

Supplementary information for

Conditional quorum-sensing induction of a cyanide-insensitive terminal oxidase stabilizes cooperating populations of *Pseudomonas aeruginosa*

Yan et al. (2019)

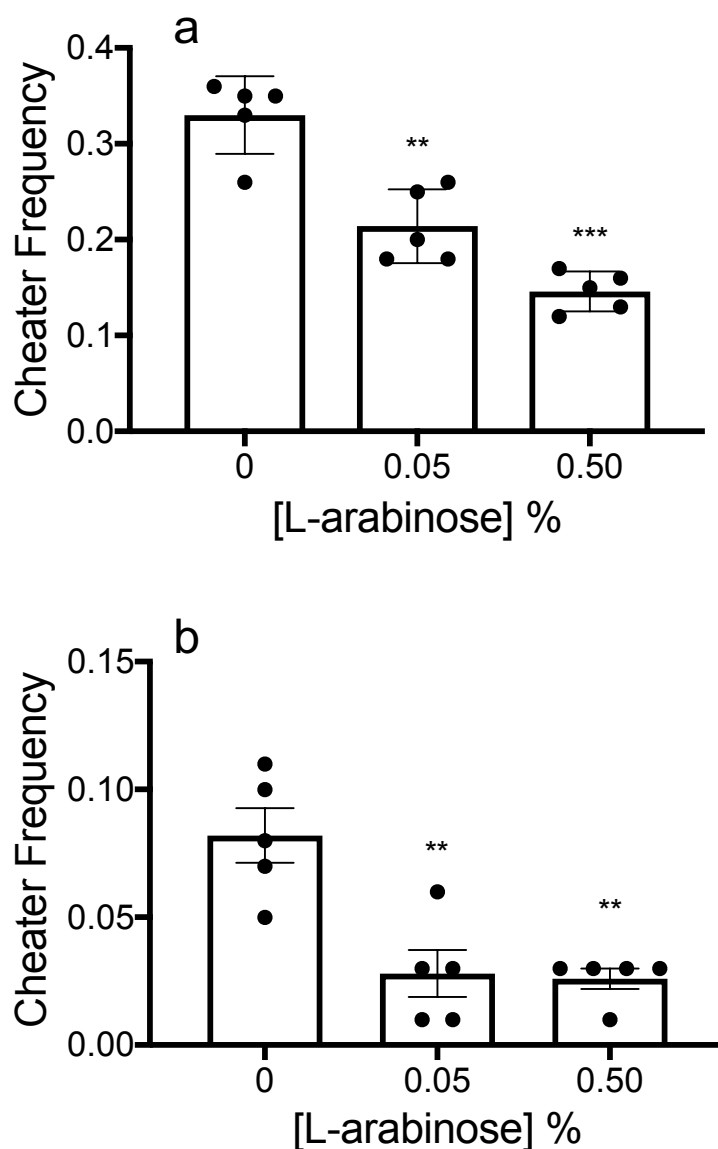
Supplementary Table 1. Bacterial strains and plasmids used in this study.

Strains or plasmids	Genotype and/or relevant characteristics	Reference or source
<i>P. aeruginosa</i>		
PAO1-UW	Wild-type	1
PAO1 $\Delta lasR$	<i>lasR</i> null mutant	2
PAO1 $\Delta rhlR$	<i>rhlR</i> null mutant	2
PAO1 $\Delta lasRrhlR$	<i>lasR rhlR</i> null double mutant	2
PAO1 $\Delta hcnC$	<i>hcnC</i> null mutant	2
PAO1 $\Delta cioA$	<i>cioA</i> null mutant	This study
PAO1 $\Delta rhdA$	<i>rhdA</i> null mutant	This study
PAO1 $\Delta PA4133$	PA4133 null mutant	This study
PAO1 ara-hcn1	Wild-type UW strain with L-arabinose-inducible <i>hcnABC</i> inserted at the <i>att</i> site	This study
PAO1 $\Delta lasR$ ara-hcn1	<i>lasR</i> null mutant with L-arabinose-inducible <i>hcnABC</i> inserted at the <i>att</i> site	This study
<i>E. coli</i>		
S17.1 ($\lambda pir+$)	Conjugal transfer strain for pEXG2 plasmids	
DH5 α	Host strain for pUC18T plasmid recombination	
Plasmids		
pEXG2	Gm ^r <i>mob</i> , <i>sacB</i> , <i>lacZα</i>	3
pEXG2- $\Delta cioA$	pEXG2 derivative carrying the <i>cioA</i> flanking regions	This study
pEXG2- $\Delta rhdA$	pEXG2 derivative carrying the <i>rhdA</i> flanking regions	This study
pEXG2- $\Delta PA4133$	pEXG2 derivative carrying the PA4133 flanking regions	This study
pUC18T-mini-Tn7T- <i>ParaBAD-1hcnABC</i> -Gm	pUC18 derivative carrying mini-Tn7T transposable cassette containing <i>araC</i> and the <i>araBAD</i> promoter immediately upstream of the 2995 bp <i>hcnABC</i> operon	This study
pTNS2	Helper plasmid for transposition of mini-Tn7T cassette	4

Supplementary Table 2. RT-PCR primers used in this study.

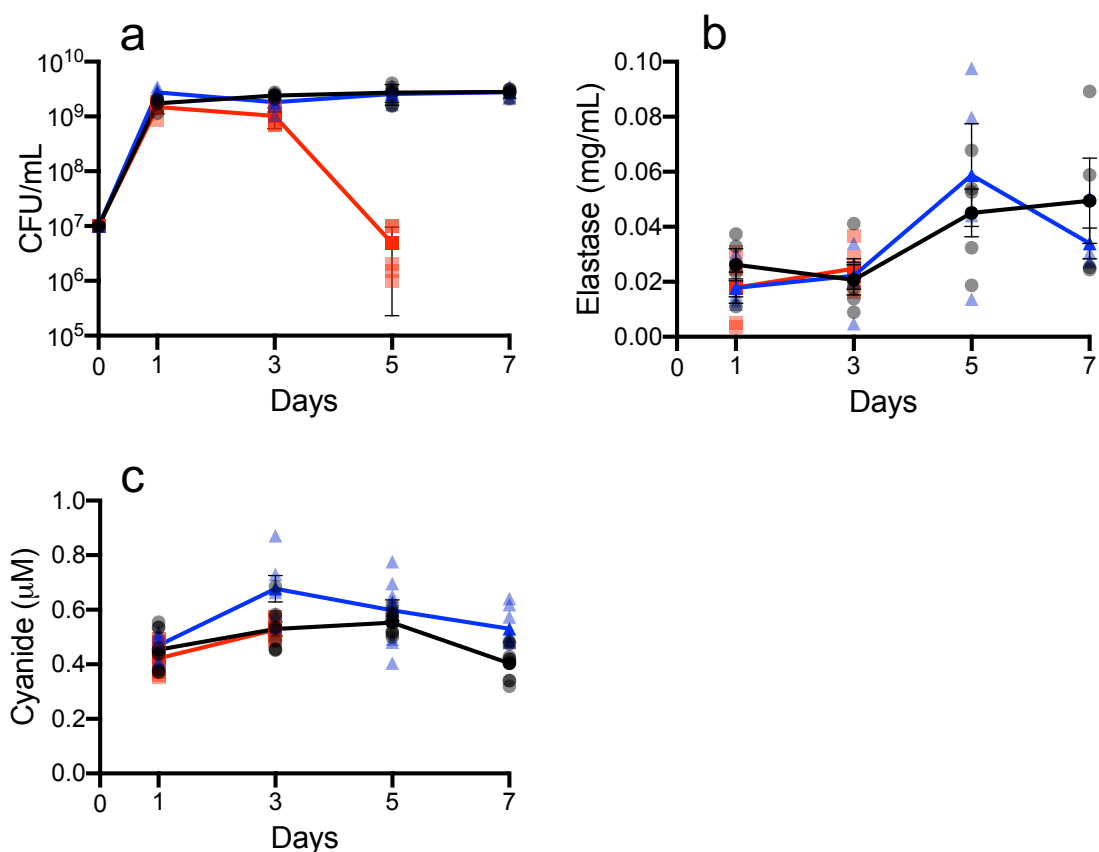
Target gene	Primer name	Sequence (5'-3')	Amplicon (bp)	T _m (°C)
<i>lasR</i>	<i>lasR</i> -F	cagaagatggcgagcgacctg	102	59.9
	<i>lasR</i> -R	cgggtagttgccgacgatgaag		59.7
<i>rhlR</i>	<i>rhlR</i> -F	gcgaccagcagaacatctccag	122	59.6
	<i>rhlR</i> -R	cgggttgacatcagcatcgg		59.6
<i>lasB</i>	<i>lasR</i> -F	cagaagatggcgagcgacctg	102	62
	<i>lasR</i> -R	cgggtagttgccgacgatgaag		61
PA4129	PA4129-F	gaggacaagaacgtcgagga	102	57
	PA4129-R	ctagcgtcggcggaacaa		58
PA4131	PA4131-F	tacatttcggcgcgacctt	125	59
	PA4131-R	caggtatagccgcaccagac		58
PA4133	PA4133-F	aactggttcttcggctcgtt	165	58
	PA4133-R	ggtcaggaagaagcctacgg		58
<i>cioA</i>	<i>cioA</i> -F	atcttcaaccgctcgttccc	143	58
	<i>cioA</i> -R	catcgagagcatcttgcgga		58
<i>cioB</i>	<i>cioB</i> -F	ctggtgaccttctatgcgct	130	58
	<i>cioB</i> -R	ggatgatgttcggccacagg		59
<i>rhdA</i>	<i>rhdA</i> -F	ctaccactatctcaacggcgg	176	58
	<i>rhdA</i> -R	catccataaccgccaggtc		58
<i>proC</i>	<i>proC</i> -F	cgtcgtggctcctgctcggtca	100	60
	<i>proC</i> -R	ggcggcgatggagacgatca		60

Supplementary Figure 1



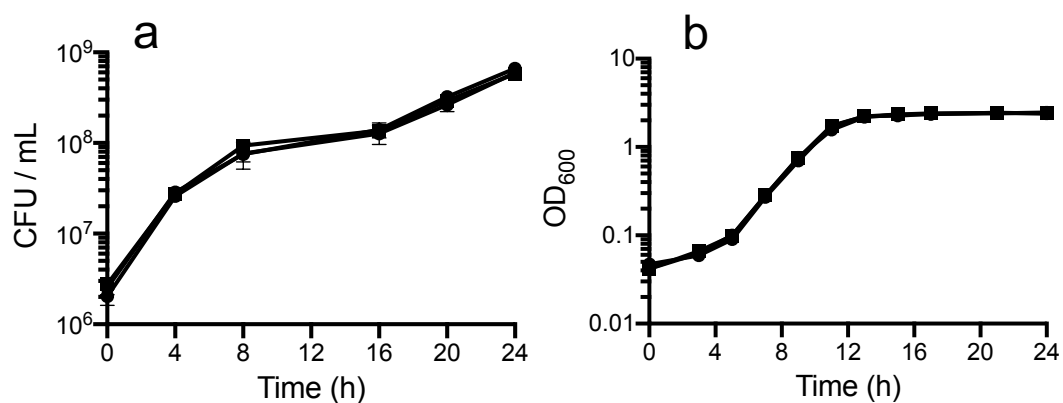
Supplementary Figure 1. Endogenous production of HCN negatively affects LasR mutant enrichment in casein broth competitions with WT. WT and LasR mutant strains harboring an L-arabinose-inducible copy of *hcnABC* were competed for 24 h in casein medium from a combined starting $OD_{600}=0.05$ and target cheater frequency of 0.1 (a) or 0.01 (b). All other experimental conditions are the same as described for other competitions in Materials and Methods of the main text. Statistical significance determined by unpaired Student's *t*-test vs 0% L-arabinose alone ($n=5$); ***, $p < 0.001$; **, $p < 0.01$. Means, s.e.m., and individual biological replicates are shown.

Supplementary Figure 2



Supplementary Figure 2. Co-culture productivity and QS-regulated cooperative behaviors. In co-culture competitions between LasR mutant cheaters and WT cooperators (black circles), PA4133-null cooperators (blue triangles), or *cioA*-null cooperators (red squares), productivity was measured as CFU/mL (**a**) at each indicated timepoint. QS-regulated cooperative behaviors were also measured at these timepoints as elastase production (**b**) or cyanide concentration (**c**). See Materials and Methods for experimental procedures. A minimum of five biological replicates were performed and means, s.e.m., and individual replicates are shown.

Supplementary Figure 3



Supplementary Figure 3. Growth rates of WT PAO1 (circle), a CioA-mutant (square), and a PA4133-mutant (triangle). Bacteria were inoculated from overnight LB-MOPS cultures into casein broth (a) or LB-MOPS (b). Cell density in casein broth was measured by colony counts and in LB-MOPS by optical density at 600 nm. Means and s.e.m. are shown ($n=5$).

References

1. Stover, C. K. et al. Complete genome sequence of *Pseudomonas aeruginosa* PAO1, an opportunistic pathogen. *Nature* **406**, 959-964 (2000).
2. Wang, M. et al. Quorum sensing and policing of *Pseudomonas aeruginosa* social cheaters. *Proc. Natl. Acad. Sci. U.S.A.* **112**, 2187-2191 (2015).
3. Rietsch, A. et al. ExsE, a secreted regulator of type III secretion genes in *Pseudomonas aeruginosa*. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 8006-8011(2005).
4. Choi, K. H. et al. mini-Tn7 insertion in bacteria with secondary, non-*glmS*-linked attTn7 sites: example *Proteus mirabilis* HI4320. *Nat. Protoc.* **1**, 170-178 (2006).