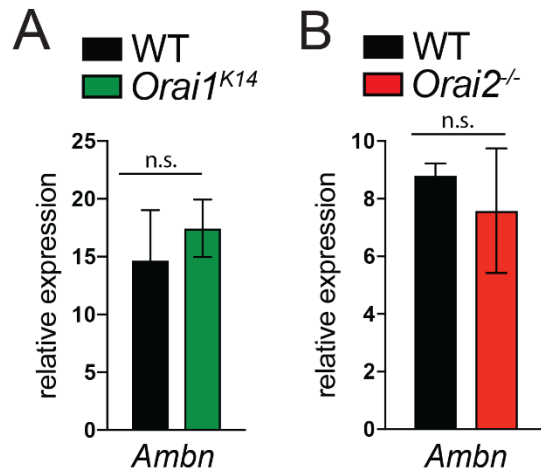
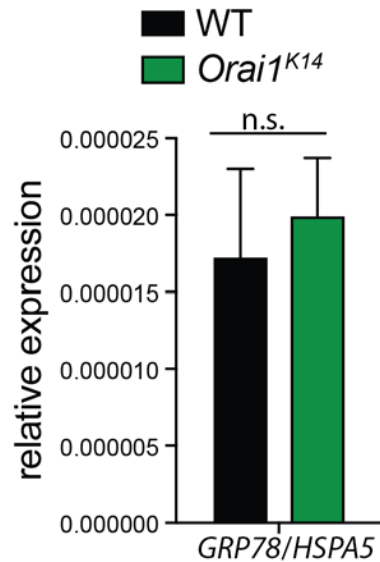


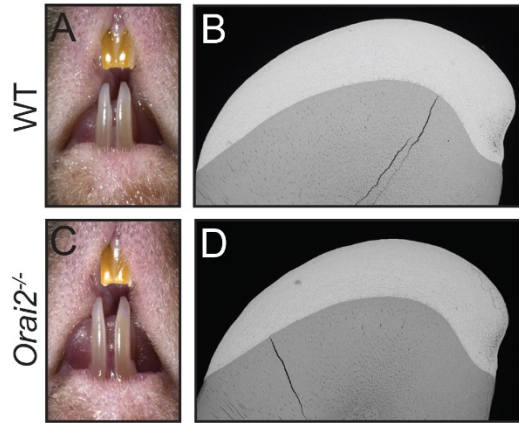
## Supplementary Figures



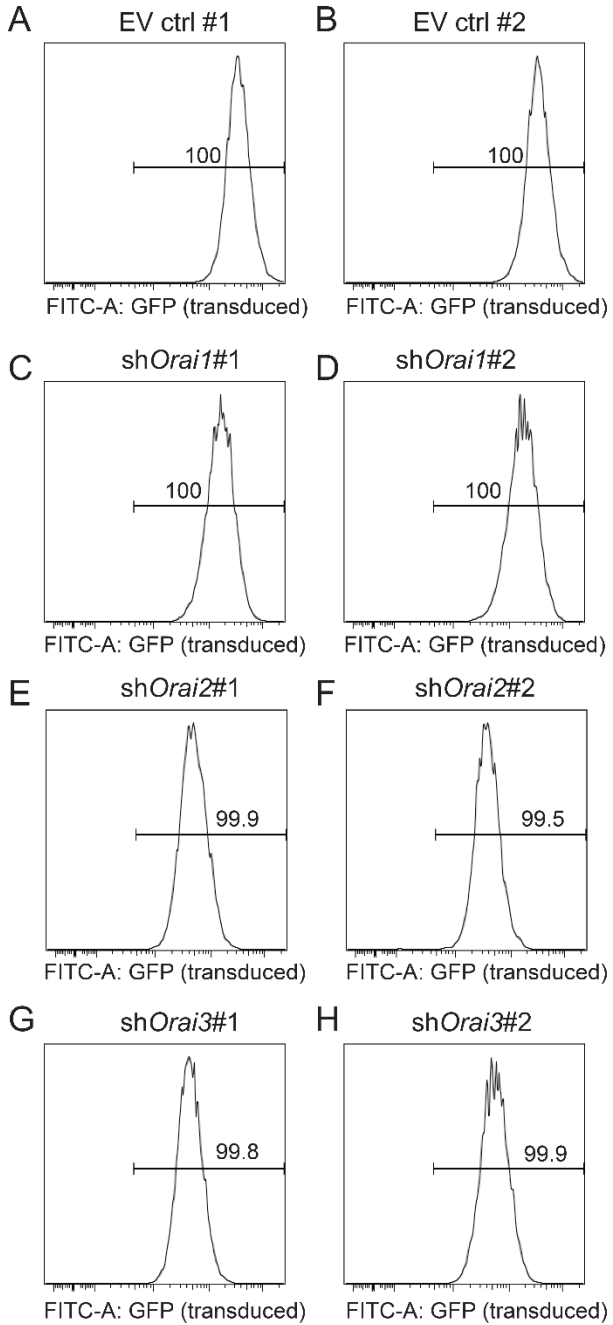
**Supplementary Figure 1. Unchanged expression of ameloblastin in ORAI-deficient cells.** The expression of the ameloblastin gene (*Ambn*) was analyzed in enamel organ cells of *Orai1<sup>K14</sup>* (A) and *Orai2<sup>-/-</sup>* mice (B) by RT-PCR showing no significant differences. Data represent averages (+/- SEM) of N=5 mice analyzed using Mann-Whitney (A) or Welch's t-test (B).



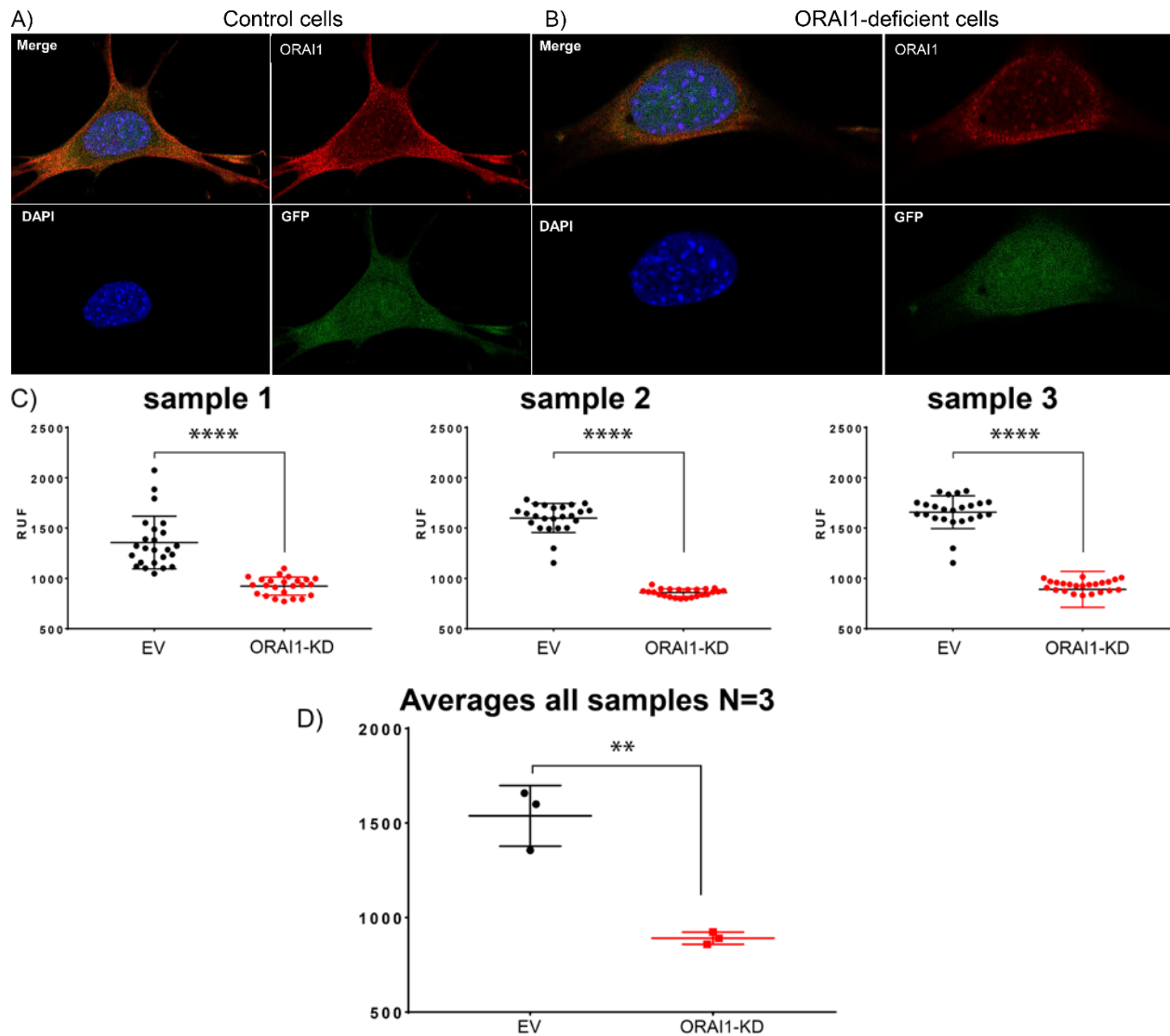
**Supplementary Figure 2. Enamel cells of *Orai1*<sup>K14</sup> mice do not show increased expression of unfolded protein response (UPR) markers.** The expression of GRP78 (gene name *HSPA5*) associated with UPR was analyzed in enamel organ cells of *Orai1*<sup>K14</sup> mice by RT-PCR showing no significant differences. Data represent averages (+/- SEM) of N=5 mice analyzed using unpaired Student's t-test.



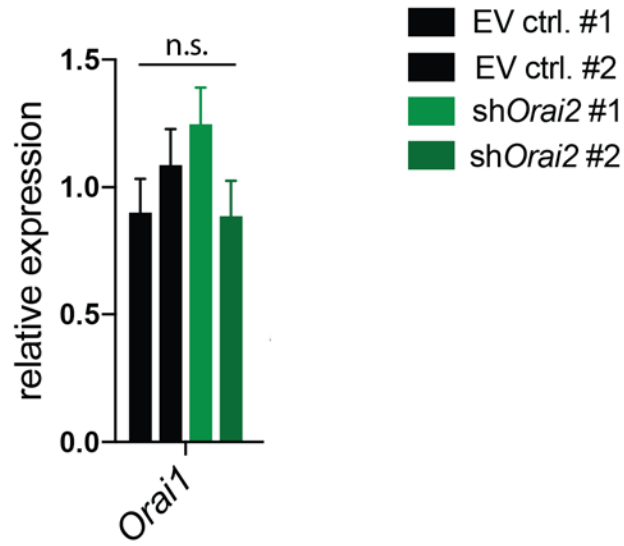
**Supplementary Figure 3. Normal dental phenotype in ORAI2-deficient mice. (A, C)** Macroscopic images of incisors of WT (A) and *Orai2*<sup>-/-</sup> (C) mice. **(B, D)** Scanning electron microscopy (SEM) images of cross-sections from incisors of WT (B) and *Orai2*<sup>-/-</sup> (D) mice imaged one millimeter from the tip. Data in (A-D) are representative of three mice per group.



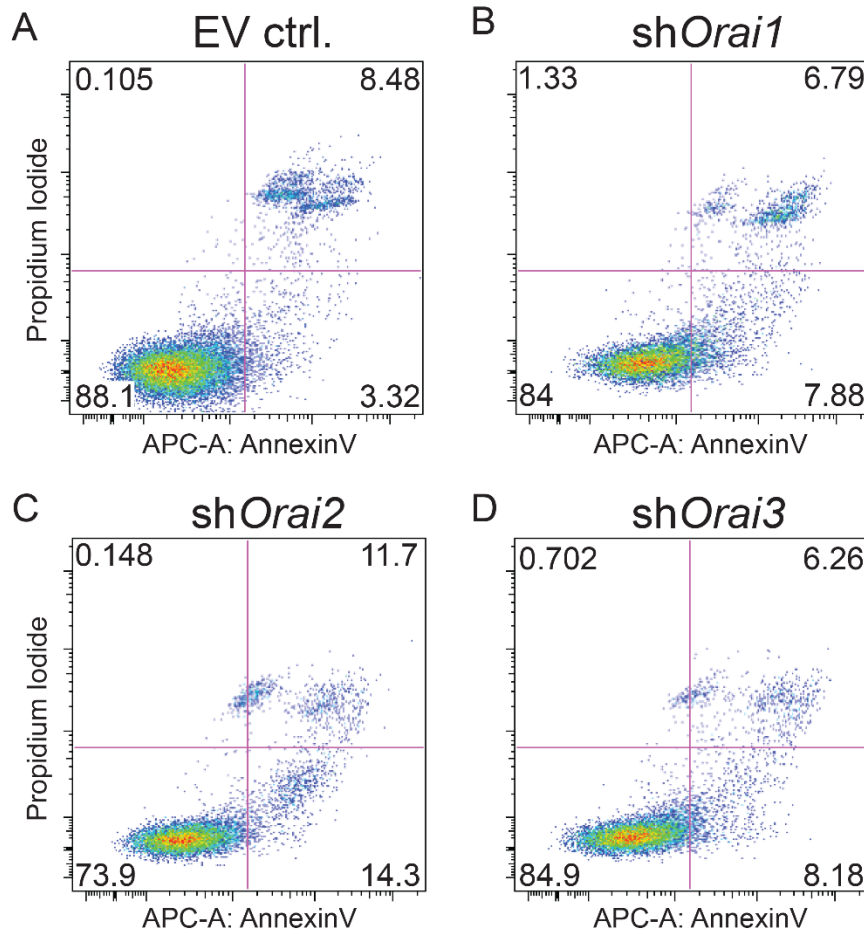
**Supplementary Figure 4. Transduction efficiency in LS8 cells.** Cells were transduced with either empty vector (EV ctrl., **A** and **B**) or shRNAs against *Orai1* (shOrai1, **C** and **D**), *Orai2* (shOrai2, **E** and **F**), and *Orai3* (shOrai3, **G** and **H**) and analyzed for GFP expression by flow cytometry. Data are representative of N=3 experiments.



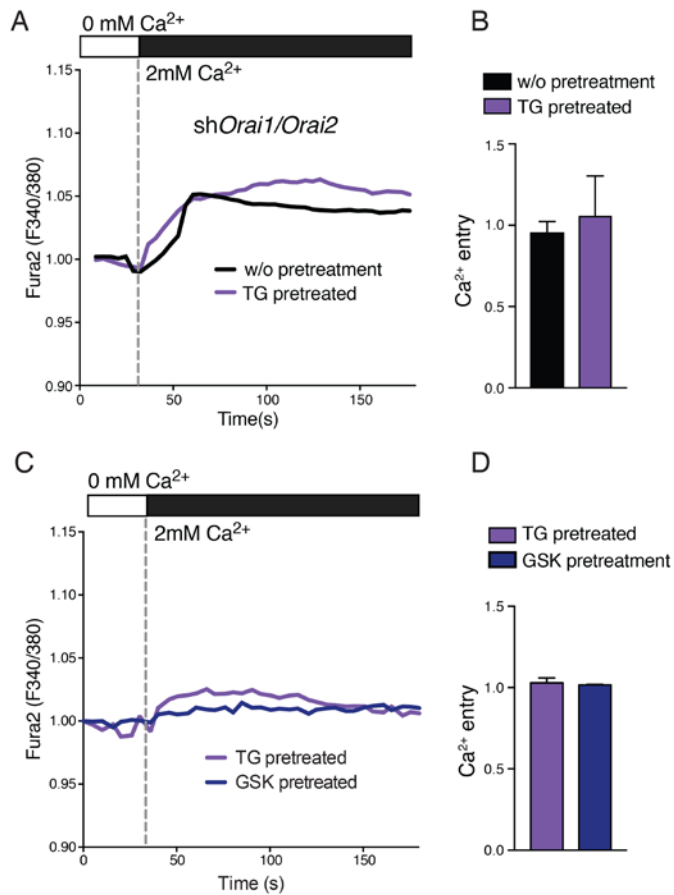
**Supplementary Figure 5. Immunofluorescence analysis showing downregulation of ORAI1 in GFP positive *shOrai1* cells.** **A-B)** Downregulation of ORAI1 protein was confirmed by blinded immunofluorescence in GFP positive control cells (A) and *shOrai1* cells (B) using anti-ORAI1 (Sigma SAB3500126) following protocols reported in (1). **C-D)** Quantification of differences in fluorescence between control and *shOrai1* cells following protocols reported in (2). Three independent experiments were performed with  $p < 0.01$  (\*\*) and  $p < 0.0001$  (\*\*\*\*) using Mann-Whitney (C) or unpaired Student's t-test (D)



**Supplementary Figure 6. *Orai1* expression is not significantly altered in sh*Orai2* cells.** The expression of *Orai1* was analyzed in sh*Orai2* cells showing no significant differences. Data represent averages ( $\pm$  SEM) of N= 3 independent experiments analyzed using unpaired Student's t-test.

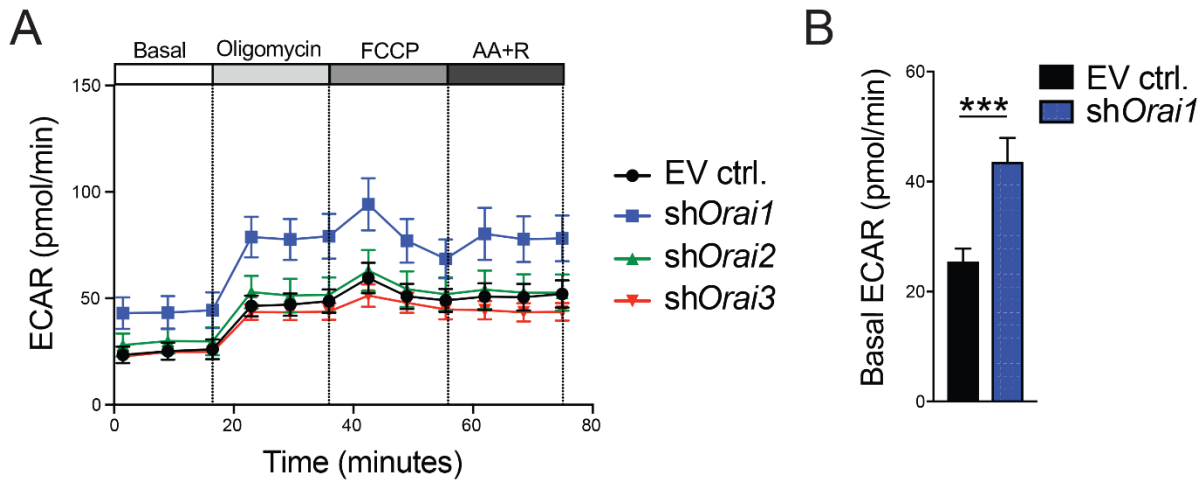


**Supplementary Figure 7. Apoptosis and cell death.** Apoptosis and cell death were analyzed in LS8 cells transduced with either empty vector (EV ctrl., **A**) or shRNAs against *Orai1* (sh*Orai1*, **B**), *Orai2* (sh*Orai2*, **C**), and *Orai3* (sh*Orai3*, **D**). Cells were stained with Annexin V and propidium iodide (PI) and analyzed by flow cytometry. Representative dot blots shown in this Figure are the basis of data shown in main Fig.4I.

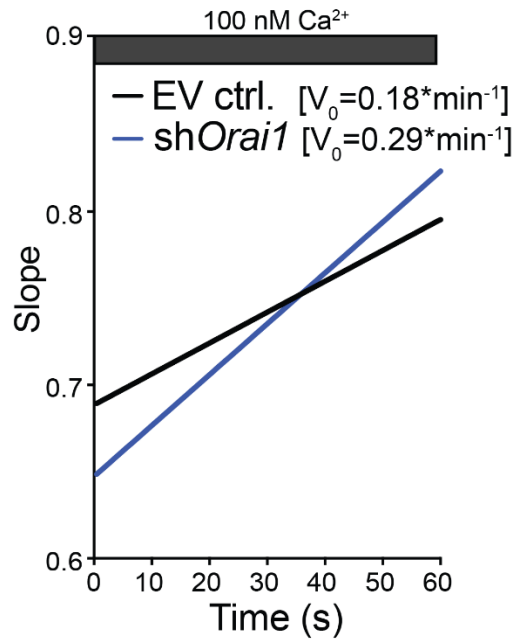


**Supplementary Figure 8. Small Ca<sup>2+</sup> leak is present in shOrai1/Orai2 cells.** (A) Cytosolic Ca<sup>2+</sup> uptake was measured in Fura2-loaded LS8 cells transduced with shRNA against *Orai1* and *Orai2* (shOrai1/Orai2) using a Flexstation III. Purple tracings are shOrai1/Orai2 cells that had been pretreated with thapsigargin as described in our SOCE measurements shown in Fig 5. Black tracings represent Ca<sup>2+</sup> uptake in shOrai1/Orai2 cells but without prior thapsigargin treatment. This minor uptake results from a change in the external Ringer solution of 0 mM Ca<sup>2+</sup> with a solution containing 2mM Ca<sup>2+</sup>, thus potentially reflecting a plasma membrane leak. (B) Statistical analyses of Ca<sup>2+</sup> leak in shOrai1/Orai2 cells. (N=3) independent experiments; data are mean +/- SEM. (C) In a separate experiment, re-addition of Ca<sup>2+</sup> to thapsigargin stimulated shOrai1/Orai2 showed a similar elevation to that observed in cells preincubated with the CRAC channel inhibitor GSK-7975A. (D) Statistical analyses of Ca<sup>2+</sup> peak in shOrai1/Orai2 cells. (N=3) independent experiments; data are mean +/- SEM.





**Supplementary Figure 9. Increased extracellular acidification rate (ECAR) in shOrai1 cells.** (A-B) Extracellular acidification rates (ECAR) in LS8 cells transduced with either empty vector (EV ctrl.) or shRNAs against *Orai1*, *Orai2* or *Orai3*. ECAR was analyzed using a Seahorse XFe analyzer and sequential treatment with Oligomycin, FCCP and Antimycin A / Rotenone. Representative ECAR traces (A) and quantification of basal ECAR values (B). Data represent 4 independent experiments (mean +/- SEM).  $p < 0.001$  (\*\*\*) using Mann-Whitney test (B).



**Supplementary Figure 10. Quantification of velocity (slope) of ER Ca<sup>2+</sup> refilling.** The slope of Ca<sup>2+</sup> refilling in EV ctrl. and sh*Orail1* cells was analyzed during the first 60s after tBHQ application in 100 nM Ca<sup>2+</sup> ( $V_0$ ). Data represent the mean  $\pm$  SEM of 3 independent experiments. sh*Orail1* cells showed a significant faster refilling ( $V_0 = 0.29 \times \text{min}^{-1} \pm 0.00137$ ) compared to ctrl cells ( $V_0 = 0.18 \times \text{min}^{-1} \pm 0.00059$ ) with  $p < 0.05$  (\*) using an unpaired Student's t-test).

**Table S1. Primer and shRNA sequences used in this study.**

Specific mouse Primers designed for Genotyping

Primer	Forward sequence	Reverse sequence
<i>Cre</i>	ACGACCAAGTGACAGCAATG	CTCGACCAGTTTAGTTACCC
<i>Orai1</i>	CAGCGTGCATAATATACCTAACT CTACCCG	GTATTGATGAGGAGAGCAAGCGT GAATC
<i>Orai2</i>	CTCGGCAGTTGCCTGTTTG	ACAGCCACCACGCTCATC

shRNA sequences for stable knockdown of Orai1-3

Target gene	shRNA sequences
<i>Orai1 #1</i>	TGCTGTTGACAGTGAGCGCGGAGATTTTCTTCTGTGAGAATAGTGAAGCCACAGATGT ATTCTCACAGAAGAAAATCTCCATGCCTACTGCCTCGGA
<i>Orai1 #2</i>	TGCTGTTGACAGTGAGCGCCCTGGCGCAAGCTCTACTTAATAGTGAAGCCACAGATGT ATTAAGTAGAGCTTGCGCCAGGATGCCTACTGCCTCGGA
<i>Orai2 #1</i>	TGCTGTTGACAGTGAGCGACGCCACAACCGTGAGATCGAATAGTGAAGCCACAGATGT ATTCGATCTCACGGTTGTGGCGCTGCCTACTGCCTCGGA
<i>Orai2 #2</i>	TGCTGTTGACAGTGAGCGCGAGCAACATCCACAACCTCAATAGTGAAGCCACAGATGTA TTGAGGTTGTGGATGTTGCTCATGCCTACTGCCTCGGA
<i>Orai3 #1</i>	TGCTGTTGACAGTGAGCGCCAGCTGGAGAACGATCATGAATAGTGAAGCCACAGATGTA TTCATGATCGTTCTCCAGCTGTTGCCTACTGCCTCGGA
<i>Orai3 #2</i>	TGCTGTTGACAGTGAGCGCCTCCCTTAGTTTAGCTTCTAATAGTGAAGCCACAGATGTAT TAGAAGCTAAACTAAGGGAGATGCCTACTGCCTCGGA

Specific mouse Primers designed for qPCR

Primer	Forward sequence	Reverse sequence
<i>Hprt1</i>	GCTGACCTGCTGGATTACAT	TTGGGGCTGTACTGCTTAAC
<i>Orai1</i>	AGACTGCCTGATCGGATGGC	TTGTCCCCGAGCCATTTCT
<i>Orai2</i>	GCAGCTACCTGGAACCTCGTC	GTTGTGGATGTTGCTCACCG
<i>Orai3</i>	CAGTCAGCACTCTCTGCGG	TGGCCACCATGGCGAAG
<i>Enam</i>	TATGGTCTTCCACCAAGGAA	TAGGCACACCATCTCCAAAT
<i>AmelX</i>	CCCTGAGCTTCAGACAGAAA	CTGCCTTATCATGCTCTGGT
<i>Atp2a2</i>	CAGCCTGTCTGAGAACCAGT	CTCCAGATAGTTCCGAGCAA
<i>Hspa5</i>	ACTTGGGGACCACCTATTCCT	ATCGCCAATCAGACGCTCC

