

Supplementary information

**Antitumor immunity augments the therapeutic effects of p53 activation
on acute myeloid leukemia.**

Hayashi et al.

Supplementary Table 1. Information of each human AML sample used in this study.

Viability of cells treated with DS-5272 was measured by WST-1 assay. Results are normalized to the viability of DMSO-treated cells, set at 1(n=3). Data are shown as the average and SD of duplicate wells.

| Compound | | Concentration (µM) | | | | | | | | | | Genomic Alterations |
|-----------|---------------|--------------------|-------|-------|-------|-------|-------|-------|-------|-------|------|---|
| DS-5272 | | 2.048 | 1.024 | 0.512 | 0.256 | 0.128 | 0.064 | 0.032 | 0.016 | 0.008 | 0 | |
| Sample ID | | | | | | | | | | | | |
| AML-(1) | Avg (%Viable) | 103% | 129% | 141% | 118% | 112% | 128% | 115% | 108% | 95% | 100% | FLT3 D835E – subclonal # , D835V – subclonal # , D835Y – subclonal # # KIT D816V ASXL1 G645fs*58 CBL deletion exon 9 CHEK2 T367fs*15 PTPN11 D61V , inv(3) |
| | St. Dev | 13.2% | 25.6% | 41.6% | 7.7% | 12.4% | 26.2% | 2.8% | 1.9% | 11.6% | 0.8% | |
| AML-(2) | Avg (%Viable) | 95% | 106% | 94% | 92% | 94% | 96% | 94% | 100% | 92% | 100% | CDKN2A/B p16INK4a loss and p14ARF loss exon 1 and CDKN2B loss |
| | St. Dev | 22.1% | 8.5% | 7.8% | 2.6% | 13.1% | 8.1% | 1.3% | 8.6% | 2.0% | 7.8% | |
| AML-(3) | Avg (%Viable) | 54% | 77% | 85% | 92% | 101% | 113% | 99% | 99% | 105% | 100% | KRAS G12A – subclonal # NRAS Q61H – subclonal # KMT2A (MLL) MLL-MLLT4 (AF6) fusion PTPN11 E76K – subclonal # , S502L – subclonal # |
| | St. Dev | 2.6% | 6.3% | 10.8% | 13.0% | 12.6% | 9.3% | 0.3% | 26.3% | 13.1% | 3.3% | |
| AML-(4) | Avg (%Viable) | 2% | 2% | 17% | 48% | 76% | 79% | 84% | 90% | 98% | 100% | t(7;12)(q36;p13) MNX1/ETV6 fusion PTPN11 p.D61V c.182A>T |
| | St. Dev | 0.3% | 1.2% | 8.6% | 15.4% | 13.3% | 4.1% | 7.9% | 1.3% | 17.7% | 6.2% | |
| AML-(5) | Avg (%Viable) | 40% | 62% | 68% | 76% | 93% | 87% | 107% | 98% | 107% | 100% | MLL MLL-MLLT3 (AF9) fusion PC R583H – subclonal # WT1 R462Q – subclonal# |
| | St. Dev | 1.0% | 7.1% | 4.9% | 0.1% | 10.9% | 17.7% | 3.9% | 10.8% | 11.5% | 9.4% | |
| AML-(6) | Avg (%Viable) | 2% | 9% | 38% | 45% | 73% | 89% | 82% | 95% | 93% | 100% | FLT3 D835A – subclonal # , D835E – subclonal # , D835Y –subclonal #, # I836del – subclonal#, V491L – subclonal#, V579A – subclonal# KRAS G13D MLL MLL-MVB12B fusion MLLT10 LL-MLLT10 (AF10) fusion |
| | St. Dev | 0.6% | 0.0% | 0.5% | 3.1% | 13.5% | 1.9% | 3.7% | 3.7% | 1.0% | 2.7% | |
| AML-(7) | Avg (%Viable) | 33% | 49% | 60% | 67% | 78% | 83% | 94% | 92% | 91% | 100% | MLL inv(11)(p11.2q23) ; HPIM- β chain regulatory sequence on 7q fusion |
| | St. Dev | 3.2% | 0.8% | 2.4% | 1.0% | 5.2% | 1.5% | 11.1% | 6.9% | 6.7% | 4.9% | |
| AML-(8) | Avg (%Viable) | 7% | 18% | 31% | 49% | 70% | 70% | 86% | 96% | 90% | 100% | CBFA2T3-GLIS2 rearrangement |
| | St. Dev | 1.0% | 1.5% | 1.9% | 3.2% | 10.9% | 2.5% | 0.6% | 8.7% | 3.8% | 4.2% | |
| AML-(9) | Avg (%Viable) | 12% | 18% | 38% | 58% | 74% | 75% | 105% | 87% | 96% | 100% | MLL MLL-MLLT3 (AF9) fusion |
| | St. Dev | 2.2% | 3.4% | 7.0% | 0.1% | 2.1% | 9.2% | 8.3% | 3.1% | 13.5% | 4.4% | |
| AML-(10) | Avg (%Viable) | 127% | 124% | 106% | 112% | 108% | 106% | 105% | 101% | 104% | 100% | NF1 Q1775* PTPN11 A461G – subclonal # CDKN2A/B loss ETV6 loss MLLT10 PICALM-MLLT10 fusion PHF6 R274Q TP53 K164E – subclonal # , Y126* |
| | St. Dev | 11.1% | 9.5% | 12.4% | 3.5% | 3.9% | 10.7% | 11.4% | 3.5% | 6.2% | 9.7% | |
| AML-(11) | Avg (%Viable) | 17% | 29% | 41% | 53% | 72% | 83% | 89% | 97% | 95% | 100% | DNMT3A R882C FLT3 L576_Q577ins17 PTPN11 D61Y NPM1 W288fs*10+ WT1 A382fs*11, A382fs*4 |
| | St. Dev | 1.6% | 3.9% | 3.9% | 0.7% | 1.7% | 2.9% | 9.8% | 2.0% | 5.0% | 0.7% | |

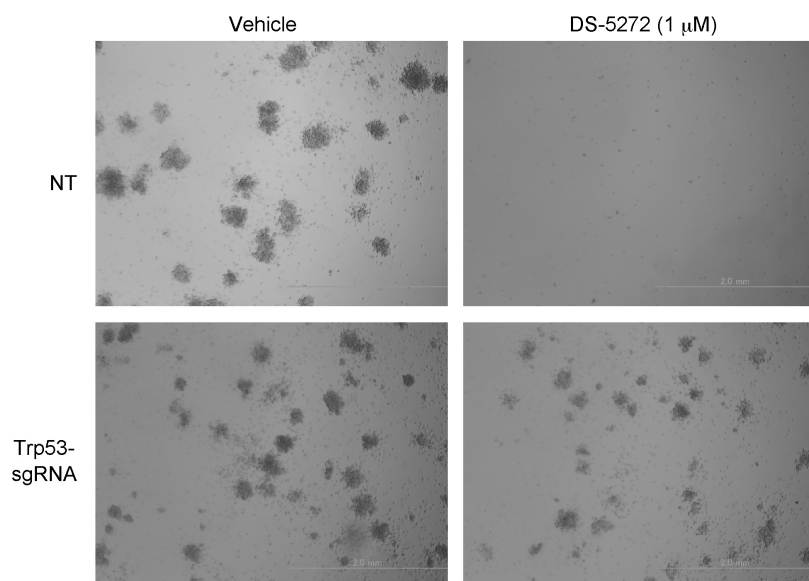
Supplemental Table 2. Sequences of primers used in this study.

| Name | Forward (5'-3') | Reverse (5'-3') | Application |
|-----------------|---------------------------|---------------------------|-------------|
| <i>Cdkn1a</i> | ATCACCAGGATTGGACATGG | CGGTGTCAGAGTCTAGGGGA | qRT-PCR |
| <i>Mdm2</i> | CTGCTCTCACTCAGCGATGT | TCTGTGAAGGAGCACAGGAA | qRT-PCR |
| <i>Bbc3</i> | TGTCGATGCTGCTCTTCTTG | GTGTGGAGGAGGAGGAGTGG | qRT-PCR |
| <i>Gadd45a</i> | AGACCGGAAAGGATGGACAC | GTACACGCCGACCGTAATG | qRT-PCR |
| <i>Gapdh</i> | TGTGTCCGTCGTCGTGGATCTGA | TTGCTGTTGAAGTCGCAGGAG | qRT-PCR |
| <i>Trp53 WT</i> | GTTATGCATCCATACAGTACA | CCGCAGGATTTACAGACACC | Genotyping |
| <i>Trp53 KO</i> | ACGTGTTTGTTACCTCTGC | TCGCCTTCTTGACGAGTTCT | Genotyping |
| <i>Hif1a</i> | AGATAGGAAGATCTCCAGTTCAGT | GAAAACTGTCTGTAACCTTCATTCC | Genotyping |
| <i>Cre-ERT2</i> | TCGATGCAACGAGTGATGAG | TTCGGCTATACGTAACAGGG | Genotyping |
| <i>NT</i> | caccgCGCTTCCGCGGCCGTTCAA | aaacTTGAACGGGCCGGAAGCGc | sgRNA |
| <i>sgTrp53</i> | caccgGAAGTCACAGCACATGACGG | aaacCCGTCATGTGCTGTGACTTCc | sgRNA |
| <i>sgPD-L1</i> | caccgGCCTGCTGTCACTTGCTACG | aaacCGTAGCAAGTGACAGCAGGCc | sgRNA |

Supplemental Table 3. Antibodies used for Mass Cytometry analysis.

| Metal label | Specificity | Clone | Vendor | Cat No. |
|--------------------|--|-----------------|---------------|----------------|
| 141Pr | Ly-6G/C(Gr1) | RB6-8C5 | Fluidigm | 3141005B |
| 143Nd | CD41 | MWreg30 | Fluidigm | 3143009B |
| 145Nd | CD4 | RM4-5 | Fluidigm | 3145002B |
| 146Nd | VCAM1 | 429 (MVCAM.A) | BioLegend | 105702 |
| 147Sm | CD45.2 | 104 | Fluidigm | 3147004B |
| 149Sm | CXCR4(CD184) | L276F12 | Biolegend | 146502 |
| 150Nd | HIF1 α | H206 | Santa Claus | sc-10790 |
| 151Eu | CD49d (Integrin alpha 4) | R1-2 | Fluidigm | 3151016B |
| 152Sm | phospho-Akt [S473] | D9E | Fluidigm | 3152005A |
| 153Eu | phospho-STAT1 [Y701] | 58D6 | Fluidigm | 3153003A |
| 156Gd | phospho-p38 MAP Kinase [T180/Y182] | D3F9 | Fluidigm | 3156002A |
| 158Gd | phospho-STAT3 [Y705] | 4/P-STAT3 | Fluidigm | 3158005A |
| 159Tb | phospho-MAPKAP Kinase2 [T334] | 27B7 | Fluidigm | 3159010A |
| 160Gd | CD45R (B220) | RA3-6B2 | Fluidigm | 3160012B |
| 161Dy | Ki-67 | B56 | Fluidigm | 3161007B |
| 162Dy | p53 (For whole protein) | X77 | Abcam | ab16465 |
| 163Dy | phospho-mTOR [S2448] | EPR426(2) | Abcam | ab109268 |
| 165Ho | beta-Catenin | D13A1 | Fluidigm | 3165027A |
| 167Er | EPCR (CD201) | eBio1560 (1560) | eBioscience | 16-2012-83 |
| 168Er | CD8a | 53-6.7 | Fluidigm | 3168003B |
| 169Tm | Ly-6A/E (Sca-1) | D7 | Fluidigm | 3169015B |
| 170Er | CD49b | HMa2 | Fluidigm | 3170008B |
| 171Yb | phospho-ERK 1/2 [T202/Y204] | D13.14.4E | Fluidigm | 3171010A |
| 172Yb | CD11b (Mac-1) | M1/70 | Fluidigm | 3172012B |
| 173Yb | CD117 (ckit) | 2B8 | Fluidigm | 3173004B |
| 174Yb | phospho-STAT4 | 38/p-Stat4 | Fluidigm | 3174005A |
| 175Lu | PGC1 alpha | polyclonal | Abcam | ab54481 |
| 191Ir | Cell-ID™ Intercalator-Ir (DNA staining) | NA | Fluidigm | 201192A |
| 193Ir | Cell-ID™ Intercalator-Ir (DNA staining) | NA | Fluidigm | 201192A |
| 198Pt | Cell-ID™ Cisplatin (Live/Dead Cell staining) | NA | Fluidigm | 201198 |
| NA: not available | | | | |

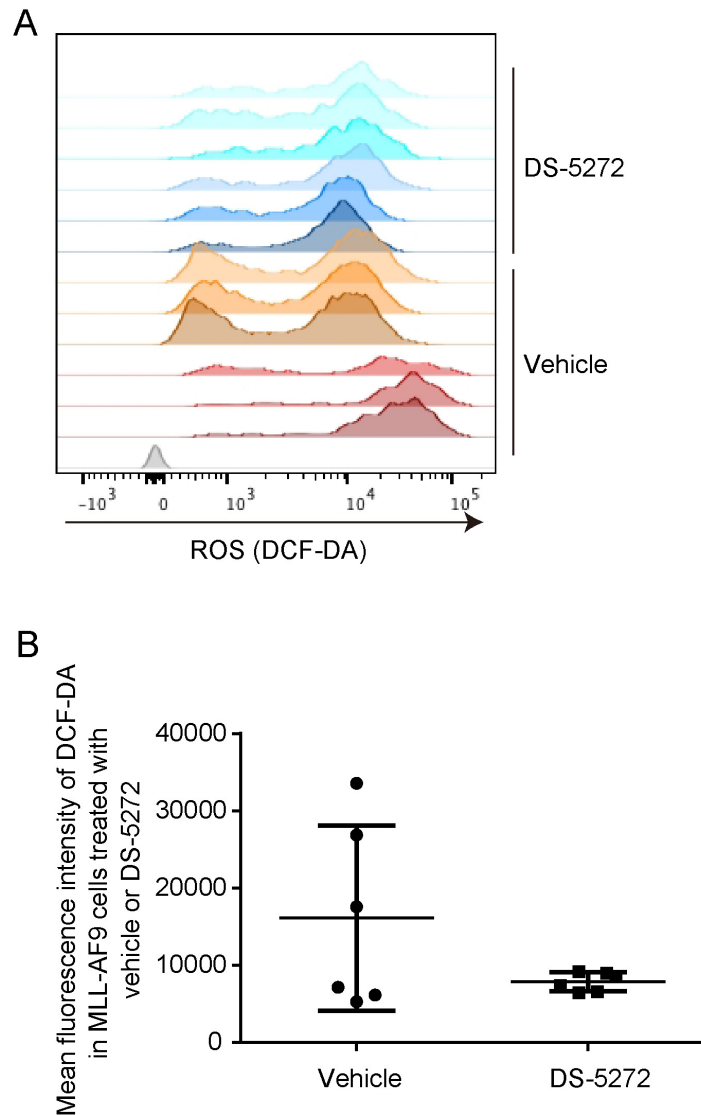
Supplementary Figure 1



Supplementary Figure 1. p53-deficient MLL-AF9 cells are resistant to DS-5272.

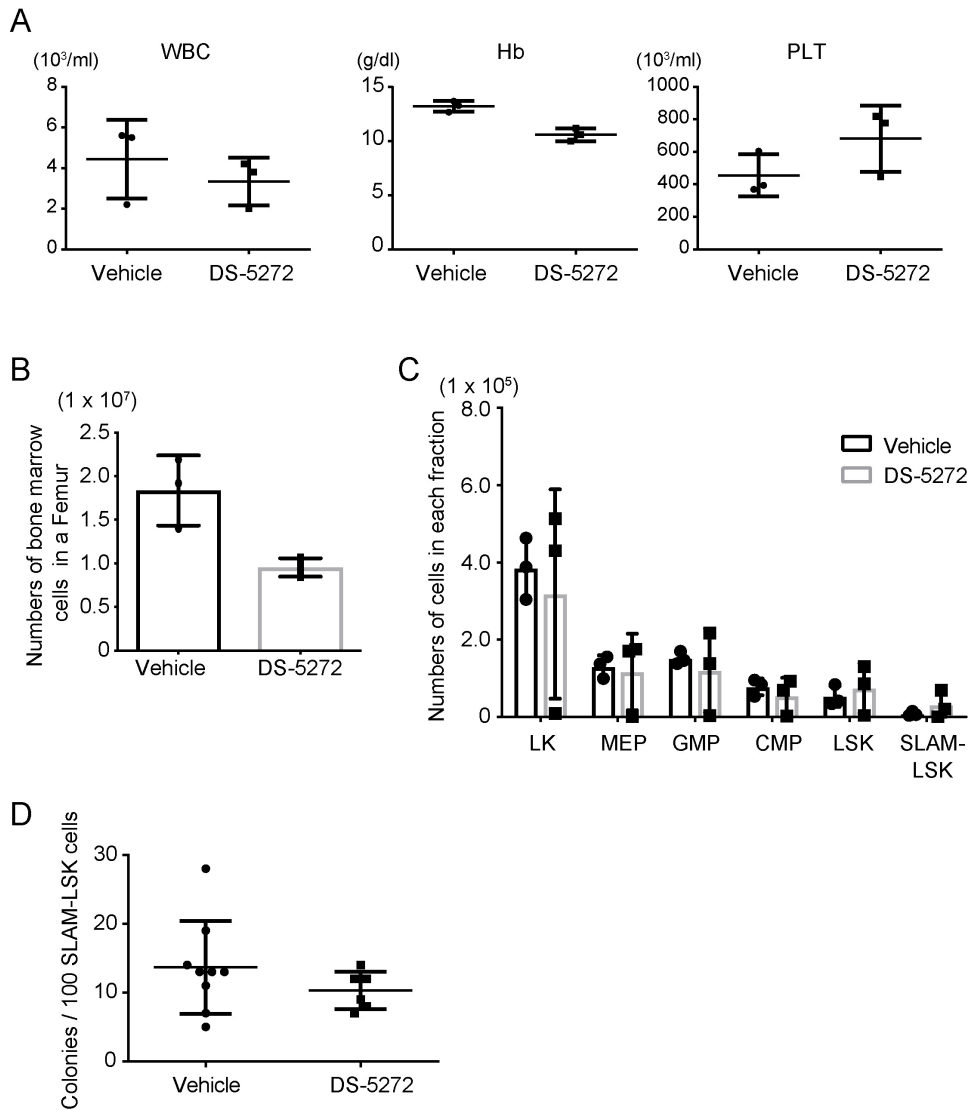
5×10^4 MLL-AF9 cells, transduced with Cas9 together with non-targeting (NT) or Trp53-targeting sgRNA, were plated in MethoCult™ M3234 supplemented with 10 ng/ml mouse SCF, 10 ng/ml, mouse GM-CSF, 10 ng/ml mouse IL-3, and 10 ng/ml mouse IL-6, with/without DS-5272 (1 mM) for 5 days. Representative photos of each plate are shown (Scale bars: 2 mm).

Supplementary Figure 2



Supplementary Figure 2. DS-5272 does not induce ROS overproduction in MLL-AF9 cells. **A.** FACS plots showing intensity of ROS (DCF-DA) in MLL-AF9 leukemia cells. Leukemia cells were isolated from bone marrow 24 hours after treatment with vehicle control or DS-5272. **B.** Quantification of MFI of ROS (DCF-DA) in MLL-AF9 leukemia cells. Data are shown as mean \pm s.d.

Supplementary Figure 3

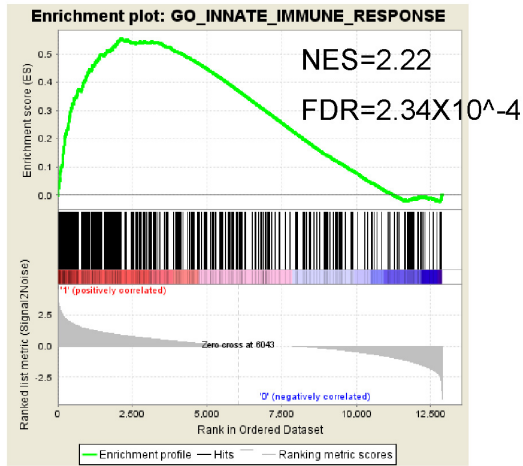


Supplementary Figure 3. DS-5272 does not significantly inhibit normal hematopoiesis.

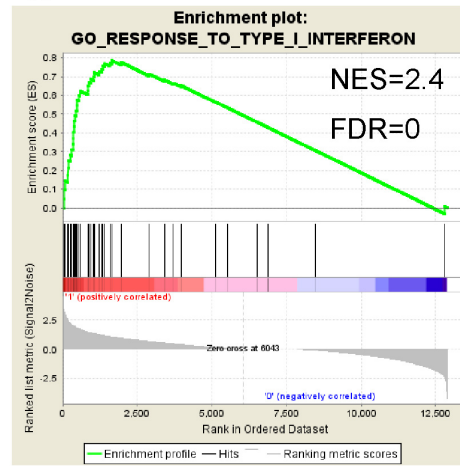
Female 8-weeks-old C57BL/6 mice were treated with vehicle or DS-5272 for 2 weeks (80mg/kg, 3 times per week). (A-C). Shown are WBC counts, Hb levels and PLT counts (A), numbers of bone marrow cells in a femur (B), and numbers of hematopoietic stem and progenitor cells (C) in mice treated with vehicle or DS-5272. D. 100 SLAM-LSK cells were sorted and were used for colony forming assay. LK: lineage⁻cKit⁺, MEP: megakaryocyte/erythroid progenitor, GMP: granulocyte/monocyte progenitor, CMP: common myeloid progenitor, LSK: lineage⁻Scal⁺cKit⁺, SLAM-LSK: CD150⁺CD48⁺lineage⁻Scal⁺cKit⁺. Data are shown as mean ± s.d.

Supplementary Figure 4

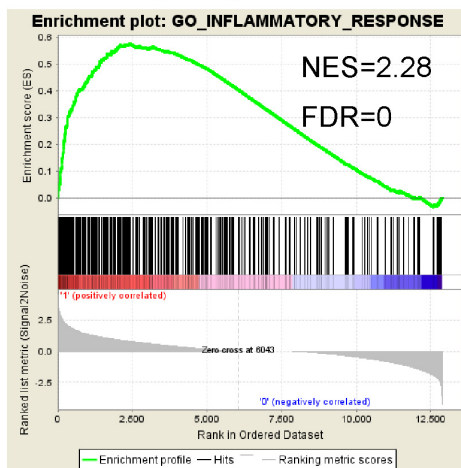
Innate Immune Response



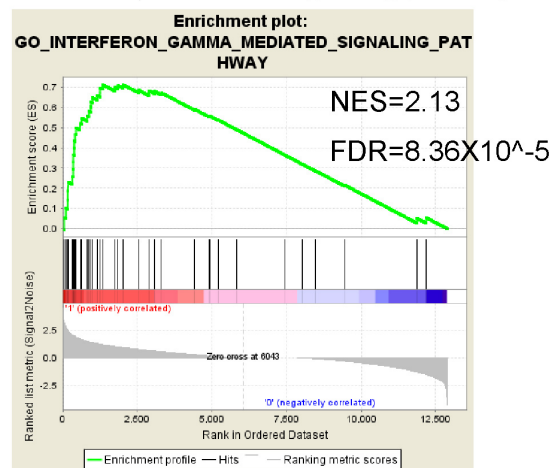
Type I Interferon



Inflammatory Response

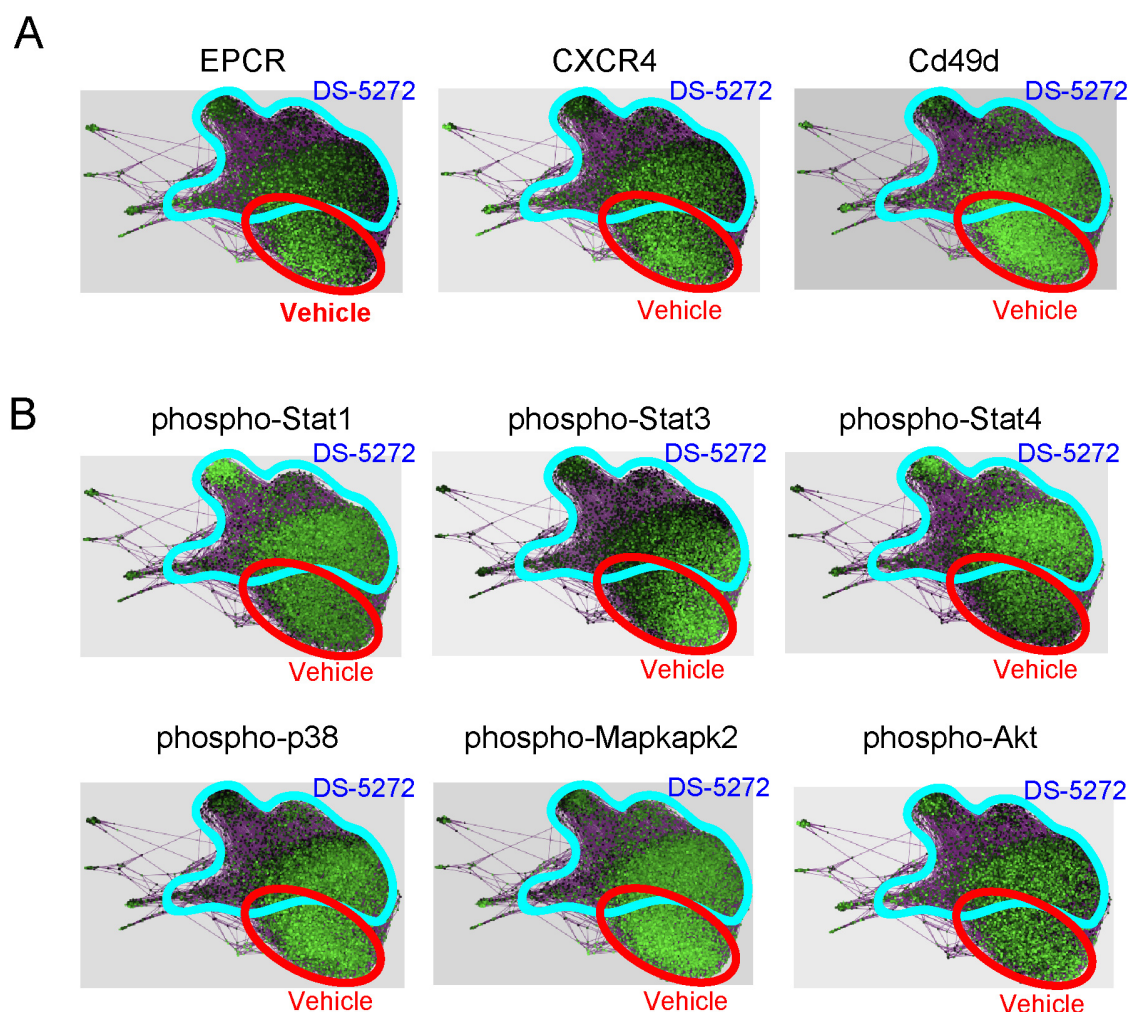


Interferon-γ-Mediated Signaling pathway



Supplementary Figure 4. GSEA of upregulated genes in MLL-AF9 cells treated with DS-5272. GSEA revealed that genes related to Innate Immune Response, Type 1 Interferon, Inflammatory Response, and Interferon- γ -Mediated Signaling Pathway were upregulated in MLL-AF9 cells treated with DS-5272 compared with those treated with vehicle.

Supplementary Figure 5

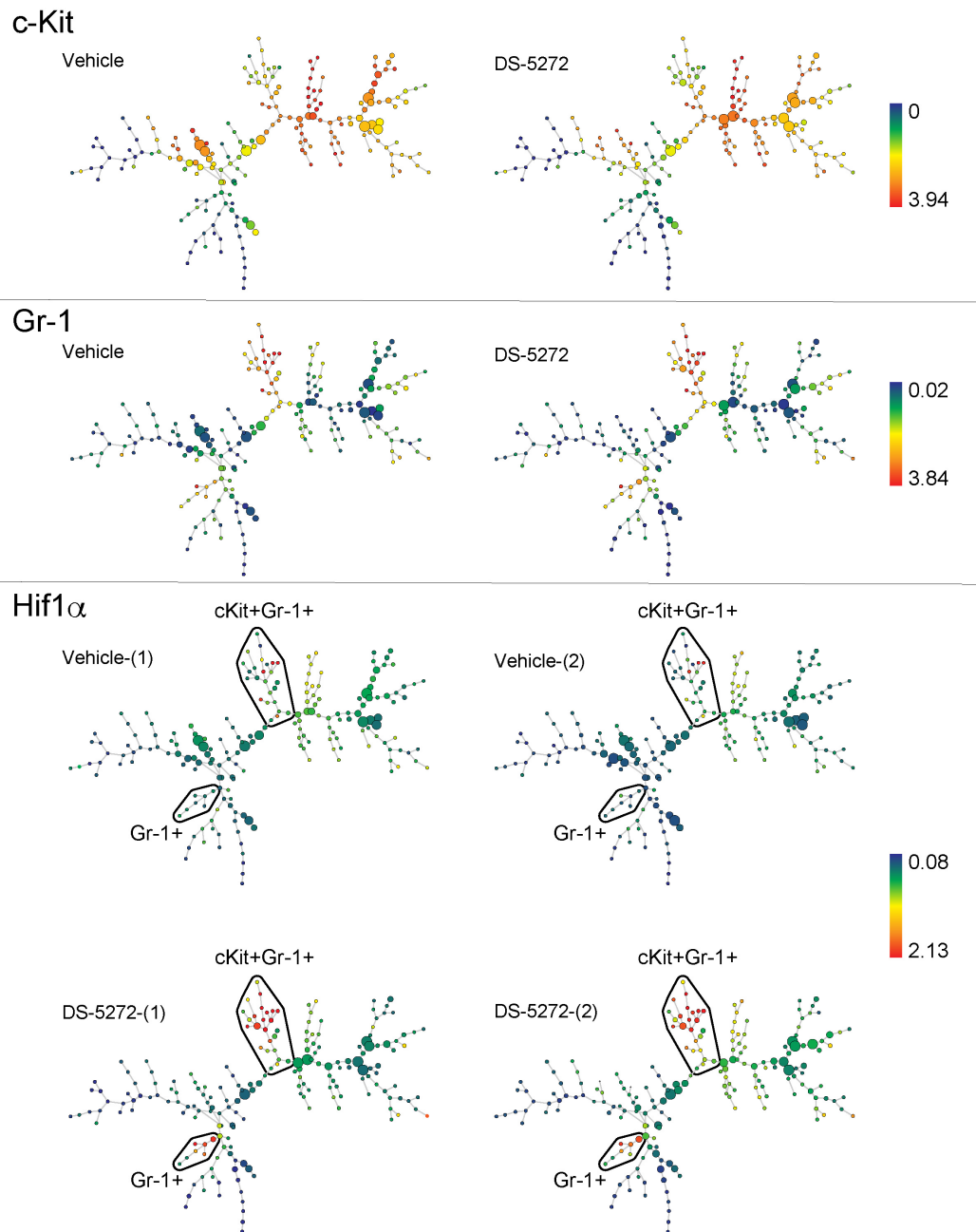


Supplementary Figure 5. SPRING plots of MLL-AF9 cells treated with Vehicle or DS-5272.

A, B. 25-dimensional analysis using SPRING

(<https://kleintools.hms.harvard.edu/tools/spring.html>) of bone marrow cells from MLL-AF9 leukemia mice treated with vehicle and DS-5272 for 24 hours (n=2/condition). Differences in the location of cells within the SPRING map resulted from changes in protein expression. The regions enclosed by the red and blue curves represent the cells treated with vehicle or DS-5272, respectively. Cell surface expression of adhesion molecules (EPCR, CXCR, Cd49d, **A**) and intracellular signaling pathway markers (phospho-Stat1, phospho-Stat3, phospho-Stat4, phospho-p38, phospho-Mapkapk2, phospho-Akt, **B**) in vehicle- and DS-5272-treated MLL-AF9 leukemia cells are shown.

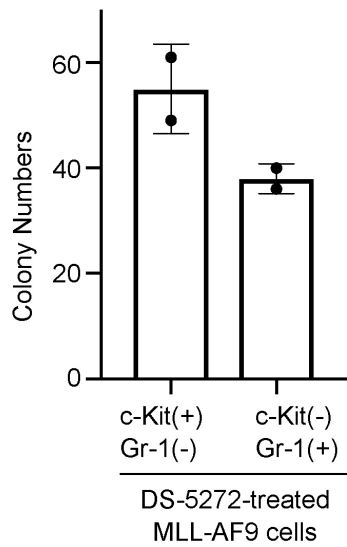
Supplementary Figure 6



Supplementary Figure 6. SPADE plots of MLL-AF9 cells treated with Vehicle or DS-5272.

SPADE plots of bone marrow cells from MLL-AF9 leukemia mice treated with vehicle or DS-5272 for 24 hours ($n=2$ /condition). MLL-AF9 leukemia cells were grouped based on cell surface expression of c-Kit and Gr-1. Protein expression of HIF1 α in vehicle- and DS-5272-treated MLL-AF9 leukemia cells are shown.

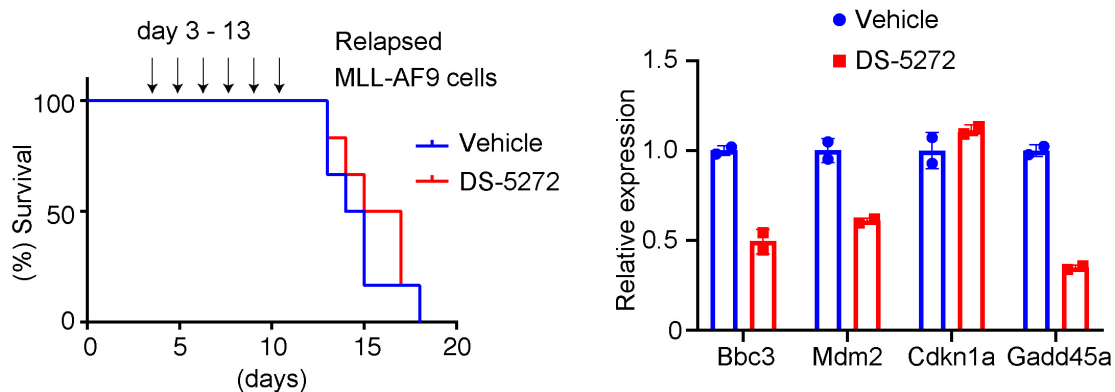
Supplementary Figure 7



Supplementary Figure 7. Gr-1⁺ MLL-AF9 cells retain clonogenic activity.

MLL-AF9 leukemia mice were treated with DS-5272, and spleen cells were collected from them 24 hours after the treatment. GFP⁺c-Kit⁺Gr-1⁻ cells and GFP⁺c-Kit⁻Gr-1⁺ cells were isolated by fluorescence-activated cell sorting and colony formation assays were performed in duplicates. Data are shown as mean \pm s.d.

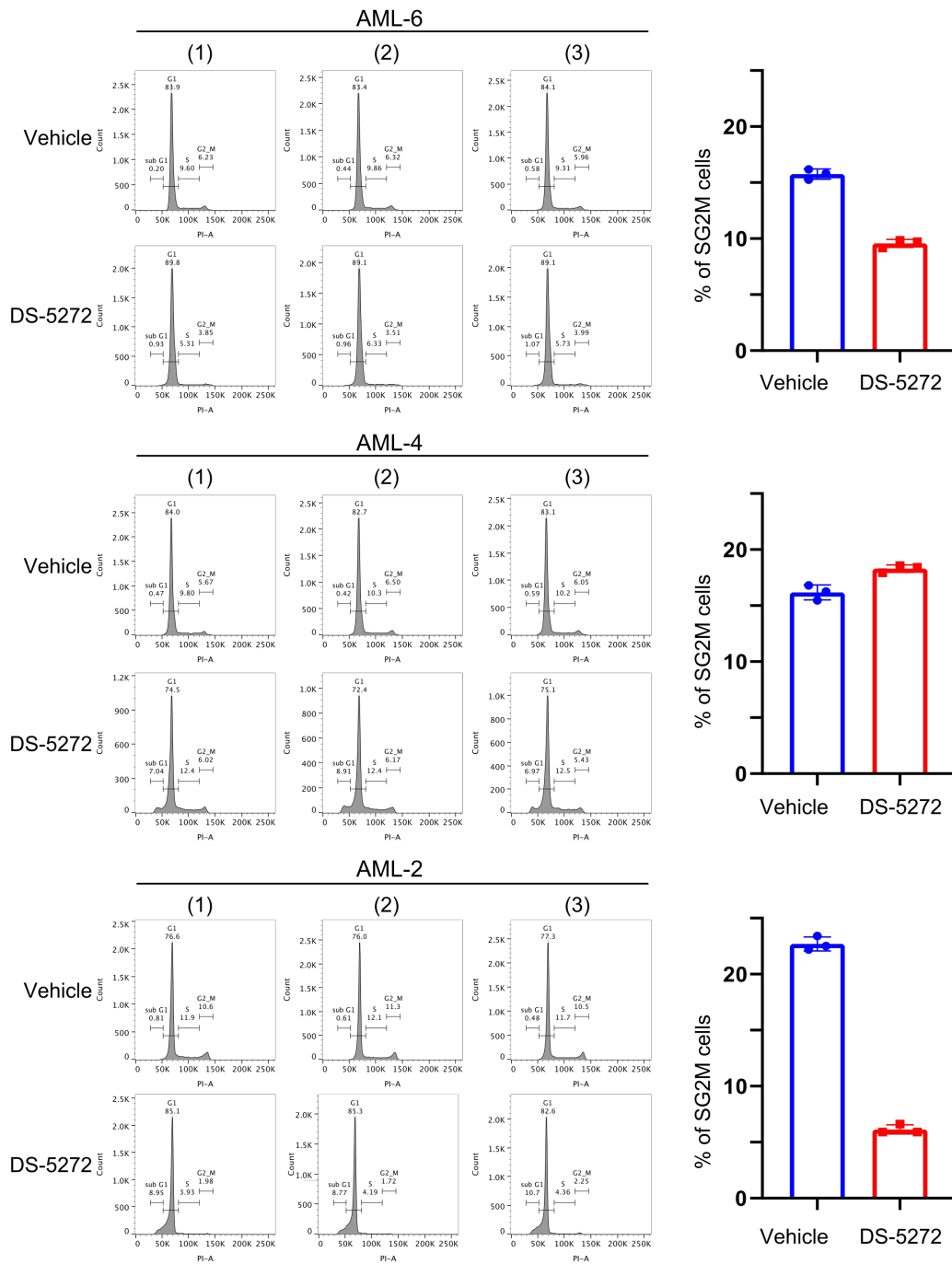
Supplementary Figure 8



Supplementary Figure 8. Relapsed MLL-AF9 cells are resistant to DS-5272.

A. MLL-AF9 cells were collected from mice that became moribund after DS-5272 treatment, and were serially transplanted into recipient mice. The mice injected with the relapsed MLL-AF9 leukemia cells after DS-5272 treatment were again treated with vehicle or DS-5272 from day 3 to day 13. Kaplan-Meier survival curves of these mice are shown. DS-5272 showed no inhibitory effects on the relapsed MLL-AF9 leukemia cells. **B.** GFP⁺ MLL-AF9 leukemia cells were collected from bone marrows of vehicle- or DS-5272-treated mice. Expression levels of p53-target genes in MLL-AF9 cells were assessed by qPCR. The relapsed MLL-AF9 cells were no longer responsive to DS-5272 treatment. Data are shown as mean \pm s.d.

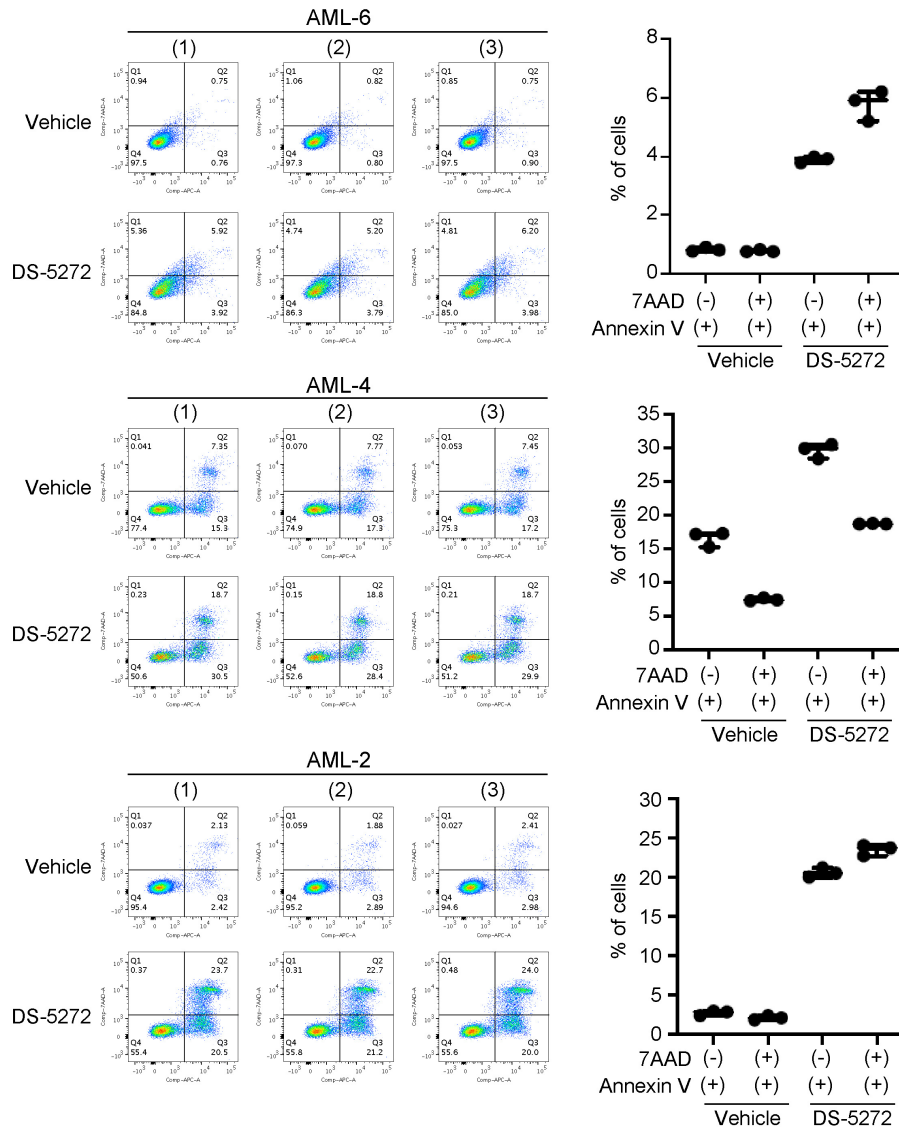
Supplementary Figure 9



Supplementary Figure 9. Cell-cycle status of human AML cells treated with DS-5272.

Cell-cycle status was assessed after 3 days culture with vehicle or DS-5272 (250 nM) using human AML-(2), (4) and (6) cells. Shown are cell cycle profiles assessed by FACS with PI staining of DNA. The numbers indicate the percentages of cells in the sub-G1, G1, S and G2/M phases. See also Figure 6B. Data are shown as mean \pm s.d.

Supplementary Figure 10

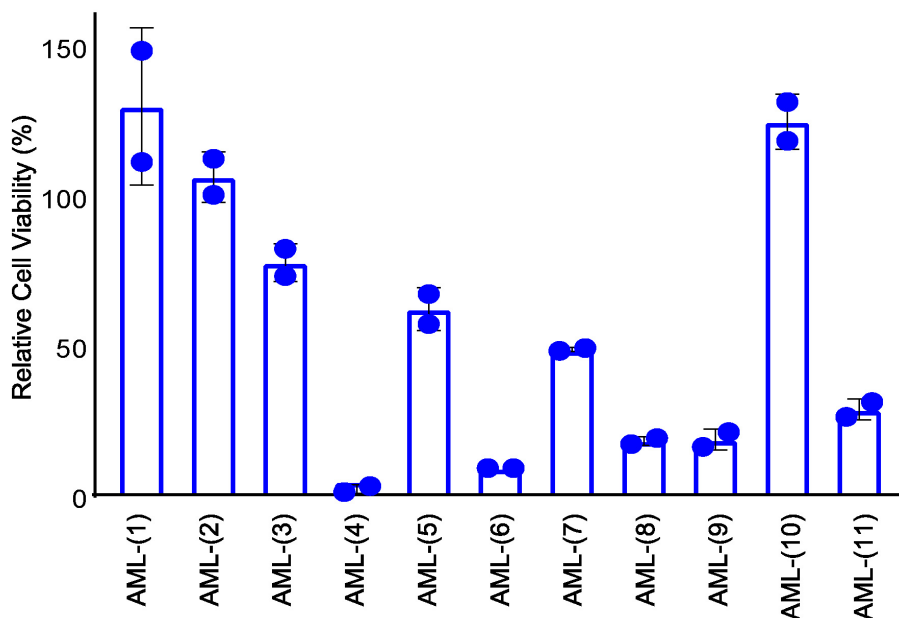


Supplementary Figure 10. Apoptosis of human AML cells treated with DS-5272.

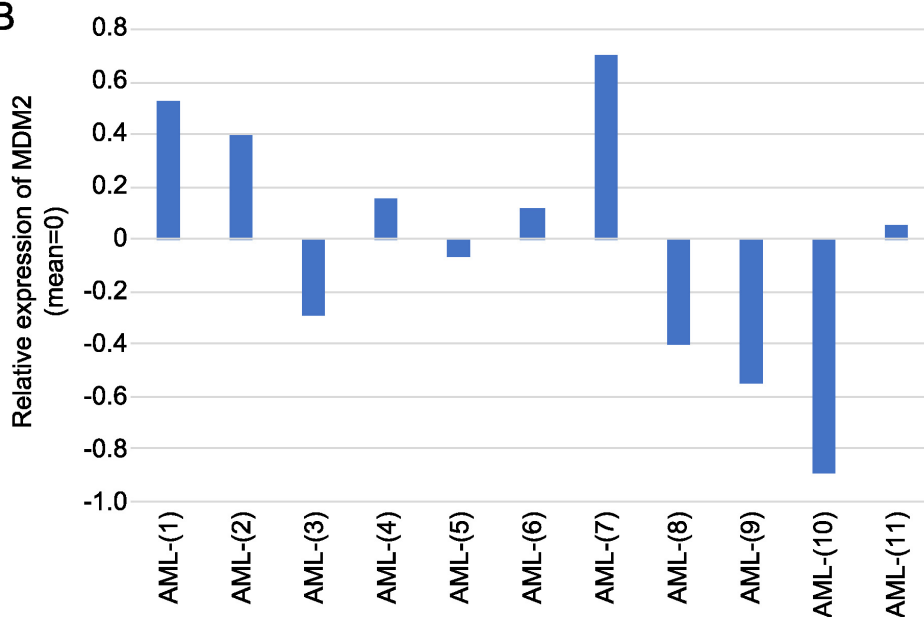
Apoptosis was assessed after 3 days culture with vehicle or DS-5272 (250 nM) using human AML-(2), (4) and (6) cells. Shown are FACS profiles of Annexin V and PI staining. The numbers indicate the percentages of Annexin V+ cells. See also Figure 6B. Data are shown as mean \pm s.d.

Supplementary Figure 11

A



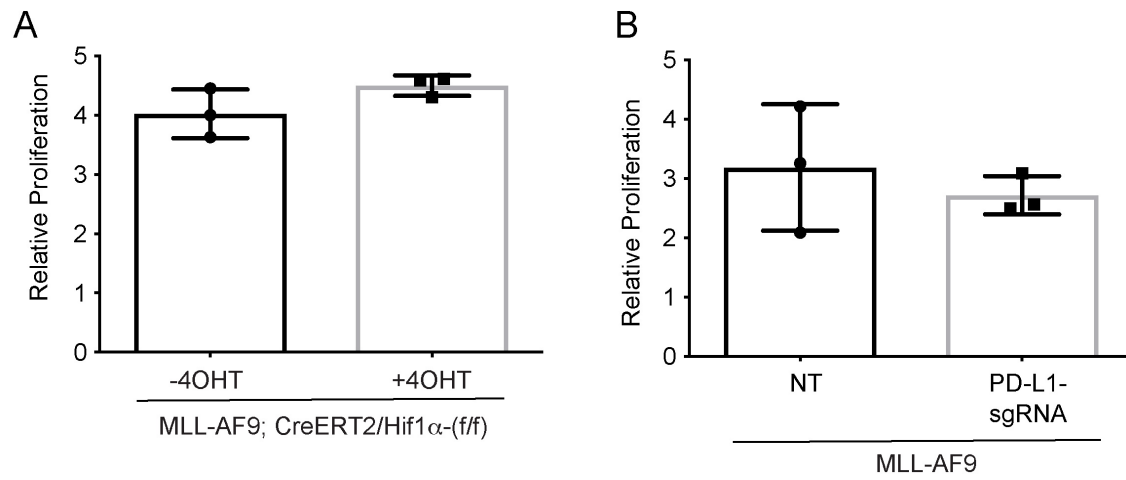
B



Supplementary Figure 11. MDM2 expression does not predict the sensitivity of MLL-AF9 cells to DS-5272.

A. Viability of eleven PDX-derived human AML cultures treated with DS-5272 (1 μ M) was measured by MTS assay. Results are normalized to the viability of DMSO-treated cells, set at 1 (n = 3). AML-(10) has a TP53 mutation. Data are shown as the mean \pm SD of triplicate wells. **B.** Relative MDM2 levels in the eleven PDX-derived human AML cells. See also Figure 6A and Supplementary Table 1.

Supplementary Figure 12

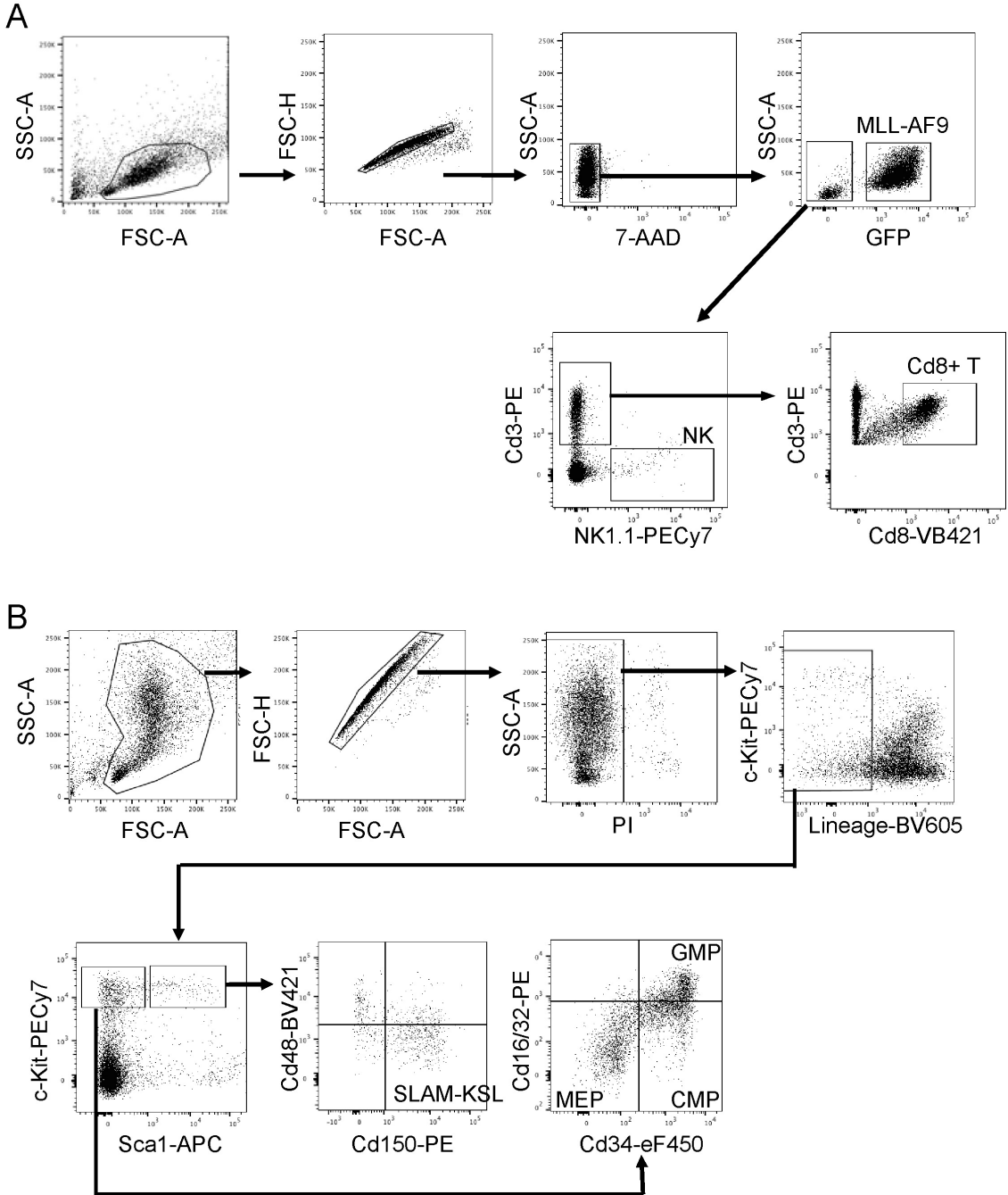


Supplementary Figure 12. Depletion of PD-L1 or Hif1 α does not alter the growth of MLL-AF9 cells.

A, B. Effects of HIF1 α (**A**) or PD-L1 (**B**) depletion on the growth of MLL-AF9 cells were measured by WST-8 assay. Depletion of these genes did not change the *in vitro* growth of MLL-AF9 cells. Data are shown as mean \pm s.d.

Supplementary Figure 13

- A. Gating strategy for MLL-AF9 cells, NK cells and Cd8+ T cells.
- B. Gating strategy for SLAM-LSK, CMP, GMP and MEP cells.



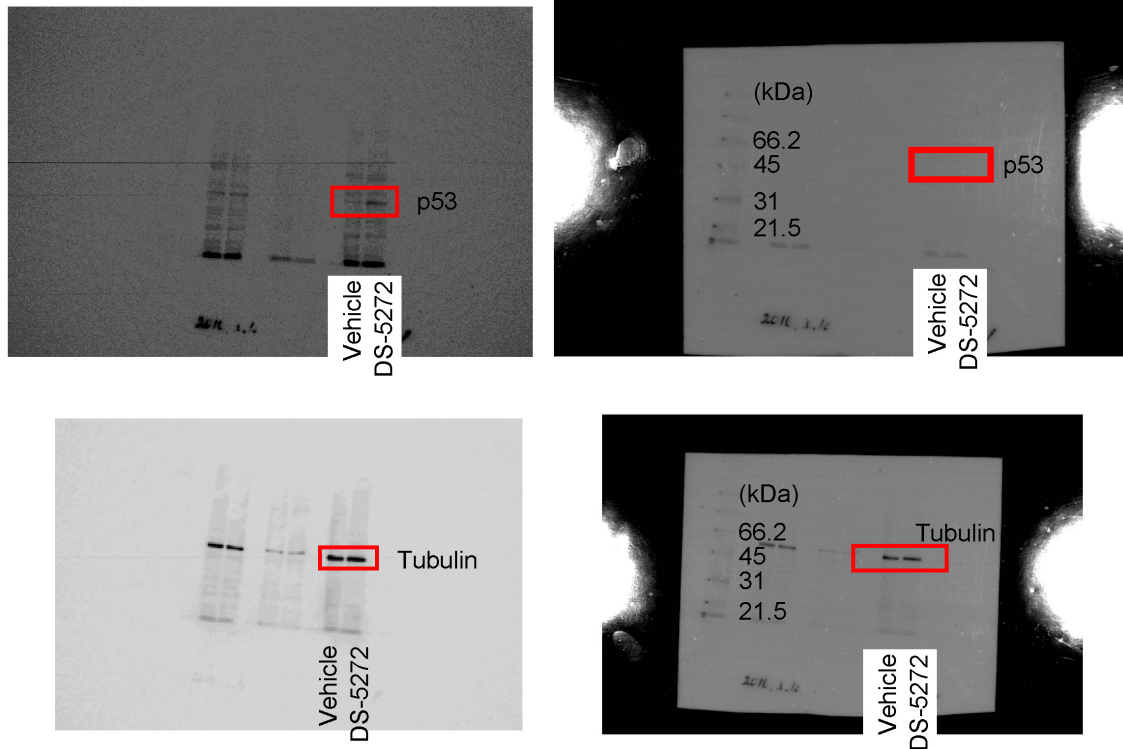
Supplementary Figure 13. Gating strategies used for flow cytometry.

Supplementary Figure 14

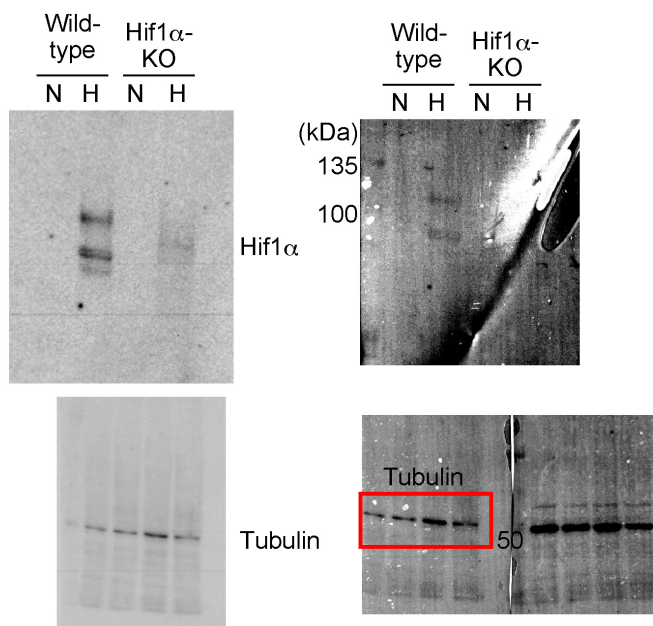
A. Full blots for Figure 1C.

B. Full blots for Figure 4B.

A



B



Supplementary Figure 14. Uncropped immunoblot images of Figure 1C and 4B.