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## **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see <u>Authors & Referees</u> and the <u>Editorial Policy Checklist</u>.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
$\boxtimes$	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
$\boxtimes$	A description of all covariates tested
X	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
$\boxtimes$	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
$\boxtimes$	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
$\boxtimes$	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

## Software and code

Policy information about availability of computer code

Data collection Provide

Provide a description of all commercial, open source and custom code used to collect the data in this study, specifying the version used OR state that no software was used.

Data analysis

bcl2fastq 2.1.16 STAR 2.5.1 tophat2 2.10 PiGx-scRNAseq 0.0.6 R 3.5 with tied versions of: BioconductoR quasR edgeR Destiny ggplot pheatmap ComplexHeatmap

Seurat 2.1 Python 3.6 Scanpy 1.3.6 Velocyto 0.17

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

## Data

Policy information about <u>availability of data</u>

Commonly misidentified lines

(See ICLAC register)

N/A

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Raw sequencing reads as well as raw readcounts for both the bulk RNA-seq and the single-cell RNA-seq, along with normalized counts and t-SNE coordinates and cell cycle information, are available in the NCBI GEO repository, accession number GSE123782.

Field-spe	ocific ro	norting		
<u> </u>		the best fit for your research. If you are not sure, read the appropriate sections before making your selection.		
Life sciences	Ве	ehavioural & social sciences Ecological, evolutionary & environmental sciences		
For a reference copy of	the document with a	all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>		
Life scier	nces stu	ıdy design		
All studies must dis	sclose on these	points even when the disclosure is negative.		
Sample size		Two independent experiments were performed with two biological replicates for four time points each. On average about 5000 cells were measured per sample. Selection of cells is a random process.		
Data exclusions	No datasets wer	atasets were excluded from the data submission.		
Replication	Described effect	ibed effects were reproducible.		
Randomization	Samples do not	o not have covariates that need to be taken into account.		
Blinding	Blinding was not relevant in this study.			
We require informati	ion from authors a	Decific materials, systems and methods about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.		
Materials & experimental systems  n/a Involved in the study		n/a Involved in the study		
Antibodies		ChIP-seq		
Eukaryotic cell lines		Flow cytometry		
Palaeontology MRI-based neuroimaging				
Animals ar	nd other organism	S		
Human research participants				
Clinical dat	ta			
Eukaryotic c	ell lines			
Policy information	about <u>cell lines</u>			
Cell line source(s)		Primary human fibroblasts were obtained from Promocell (C-12300), HEK 293 cells from Thermo Fisher (R78007)		
Authentication Cell lines were no		Cell lines were not validated further in our laboratory after purchase.		
Mycoplasma contamination Cells were regular		Cells were regulary tested for mycoplasm contamination using the Lonza MycoAlert kit		