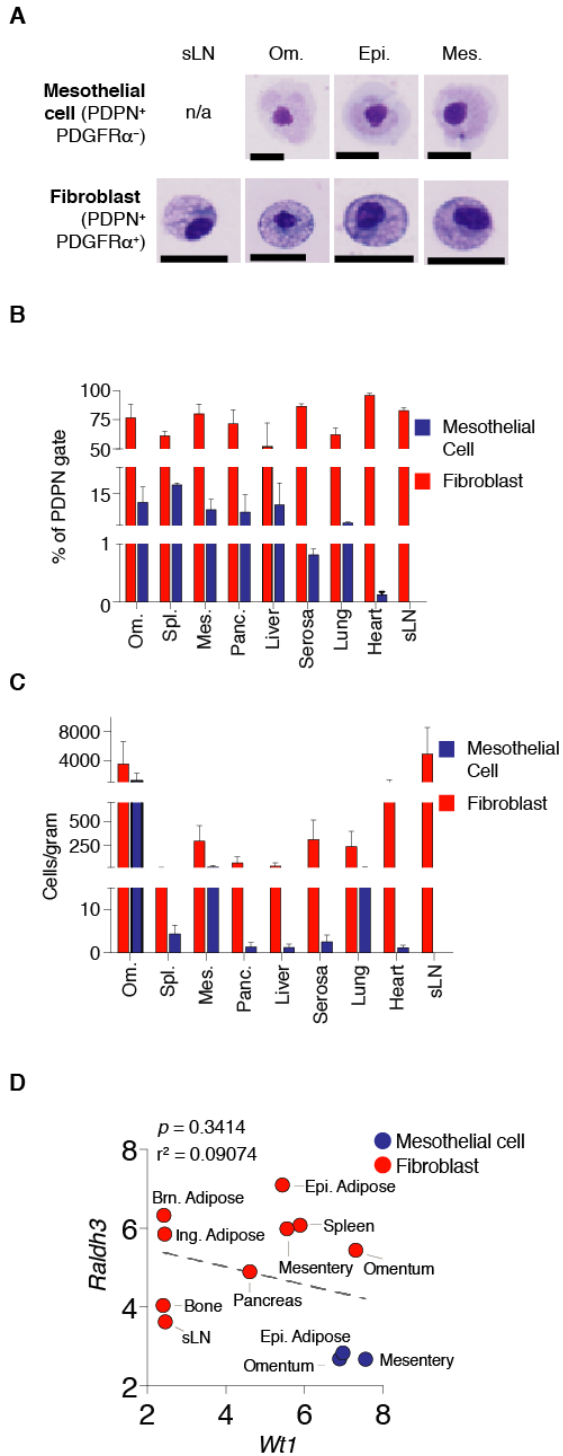


Figure S2. *Wt1*<sup>+</sup> mesothelial and fibroblastic stromal cell identification and expression of *Wt1* and *Raldh3*.

*Related to Figure 3*

(A) FACS sorted samples after cytopspin and stained with Wright-Giemsa stain. Scale bar indicates 25 $\mu$ M. (B) Frequency of mesothelial cells and fibroblasts in PDPN<sup>+</sup> gate in cavity and non-cavity tissues. (C) Mesothelial cells and fibroblasts per gram of tissue. (D) Correlation between *Raldh3* and *Wt1* in FACS-sorted mesothelial cells (blue dots) and fibroblasts (red dots). Gene expression across biological replicates (TPM + 5 (Log<sub>2</sub>)) were averaged to generate data points. (A) Data are representative of 2 samples. (B, C) Data are from 3 experiments with 1 mouse per experiment. (D) 3 independent biological replicates were sequenced for each cell type, except spleen and pancreas fibroblasts (n = 2). (B-C) Mean values + SEM are shown. (D) p value and r-squared values determined by linear regression.

Buechler...Turley, Figure S3

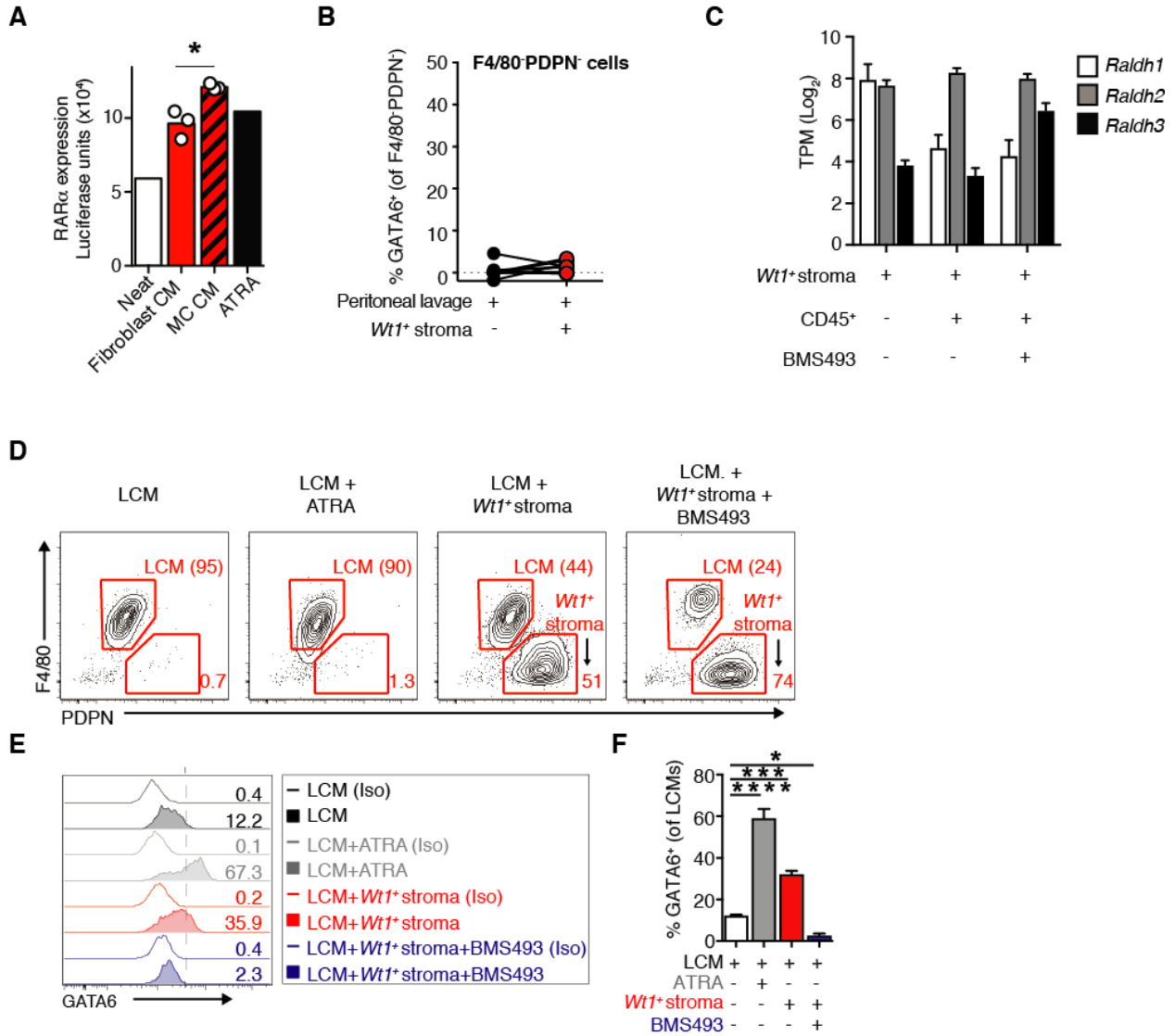


Figure S3. *Wt1*<sup>+</sup> mesothelial and fibroblastic stromal cells produce active retinoic acid and maintain GATA6 in LCM.

Related to Figures 4 and 5

(A) Luciferase expression in RAR $\alpha$ -luciferase cells after 24 hours of stimulation with media or conditioned media from mesothelial cells, fibroblasts or ATRA. (B) Quantification of the frequency of live GATA6<sup>+</sup> F4/80<sup>+</sup> PDPN<sup>-</sup> cells after culture with autologous omental *Wt1*<sup>+</sup> stromal cells. (C) Expression (TPM; transcripts per million) of *Raldh1*, *Raldh2*, *Raldh3* in FACS-sorted omental *Wt1*<sup>+</sup> stromal cells after 1 week in culture as noted. (D) Representative gating FACS-sorted co-culture assays. Numbers in parentheses represent frequency. (E) Representative GATA6 expression on peritoneal LCMs. Numbers represent frequency of GATA6<sup>+</sup> LCMs. Shaded histograms represent isotype staining. (F) Frequency of GATA6<sup>+</sup> peritoneal LCM after treatment with ATRA, co-cultured with omental *Wt1*<sup>+</sup> stromal cells with cell-cell contact or co-cultured with *Wt1*<sup>+</sup> stromal cells and BMS493. (A) Each dot represents 1 conditioned media lot. (B, E-F) GATA6 expression was determined by GATA6 staining minus isotype. Data are from  $\geq 3$  (B-C, F) or representative of 3 (D-E) experiments. 5 mice were pooled for FACS-sorted to generate cells in D-F. \* $p < 0.05$ , \*\*\* $p < 0.0005$ , \*\*\*\* $p < 0.0001$ , as determined by unpaired student's t-test (A) or Dunnett's multiple comparison post-test (F). Data are represented as mean (A-B), + SEM (C, F).

Buechler...Turley, Figure S4

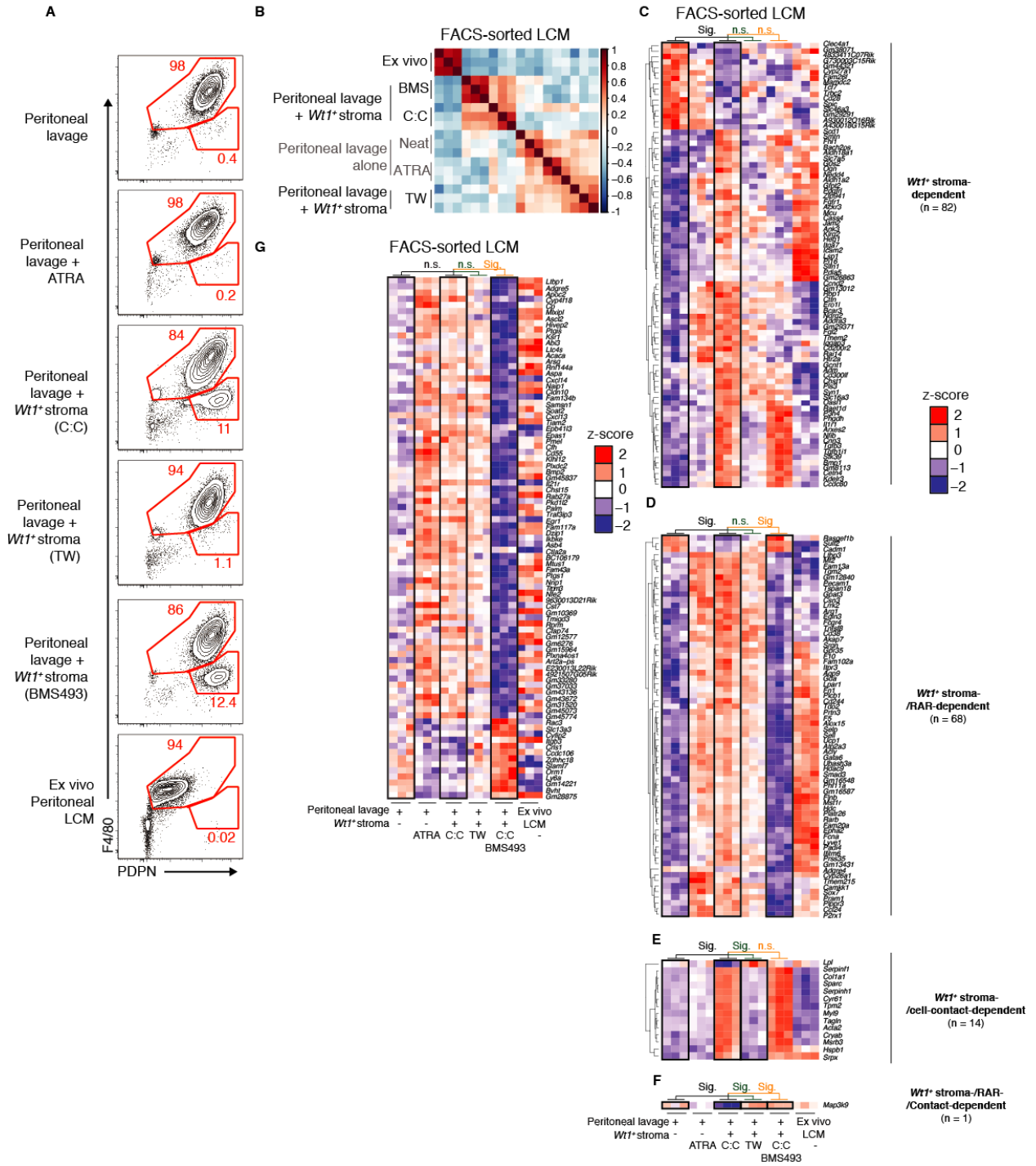


Figure S4. *Wt1*<sup>+</sup> stromal cells program LCM identity.

Related to Figure 5.

(A) Representative gating for FACS sorting of LCM or *Wt1*<sup>+</sup> stromal cells from co-cultures or ex vivo. Numbers represent cell frequency. (B) Sample similarity heatmap showing spearman correlation between samples calculated from normalized, log transformed, mean-centered expression of top 500 most variable transcripts. (C) *Wt1*<sup>+</sup> stromal cell-dependent genes were significantly different in LCM sorted from peritoneal lavage alone compared to LCMs sorted from peritoneal lavage:*Wt1*<sup>+</sup> stromal cell co-culture (66 genes went up in co-culture with *Wt1*<sup>+</sup> stromal cells; 16 down). (D) *Wt1*<sup>+</sup> stromal cell-/RAR-dependent genes were those significantly different in (C) and between LCMs sorted from peritoneal lavage:*Wt1*<sup>+</sup> stromal cells and LCMs sorted from peritoneal lavage:*Wt1*<sup>+</sup> stromal cells with BMS493 treatment (65 genes from went up in co-culture with *Wt1*<sup>+</sup> stromal cells and significantly changed when cultured with *Wt1*<sup>+</sup> stromal cells with BMS493; 3 went down in these comparisons). *Wt1*<sup>+</sup> stromal cell-/cell-contact-dependent genes were those significantly different in (C) and significantly different between LCMs sorted from peritoneal lavage:*Wt1*<sup>+</sup> stromal cell co-cultures in a transwell format (13 genes from went up in in co-culture with *Wt1*<sup>+</sup> stromal cells and significantly changed when cultured with *Wt1*<sup>+</sup> stromal cells in transwell; 1 went down in these comparisons). *Wt1*<sup>+</sup> stromal cell-/RAR-/cell-contact-dependent genes were those significantly different in comparisons made in (C), (D) and (E) (n = 1). (F) Differentially expressed genes in FACS-sorted LCM (same threshold as (C)) between peritoneal lavage:*Wt1*<sup>+</sup> stromal cells (C:C) compared to peritoneal lavage:*Wt1*<sup>+</sup> stromal cells (C:C) with BMS493 (n = 85). (A-G) Co-culture with cell contact (C:C), Co-culture with 0.4uM transwell membrane separation (TW), ATRA, BMS493. Cells were cultured at a ratio of 4-11:1(Peritoneal



lavage: *Wt1*<sup>+</sup> stromal cells) for 6 days. Three independent biological replicates were sorted for each treatment.

Buechler...Turley, Figure S5

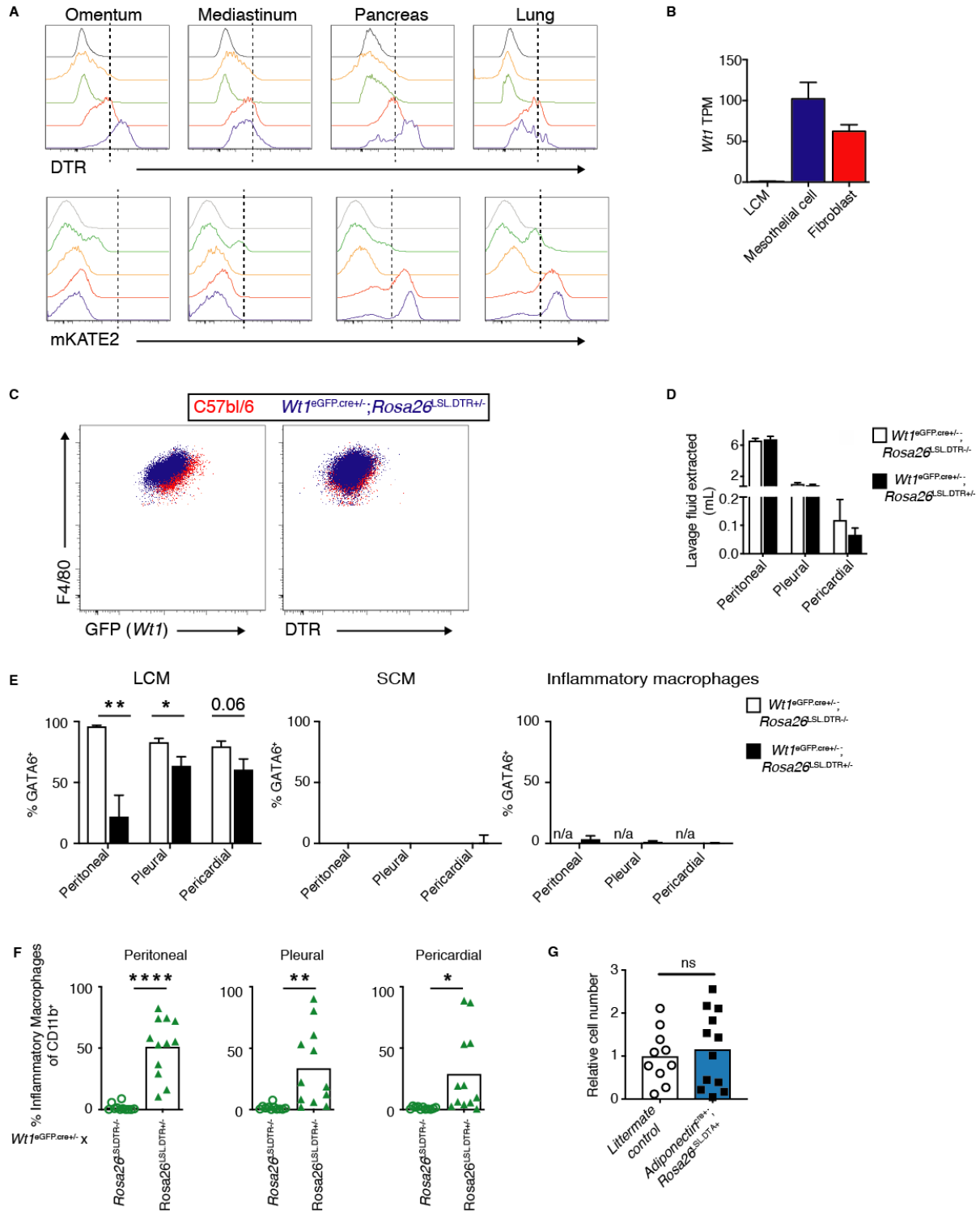


Figure S5. *Wt1*<sup>+</sup> stromal cells are critical for LCM homeostasis.

Related to Figure 6.

(A) GFP, DTR and mKATE2 gating in *Wt1*<sup>eGFP.cre</sup>;*Rosa26*<sup>LSL.DTR</sup> (top) and *Wt1*<sup>eGFP.cre+/-</sup>;*Rosa26*<sup>LSL.mKATE2+/-</sup> (bottom). CD45<sup>-</sup>EpCAM<sup>-</sup>CD31<sup>-</sup>PDPN<sup>+</sup>PDGFR $\alpha$ <sup>-</sup> mesothelial cells (blue), CD45<sup>-</sup>EpCAM<sup>-</sup>CD31<sup>-</sup>PDPN<sup>+</sup>PDGFR $\alpha$ <sup>+</sup> fibroblasts (red), CD45<sup>-</sup>EpCAM<sup>-</sup>PDPN<sup>-</sup>/+CD31<sup>+</sup> endothelial cells (orange) and CD45<sup>-</sup>EpCAM<sup>-</sup>PDPN<sup>-</sup>CD31<sup>-</sup> double negative cells (green) and SSC<sup>low</sup>CD45<sup>+</sup> hematopoietic cells (black). (B) *Wt1* expression in LCMs, omental mesothelial cells and omental fibroblasts. TPM = transcripts per million reads. (C) DTR expression in LCM. Cells gated on peritoneal LCM. C57bl/6 control used to account for autofluorescence in GFP channel. (D) Lavage fluid extracted. (E) Frequency of GATA6<sup>+</sup> LCM, SCM and inflammatory macrophages in *Wt1*<sup>eGFP.cre+/-</sup>;*Rosa26*<sup>LSL.DTR-/-</sup> and *Wt1*<sup>eGFP.cre+/-</sup>;*Rosa26*<sup>LSL.DTR+/-</sup> mice 24 hours after DT-administration. Inflammatory macrophages were excluded from analysis of *Wt1*<sup>eGFP.cre+/-</sup>;*Rosa26*<sup>LSL.DTR-/-</sup> mice due to low cell number (n/a) (F) Frequency of inflammatory macrophages in cavity spaces 24 hours after administration of DT in *Wt1*<sup>eGFP.cre+/-</sup>;*Rosa26*<sup>LSL.DTR-/-</sup> and *Wt1*<sup>eGFP.cre+/-</sup>;*Rosa26*<sup>LSL.DTR+/-</sup> mice 24 hours. LCM were gated as in Figure 6, SCM were gated as Live CD45<sup>+</sup>Gr1<sup>-</sup>SiglecF<sup>-</sup>CD11b<sup>+</sup>MHCII<sup>+</sup>F4/80<sup>-</sup>, inflammatory macrophages were gated as in CD45<sup>+</sup>Gr1<sup>-</sup>SiglecF<sup>-</sup>CD11b<sup>+</sup>MHCII<sup>-</sup>F4/80<sup>+</sup>. (G) Relative number of LCMs in littermate and *Adiponectin*<sup>cre+</sup>;*Rosa26*<sup>LSL.DTA+</sup> mice. LCMs were gated as CD11b<sup>+</sup>ICAM2<sup>+</sup>. (B) LCMs and omental *Wt1*<sup>+</sup> stromal cells were FACS sorted and RNAseq was performed. Data are from three biological replicates. Data are from 4 (D, E (pleural and pericardial cavity) or 2 experiments peritoneal cavity (E) or 3 experiments (G) or representative of 2-4 experiments (C). Each dot represents 1 mouse (F, G). Relative number was determined by multiplying the total cell number in the lavage by the percentage of CD11b<sup>+</sup>ICAM2<sup>+</sup>

cells. \* $p < 0.05$ , \*\* $p < 0.005$ , \*\*\*\* $p < 0.0001$ , as determined by unpaired student's t-test (E-G). Mean (F, G) or mean +SEM (B, D, E) are shown.

Buechler...Turley. Table S1

Related to Figure 5

<b>Peritoneal LCM hallmark genes</b>	<b><i>Wt1</i><sup>+</sup> stromal regulation</b>	<b>References</b>
<i>Gata6</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014)
<i>Rarb</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014)
<i>Padi4</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014)
<i>Cd93</i>		Gautier (2012), Okabe (2014), Gautier (2014)
<i>Serpib2</i>		Gautier (2012), Okabe (2014), Rosas (2014)
<i>F5</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014), Rosas (2014)
<i>Tgfb2</i>		Gautier (2012), Okabe (2014), Rosas (2014)
<i>Arg1</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014), Gundra (2014)
<i>Lyz1</i>		Gautier (2012), Okabe (2014), Gautier (2014)
<i>Flnb</i>		Gautier (2012), Okabe (2014), Rosas (2014)
<i>Ank</i>		Gautier (2012), Okabe (2014), Rosas (2014), Gundra (2014)
<i>Rai14</i>	<i>Wt1</i> <sup>+</sup> stroma-dependent	Gautier (2012), Okabe (2014), Rosas (2014)
<i>Hdc</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014), Gautier (2014)
<i>Wnt2</i>		Gautier (2012), Okabe (2014), Rosas (2014)
<i>Vmn2r26</i>		Gautier (2012), Okabe (2014), Rosas (2014)
<i>Apoc2</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014), Gundra (2014)
<i>Aqp9</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014), Rosas (2014)
<i>F13a1</i>		Gautier (2012), Okabe (2014), Gundra (2014)
<i>Egln3</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014), Rosas (2014)
<i>Stard13</i>		Gautier (2012), Okabe (2014), Rosas (2014)
<i>Marco</i>		Gautier (2012), Okabe (2014), Gundra (2014)
<i>Nfe2</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014)
<i>Fn1</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014)
<i>Prg4</i>		Gautier (2012), Okabe (2014)
<i>Icam2</i>	<i>Wt1</i> <sup>+</sup> stroma-dependent	Gautier (2012), Okabe (2014)

<i>Lrg1</i>		Gautier (2012), Okabe (2014)
<i>Emilin2</i>		Gautier (2012), Okabe (2014)
<i>Thbs1</i>		Gautier (2012), Okabe (2014)
<i>Saa3</i>		Gautier (2012), Okabe (2014)
<i>Calml4</i>		Gautier (2012), Okabe (2014)
<i>Mstr1</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Okabe (2014)
<i>Cgnl1</i>		Gautier (2012), Okabe (2014)
<i>Lama3</i>		Gautier (2012), Gundra (2014)
<i>Cxcl13</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Gundra (2014)
<i>Clec4d</i>		Gautier (2012), Gundra (2014)
<i>Cmah</i>		Gautier (2012), Rosas (2014)
<i>Fgfr1</i>	<i>Wt1</i> <sup>+</sup> stroma-dependent	Gautier (2012), Rosas (2014)
<i>Garnl3</i>		Gautier (2012), Rosas (2014)
<i>Selp</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Rosas (2014)
<i>1110032E23Rik</i>		Gautier (2012), Rosas (2014)
<i>Naip1</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Rosas (2014)
<i>Dnahc12</i>		Gautier (2012), Gautier (2014)
<i>Gbp1</i>		Gautier (2012), Gautier (2014)
<i>Car6</i>		Gautier (2012), Gautier (2014)
<i>Slfn4</i>		Gautier (2012), Gautier (2014)
<i>Sell</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2012), Gautier (2014)
<i>Hpse</i>		Gautier (2012), Gautier (2014)
<i>S100a4</i>		Gautier (2012), Gautier (2014)
<i>Slfn1</i>	<i>Wt1</i> <sup>+</sup> stroma-dependent	Gautier (2012), Gautier (2014)
<i>Tspan32</i>		Gautier (2012), Gautier (2014)
<i>F7</i>		Gautier (2012), Gautier (2014)
<i>Cav1</i>		Gautier (2012), Gautier (2014)
<i>Hp</i>		Gautier (2012), Okabe (2014), Gundra (2014)
<i>Gm1673</i>		Okabe (2014), Gundra (2014)
<i>Ltbp1</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Okabe (2014), Rosas (2014)
<i>Kcnn4</i>		Gautier (2014), Gundra (2014)
<i>Nuf2</i>		Gautier (2014), Gundra (2014)
<i>Apoc1</i>		Gautier (2014), Gundra (2014)
<i>Rgs2</i>		Gautier (2014), Gundra (2014)
<i>Ldhd</i>		Gautier (2014), Gundra (2014)
<i>Aspa</i>	<i>Wt1</i> <sup>+</sup> stroma-/RAR-dependent	Gautier (2014)

Table S1. List of peritoneal LCM hallmark genes. Gene name, whether expression of this gene in LCM requires *Wt1*<sup>+</sup> stromal cells, RAR signaling or both and references are listed.

Hallmark genes were derived from the references listed.