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Supplemental Information

**m⁶A RNA Methylation Maintains Hematopoietic
Stem Cell Identity and Symmetric Commitment**

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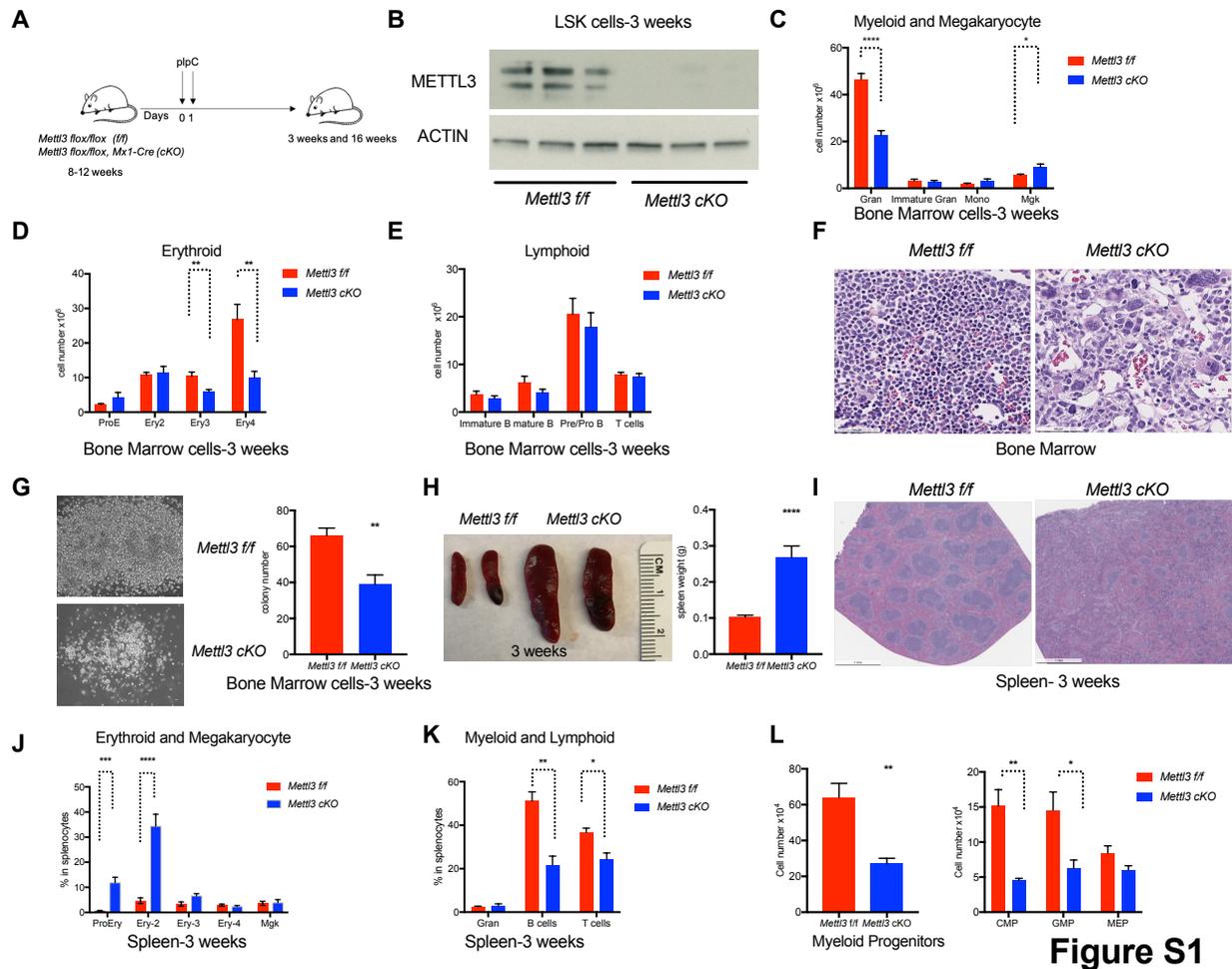


Figure S1

Supplementary Figure 1. m⁶A is essential for normal hematopoiesis. Related to Figure 1.

All samples were collected from mice at 3 weeks post plpC administration. **(A)** Schematic diagram of the experimental procedure for analyzing *Mettl3* *f/f* and *Mettl3* *cKO* mice. **(B)** Immunoblot to determine METTL3 expression using sorted LSK cells. **(C-E)** Myeloid, megakaryocytes, erythroid, and lymphoid lineage differentiation in BM was determined by flow cytometry base on different cell surface markers. Erythroblast (ProE, Ter119-CD71+; Ery2, Ter119+CD71+; Ery3, Ter119+CD71^{mid}; Ery4, Ter119+CD71-), Gran (Granulocyte, Gr1+Mac1+), Immature Granulocyte(Mac1+Gr1^{mid}), Monocyte(Mac1+Gr1-), Megakaryocytes (Mgk, CD41+), Immature B cells(B220+IgM+), Pre/Pro B cells (B220+IgM-), Mature B cells (B220^{hi}IgM+), T cells (CD3+). n=11. **(F)** Representative images show H&E-stained cross sections of BM isolated from the *Mettl3* *f/f* and *Mettl3* *cKO* mice. **(G)** *Mettl3* depleted BM formed fewer and smaller colonies. *Mettl3* *f/f* and METTL3 depleted BM cells were plated in methylcellulose cultures, supplemented with cytokines. Left: Representative images of flox/flox and cKO colony. Right: Colonies were scored 10 days after plating. n =9,7. **(H)** Splenomegaly is observed in cKO mice three weeks post plpC administration. Left: representative image of spleens from indicated mice. Right: measurement of spleen weight from *Mettl3* *f/f* and *Mettl3* *cKO* mice. n=11. **(I)** Representative images show H&E-stained cross sections of spleens isolated from the *Mettl3* *f/f* and *Mettl3* *cKO* mice. **(J, K)** Erythroid, myeloid and lymphoid cells were determined by flow cytometry in *Mettl3* *f/f* and *Mettl3* *cKO* spleens. Cell population was determined by cell surface marker. n=8. **(L)** Absolute numbers of myeloid progenitor (MP, Lin-ckit+Sca1-), CMP(Lin-cKit+Sca1-CD34+FcrR-), GMP (Lin-cKit+Sca1-CD34+FcrR+) and MEP(Lin-cKit+Sca1-CD34-FcrR-) in *Mettl3* *f/f* and *Mettl3* *cKO* BM. n=10, 6. Mean and SEM are shown (*, P < 0.05; **, P < 0.01; ***, P < 0.001, ****, P < 0.0001).

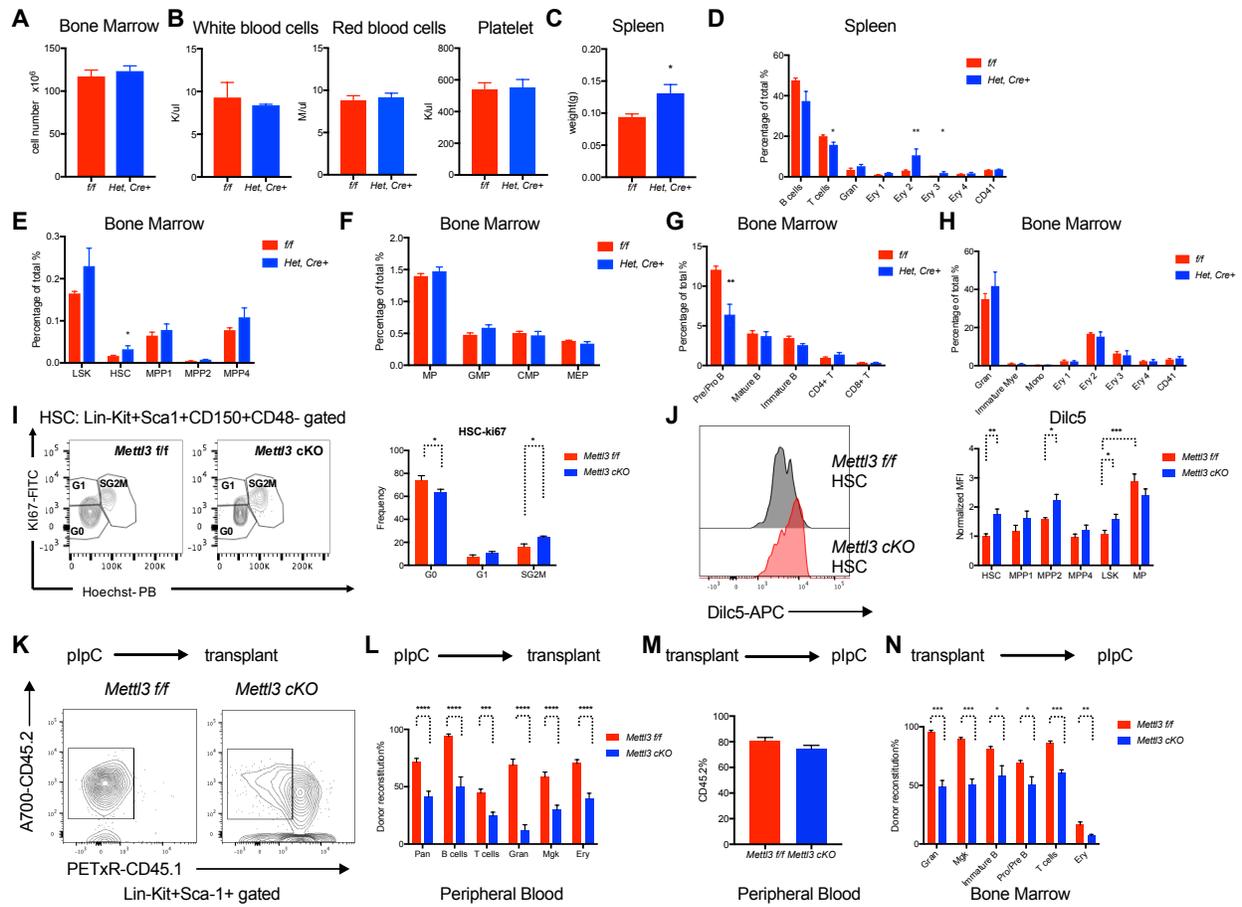


Figure S2

Supplementary Figure 2. *Mettl3* required for HSC reconstitution capacity. Related to Figure 2.

(A) Whole bone marrow cellularity in *Mettl3 f/f* and *Het, Cre+* (*Mettl3 f/-, Mx1-Cre*) mice. n=4. **(B)** Whole blood counts of white blood cell (WBCs), red blood cells (RBCs), platelets (PLT) of *Mettl3 f/f* and *Het, Cre+* mice. n=4. **(C)** Spleen weight of *Mettl3 f/f* and *Het, Cre+* mice. n=4. **(D)** Mature lineage cells were determined by flow cytometry in *Mettl3 f/f* and *Het, Cre+* spleens base on cell surface marker as shown in **Supplementary Figure 1**. n=4. **(E, F)** Frequency of hematopoietic stem and progenitor compartments (LSK, HSC, MPP1, MPP2 and MPP4) and myeloid progenitors (MP, CMP, GMP and MEP) in *Mettl3 f/f* and *Het, Cre+* mice bone marrow. n=4. **(G, H)** Erythroid, myeloid, megakaryocytes and lymphoid lineage differentiation in *Mettl3 f/f* and *Het, Cre+* mice were assessed by flow cytometry base on cell surface markers. n=4. **(I)** *Mettl3 cKO* HSC cells are more proliferative. Representative flow cytometry plots and quantification of *Mettl3 f/f* and *Mettl3 cKO* HSC cell cycle by Ki67 staining. n=5. **(J)** Increased mitochondrial activity in *Mettl3 cKO* HSCs. Left: representative histograms of Dilc5 staining in *Mettl3 f/f* and *Mettl3 cKO* HSCs. Right: Mitochondrial membrane potential in HSPC compartments was assessed by Dilc5 staining quantified by flow cytometry. n=5. **(K)** Representative flow cytometry of surface markers to indicate engraftment of CD45.2 donor cells in LSK population from non-competitive transplanted recipient mice. **(L)** *Mettl3 cKO* HSCs have impaired reconstitution capacity. Chimerism of CD45.2 in different lineage cell populations in peripheral blood from non-competitive transplanted recipient mice. n=10. **(M,N)** Cell autonomous transplant experiments. **(M)** Comparable engraftment of *Mettl3 flox/flox, Mx1-Cre-* and *Mx1-Cre+* recipient pre-plpC. Chimerism of CD45.2 donor cells in peripheral blood from recipient mice was analyzed before plpC injection. n=15. **(N)** CD45.2 chimerism analysis of myeloid, erythroid and lymphoid cells from recipient mice. Cell compartment is determined by cell surface marker as showed previously. n=5. Mean and SEM are shown (*, P < 0.05; **, P < 0.01; ***, P < 0.001, ****, P < 0.0001).

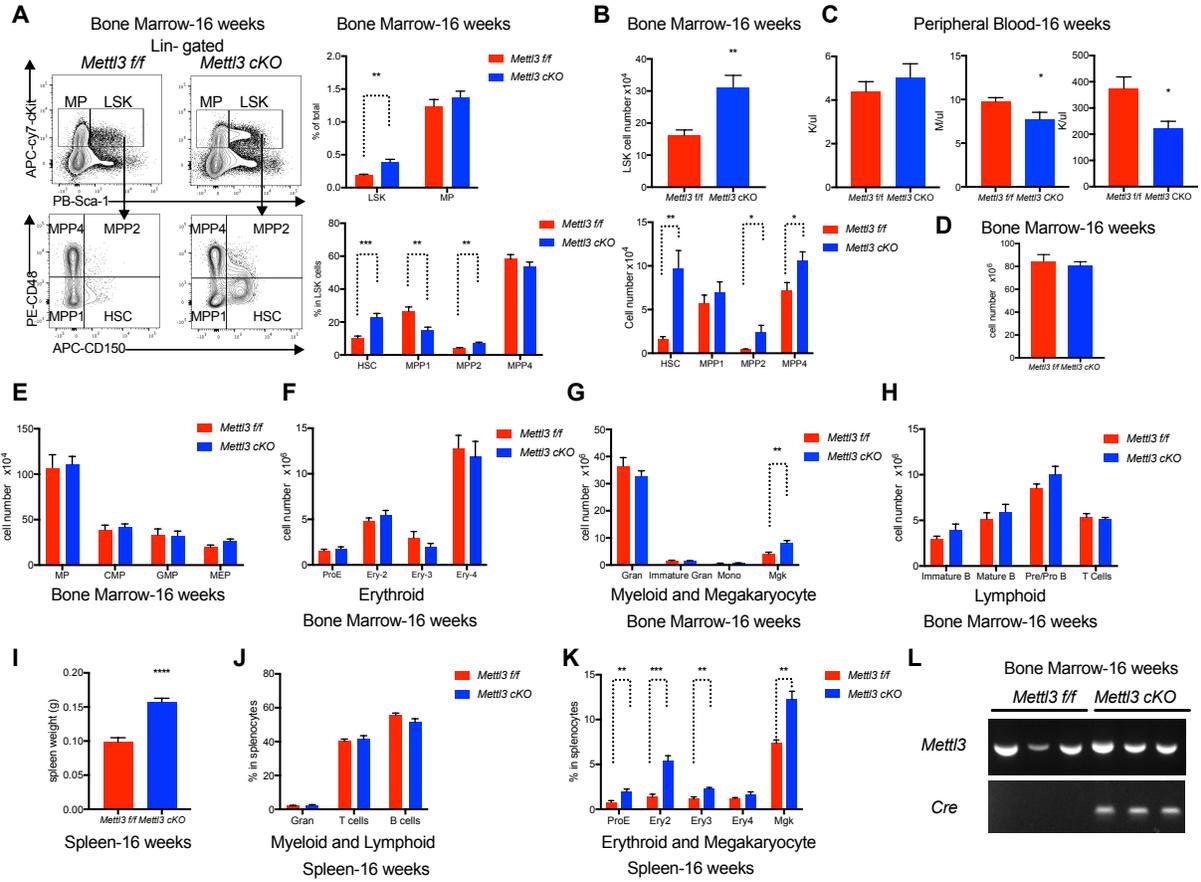


Figure S3

Supplementary Figure 3. HSPC expansion persists in *Mettl3* cKO mice 16 weeks post plpC. Related to Figure 3.

All samples used were from *Mettl3* f/f and *Mettl3* cKO mice at 16 weeks post plpC. **(A)** Left: Representative flow cytometry plots of surface markers to indicate the stem and progenitor cell compartments in BM. Right: Frequency of LSK and MP in BM cells and percentage of HSC, MPP1, MPP2 and MPP4 in LSK population. n=6. **(B)** Absolute cell numbers of LSK, HSC and MPPs, from indicated mice at 16 weeks post plpC as assessed by flow cytometry. n=6. **(C)** Whole blood counts of white blood cell (WBCs), red blood cells (RBCs), platelets (PLT) of *Mettl3* f/f and *Mettl3* cKO mice. **(D)** Bone marrow cellularity of *Mettl3* f/f and *Mettl3* cKO mice. n=6. **(E)** Absolute cell numbers of progenitor, CMP, GMP and MEP in BM cells. n=6. **(F-H)** Erythroid, myeloid, megakaryocytes and lymphoid lineage differentiation in long-term METTL3 deleted mice were assessed by flow cytometry base on cell surface markers as shown in **Supplementary Figure 1**. n=6. **(I)** Measurement of spleen weight from *Mettl3* f/f and *Mettl3* cKO mice. n=6. **(J,K)** Erythroid, myeloid and lymphoid cells were evaluated by flow cytometry in splenocytes. n=6. **(L)** Genotyping PCR using genomic DNA from *Mettl3* f/f and *Mettl3* cKO mice bone marrow cells at 16 weeks post plpC. Mean and SEM are shown (*, P < 0.05; **, P < 0.01; ***, P < 0.001, ****, P < 0.0001).

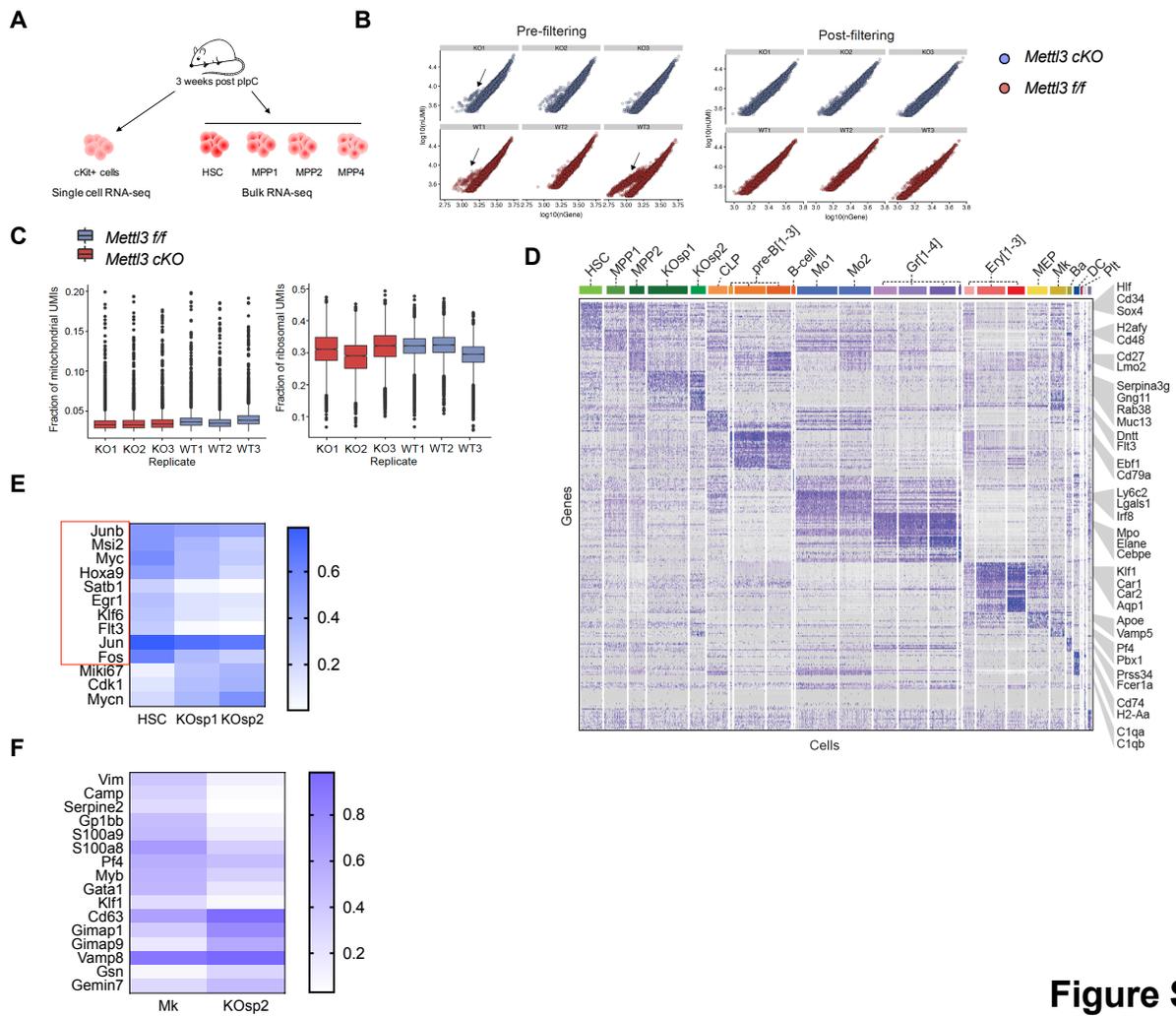


Figure S4

Supplementary Figure 4. Loss of *Mettl3* results in new HSC-like clusters base on scRNA-seq. Related to Figure 4.

(A) Scheme of experiment strategy for single cell RNA-seq and bulk RNA-seq. **(B)** Filter strategy in three replicates of single cell RNA-seq. **(C)** Number of ribosomal or mitochondrial reads detected for each cell in each replicate. **(D)** Single-cell transcriptome landscape defined by gene expression. Different cell cluster was labeled. **(E)** Heatmap to show differentially expressed genes in *Mettl3* cKO specific clusters compared to HSC cluster. **(F)** Representative differentially expressed genes between KOsp2 cluster and Megakaryocytes cluster were shown as heat map.

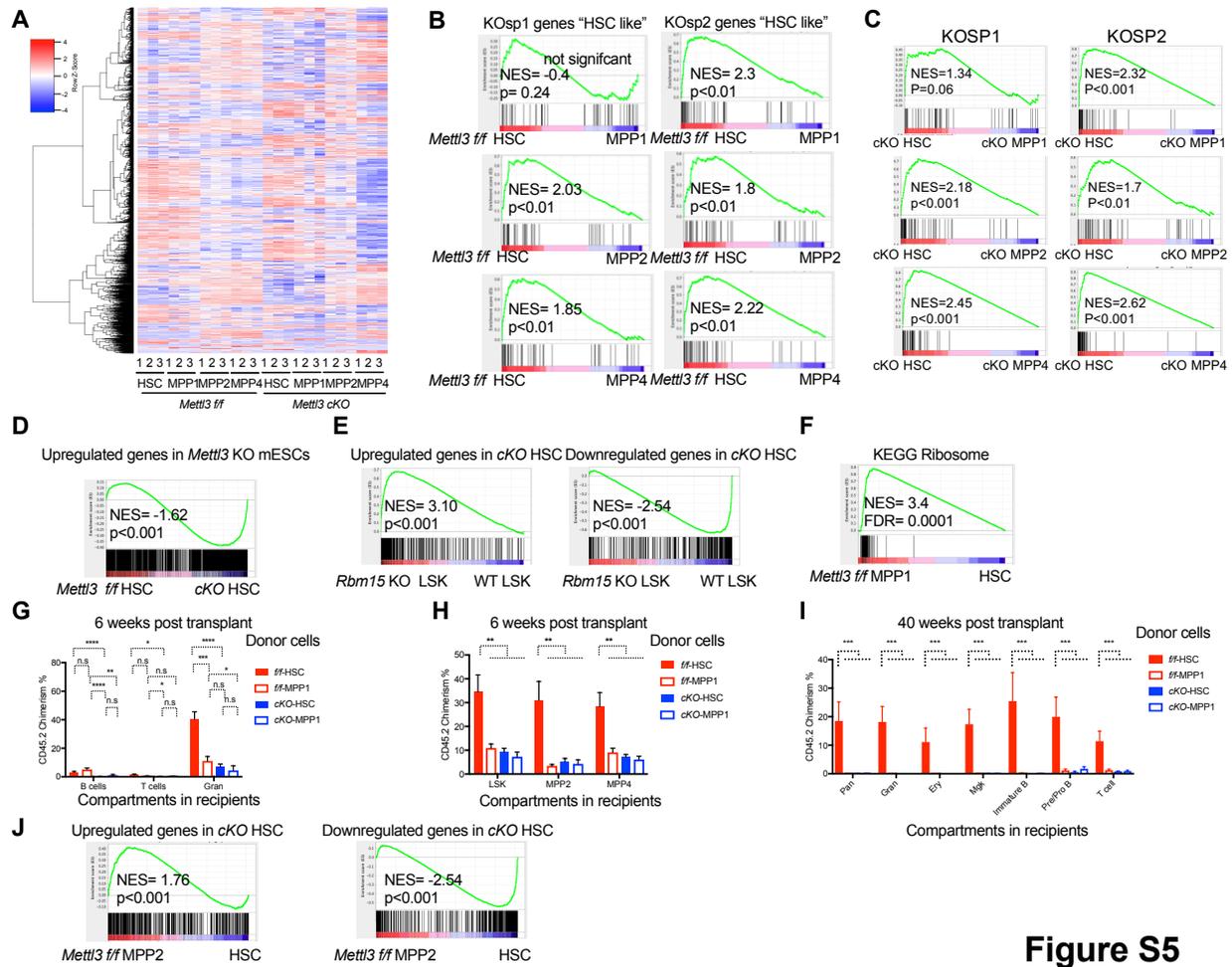


Figure S5

Supplementary Figure 5. HSC identity is altered in *Mettl3* cKO mice. Related to Figure 5.

(A) Unsupervised clustering of differentially expressed genes in three replicates of *Mettl3* *f/f* and *Mettl3* cKO HSC, MPP1, MPP2 and MPP4 cells were shown. **(B)** *Mettl3* cKO specific clusters are cKO HSC-like based on gene expression. GSEA analysis of cKO specific cell clusters gene sets (top 50 expressed genes from scRNA-seq) against rank lists of differentially expressed genes between *Mettl3* cKO HSCs and cKO MPPs. **(C)** GSEA analysis with top expressed genes in cKO specific cell clusters from scRNA-seq showing the signature of genes enriched in *Mettl3* *f/f* HSCs. **(D)** *Mettl3* KO ESC signature is enriched in *Mettl3* cKO HSCs. Gene Set Enrichment Analysis of upregulated genes in *Mettl3* KO mESCs against the ranklist of differentially expressed genes between *Mettl3* *f/f* and *Mettl3* cKO HSCs. Normalized enrichment score and p value are shown. **(E)** Gene-set enrichment for up or down regulated genes in *Mettl3* cKO HSCs, as compared to differentially expressed genes in RBM15 KO LSK cells. Normalized enrichment score and FDR are shown. **(F)** *Mettl3* *f/f* MPP1 was enriched in KEGG ribosome gene set in regulating translation compared to *Mettl3* *f/f* HSCs. **(G-I)** HSCs and MPP1s from cKO mice fail to engraft in recipient mice in a competitive manner. Sorted HSCs and MPP1s from *Mettl3* *f/f* and *Mettl3* cKO mice were injected into CD45.1 recipient mice competitively with normal CD45.1 BM cells. **(G)** CD45.2 chimerism was shown in peripheral blood at 4 weeks post transplantation. n=10. **(H)** CD45.2 chimerism analysis to show donor engraftment in LSK, MPP2 and MPP4 compartments. n=10. **(I)** Engraftment of CD45.2 donor cells was analyzed in lineage compartments from recipient mice at 40 weeks. n=10. **(J)** Gene-set enrichment analysis plots of up or down regulated genes in *Mettl3* cKO HSC identified by bulk RNA-seq, against the rank list of differentially expressed genes between *Mettl3* *f/f* HSC and *Mettl3* *f/f* MPP2. Mean and SEM are shown (*, P < 0.05; **, P < 0.01; ***, P < 0.001, ****, P < 0.0001).

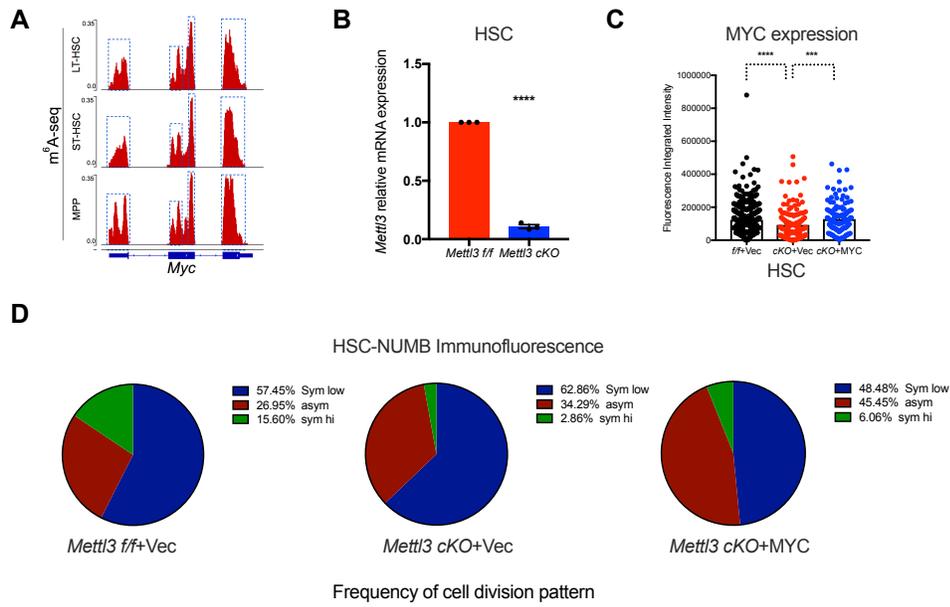


Figure S6

Supplementary Figure 6. m⁶A-mediated *Myc* regulation is essential for HSC asymmetric and symmetric cell division. Related to Figure 6.

(A) m⁶A marks on *Myc* transcripts in HSCs and MPPs. m⁶A peaks were indicated in dot line square. **(B)** qRT-PCR of *Mettl3* in *Mettl3 f/f* and *Mettl3 cKO* HSCs. **(C,D)** Overexpression of MYC in *Mettl3 cKO* HSCs partially rescued HSC symmetric commitment defect in pair daughter assay as quantified by immunofluorescence. Number of daughter pairs assessed: *Mettl3 f/f* +Vec, n=141; *cKO*+Vec, n=35; *cKO*+MYC, n=33.

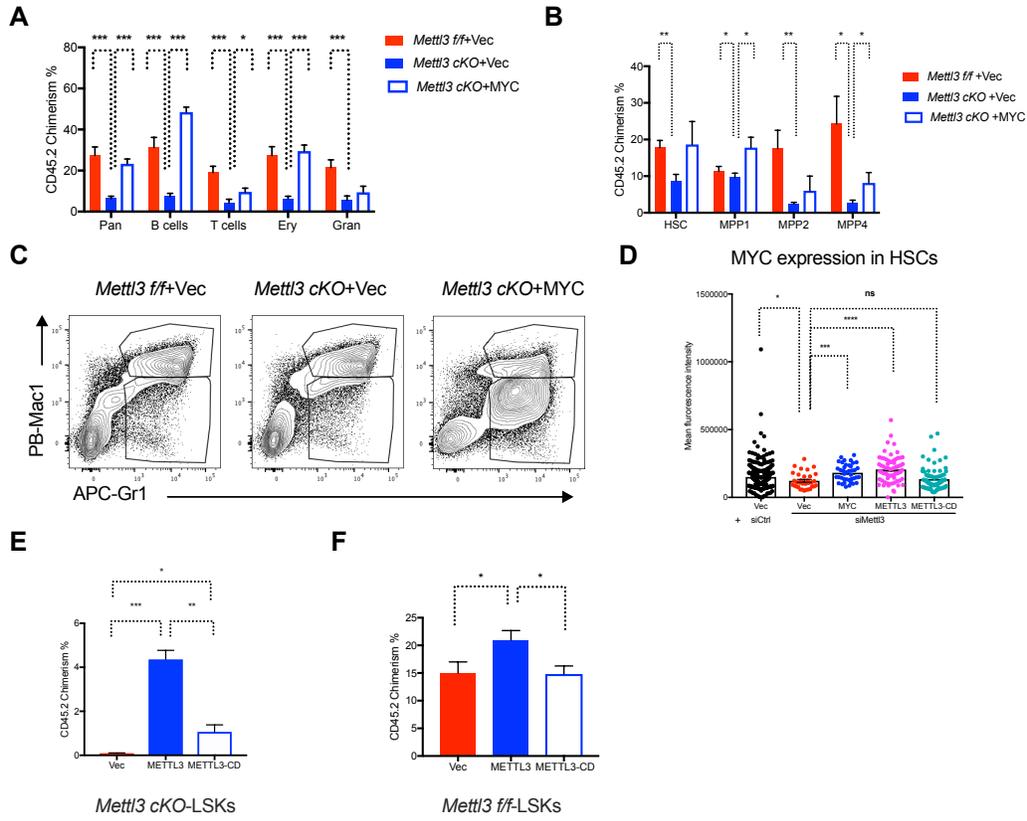


Figure S7

Supplementary Figure 7. METTL3 and MYC partially rescue defect in *Mettl3* cKO HSCs. Related to Figure 7.

(A-C) Sorted control or *Mettl3* cKO LSK cells transduced with control or MYC overexpression retrovirus and then transplanted into recipient mice. **(A)** Engraftment of donor-derived CD45.2 cells in different lineage cells including B cells(B220+), T cells (CD3+), Erythroblast (Ter119+) and Granulocyte (Gr1+). n=10. **(B)** Engraftment of donor-derived CD45.2 cells in HSC and MPP compartments in recipient mice BM. n=10. **(C)** Representative flow cytometry plots to show abnormal myeloblast in recipient mice transplanted with MYC overexpressed *Mettl3* cKO LSKs. **(D)** Sorted HSCs were co-transfected with control or *Mettl3* siRNA together with empty vector or MYC, METTL3, METTL3-CD constructs as indicated in the figure. Overall MYC expression is quantified by immunofluorescence. **(E)** LSK cells were sorted from *Mettl3* cKO mice and transduced with control or wild-type METTL3 or catalytically dead METTL3 (METTL3-CD) overexpression retrovirus. Donor cells were then transplanted into CD45.1 recipient mice with CD45.1 competitor BM cells. CD45.2 chimerism was shown in peripheral blood at 4 weeks post transplantation. n=5. **(F)** Sorted *Mettl3* f/f LSK cells were transduced with a retrovirus expressing GFP together with an empty vector as control, or a retrovirus expressing METTL3 or METTL3-CD and then transplanted into recipient mice. CD45.2 donor engraftment was analyzed at 4 weeks post-transplant. n=10. Mean and SEM are shown (*, P < 0.05; **, P < 0.01; ***, P < 0.001, ****, P < 0.0001).