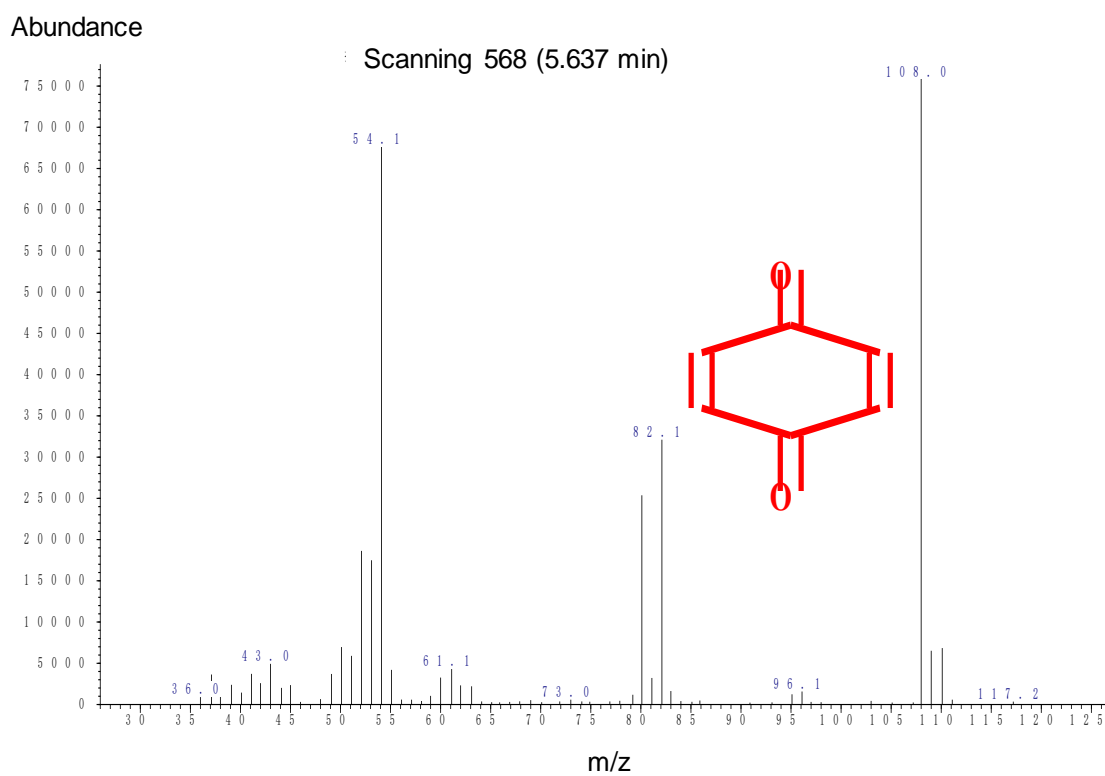
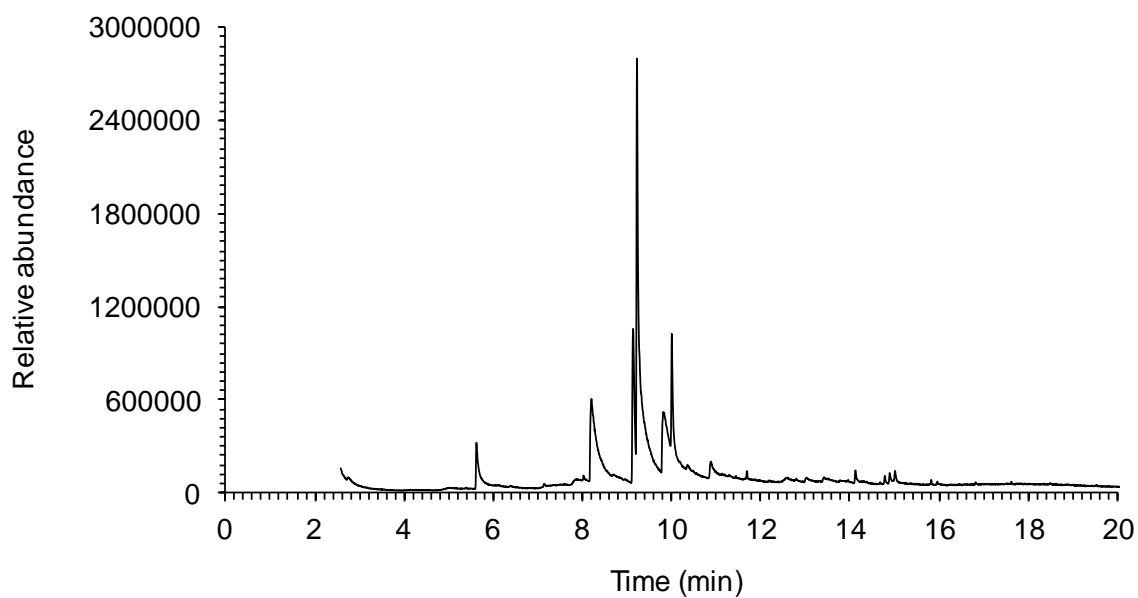
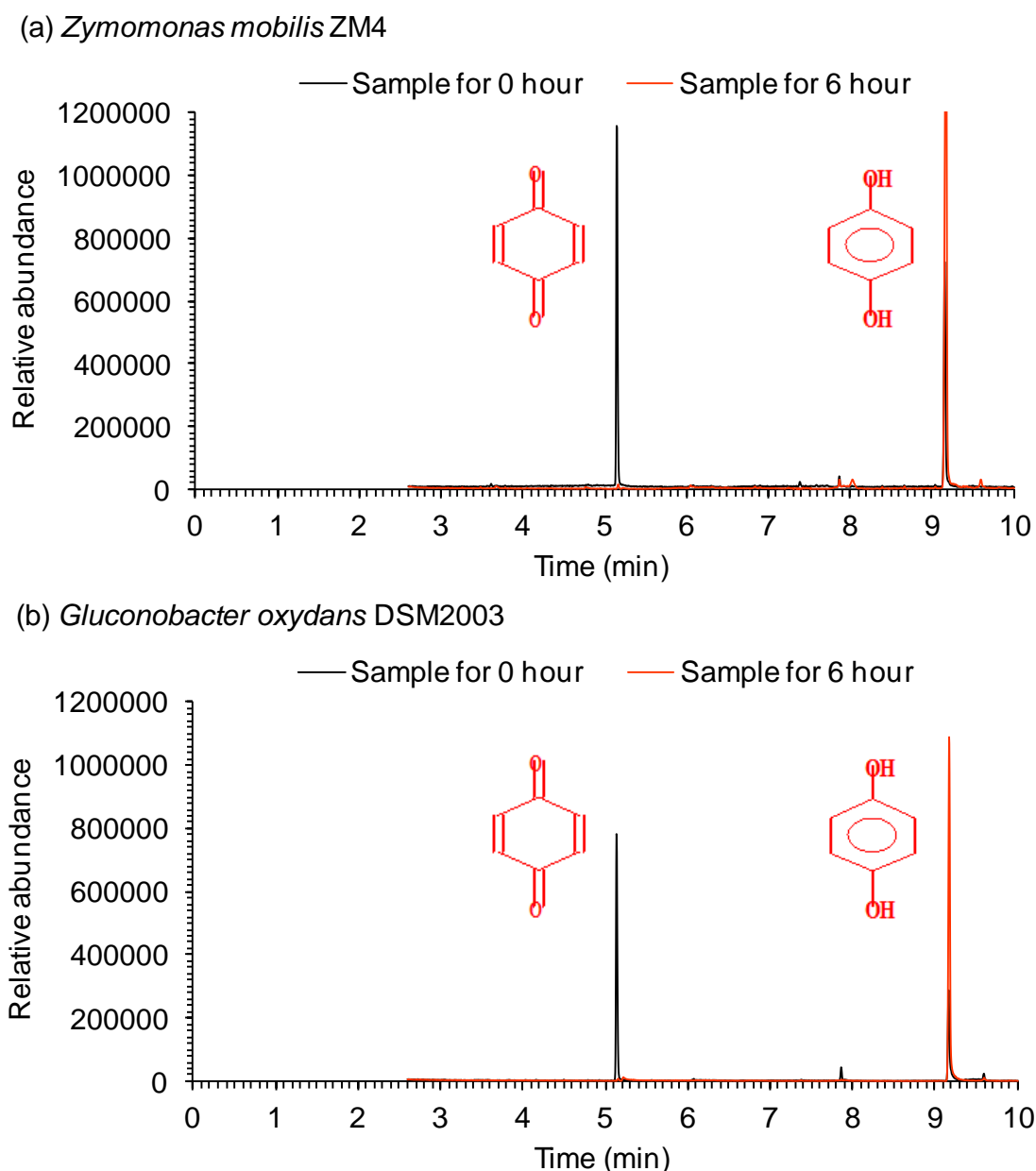


## SUPPLEMENTAL MATERIALS

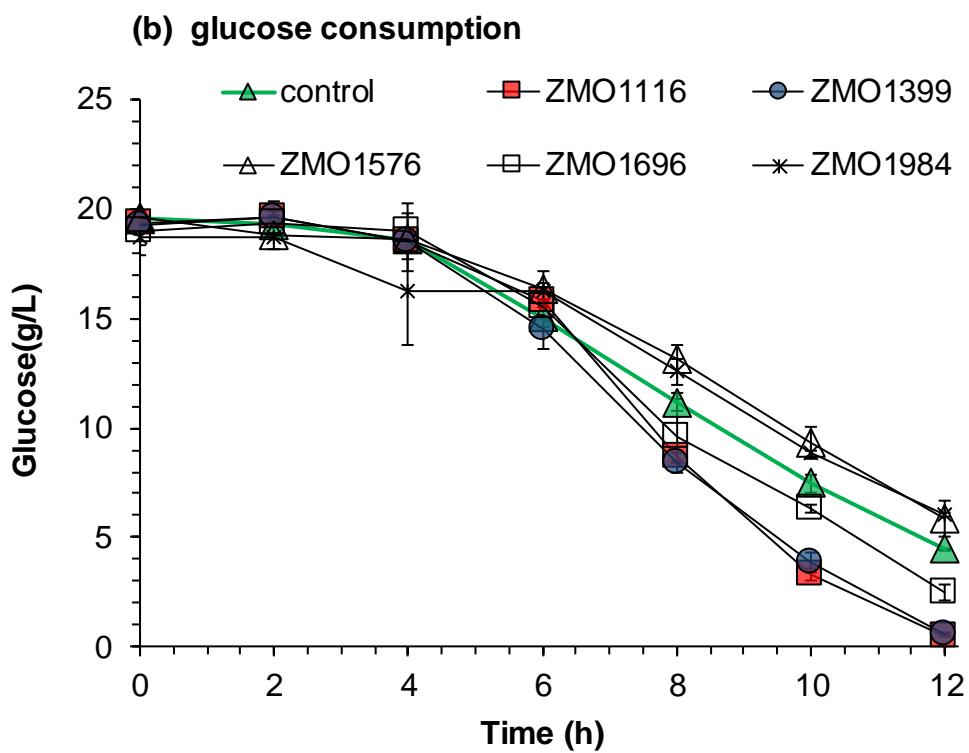
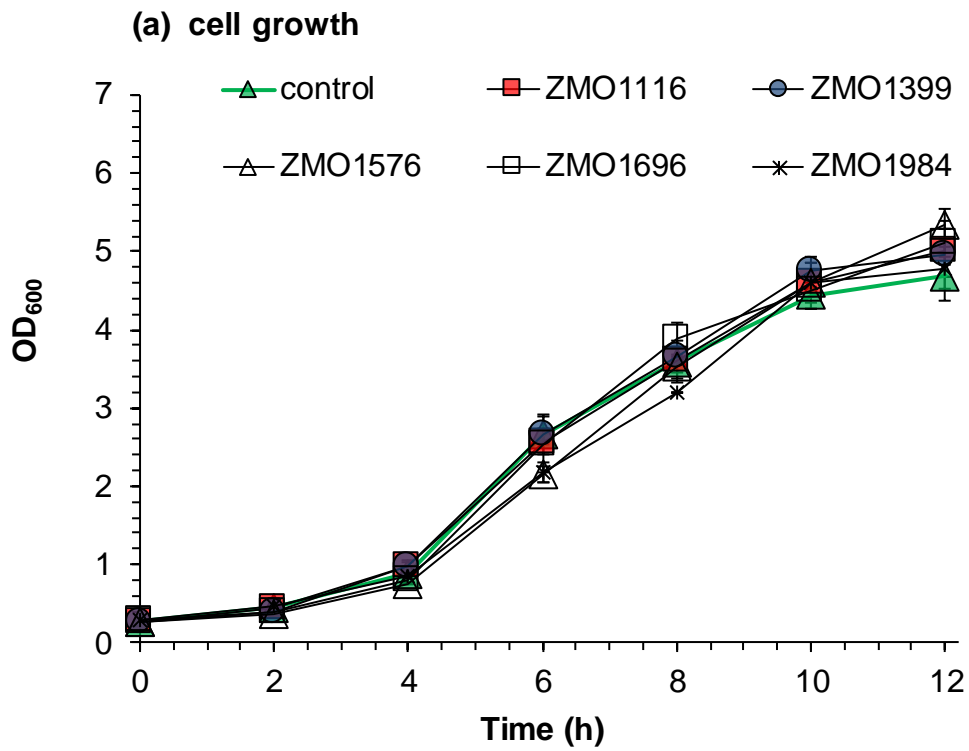


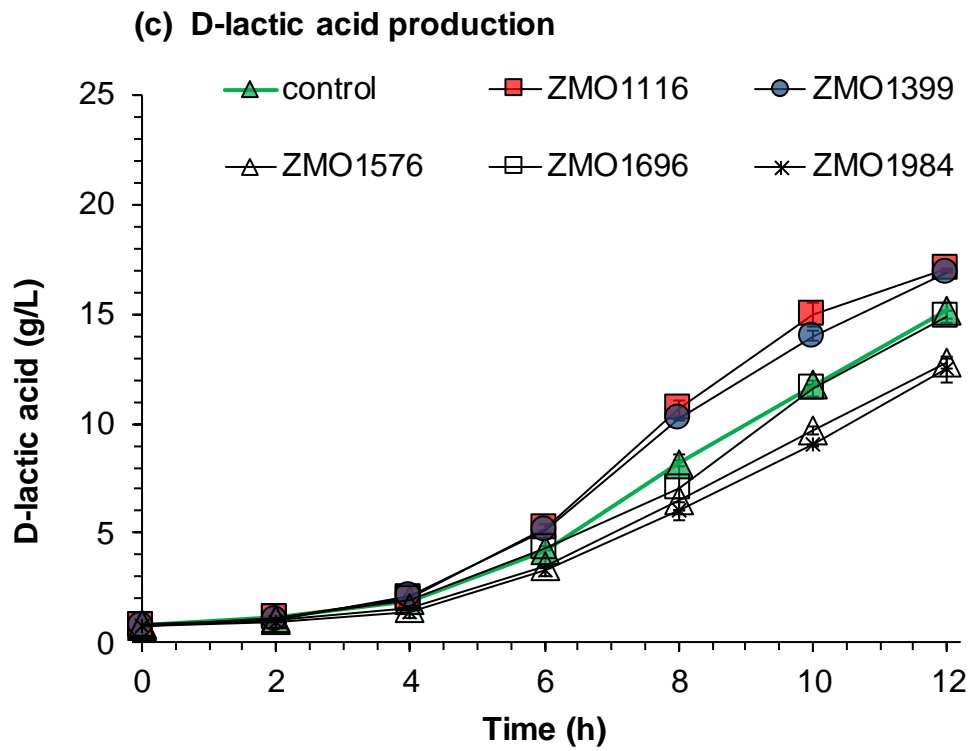
**Figure S1.** GC-MS identification of the formation of BQ during dry acid pretreatment of the representative lignocellulose feedstock of corn stover. The compound with retention time at 5.64 min was identified as BQ (at m/z 108).



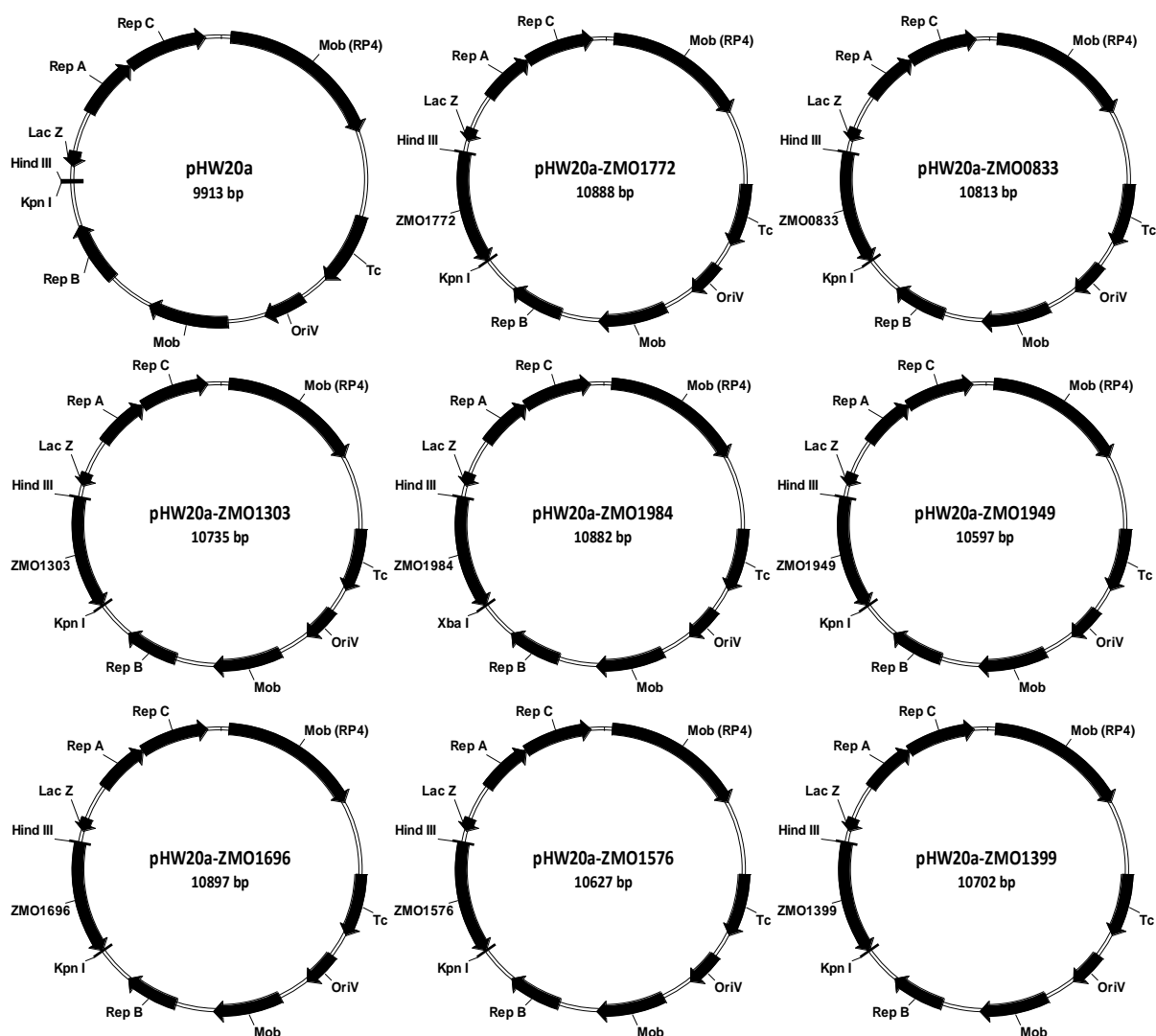
**Figure S2.** GC-MS identification of the intermediates generated during degradation of p-benzoquinone (BQ) by *Z. mobilis* ZM4 and *G. oxydans* DSM2003. (a) *Z. mobilis* ZM4; and (b) *G. oxydans* DSM2003. The whole flask of the *Z. mobilis* ZM4 and *G. oxydans* DSM2003 cell mass in exponential growth were harvested, washed by phosphate buffered saline (PBS, pH 7.0) and resuspended in the same volume of PBS, then inoculated into the fresh PBS containing 20 g/L glucose and 200 mg/L BQ. The cell cultures were collected at 0 h and 6 h for GC-MS analysis. Conditions: 10% (v/v) inoculum size, 30 °C, in static state culture for *Z. mobilis* ZM4; and 30 °C, 220 rpm for *G. oxydans* DSM2003. The compound with retention

time at 5.13 min and 9.17 min was identified as BQ (at m/z 108) and hydroquinone (HQ) (at m/z 110), respectively.





**Figure S3.** Cell growth, glucose consumption and D-lactic acid fermentation of *P. acidilactici* ZY15 recombinant strains with different genes in MRS medium containing 35 mg/L BQ.



**Figure S4.** Construction of the plasmids for overexpression of key genes with up-regulations by more than twofold in response to p-benzoquinone. pHW20a represents the empty plasmid vector without gene overexpression. The eight genes include one oxidoreductase gene *ZMO1772* encoding NAD(P)H quinone oxidoreductase; three reductase genes *ZMO0833* encoding UDP-N-acetylenolpyruvoylglucosamine reductase, *ZMO1303* encoding pyrroline-5-carboxylate reductase, and *ZMO1984* encoding aldo/keto reductase; three dehydrogenase genes *ZMO1949* encoding NAD(P)H dehydrogenase, *ZMO1696* encoding Zinc-binding alcohol dehydrogenase, and *ZMO1576* encoding short-chain dehydrogenase/reductase; and one fatty acid hydroxylase gene *ZMO1399*.

**Table S1** Genes and primers used in qRT-PCR assay.

Gene	Annotation	Primer sequence (5'-3')	
		Forward	Reverse
<b>Primers for the oxidoreductase gene</b>			
<i>ZMO1116</i>	Oxidoreductase	TGTGGTTTGGGCCATCCG	TGTCGGTGCGTCCTGTTTGT
<i>ZMO1772</i>	NAD(P)H quinone oxidoreductase	GGTCGCCTTGTCATTGTCG	GGCTGTTCTGGCACGCAT
<i>ZMO1885</i>	NADH flavin oxidoreductase	TGGAGTGATGCCCAAGTAGAAG	CACTGACATTAGACGGCACCATA
<b>Primers for the reductase gene</b>			
<i>ZMO1993</i>	NADPH quinone reductase	GCGGTGTCGGTAGCTTGTT	GCCTTCGCCGTGATTCTG
<i>ZMO0833</i>	UDP-N-acetylenolpyruvoylglucosamine reductase	ATCGCCTGCGTTGTGGTG	GCATTCATGCGGATCATACCA
<i>ZMO1222</i>	3-oxoacyl-(acyl-carrier-protein) reductase	TTAGCCGTGCCGTCATCAGA	CGATCATGCCTGCCTTTGC
<i>ZMO1303</i>	Pyrroline-5-carboxylate reductase	ACAACCCTGATTTCTATTCTTGCC	AACAACGCCCTTCCCTAACG
<i>ZMO1984</i>	Aldo/keto reductase	TCGCCATTTGTCAGCCTATC	CAGCAAAGTTACTGCTACCCAC
<i>ZMO1254</i>	Redoxin domain-containing protein	CGCTATGGTAATCCCTATCAGGC	GCTCATAAGCTGTGGCAAATCC
<b>Primers for the dehydrogenase gene</b>			

<i>ZMO1335</i>	NAD(P)H dehydrogenase (quinone)	CCCGATTTGGGCGTTTG	AGACCGCTCCGACCTTTCC
<i>ZMO1949</i>	NAD(P)H dehydrogenase (quinone)	TGGAGGCAAGCGGTTCTAC	GCAACAGCGGTTTGAGCAT
<i>ZMO0157</i>	D-isomer specific 2-hydroxyacid dehydrogenase	TATGGCTGTCGGGTTATTCC	TTCGGCGATGGCTTGG
<i>ZMO0788</i>	D-sorbitol dehydrogenase	CAGTTGTCACGGTCGCAATG	GCCGCAATCTGACTATCGTTTA
<i>ZMO1576</i>	Short-chain dehydrogenase/reductase	TCTATCGTCGCTAAGGGTGGTC	GCTGCAATCCCATAAAGAACG
<i>ZMO1696</i>	Zinc-binding alcohol dehydrogenase	TCCGAAGACAGGCGAAGAATG	CCTCGTCACCGACCTTAAATAGC

#### **Primers for other genes**

<i>ZMO1399</i>	Fatty acid hydroxylase	GAAACGGATTCACTATGACCAC	CCGACAGCATAACCGATAACA
<i>ZMO0020</i>	Hypothetical protein	CCGATCTAGTAAGCCAATTCACC	TTTCAAATCTGTTGGTTGGGTGT
<i>ZMO0021</i>	Hypothetical protein	CCACTTCATATCGCTTCTGTCG	GCGTAATCGGTGATCCCAA
<i>ZMO0074</i>	Hypothetical protein	CGATAAAGACCGCCCGACC	TCGCCAAGCAGGCATTCCG
<i>ZMO1821</i>	Hypothetical protein	AGAAAGCCGCCGCCATC	CACCATCATACCAACTGTCAACG
<i>ZMOr003</i>	16S rRNA	TTAAGTTGGGCACTTTAGAGGAAC	TGTCACCGCCATTGTAGCAC

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Note: The 16S rRNA gene ZMOr003 was used as the internal control to normalize the difference of the data.



**Table S2** Microbial strains, plasmids and primers used in this study.

<b>Strains</b>	<b>Genotype</b>	<b>Sources/references</b>
<i>Zymomonas mobilis</i> ZM4	Wild-type strain, ATCC31821	ATCC
<i>Gluconobacter oxydans</i> DSM2003	Wild-type strain, DSM2003	DSMZ
<i>Saccharomyces cerevisiae</i> XH7	Co-metabolize glucose and xylose into ethanol	Li et al., 2016 <sup>[1]</sup>
<i>Saccharomyces cerevisiae</i> DQ1	Thermal tolerant mutant	Chu et al., 2012 <sup>[2]</sup>
<i>Pediococcus acidilactici</i> TY112	Co-fermentation of L-lactic acid from glucose and xylose	Yi et al., 2016 <sup>[3]</sup>
<i>Zymomonas mobilis</i> 8b	Co-fermentation of ethanol from glucose and xylose	Zhang et al., 1995 <sup>[4]</sup>
<i>Escherichia coli</i> S 17-1 $\lambda\pi$	<i>Pro</i> , <i>res</i> <sup>-</sup> , <i>mod</i> <sup>+</sup> ; chromosomal integrated RP4, 2- <i>Tc</i> ::Mu- <i>Km</i> :: <i>Tn7</i> ; <i>Tp</i> , <i>sm</i>	Simon et al., 1983 <sup>[5]</sup>
<i>Amorphotheca resinae</i> ZN1	Wild-type strain	Zhang et al., 2010 <sup>[6]</sup>
<b>Plasmids</b>	<b>Description</b>	<b>Sources/references</b>
pHW20a	<i>Tc</i> <sup>r</sup> , <i>mob</i> (RP4), <i>mob</i> (RSF1010), <i>lacZ</i> $\alpha$ , MCS, and <i>oriV</i>	Dong et al., 2011 <sup>[7]</sup>
pHW20a- <i>ZMO1772</i>	NAD(P)H quinone oxidoreductase gene <i>ZMO1772</i> from ZM4	This study
pHW20a- <i>ZMO0833</i>	UDP-N-acetylenolpyruvoylglucosamine reductase gene <i>ZMO0833</i> from ZM4	This study
pHW20a- <i>ZMO1303</i>	Pyrroline-5-carboxylate reductase gene <i>ZMO1303</i> from ZM4	This study
pHW20a- <i>ZMO1984</i>	Aldo/keto reductase gene <i>ZMO1949</i> from ZM4	This study

pHW20a- <i>ZMO1949</i>	NAD(P)H dehydrogenase (quinone) gene <i>ZMO1949</i> from ZM4	This study
pHW20a- <i>ZMO1696</i>	Zinc-binding alcohol dehydrogenase gene <i>ZMO1696</i> from ZM4	This study
pHW20a- <i>ZMO1576</i>	Short-chain dehydrogenase/reductase gene <i>ZMO1576</i> from ZM4	This study
pHW20a- <i>ZMO1399</i>	Fatty acid hydroxylase gene <i>ZMO1399</i> from ZM4	This study

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**Primers for gene amplification**
**Sequence (5'-3')**

<i>ZMO1772</i> -F	CCCA <u>AAGCTT</u> ATGGCAGGCAACATGATGAAAG
<i>ZMO1772</i> -R	CGGGGTACCTCAAAAAAGCGCCTCTGGC
<i>ZMO0833</i> -F	CCCA <u>AAGCTT</u> ATGACGACTGCTACATCTTCC
<i>ZMO0833</i> -R	CGGGGTACCTCATGCCTTATCCCCATC
<i>ZMO1303</i> -F	CCCA <u>AAGCTT</u> ATGAGTGATACCGCATCAGATTC
<i>ZMO1303</i> -R	CGGGGTACCTTATTGCGCGATAGTCTCTTTG
<i>ZMO1984</i> -F	CCCA <u>AAGCTT</u> ATGGATTATACGTATTTGGGTCG
<i>ZMO1984</i> -R	GCT <u>CTAGACT</u> ACCATGCATAGGCTTCAGG
<i>ZMO1949</i> -F	CCCA <u>AAGCTT</u> ATGAAAGTATTGATCGTTCACGC
<i>ZMO1949</i> -R	CGGGGTACCTCATGGTTGTTGTTTCCTCAAAC
<i>ZMO1696</i> -F	CCCA <u>AAGCTT</u> ATGCGCGCCATAGGTTATC

ZMO1696-R	CGGGGTACCTTAGAAGCCTTCTAAGACGATTTTA
ZMO1576-F	CCCAAGCTTATGAACCAGAATATCCGCAA
ZMO1576-R	CGGGGTACCTTATAATGCCTGTTTTGTCGG
ZMO1399-F	CCCAAGCTTATGAACACAACACTGATGCCAAGAC
ZMO1399-R	CGGGGTACCTCAGCGGATATCCTGTATTTTATC

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Note: The underline indicates the digestion sites.

## REFERENCES

1. Li H, Shen Y, Wu M, Hou J, Jiao C, Li Z, Liu X, Bao X. 2016. Engineering a wild-type diploid *Saccharomyces cerevisiae* strain for second-generation bioethanol production. *Bioresour Bioprocess* 3:51.
2. Chu D, Zhang J, Bao J. 2012. Simultaneous saccharification and ethanol fermentation of corn stover at high temperature and high solids loading by a thermotolerant strain *Saccharomyces cerevisiae* DQ1. *Bioenergy Res* 5:1020-1026.
3. Yi X, Zhang P, Sun J, Tu Y, Gao Q, Zhang J, Bao J. 2016. Engineering wild-type robust *Pediococcus acidilactici* strain for high titer l- and d-lactic acid production from corn stover feedstock. *J Biotechnol* 217:112-121.
4. Zhang M, Eddy C, Deanda K, Finkelstein M, Picataggio S. 1995. Metabolic Engineering of a pentose metabolism pathway in ethanologenic *Zymomonas mobilis*. *Science* 267:240-243.
5. Simon R, Prierer U, Puhler A. 1983. A broad host range mobilization system for in vivo genetic engineering: Transposon mutagenesis in

gram negative bacteria. *Bio Technol* 1:784-791.

6. Zhang J, Zhu Z, Wang X, Wang N, Wang W, Bao J. 2010. Biodetoxification of toxins generated from lignocellulose pretreatment using a newly isolated fungus, *Amorphotheca resinae* ZN1, and the consequent ethanol fermentation. *Biotechnol Biofuels* 3:26.
7. Dong HW, Bao J, Ryu DDY, Zhong JJ. 2011. Design and construction of improved new vectors for *Zymomonas mobilis* recombinants. *Biotechnol Bioeng* 108:1616-1627.