

## Supporting Information

### **A comparative assessment of conventional and molecular methods, including MinION nanopore sequencing, for surveying water quality**

*Kishor Acharya<sup>†</sup>, Santosh Khanal<sup>‡</sup>, Kalyan Pantha<sup>#‡</sup>, Niroj Amatya<sup>¥‡</sup>, Russell J. Davenport<sup>†</sup>,  
David Werner<sup>†\*</sup>*

<sup>†</sup> School of Engineering, Newcastle University, Newcastle upon Tyne, NE1 7RU, United Kingdom

<sup>‡</sup> Department of Pharmacology, School of Medicine, University of Colorado, Aurora, Colorado 80045, USA

<sup>#</sup> Group for Rural Infrastructure Development, Wise use House, Jwagal, Lalitpur, Nepal

<sup>†</sup> Faculty of Chemistry, University Duisburg-Essen, Universitätsstr. 5, D-45141 Essen, Germany

<sup>¥</sup> Department of Medical Microbiology, Nobel College, Pokhara University, Kathmandu, Nepal

\*Corresponding author: [david.werner@ncl.ac.uk](mailto:david.werner@ncl.ac.uk)

## Chemical water quality assessment for different water sources in the Kathmandu Valley

### Methods

Grab water samples were analysed for pH, dissolved oxygen, electrical conductivity, and temperature on-site using a portable probe from EXTECH INSTRUMENTS (Boston, USA). For chemical analysis, water samples were filtered on-site through 0.22 µm PVDF syringe filters (Gilson Scientific Ltd, Dunstable UK), collected in two 50 mL sterile polyethylene vials and then the vials were frozen for analysis at NU. HACH cuvette test kits were used to measure nitrite ( $\text{NO}_2^-$ -N), nitrate ( $\text{NO}_3^-$ -N), ammonia ( $\text{NH}_3$ -N), and fluoride ( $\text{F}^-$ ) in the filtered samples, following the manufacturer's instructions (HACH LANGE LTD, Manchester, UK). Similarly, alkalinity was measured using a HACH digital titrator, following the manufacturer's instructions (HACH LANGE LTD, Manchester, UK). Analysis of filtered metals was undertaken using a Varian Vista-MPX ICP-OES or Agilent 770 Series ICP-MS, as appropriate for the metal concentration. Anion analysis was conducted using a Dionex DX320 ion chromatograph. Blanks and standards were used throughout and duplicate samples were analysed for HACH cuvette test. For quality assurance, charge balances were calculated to check results for electric neutrality considering  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{HCO}_3^-$ ,  $\text{Cl}^-$ ,  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$  charges <sup>1</sup>.

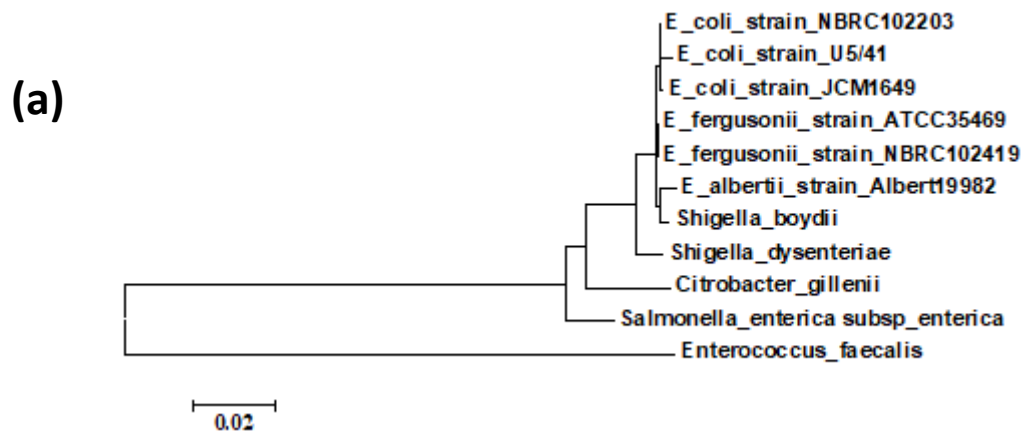
### Results

In addition to microbial water quality it is important to not overlook chemical water quality issues. **Table S8** compares concentrations of different relevant ions in the water from the 13 different sources. The Nepali national drinking-water quality standard and the WHO guideline for nitrate concentration in drinking water is 50 mg/L, <sup>2,3</sup>; water samples from two sources, the stone spout (location 11) and a tube well (location 8), contained concentrations at or slightly above this guideline value. Chloride, nitrite and fluoride concentrations were

below WHO guidelines of 250 mg/L, 3 mg/L and 1.5 mg/L, respectively <sup>2</sup>, in all the water samples (Nepali standard for chloride, fluoride and nitrite are not available), while sulphate concentrations were below the Nepali standard of 250 mg/L in all the waters. The Nepali standard for ammonia of 1.5 mg/L was violated in one water sample (location 7; tube well) with a concentration of 1.779 mg/L. The pH of water samples ranged from 6.2 to 8.16 (**Table S9**). The Nepali national water quality guideline requires pH to be between 6.5 and 8.5 <sup>3</sup>. The pH was < 6.5 in three of the sources (location 7, 8 and 13). None of the water sources violated the Nepali standard of 1500 uS/cm specific conductance (**Table S9**).

**Table S10** shows concentrations of different metals analysed in the water from the 13 sources. A study by *Shrestha et. al.* reported heavy metals contamination in deep groundwater sources in the Kathmandu Valley, especially arsenic, zinc, lead, iron and manganese <sup>4</sup>. However, none of the filtered samples collected in this study exceeded the Nepali water-quality standard for arsenic, zinc and lead of 50 ug/L, 3 mg/L and 0.01 mg/L, respectively. Exceedance of the Nepali standard for iron of 0.3 mg/L (WHO guideline for iron not available) was observed in the water from two sources; location 5 (deep borehole) and location 7 (tube well). Likewise, samples from three sources; location 5 (deep borehole), location 6 (tube well) and location 7 (tube well), violated the WHO (0.1 mg/L) drinking water quality guideline for manganese. Locations 5 and 7 violated the Nepali manganese standard (0.2 mg/L). The WHO and Nepali standard for aluminium of 0.2 mg/L was violated in one water sample (location 4; tube well) with a concentration of 0.214 mg/L.

Concentrations of all other metals, including Ca, Mg, Na, K, Ba, Cu, Cd, Sb, Si, Ni, Cr and Sr were well below the Nepali and WHO guidelines.



(b)

Percent Identity Matrix - created by Clustal2.1

1: <i>Enterococcus faecalis</i>	100.00	75.28	76.72	76.38	75.50	75.82	76.03	75.58	76.14	75.72	76.87
2: <i>Citrobacter gillenii</i>	75.28	100.00	96.07	96.23	95.31	95.72	95.35	96.18	95.61	96.25	96.40
3: <i>Salmonella enterica</i>	76.72	96.07	100.00	97.11	96.28	96.92	97.16	96.86	97.33	96.93	96.40
4: <i>Shigella dysenteriae</i>	76.38	96.23	97.11	100.00	98.05	98.31	98.65	98.77	98.99	98.84	98.92
5: <i>E_coli_strain_U5/41</i>	75.50	95.31	96.28	98.05	100.00	98.18	98.39	98.53	98.83	98.81	99.37
6: <i>E_albertii_strain_Albert19982</i>	75.82	95.72	96.92	98.31	98.18	100.00	99.20	98.98	99.13	99.05	99.28
7: <i>Shigella boydii</i>	76.03	95.35	97.16	98.65	98.39	99.20	100.00	99.32	99.60	99.18	99.37
8: <i>E_fergusonii_strain_NBRC102419</i>	75.58	96.18	96.86	98.77	98.53	98.98	99.32	100.00	99.59	99.32	99.73
9: <i>E_fergusonii_strain_ATCC35469</i>	76.14	95.61	97.33	98.99	98.83	99.13	99.60	99.59	100.00	99.59	99.73
10: <i>E_coli_strain_NBRC102203</i>	75.72	96.25	96.93	98.84	98.81	99.05	99.18	99.32	99.59	100.00	99.91
11: <i>E_coli_strain_JCM1649</i>	76.87	96.40	96.40	98.92	99.37	99.28	99.37	99.73	99.73	99.91	100.00

**Figure S1** (a) Phylogenetic distance tree (Neighbour-Joining) generated with 16S rRNA sequences of *E coli*, *E fergusonii*, closely related other strains and a distant strain. The 16S rRNA sequences were obtained from the NCBI 16S rRNA database. (b) The percent identity matrix of the 16S rRNA sequences used for creating the phylogenetic distance tree.

**Table S1** Actual composition of different bacterial species present in the Mock Community based on genomic DNA<sup>5</sup>.

Microorganisms	Percentage DNA composition (%)	16S rRNA gene composition (%)
<i>Listeria monocytogenes</i>	12	14.1
<i>Pseudomonas aeruginosa</i>	12	4.2
<i>Bacillus subtilis</i>	12	17.4
<i>Escherichia coli</i>	12	10.1
<i>Salmonella enterica</i>	12	10.4
<i>Lactobacillus fermentum</i>	12	18.4
<i>Enterococcus faecalis</i>	12	9.9
<i>Staphylococcus aureus</i>	12	15.5
<i>Saccharomyces cerevisiae</i>	2	na
<i>Cryptococcus neoformans</i>	2	na

**Table S2** MinION 16S rRNA sequencing result for the MOCK Community at species level. The 16S rRNA amplicon sequencing was performed in triplicate. The species highlighted in blue are the actual bacteria who's DNA are present in the MOCK community.

Species	Average number of reads	Standard deviation	16S rRNA gene percentage (%)	Standard deviation
<i>Lactobacillus fermentum</i>	13446.33	3666.82	9.25	0.13
<i>Staphylococcus aureus</i>	12443.67	4241.45	8.47	0.54
<i>Enterococcus faecalis</i>	7831.33	2077.18	5.39	0.10
<i>Salmonella enterica</i>	7177.33	2420.26	4.89	0.28
<i>Bacillus mojavensis</i>	5848.00	1884.89	3.99	0.18
<i>Pseudomonas aeruginosa</i>	5373.33	1811.16	3.66	0.32
<i>Bacillus halotolerans</i>	4772.33	1403.69	3.27	0.06
<i>Bacillus subtilis</i>	4509.00	1242.45	3.10	0.05
<i>Listeria innocua</i>	3211.00	1002.29	2.19	0.07
<i>Bacillus vallismortis</i>	1675.67	305.03	1.17	0.12
<i>Staphylococcus simiae</i>	1669.00	465.84	1.15	0.06
<i>Listeria seeligeri</i>	1412.00	370.07	0.97	0.04
<i>Listeria monocytogenes</i>	1096.33	68.09	0.78	0.15
<i>Escherichia fergusonii</i>	940.00	198.87	0.65	0.06
<i>Bacillus atrophaeus</i>	858.67	251.42	0.59	0.03
<i>Listeria ivanovii</i>	775.00	288.97	0.52	0.05
<i>Lactobacillus gastricus</i>	746.33	246.02	0.51	0.04
<i>Bacillus amyloliquefaciens</i>	584.00	137.07	0.40	0.02
<i>Staphylococcus capitis</i>	602.67	192.27	0.41	0.02
<i>Shigella sonnei</i>	449.67	93.35	0.31	0.06
<i>Staphylococcus haemolyticus</i>	452.67	117.50	0.31	0.02
<i>Bacillus altitudinis</i>	456.00	89.54	0.32	0.04
<i>Shigella flexneri</i>	421.33	143.54	0.29	0.02
<i>Staphylococcus hominis</i>	339.33	88.93	0.23	0.00
<i>Staphylococcus auricularis</i>	211.00	24.52	0.15	0.04

**Table S2 Continued**

<i>Citrobacter koseri</i>	250.67	34.30	0.18	0.02
<i>Escherichia coli</i>	247.33	31.79	0.17	0.02
<i>Bacillus nematocida</i>	257.00	45.90	0.18	0.02
<i>Bacillus velezensis</i>	238.00	55.07	0.16	0.01
<i>Staphylococcus caprae</i>	251.33	46.06	0.18	0.02
<i>Bacillus nakamurai</i>	213.33	13.43	0.15	0.03
<i>Lactobacillus equigenerosi</i>	221.67	39.50	0.16	0.02
<i>Staphylococcus saccharolyticus</i>	185.33	57.83	0.13	0.01
<i>Listeria marthii</i>	190.33	33.50	0.13	0.02
<i>Pseudomonas otitidis</i>	182.67	73.16	0.12	0.02
<i>Bacillus licheniformis</i>	177.33	44.74	0.12	0.01
<i>Staphylococcus petrasii</i>	148.00	18.33	0.10	0.01
<i>Listeria grayi</i>	147.33	41.79	0.10	0.01
<i>Enterococcus wangshanyuanii</i>	132.00	14.93	0.09	0.01
<i>Bacillus cereus</i>	143.00	28.62	0.10	0.01
<i>Bacillus shackletonii</i>	153.00	24.25	0.11	0.02
<i>Enterococcus rivorum</i>	163.33	42.02	0.11	0.02
<i>Pseudomonas resinovorans</i>	144.67	48.09	0.10	0.01
<i>Staphylococcus sciuri</i>	123.67	9.07	0.09	0.02
<i>Enterococcus faecium</i>	147.67	30.57	0.10	0.02
<i>Staphylococcus delphini</i>	121.33	21.39	0.08	0.01
<i>Lactobacillus gorillae</i>	137.33	35.92	0.09	0.01
<i>Bacillus stratosphericus</i>	95.33	8.02	0.07	0.02
<i>Lactobacillus reuteri</i>	124.67	59.00	0.08	0.02
<i>Bacillus siamensis</i>	109.00	28.93	0.08	0.00
<i>Bacillus zhangzhouensis</i>	99.33	6.81	0.07	0.01
<i>Enterococcus hirae</i>	106.33	27.65	0.07	0.00
<i>Bacillus firmus</i>	111.00	27.87	0.08	0.00
<i>Staphylococcus piscifermentans</i>	95.67	13.87	0.07	0.01
<i>Macrococcus caseolyticus</i>	99.67	15.89	0.07	0.01
<i>Listeria aquatica</i>	79.67	9.02	0.06	0.02
<i>Staphylococcus epidermidis</i>	115.33	42.44	0.08	0.01
<i>Bacillus pumilus</i>	106.33	24.58	0.07	0.01
<i>Bacillus safensis</i>	109.67	27.50	0.08	0.01
<i>Pseudomonas alcaligenes</i>	82.67	18.77	0.06	0.01
<i>Lactobacillus mucosae</i>	112.00	42.53	0.08	0.01
<i>Bacillus swezeyi</i>	107.67	31.88	0.07	0.01
<i>Bacillus haynesii</i>	72.33	2.08	0.05	0.01
<i>Klebsiella pneumoniae</i>	96.67	25.54	0.07	0.01
<i>Staphylococcus devriesei</i>	89.33	18.61	0.06	0.01
<i>Bacillus acidicola</i>	81.67	10.50	0.06	0.01
<i>Pseudomonas stutzeri</i>	85.33	19.86	0.06	0.00

**Table S2 Continued**

<i>Staphylococcus massiliensis</i>	80.67	11.93	0.06	0.02
<i>Enterococcus sulfureus</i>	94.33	26.41	0.06	0.01
<i>Escherichia marmotae</i>	86.33	20.98	0.06	0.00
<i>Lactobacillus ingluviei</i>	88.67	31.56	0.06	0.00
<i>Enterobacter cloacae</i>	69.00	5.29	0.05	0.01
<i>Enterococcus saccharolyticus</i>	93.67	35.53	0.06	0.01
<i>Bacillus pseudofirmus</i>	84.00	32.05	0.06	0.01
<i>Staphylococcus kloosii</i>	75.33	24.21	0.05	0.00
<i>Bacillus ginsengihumi</i>	66.67	5.69	0.05	0.01
<i>Listeria floridensis</i>	79.67	22.50	0.05	0.00
<i>Pseudomonas pseudoalcaligenes</i>	60.33	13.50	0.04	0.01
<i>Bacillus sporothermodurans</i>	65.00	12.17	0.05	0.00
<i>Staphylococcus carnosus</i>	65.33	7.09	0.05	0.01
<i>Staphylococcus felis</i>	62.33	4.73	0.04	0.01
<i>Shigella dysenteriae</i>	65.67	17.67	0.05	0.00
<i>Bacillus cohnii</i>	64.00	11.00	0.04	0.01
<i>Pseudomonas argentinensis</i>	48.33	3.51	0.04	0.01
<i>Enterococcus termitis</i>	56.67	5.77	0.04	0.01
<i>Citrobacter farmeri</i>	71.33	32.01	0.05	0.01
<i>Bacillus songklensis</i>	67.67	20.01	0.05	0.01
<i>Bacillus paralicheniformis</i>	48.00	10.00	0.04	0.01
<i>Staphylococcus simulans</i>	53.00	11.27	0.04	0.00
<i>Escherichia albertii</i>	67.33	19.14	0.05	0.01
<i>Bacillus herbersteinensis</i>	39.00	6.24	0.03	0.01
<i>Shigella boydii</i>	50.67	10.79	0.04	0.00
<i>Bacillus carboniphilus</i>	49.67	3.51	0.04	0.01
<i>Bacillus halmapalus</i>	52.67	10.26	0.04	0.00
<i>Bacillus luciferensis</i>	38.33	5.51	0.03	0.01
<i>Lactobacillus siliginis</i>	42.33	3.51	0.03	0.01
<i>Bacillus mycoides</i>	52.33	17.21	0.04	0.00
<i>Bacillus galliciensis</i>	49.67	15.95	0.03	0.00
<i>Pseudomonas jinjuensis</i>	41.33	2.08	0.03	0.01
<i>Bacillus thuringiensis</i>	35.00	3.46	0.02	0.01
<i>Bacillus aquimaris</i>	43.00	8.89	0.03	0.00
<i>Listeria weihenstephanensis</i>	35.33	11.02	0.02	0.01
<i>Staphylococcus chromogenes</i>	50.33	14.01	0.03	0.01
<i>Bacillus flexus</i>	43.00	10.58	0.03	0.00
<i>Enterococcus pseudoavium</i>	43.00	7.55	0.03	0.00
<i>Shimwellia blattae</i>	42.33	10.21	0.03	0.00
<i>Bacillus cytotoxicus</i>	36.00	11.14	0.02	0.00
<i>Bacillus isabelliae</i>	34.33	4.51	0.03	0.01
<i>Kosakonia sacchari</i>	32.33	10.60	0.02	0.01
<i>Enterococcus avium</i>	37.33	9.29	0.03	0.00

**Table S2 Continued**

<i>Bacillus gottheilii</i>	36.33	5.86	0.03	0.00
<i>Cronobacter sakazakii</i>	25.33	5.86	0.02	0.01
<i>Pseudomonas guguanensis</i>	37.67	5.77	0.03	0.01
<i>Bacillus aerius</i>	49.33	16.26	0.03	0.01
<i>Staphylococcus gallinarum</i>	39.00	8.54	0.03	0.00
<i>Bacillus xiamenensis</i>	39.00	8.19	0.03	0.01
<i>Bacillus zhanjiangensis</i>	31.33	2.31	0.02	0.00
<i>Pseudomonas mendocina</i>	32.00	2.00	0.02	0.01
<i>Bacillus deserti</i>	29.67	2.52	0.02	0.00
<i>Enterococcus moraviensis</i>	41.67	17.47	0.03	0.01
<i>Staphylococcus lugdunensis</i>	31.33	4.51	0.02	0.01
<i>Enterococcus dispar</i>	32.00	6.24	0.02	0.00
<i>Bacillus mediterraneensis</i>	26.67	3.06	0.02	0.00
<i>Bacillus timonensis</i>	29.67	3.51	0.02	0.01
<i>Enterococcus haemoperoxidus</i>	32.00	6.00	0.02	0.00
<i>Enterococcus canis</i>	41.67	16.04	0.03	0.01
<i>Staphylococcus xylosus</i>	26.33	3.21	0.02	0.00
<i>Staphylococcus stepanovicii</i>	30.67	6.11	0.02	0.00
<i>Staphylococcus cohnii</i>	27.33	3.51	0.02	0.00
<i>Bacillus fastidiosus</i>	23.67	7.02	0.02	0.00
<i>Enterococcus asini</i>	25.67	6.43	0.02	0.01
<i>Lactobacillus vaccinostrercus</i>	33.00	14.00	0.02	0.00
<i>Listeria rocourtiae</i>	28.00	4.58	0.02	0.00
<i>Others</i>	58901.33	15512.66	40.59	0.42
<b>Total number of reads</b>	<b>145392</b>	<b>39928.05</b>		



**Table S3** Plate count results for Fecal *E.coli* and Total coliform in the water samples from 13 different sites. Data points are an average of duplicate biological samples and the number in parentheses indicates the standard deviation. Where required, the dilution was performed with normal saline solution (NS). Two blanks; membrane filter (MF) and NS, were also tested for Fecal *E.coli* and Total coliform in duplicate.

Sample Site ID	1	2	3	4	5	6	7	8	9	10	11	12	13	Blank (MF)	Blank (NS)
<b>Faecal <i>E coli</i> (CFU/100 mL)</b> <b>WHO and Nepalese guideline value: 0 CFU/100 mL</b>	0	0	0	0	0	0	0	0.5 (0.7)	215 (134.4)	0	1.5 (0.7)	3 (1.4)	0	0	0
<b>Total Coliform (CFU/100 mL)</b> <b>WHO and Nepalese guideline value: 0 CFU/100 mL</b>	0	0	0	0	0	330 (14.14)	>20000	>200	>2000	>2000	88.5 (2.12)	39.5 (14.85)	0	0	0

**Table S4** Relative abundances of different putative genera estimated from 16S rRNA reads sequenced with the Illumina NGS platform. The numbers are an average of duplicate samples and the numbers in parenthesis indicate the standard deviation. n.d indicated not detected.

<b>Location</b>	<i><b>Clostridium</b></i>	<i><b>Vibrio</b></i>	<i><b>Legionella</b></i>	<i><b>Streptococcus</b></i>	<i><b>Prevotella</b></i>	<i><b>Bacteroides</b></i>	<i><b>Pseudomonas</b></i>	<b>Total Reads</b>
<b>Location 2</b>	1.70E-02 (1.20E-02)	2.28E-02 (4.62E-03)	n.d	6.98E-04 (2.32E-04)	7.59E-04 (7.61E-04)	n.d	2.14E-03 (3.25E-04)	42258 (4162)
<b>Location 3</b>	n.d	8.38E-04 (8.32E-05)	2.96E-05 (4.19E-05)	4.50E-04 (8.61E-05)	3.24E-04 (8.11E-05)	n.d	2.22E-04 (3.24E-05)	130401 (63480)
<b>Location 4</b>	1.08E-02 (8.03E-04)	9.48E-05 (2.79E-07)	n.d	6.49E-04 (8.61E-06)	4.60E-04 (8.85E-05)	1.04E-04 (1.47E-04)	1.68E-02 (3.52E-03)	79055 (6469)
<b>Location 5</b>	3.69E-03 (2.01E-03)	2.18E-04 (1.36E-04)	6.17E-05 (8.72E-05)	n.d	7.58E-05 (4.38E-05)	n.d	n.d	77394 (51602)
<b>Location 6</b>	1.82E-02 (1.61E-03)	1.90E-04 (3.86E-05)	4.75E-04 (3.02E-05)	n.d	n.d	1.60E-05 (2.26E-05)	1.95E-04 (5.85E-05)	139606 (37060)
<b>Location 7</b>	1.42E-02 (7.22E-03)	1.60E-05 (2.26E-05)	n.d	n.d	n.d	3.16E-04 (4.12E-05)	3.76E-05 (5.31E-05)	144883 (11351)
<b>Location 8</b>	2.10E-02 (1.04E-02)	5.69E-04 (1.75E-05)	6.04E-04 (1.12E-04)	n.d	n.d	6.39E-05 (5.46E-05)	4.85E-04 (1.04E-04)	122936 (16706)
<b>Location 9</b>	9.39E-02 (1.40E-02)	n.d	3.31E-04 (2.21E-05)	n.d	2.09E-03 (4.43E-04)	1.38E-03 (1.52E-04)	1.34E-02 (4.83E-05)	115934 (64222)
<b>Location 10</b>	2.25E-01 (4.66E-02)	8.38E-05 (1.27E-05)	4.46E-04 (4.88E-05)	4.22E-05 (5.96E-05)	4.53E-03 (1.29E-03)	3.51E-03 (5.70E-04)	2.99E-02 (7.50E-04)	112765 (23757)
<b>Location 11</b>	1.59E-02 (3.21E-03)	1.57E-03 (1.79E-04)	2.67E-04 (3.78E-04)	5.55E-05 (7.85E-05)	1.59E-03 (5.97E-04)	4.17E-04 (2.35E-04)	8.15E-04 (5.75E-04)	33930 (7285)
<b>Location 12</b>	2.33E-01 (3.29E-01)	2.92E-04 (2.07E-04)	1.32E-04 (1.86E-04)	2.83E-03 (9.55E-04)	2.75E-02 (6.75E-03)	5.22E-03 (4.29E-05)	3.63E-03 (2.03E-03)	156004 (48730)
<b>Location 13</b>	n.d	3.96E-04 (1.73E-04)	3.68E-03 (2.65E-04)	n.d	2.76E-04 (2.38E-06)	6.80E-05 (2.63E-05)	4.21E-04 (3.72E-05)	115666 (49772)

**Table S5** Relative abundances of different putative genera estimated from 16S rRNA reads sequenced with the MinION NGS platform. n.d indicated not detected.

<b>Location</b>	<i>Clostridium</i>	<i>Vibrio</i>	<i>Legionella</i>	<i>Streptococcus</i>	<i>Prevotella</i>	<i>Bacteroides</i>	<i>Pseudomonas</i>
<b>Location 2</b>	6.61E-04	3.21E-03	n.d	5.19E-04	n.d	n.d	1.39E-03
<b>Location 3</b>	2.64E-05	2.64E-05	4.40E-05	2.20E-04	1.76E-05	n.d	5.02E-04
<b>Location 4</b>	1.44E-04	1.29E-05	2.14E-05	4.40E-04	3.22E-05	6.43E-06	8.33E-02
<b>Location 5</b>	4.08E-05	3.50E-05	1.28E-04	4.08E-05	3.50E-05	1.17E-05	6.94E-04
<b>Location 6</b>	6.42E-04	6.52E-05	1.13E-03	1.36E-04	2.77E-05	6.91E-05	7.29E-04
<b>Location 7</b>	2.21E-04	9.75E-05	1.28E-04	3.75E-05	1.50E-05	3.38E-05	3.51E-03
<b>Location 8</b>	7.95E-04	1.32E-04	1.84E-03	1.54E-04	1.72E-05	2.29E-05	1.60E-03
<b>Location 9</b>	1.47E-03	6.51E-05	1.38E-03	1.86E-04	2.86E-04	2.17E-04	1.74E-02
<b>Location 11</b>	4.49E-03	1.88E-04	8.26E-03	2.37E-04	6.43E-04	3.16E-04	1.98E-02
<b>Location 12</b>	5.30E-05	1.44E-04	1.98E-03	4.00E-03	9.44E-03	3.91E-03	5.23E-04
<b>Location 13</b>	n.d	4.55E-05	3.52E-03	2.27E-05	3.03E-05	3.79E-05	3.93E-02

**Table S6** Results of the Spearman rank correlation analysis (n = 12) between faecal indicator bacterial families or genera across all the water samples analysed with Illumina (Table S4) and MinION (Table S5).

<b>Putative pathogenic genera or family</b>	<b>Spearman rank Correlation coefficient</b>	<b>p-value</b>
Clostridium_MinION vs Clostridium_Illumina	0.96	< 0.05
Legionella_MinION vs Legionella_Illumina	0.82	< 0.05
Pseudomonas_MinION vs Pseudomonas_Illumina	0.51	< 0.05
Streptococcus_MinION vs Streptococcus_Illumina	0.89	< 0.05
Vibrio_MinION vs Vibrio_Illumina	0.52	< 0.05
Bacteroides_MinION vs Bacteroides_Illumina	0.84	< 0.05
Prevotella_MinION vs Prevotella_Illumina	0.66	< 0.05
Enterobacteriaceae_MinION vs Enterobacteriaceae_Illumina	0.81	< 0.05

**Table S7** 16S rRNA gene sequencing reads from MinION NGS platform matching the putative pathogens listed by HSE <sup>6</sup>, and total number of reads per samples are reported in last row.

Potential Pathogens <sup>6</sup>	Sampling Site ID										
	2	3	4	5	6	7	8	9	11	12	13
<i>Actinobacillus actinomycetemcomitans</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Actinomadura madurae</i>	0	0	0	0	1	0	0	0	0	1	0
<i>Actinomadura pelletieri</i>	0	0	0	0	0	0	0	0	0	2	0
<i>Actinomyces gerencseriae</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Actinomyces israelii</i>	0	0	0	0	0	0	0	0	0	1	0
<i>Actinomyces pyogenes</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Arcanobacterium haemolyticum</i>	0	0	0	0	1	0	0	1	0	2	0
<i>Arcanobacterium pyogenes</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Arcobacter butzleri</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Bacillus anthracis</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Bacillus cereus</i>	0	0	0	0	0	0	2	0	4	0	3
<i>Bacteroides fragilis</i>	0	0	0	0	0	1	0	0	1	10	0
<i>Bartonella bacilliformis</i>	1	0	4	0	2	0	2	4	0	0	0
<i>Bartonella quintana</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Bordetella bronchiseptica</i>	0	0	0	2	23	2	0	0	0	0	0
<i>Bordetella parapertussis</i>	0	2	0	1	40	6	0	0	2	0	5
<i>Bordetella pertussis</i>	0	0	0	0	1	0	0	0	0	0	1
<i>Borrelia burgdorferi</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Borrelia duttonii</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Borrelia recurrentis</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Brucella abortus</i>	0	0	309	1	0	0	0	0	0	0	0
<i>Brucella canis</i>	0	0	185	0	1	0	0	1	0	0	2
<i>Brucella melitensis</i>	0	1	10	0	0	0	0	0	0	0	0
<i>Brucella suis</i>	0	0	17	0	0	0	0	0	0	0	0
<i>Burkholderia cepacia</i>	0	0	0	2	1	1	1	2	0	0	0

**Table S7 Continued**

<i>Burkholderia mallei</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Burkholderia pseudomallei</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Campylobacter fetus</i>	0	0	0	0	2	0	0	0	0	0	0
<i>Campylobacter jejuni</i>	0	0	0	0	3	0	0	0	1	0	0
<i>Cardiobacterium hominis</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Chlamydophila pneumoniae</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Chlamydophila psittaci</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Chlamydophila trachomatis</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Clostridium botulinum</i>	0	0	0	0	3	0	0	0	1	1	0
<i>Clostridium perfringens</i>	0	0	1	0	1	0	1	4	0	3	0
<i>Clostridium tetani</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Corynebacterium diphtheriae</i>	0	7	0	0	1	0	0	0	0	0	0
<i>Corynebacterium haemolyticum</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Corynebacterium minutissimum</i>	0	0	1	0	0	0	0	2	0	0	0
<i>Corynebacterium pseudotuberculosis</i>	0	3	0	0	0	0	0	1	0	0	0
<i>Corynebacterium pyogenes</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Corynebacterium ulcerans</i>	0	1	0	0	0	0	0	1	0	0	0
<i>Coxiella burnetti</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Edwardsiella tarda</i>	0	0	0	0	1	0	0	1	0	0	0
<i>Ehrlichia sennetsu</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Eikenella corrodens</i>	0	0	0	1	0	0	0	0	0	0	0
<i>Elizabethkingia meningoseptica</i>	0	0	0	0	2	0	1	0	0	4	0
<i>Enterobacter cloacae</i>	1	0	0	0	1	1	3	4	10	4	0
<i>Erysipelothrix rhusiopathiae</i>	0	0	0	0	46	0	0	1	0	0	0
<i>Escherichia coli</i>	0	0	0	0	0	0	0	0	0	1	0
<i>Flavobacterium meningosepticum</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Fluoribacter bozemanii</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Francisella tularensis</i>	0	0	0	3	0	0	1	0	0	0	0



**Table S7 Continued**

<i>Mycobacterium xenopi</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Mycoplasma caviae</i>	0	0	0	0	0	0	0	0	1	0	2
<i>Mycoplasma hominis</i>	0	0	0	0	2	0	0	0	1	0	0
<i>Mycoplasma pneumoniae</i>	0	0	0	0	0	1	0	0	0	0	0
<i>Neisseria gonorrhoeae</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Neisseria meningitidis</i>	0	0	0	0	1	0	1	0	0	1	0
<i>Nocardia asteroides</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Nocardia braziliensis</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Nocardia farcinica</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Nocardia nova</i>	0	0	0	0	0	0	0	1	0	0	0
<i>Nocardia otitidiscaviarum</i>	0	0	0	0	2	0	0	1	0	0	0
<i>Pasteurella multocida</i>	0	0	0	0	0	1	0	0	0	0	0
<i>Peptostreptococcus anaerobius</i>	0	0	0	0	1	0	0	0	0	0	0
<i>Plesiomonas shigelloides</i>	0	0	1	0	0	1	0	0	0	0	0
<i>Proteus mirabilis</i>	0	0	0	1	0	0	0	0	11	0	0
<i>Proteus penneri</i>	0	0	0	0	0	1	0	0	0	0	0
<i>Proteus vulgaris</i>	0	0	0	0	0	0	0	0	1	0	0
<i>Providencia alcalifaciens</i>	0	0	1	0	0	0	0	0	0	1	0
<i>Providencia rettgeri</i>	0	0	0	0	0	0	0	3	0	1	0
<i>Pseudomonas aeruginosa</i>	2	0	1434	2	5	14	12	47	8	43	0
<i>Pseudomonas mallei</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Pseudomonas pseudomallei</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Rhodococcus equi</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Rickettsia akari</i>	0	0	0	0	7	0	0	0	0	0	0
<i>Rickettsia canada</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Rickettsia conorii</i>	0	0	0	0	1	1	2	2	7	3	3
<i>Rickettsia montana</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Rickettsia mooseri</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Rickettsia prowazekii</i>	0	0	0	0	1	0	1	0	3	0	0

**Table S7 continued**

<i>Rickettsia rickettsia</i>	0	0	0	0	1	0	0	0	1	0	0
<i>Rickettsia sennetsu</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Rickettsia tsutsugamushi</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Rickettsia typhi</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Rochalimaea quintana</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Salmonella arizonae</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Salmonella enterica</i>	0	0	0	3	2	0	1	1	0	2	1
<i>Salmonella paratyphi</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Salmonella typhi</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Shigella boydii</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Shigella dysenteriae</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Shigella flexneri</i>	0	0	0	0	2	0	0	0	0	1	0
<i>Shigella sonnei</i>	0	0	0	0	0	0	0	1	0	0	0
<i>Staphylococcus aureus</i>	0	0	0	0	0	0	0	0	1	0	0
<i>Streptobacillus moniliformis</i>	0	0	0	0	2	0	0	1	0	1	0
<i>Streptococcus agalactiae</i>	0	0	0	0	1	0	0	0	0	0	0
<i>Streptococcus dysgalactiaeaequisimilis</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Streptococcus pneumoniae</i>	0	1	14	0	1	2	0	3	0	1	0
<i>Streptococcus pyogenes</i>	0	0	0	0	0	0	0	0	0	1	0
<i>Streptococcus suis</i>	2	0	5	1	2	3	1	1	1	49	0
<i>Treponema carateum</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Treponema pallidum</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Treponema pertenue</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Ureaplasma parvum</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Ureaplasma urealyticum</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Vibrio cholerae</i>	0	0	0	0	1	3	2	0	0	0	0
<i>Vibrio parahaemolyticus</i>	0	0	1	0	0	2	1	3	2	0	0
<i>Yersinia enterocolitica</i>	0	0	0	0	0	0	0	1	0	0	0



**Table S7 continued**

<i>Yersinia pestis</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Yersinia pseudotuberculosis</i>	0	0	0	0	0	0	0	0	1	0	0
<b>Total Number of Reads</b>	<b>42370</b>	<b>113607</b>	<b>466260</b>	<b>171502</b>	<b>506157</b>	<b>266619</b>	<b>174893</b>	<b>230575</b>	<b>101137</b>	<b>76247</b>	<b>131952</b>

**Table S8** Anions in the filtered water samples from 13 different sources, along with WHO and Nepalese guideline values where applicable. The water was analysed with two different methods; HACH Cuvette Test and Ion Chromatography. The analysis was performed in duplicate for HACH Cuvette Test and the number in parenthesis is the standard deviation.

Sample Site ID	HACH Cuvette Test				Ion Chromatography						
	Fluoride (mg/L)	Ammonium (mg/L)	Nitrite (mg/L)	Nitrate (mg/L)	Fluoride (mg/L)	Chloride (mg/L)	Bromide (mg/L)	Nitrate (mg/L)	Phosphate (mg/L)	Nitrite (mg/L)	Sulphate (mg/L)
1	<0.1	<0.019	<0.05	3.1501 (0.0726)	0.078	16.780	n.d.	2.083	0.061	0.062	9.670
2	<0.1	0.0492 (0.0015)	<0.05	<1	0.157	0.202	0.044	0.201	0.072	0.018	0.201
3	<0.1	<0.019	<0.05	<1	0.053	0.921	n.d.	0.513	0.033	0.014	0.584
4	<0.1	<0.019	<0.05	2.7228 (0.1560)	0.054	6.156	n.d.	1.979	0.082	0.021	7.730
5	0.378 (0.028)	<0.019	<0.05	1.4004 (0.0229)	0.369	2.048	0.012	0.036	17.162	0.015	0.366
6	0.22 (0.011)	0.2808 (0.015)	<0.05	<1	0.400	31.690	n.d.	0.155	0.240	0.092	62.580
7	<0.1	1.7791 (0.0316)	<0.05	2.1966 (0.0389)	0.091	38.488	0.713	0.877	0.054	0.136	20.246
8	<0.1	<0.019	<0.05	59.0741 (1.5943)	0.025	55.814	0.196	52.550	0.068	0.184	61.483
9	<0.1	<0.019	<0.05	1.5425 (0.1155)	0.035	0.995	n.d.	1.340	0.192	0.030	1.994
10	<0.1	0.03122 (0.0041)	<0.05	1.2926 (0.0125)	0.162	1.510	n.d.	0.710	0.144	0.025	2.609
11	<0.1	<0.019	0.0707 (0.002)	78.9677 (1.8065)	0.247	78.279	0.754	149.82 1	1.798	0.437	0.002
12	<0.1	0.0452 (0.0016)	<0.05	2.7410 (0.2453)	0.056	1.423	n.d.	1.801	0.055	0.010	5.098

**Table S8 Continued**

13	<0.1	<0.019	<0.05	2.01 (0.0879)	0.165	4.822	n.d.	1.462	0.296	0.035	6.826
WHO guidance value	1.5		3	50	1.5	250		50		3	
Nepalese guidance value		1.5		50				50			250

**Table S9** Anions and cations used for charge balance error calculation. Total negative and positive charges are expressed as equivalent per liter.

Sample Site ID	Conductivity ( $\mu\text{S}/\text{cm}$ )	Alkalinity ( $\text{mgCaCO}_3/\text{L}$ )	$\text{Cl}^-$ ( $\text{mg}/\text{L}$ )	pH	$\text{NO}_3^-$ ( $\text{mg}/\text{L}$ )	$\text{SO}_4^{2-}$ ( $\text{mg}/\text{L}$ )	$\text{Ca}^{2+}$ ( $\text{mg}/\text{L}$ )	$\text{Mg}^{2+}$ ( $\text{mg}/\text{L}$ )	$\text{K}^+$ ( $\text{mg}/\text{L}$ )	$\text{Na}^+$ ( $\text{mg}/\text{L}$ )	Positive charges ( $\text{eq.}/\text{L}$ )	Negative charges ( $\text{eq.}/\text{L}$ )	Charge balance error (%)
1	216	41	16.780	8.2	3.099	9.670	33.338	2.916	1	2.434	0.002035	0.001544	13.724
2	8.1	4	0.202	6.5	1.000	0.201	0.493	0.100	1	0.492	0.000080	0.000106	-14.058
3	38.7	11	0.921	6.8	1.000	0.584	3.212	0.371	1	0.881	0.000255	0.000274	-3.659
4	268	98	6.156	7.8	2.833	7.730	40.195	4.378	1.2	4.232	0.002581	0.002339	4.926
5	1139	490	2.048	6.6	1.417	0.366	86.760	13.374	5.834	42.487	0.007427	0.009880	-14.170
6	805	340	31.690	6.7	1.000	62.580	98.192	14.267	15.994	32.686	0.007905	0.009007	-6.517
7	531	200	38.488	6.3	2.169	20.246	47.894	7.501	9.641	19.236	0.004091	0.005539	-15.039
8	662	130	55.814	6.2	57.947	61.483	57.293	12.639	15.990	33.802	0.005778	0.006387	-5.001
9	241	170	0.995	8.0	1.461	1.994	51.115	2.478	1	1.064	0.002827	0.003490	-10.505
10	250	126	1.510	7.9	1.284	2.609	48.745	2.916	1	1.236	0.002752	0.002635	2.160
11	670	91	78.279	6.7	77.690	0.002	59.175	12.923	27.984	56.250	0.007179	0.005279	15.247
12	217	104	1.423	7.5	2.568	5.098	35.122	6.952	1	0.902	0.002390	0.002266	2.658
13	62.2	14	4.822	6.2	1.948	6.826	4.168	1.117	1	4.661	0.000528	0.000589	-5.470

**Table S10** Metals in the filtered water samples from 13 different sources, along with WHO and Nepalese guideline values where applicable.

Cations	Sample Site ID													Guidance Value	
	1	2	3	4	5	6	7	8	9	10	11	12	13	WHO Guidance Value	Nepal Guidance Value
<b>Total Ca (mg/L)</b>	33.338	0.493	3.212	40.195	86.760	98.192	47.894	57.293	51.115	48.745	59.175	35.122	4.168	300	200
<b>Total Mg (mg/L)</b>	2.916	<0.1	0.371	4.378	13.374	14.267	7.501	12.639	2.478	2.916	12.923	6.952	1.117		
<b>Total Na (mg/L)</b>	2.434	0.492	0.881	4.232	42.487	32.686	19.236	33.802	1.064	1.236	56.250	0.902	4.661		
<b>Total K (mg/L)</b>	<1	<1	<1	1.2	5.834	15.994	9.641	15.990	<1	<1	27.984	<1	<1		
<b>Total Ba (mg/L)</b>	0.026	0.002	0.004	0.036	0.095	0.186	0.118	0.121	0.0375	0.0331	0.1236	0.0487	0.0102	0.7	
<b>Total Fe (mg/L)</b>	0.004	<0.001	<0.001	0.003	0.6656	0.1973	1.4810	0.1617	0.0426	0.0565	0.0036	0.0083	0.0022		0.3
<b>Total Mn (mg/L)</b>	0.002	<0.001	<0.001	0.001	0.5281	0.1192	0.6568	0.0501	0.0086	0.0062	<0.001	0.0013	0.0025	0.1	0.2
<b>Total Al (mg/L)</b>	0.1379	<0.005	<0.005	0.214	<0.005	0.0055	0.0062	0.0071	0.0116	0.0096	<0.005	0.0078	0.0106	0.2	0.2
<b>Total Zn (mg/L)</b>	0.004	<0.002	<0.002	0.027	0.0034	0.0234	0.0027	<0.002	<0.002	0.0035	0.0021	0.1374	0.0099		3

**Table S10 Continued**

<b>Total Pb (mg/L)</b>	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.01	0.01
<b>Total Cu (mg/L)</b>	<0.001	0.003	<0.001	<0.001	<0.001	0.0056	<0.001	<0.001	<0.001	<0.001	0.0021	0.0019	<0.001	2	1
<b>Total As (mg/L)</b>	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	0.01	0.05
<b>Total Cd (mg/L)</b>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	0.003	0.003
<b>Total Sb (mg/L)</b>	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	0.02	
<b>Total Si (mg/L)</b>	4.3374	0.1797	0.5792	5.7317	35.2107	14.3344	22.1596	10.2059	4.1522	4.1550	14.0731	3.6717	5.3907		
<b>Total Ni (mg/L)</b>	<0.001	<0.001	<0.001	<0.001	0.0015	0.0017	0.0016	0.0012	<0.001	<0.001	<0.001	<0.001	<0.001	0.07	
<b>Total Cr (mg/L)</b>	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.05	0.05
<b>Total Sr (mg/L)</b>	0.044	<0.001	0.003	0.064	0.2271	0.3163	0.1870	0.1952	0.0402	0.0392	0.1290	0.0847	0.0247		

**Table S11** List of water borne diseases endemic in the Kathmandu Valley and their causative agents.

<b>Waterborne Disease</b>	<b>Causative Agents</b>	<b>References</b>
Diarrhoea	<i>Campylobacter, Clostridium difficile, Escherichia coli, Salmonella, Shigella, Vibrio cholera</i>	7-9
Cholera	<i>Vibrio cholerae</i>	
Typhoid	<i>Salmonella enterica, Salmonella typhi, Salmonella paratyphi</i>	
Dysentery	<i>Campylobacter, Escherichia coli, Shigella, Salmonella</i>	
Gastroenteritis	<i>Campylobacter jejuni, Escherichia coli, Salmonella, Shigella, Clostridium difficile, Staphylococcus aureus</i>	

**Table S12** Real-time qPCR primers for different genetic markers

Target Organisms	Primer	Sequence (5'>>>3')	Annealing Temperature (T <sub>a</sub> )	Amplicon size	Reference
<i>Vibrio cholera</i>	EpsM -F	GAATTATTGGCTCCTGTGCA GG	57 °C	248	10
	EpsM -R	ATCGCTTGGCGCATCACTGC CC			
Total Coliform	Eco1457F	CATTGACGTTACCCGCAGAA GAAGC	60 °C	190	11
	Eco1652R	CTCTACGAGACTCAAGCTTG C			
Total <i>E.coli</i>	rodA -F	GCAAACCACCTTTGGTGC	60 °C	194	12
	rodA -R	CTGTGGGTGTGGATTGACAT			
Human <i>E.coli</i>	Hu 8 -F	ACAGTCAGCGAGATTCTTC	60 °C	177	13
	Hu 8 -R	GAACGTCAGCACCA			
	Hu 100 -F	ACGGTTATCAGCTCACGTCG	60 °C	98	(Robson and Davenport, in preparation)
	Hu 100 -R	TCGCCCCTCGAAAAGCATTA			
Total Bacteria (16S rRNA); qPCR	1055 F	ATGGCTGTCGTCAGCT	60 °C	337	14
	1392 R	ACGGGCGGTGTGTAC			
16S rRNA amplicon sequencing; MinION	27F	AGAGTTTGATCMTGGCTCAG	55 °C	1500	Oxford Nanopore Technologies, UK
	1492R	CGGTTACCTTGTTACGACTT			
16S rRNA amplicon sequencing; Illumina	515 F	GTGCCAGCMGCCGCGGTAA	55 °C	291	15
	806 R	TAATCTWTGGGVHCATCAG G			



## References

- 1 APHA. Standard Methods for the Examination of Water and Wastewater. 18th edition. American Public Health Association. (1992).
- 2 WHO. Guidelines for drinking-water quality: fourth edition incorporating the first addendum. Geneva: World Health Organization (2017).
- 3 DWSS. National Drinking Water Quality Standards. Kathmandu: Department of Water Supply and Sewerage (2005).
- 4 Shrestha, S. M., Rijal, K. & Pokhrel, M. R. Assessment of Heavy Metals in Deep Groundwater Resources of the Kathmandu Valley, Nepal. *Journal of Environmental Protection* **7**, 516 (2016).
- 5 Zymo Research. ZymoBIOMICS™ Microbial Community Standard, Catalog No. D6300 (2019).
- 6 HSE. The Approved List of biological agents. Third edition. Merseyside: Health and Safety Executive. (2013).
- 7 Leclerc, H., Schwartzbrod, L. & Dei-Cas, E. Microbial agents associated with waterborne diseases. *Critical reviews in microbiology* **28**, 371-409 (2002).
- 8 Karkey, A. *et al.* The ecological dynamics of fecal contamination and Salmonella Typhi and Salmonella Paratyphi A in municipal Kathmandu drinking water. *PLoS neglected tropical diseases* **10**, e0004346 (2016).
- 9 <http://www.waterwise.co.za/site/water/diseases/waterborne.html> (accessed 2019).
- 10 Kong, R. Y. C., Lee, S. K. Y., Law, T. W. F., Law, S. H. W. & Wu, R. S. S. Rapid detection of six types of bacterial pathogens in marine waters by multiplex PCR. *Water research* **36**, 2802-2812 (2002).
- 11 Bartosch, S., Fite, A., Macfarlane, G. T. & McMurdo, M. E. T. Characterization of bacterial communities in feces from healthy elderly volunteers and hospitalized elderly patients by using real-time PCR and effects of antibiotic treatment on the fecal microbiota. *Applied and environmental microbiology* **70**, 3575-3581 (2004).
- 12 Chandrashekar, K. M. *et al.* Limit of detection of genomic DNA by conventional PCR for estimating the load of Staphylococcus aureus and Escherichia coli associated with bovine mastitis. *Folia microbiologica* **60**, 465-472 (2015).
- 13 Gomi, R., Matsuda, T., Matsui, Y. & Yoneda, M. Fecal source tracking in water by next-generation sequencing technologies using host-specific Escherichia coli genetic markers. *Environmental science & technology* **48**, 9616-9623 (2014).
- 14 Harms, G. *et al.* Real-time PCR quantification of nitrifying bacteria in a municipal wastewater treatment plant. *Environmental science & technology* **37**, 343-351 (2003).
- 15 Kozich, J. J., Westcott, S. L., Baxter, N. T., Highlander, S. K. & Schloss, P. D. Development of a dual-index sequencing strategy and curation pipeline for analyzing amplicon sequence data on the MiSeq Illumina sequencing platform. *Appl. Environ. Microbiol.* **79**, 5112-5120 (2013).