# **Supporting Information**

Modular Synthesis of Di- and Trisubstituted Imidazoles from Ketones and Aldehydes: A Route to Kinase Inhibitors

Ian de Toledo<sup>1</sup>, Thiago A. Grigolo<sup>1</sup>, James M. Bennett<sup>2</sup>, Jonathan M. Elkins<sup>2,3</sup>, Ronaldo A. Pilli<sup>1</sup>\*.

<sup>1</sup> Department of Organic Chemistry, Institute of Chemistry, University of Campinas, UNICAMP, Campinas, CEP 13083-970 (Brazil)

<sup>2</sup> Structural Genomics Consortium, Nuffield Department of Medicine, University of Oxford, Old Road Campus Research Building, Roosevelt Drive, Oxford, OX3 7DQ, UK

<sup>3</sup> Structural Genomics Consortium, Departamento de Genética e Evolução, Instituto de Biologia, UNICAMP, Campinas, SP, 13083-886, Brazil

\*e-mail: pilli@iqm.unicamp.br

## **Table Of Contents**

1.	Optimization Data	S1
2.	NMR Considerations	S4
3.	Synthesis of compounds S1-S16	S7
4.	TR-FRET binding displacement assay	S14
5.	References	S14
6.	NMR Spectra	S16

### 1. Optimization Data

**Optimization of the reaction conditions.** 5-phenyl-2-(p-tolyl)-1H-imidazole (**5**). A 6 mL vial was charged with acetophenone **2** (46 mg, 0.375 mmol, 1.25 equiv.), DMSO (0.75 mL, 0.5 M), concentrated aqueous HBr (48% w/w, 8.9 M) (4.24  $\mu$ L, 0.03 mmol, 10 mol%), deionized water (71  $\mu$ L) and a magnetic stirrer bar under air. The reaction mixture was stirred in a pre-heated aluminum block at 85 °C and was followed by TLC analysis (30% AcOEt/Hex, p-ASD). After consumption of starting material, the reaction mixture was cooled to room temperature and diluted with MeOH (1.25 mL, 0.19 M, final concentration relative to acetophenone 2, 2:8 mixture of DMSO:MeOH). This stock DMSO:MeOH solution was added dropwise over 30 minutes via syringe to a 6 mL vial containing *p*-tolualdehyde **4** (37 mg, 0.30 mmol, 1.00 equiv.), NH<sub>4</sub>OAc (116 mg, 1.50

mmol, 5.00 equiv.) and MeOH (1.5 mL, 0.2 M in relation to **4**). The reaction mixture was stirred at room temperature for 18h and then poured directly into a separatory funnel containing a mixture of satd. NaHCO<sub>3</sub> and satd. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (1:1, 1x 20 mL) and AcOEt (10 mL). The phases were separated and the aqueous phase was extracted with AcOEt (5x 5 mL). The organic phases were combined, washed with satd. NaCl solution (1x 5 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in the rotaevaporator. The residue was diluted with AcOEt (5 mL) and a 1 mL aliquot was taken and concentrated in vacuo. To this, 1,3,5-trimethoxybenzene (10.2 mg, 0.06 mmol) and acetone-*d*<sub>6</sub> (0.6 mL) was added and the sample was analyzed by <sup>1</sup>H NMR. The crude mixtures were combined and purification of the residue by silica gel chromatography, eluting with AcOEt in hexanes (19 cm x 20 mm, gradient elution, 0%  $\rightarrow$  30%, 5% increases, 50 mL runs, 5-10 mL fractions) yielded **5** as a white solid (69% yield, 48 mg, 0.21 mmol).

Table S1. Optimization of the reaction conditions for the synthesis of the disubstituted imidazole 5



Entry	HBr loading (mol%)	Temperature (°C)	Time Oxidation (h)	Solvent	Acetophenone (equiv.)	Yield <sup>b</sup> (%)
1	200	60	24	MeOH:DMSO (8:2)	1.00	48
2	100	60	48	MeOH:DMSO (8:2)	1.00	54
3	50	60	72	MeOH:DMSO (8:2)	1.00	57
4	50	85	12	MeOH:DMSO (8:2)	1.00	55
5	30	85	18	MeOH:DMSO (8:2)	1.00	60
6	10	85	18	MeOH:DMSO (8:2)	1.00	61
7	10	85	18	EtOH:DMSO (8:2)	1.00	49
8	10	85	18	n-PrOH:DMSO (8:2)	1.00	56
9	10	85	18	i-PrOH:DMSO (8:2)	1.00	44
10	10	85	18	MeOH:DMSO:MeCN (5:2:3)	1.00	45
11	10	85	18	MeOH:DMSO:DCM (5:2:3)	1.00	44
12	10	85	18	MeOH:DMSO:DMF (5:2:3)	1.00	45
13	10	85	18	MeOH:DMSO:PhMe (5:2:3)	1.00	47
14	10	85	18	MeOH:DMSO:THF (5:2:3)	1.00	44
15	10	85	18	MeOH:DMSO:AcOEt (5:2:3)	1.00	43
16	10	85	18	MeOH:DMSO (3:7)	1.00	45
17	10	85	18	MeOH:DMSO (8:2)	1.25	69 (69)
18	10	85	18	MeOH:DMSO (8:2)	1.50	66
19	10	85	18	MeOH:DMSO (8:2)	1.75	73
20	10	85	18	MeOH:DMSO (8:2)	2.00	67
21°	10	85	18	DMSO then MeOH	1.25	(52)

<sup>a</sup>Oxidation step performed using acetophenone **2 (Table 1)**, aqueous HBr (48% w/w, 8.9 M) (**Table 1**) and DMSO (0.50 M). Condensation step performed by slow addition (30 min) of glyoxal **3** solution in MeOH:DMSO (8:2, 0.19 M relative to acetophenone **2**) to a mixture of tolualdehyde **4** (0.3 mmol) and NH<sub>4</sub>OAc (1.5 mmol) in MeOH (1 mL, 0.20 M). <sup>b</sup>Yield after work-up as determined by <sup>1</sup>H NMR analysis of the crude reaction mixture with 1,3,5-trimethoxybenzene as the internal standard. Isolated yield given in parentheses. <sup>c</sup> **Performed under stepwise procedure. Oxidation step**: Acetophenone **2** (**Table 51**), aqueous HBr (48% w/w, 8.9 M) (**Table 51**) and DMSO (0.50 M). G6% isolated yield). Condensation step performed by slow addition (30 min) of glyoxal **3** (1.25 equiv.) solution in MeOH (0.19 M relative to glyoxal **3**) to a mixture of tolualdehyde **4** (0.168 mmol) and NH<sub>4</sub>OAc (5.00 equiv.) in MeOH (0.20 M) (**79**% isolated yield).

Optimization of the reaction conditions. 4-(2-(2-methyl-4-(methylsulfonyl)phenyl)-1H-imidazol-5-yl)pyridine (32). A 10 mL round-bottom flask was charged with 4-acetylpiridine (219 mg, 1.70 mmol, 1.70 equiv.), magnetic stirrer bar and DMSO (3.5 mL, 0.5 M) under air and concentrated HBr aqueous (48% w/w, 8.9 M) (595 mL, 5.25 mmol, 3.0 equiv.) was added dropwise. The reaction mixture was stirred in preheated oil bath at 60 °C for 8h. After consumption of the starting material, indicated by TLC analysis (AcOEt, *p*-ASD), the reaction mixture was left to reach room temperature and MeOH (5.7 mL, 0.19 M) was added. This reaction mixture was added dropwise over 30 minutes via syringe to a solution of S5 (198 mg, 1.00 mmol, 1.00 equiv.) and NH<sub>4</sub>OAc (771 mg, 10.0 mmol, 10.0 equiv.) in MeOH (5 mL, 0.2 M in relation to S5) at room temperature. The reaction mixture was stirred at room temperature for 18h and the solvent was removed in the rotaevaporator, the residue was diluted with 10% MeOH/DCM (10 mL) and poured into separatory funnel containing satd. NaHCO<sub>3</sub> (1x 40 mL) and 10% MeOH/DCM (1x 15 mL). The phases were separated, and the aqueous phase was extracted with 10% MeOH/DCM (7x 10 mL). The organic phases were combined, dried over MgSO<sub>4</sub>, filtered and concentrated in the rotaevaporator. Purification by silica gel chromatography, eluting with MeOH in DCM (gradient elution  $5\% \rightarrow 9\%$ ) yielded **32** as a pale yellow solid (67% yield, 210 mg, 0.67 mmol).

	4-acetylpyrid (Table S2)	i) Oxidation Meth (Table S2) ii) Base, MeOH dine	od O O O O O O O O O O O O O	Me O D25 S5 (1.00 equiv.) Ammonia Source MeOH, rt 24h 32	P₂Me Me N	
Entry	Oxidation Method	Base	Time Oxidation	Ammonia Source	4-acetylpyridine (equiv.)	Yield <sup>a</sup> (%)
1 <sup>b</sup>	DMSO/48% HBr aq.	Na <sub>2</sub> CO <sub>3</sub>	12h	NH <sub>4</sub> OAc (10 equiv.)	1.00	45
2c	DMSO/NaBr/H <sub>2</sub> SO <sub>4</sub>	Na <sub>2</sub> CO <sub>3</sub>	10 min	NH <sub>4</sub> OAc (10 equiv.)	1.00	19
3d	DMSO/I <sub>2</sub>	Na <sub>2</sub> CO <sub>3</sub>	8h	NH₄OAc (10 equiv.)	1.00	-
4 <sup>b</sup>	DMSO/48% HBr aq.	K <sub>2</sub> CO <sub>3</sub>	8h	NH₄OAc (10 equiv.)	1.50	57
5 <sup>b</sup>	DMSO/48% HBr aq.	Et <sub>3</sub> N	8h	NH <sub>4</sub> OAc (10 equiv.)	1.50	54
6 <sup>b</sup>	DMSO/48% HBr aq.	Et <sub>3</sub> N	24h	NH OAc (10 equiv.)	1.50	48
7 <sup>e</sup>	DMSO/48% HBr aq.	Et <sub>3</sub> N	8h		1.50	40
8 <sup>b</sup>	DMSO/48% HBr aq.	(NH <sub>4</sub> ) <sub>2</sub> CO <sub>3</sub>	8h	NH <sub>4</sub> OAc (10 equiv.)	1.50	53
9 <sup>b</sup>	DMSO/48% HBr aq.	-	8h	(NH <sub>4</sub> ) <sub>2</sub> CO <sub>3</sub> (10 equiv.)	1.50	51
10 <sup>b</sup>	DMSO/48% HBr aq.	-	8h	NH <sub>4</sub> OAc (10 equiv.)	1.50	66
11 <sup>b</sup>	DMSO/48% HBr aq.	-	8h	NH <sub>4</sub> OAc (10 equiv.)	1.75	67

<b>Table 32.</b> Optimization of the reaction conditions for the synthesis of the disubstituted initiazore	Table S	<b>S2</b> . Optimization of the	e reaction conditions	for the synthesis	of the disubsti	ituted imidazole <b>3</b>
--	---------	---------------------------------	-----------------------	-------------------	-----------------	---------------------------

<sup>&</sup>lt;sup>a</sup>Oxidation step performed using 4-acetylpyrydine (**Table S2**), aqueous HBr (48% w/w, 8.9 M) (**300 mol%**) and DMSO (0.50 M). Condensation step performed by slow addition (30 min) of glyoxal solution in MeOH:DMSO (8:2, 0.19 M relative to 4-acetylpyridine) to a mixture of aldehyde (1.0 mmol) and NH<sub>4</sub>OAc (10 mmol) in MeOH (3.3 mL, 0.30 M). <sup>a</sup>Isolated yield after column cromatography <sup>b</sup>300 mol% of HBr aq. was employed and the reaction was stirred at 60 °C <sup>c</sup> Oxidation step performed according conditions reported by Karpov and colaborators<sup>1</sup> <sup>d</sup> Oxidation step performed according conditions reported by Zhu and colaborators<sup>2</sup> <sup>e</sup>Performed under reduced pressure (~60 mmHg)

Table S3. Optimization of the reaction conditions for the synthesis of the disubstituted imidazole 8



<sup>a</sup>Oxidation step performed using 4-acetylpyridine (**1.25 equiv.**), aqueous HBr (48% w/w, 8.9 M) (**Table S3**) and DMSO (0.50 M). Condensation step performed by slow addition (30 min) of glyoxal **3** solution in MeOH:DMSO (8:2, 0.19 M relative t o4-acetylpyridine) to a mixture of benzaldehyde (0.3 mmol) and NH<sub>4</sub>OAc (1.5 mmol) in MeOH (1.5 mL, 0.20 M) <sup>b</sup> Isolated yield.

#### 2. NMR Considerations

It was noted during the course of this project that addition of TFA and D<sub>2</sub>O to the sample in DMSO-*d*<sub>6</sub> for NMR characterization provided enhancement in resolution due to conversion of both tautomers present in the mixture to the corresponding protonated TFA salt. For substrates that did not provided a clear NMR spectrum, such as the three substituted imidazoles and some disubstituted imidazoles, the mixture of 0.5 mL of DMSO-*d*<sub>6</sub>, 0.1 mL of D<sub>2</sub>O and 8  $\mu$ L of TFA was the solvent of choice for carrying NMR analysis when the substrate was not acid labile. The improvement in the spectra resolution can be seen in **Figures 1-4** for the trisubstituted imidazole **59** in both <sup>1</sup>H and <sup>13</sup>C NMR. When this mixture of solvents was used, a broad signal corresponding to peak of HOD at 4.20 – 4.00 ppm appears in the <sup>1</sup>H NMR spectra and two quartets at  $\delta$  159.5 (q, *J*<sub>CF</sub> = 36.8 Hz) e 116.4 (q, *J*<sub>CF</sub> = 290.8 Hz), corresponding to two carbons present in the TFA, are seen in the <sup>13</sup>C NMR spectra.



Figure S2. <sup>1</sup>H NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 600 MHz) of 59





#### 3. Synthesis of compounds S1-S16



1-Tosyl-1H-indole (**S1**) and 1-(1-tosyl-1H-indol-3-yl)ethenone (**S2**) were prepared according to literature procedure<sup>1</sup>



2-Methyl-4-(methylthio)benzonitrile (S3). Following a modified literature procedure<sup>2,3</sup>. A 250 mL round-bottom flask was charged with 4-bromo-2methylbenzonitrile (3.03 g, 15.0 mmol, 1.00 equiv.), a magnetic stirrer bar and dry DMF (30 mL, 0.5 M) under inert atmosphere. Then, sodium thiomethoxide (1.22 g, 16.5 mm), 1.10 equiv.) was added in portions by briefly removing the Suba seal and the reaction mixture was stirred for 15h at room temperature. After consumption of the starting material, indicated by TLC analysis (20% AcOEt/Hex), water (1x 150 mL) was added and the aqueous phase was extracted with Et<sub>2</sub>O (3x 90 mL). The organic phases were combined, washed with a mixture of satd. NaCl:H<sub>2</sub>O (1:1, v/v) (3x 75 mL), dried over MgSO<sub>4</sub>, filtered and concentrated in the rotaevaporator to afford a pale yellow solid (quantitative yield, 2.45 g, 15.0 mmol). The crude solid was used in the next step without further purification. A small amount (~20 mg) was purified by column flash chromatography (0  $\rightarrow$  20% AcOEt/Hex) for further characterization. R<sub>f</sub> = 0.52 (20% AcOEt/Hex, UV, KMnO4,  $I_2/SiO_2$ ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.46 (d, J = 8.2 Hz, 3H), 7.10 (s, 1H), 7.06 (dd, J = 8.1, 2.0 Hz, 1H), 2.50 (s, 3H), 2.49 (s, 3H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) § 145.8, 142.2, 132.6, 126.5, 122.9, 118.4, 108.4, 20.6, 14.8; v<sub>max</sub> (cm<sup>-1</sup>, thin film, ATR): 2920 (w), 2900 (w), 2217 (m), 1595 (s), 1584 (w), 1486 (w), 1440 (w), 1397 (w), 1380 (w), 1220 (m), 1206 (w), 1189 (w), 1083 (m), 966 (w), 888 (m), 872 (w), 813 (m), 806 (s); mp: 60.0 - 62.0 °C (AcOEt). Spectroscopic data are in accordance with the literature<sup>2</sup>

2-Methyl-4-(methylsulfonyl)benzonitrile (S4) A 100 mL round-bottom flask was

charged with the crude product (2.45 g, 15.0 mmol, 1.00 equiv.), a large magnetic stirrer bar (2 cm x 1 cm) and DCM (55 mL, 0.27 M) under air. The solution was cooled to 0 °C in ice/water bath and solid m-CPBA (77% peracid, 8.07 g, 36.0 mmol, 2.40 equiv.) was added in small portions over 10 minutes, under vigorous stirring. After the addition, the mixture was stirred at 0 °C for 20 minutes and then for 15h at room temperature. After consumption of the starting material, indicated by TLC analysis (20% AcOEt/Hex), the reaction mixture was poured in a separation funnel containing Et<sub>2</sub>O (350 mL) and NaOH 2M aqueous (75 mL) and the phases were separated. The organic phase was washed with a mixture of satd. NaHCO<sub>3</sub> and satd. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (1:1, v/v) (3x 75 mL), followed by washing with satd. NaCl (1x 75 mL). The organic phase was dried over MgSO<sub>4</sub>, filtered and concentrated in the rotaevaporator. The residue was purified by recrystallization with AcOEt and hexanes to afford S4 (85% for two steps, 2.49 g, 12.7 mmol) as colorless needles.  $R_f = 0.54$  (50% AcOEt/Hex, KMnO<sub>4</sub>, Dragendorff); <sup>1</sup>H NMR (500 MHz, **CDCl**<sub>3</sub>)  $\delta$  7.92 (d, J = 0.5 Hz, 1H), 7.86 (dd, J = 8.1, 1.2 Hz, 1H), 7.81 (d, J = 8.1 Hz, 1H), 3.07 (s, 3H), 2.67 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 144.3, 144.0, 133.7, 129.1, 125.3, 118.2, 116.5, 44.4, 20.8; v<sub>max</sub> (cm<sup>-1</sup>, thin film, ATR): 3010 (w), 2932 (w), 2231 (w), 1400 (w), 1389 (w), 1322 (w), 1303 (s), 1287 (s), 1178 (w), 1144 (m), 1120 (s), 1089 (m), 968 (s), 898 (w), 888 (w), 832 (m), 763 (s); mp: 147.9 - 148.9 °C (AcOEt/Hex).

2-Methyl-4-(methylsulfonyl)benzaldehyde (S5) A 250 mL round-bottom flask was charged with 2-methyl-4-(methylsulfonyl)benzonitrile (2.48 g, 12.7 mmol, 1.00 equiv.), a magnetic stirrer bar and dry DCM (51 mL, 0.25 M) under inert atmosphere. The mixture was stirred and cooled to -15 °C in an ice/brine bath for 15 minutes and then DIBAL-H (99%, 2.9 mL, 15.2 mmol, 1.20 equiv.) in dry DCM (15.2 mL) was added dropwise along the flask wall over 30 minutes using syringe pump. The reaction mixture was left stirring at -15 °C for 1h, and then, at 0 °C for 3h. After consumption of the starting material, indicated by TLC analysis (50% AcOEt/Hex, Dragendorff stain), MeOH (1x 10 mL) and HCl 1 M aqueous (1x 10 mL) were added at 0 °C, the ice/water bath was removed, and the reaction mixture was left to reach room temperature. Then, HCl 1 M aqueous (1x 100 mL) was added and the phases were separated. The aqueous phase was extracted with Et<sub>2</sub>O (5x 100 mL) and the organic phases were combined, washed with satd. NaCl solution (1x 75 mL), dried over MgSO<sub>4</sub>, filtered and concentrated in the rotaevaporator to afford **S5** as a white solid (91% yield, 2.13 g, 10.7 mmol) which was used without

further purification.  $\mathbf{R}_{f} = 0.52$  (50% AcOEt/Hex, UV, KMnO<sub>4</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  10.36 (s, 1H), 7.98 (d, J = 8.0 Hz, 1H), 7.91 (dd, J = 8.0, 1.3 Hz, 1H), 7.85 (s, 1H), 3.07 (s, 3H), 2.75 (s, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  191.4, 144.4, 142.1, 137.6, 132.4, 130.6, 125.3, 44.3, 19.6; **v**<sub>max</sub> (cm<sup>-1</sup>, thin film, ATR): 3010 (w), 2952 (w), 2862 (w), 1694 (m), 1599 (w), 1457 (w), 1394 (w), 1310 (s), 1292 (s), 1184 (w), 1147 (s), 1112 (w), 1089 (w), 965 (m), 826 (w), 796 (w), 759 (s); HRMS (ESI+/TOF) m/z: [M+H]<sup>+</sup> calcd. for C<sub>9</sub>H<sub>11</sub>O<sub>3</sub>S 199.0423; Found 199.0418; mp: 118.2 – 120.9 °C (AcOEt). Spectroscopic data are in accordance with the literature<sup>4</sup>

**Miyaura Borylation: General Procedure A.** A sealed tube was charged with the corresponding bromide (1.00 equiv.),  $B_2(pin)_2 (1.05 - 1.50 \text{ equiv.})$ ,  $Pd(dppf)Cl_2 \cdot DCM$  (5 mol%) and KOAc (3.00 equiv.) and a magnetic stirrer bar under inert atmosphere. Then, degassed DME (0.4 M) was added and the tube glass tube was sealed. The reaction mixture was stirred in a pre-heated oil bath at 80 °C for 14-18h. After consumption of the starting material, indicated by TLC analysis, the reaction mixture was allowed to reach room temperature and it was diluted with DCM, filtered in pad silica gel (3 cm) which was washed with DCM until all product was eluted and the filtrate was concentrated under in the reduced pressure. The crude product was purified by silica gel column chromatography.



2-(Benzo[b]thiophen-5-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (S6). The title compound was prepared according to general procedure A, using 5bromobenzo[b]thiophene (330 mg, 1.5 mmol, 1.00 equiv.), B<sub>2</sub>(pin)<sub>2</sub> (589 mg, 2.25 mmol, 1.50 equiv.), Pd(dppf)Cl<sub>2</sub>•DCM (56 mg, 5 mol%) and KOAc (442 mg, 4.50 mmol, 3.00 equiv.). Purification by silica gel chromatography, eluting with Et<sub>2</sub>O in hexanes (21 cm x 20 mm, gradient elution, 0% → 10%, 1% increases, 50 mL runs, 15 mL fractions) yielded S6 as white solid (92% yield, 359 mg, 1.38 mmol).  $\mathbf{R}_{f} = 0.20$  (5% Et<sub>2</sub>O/Hex, UV, Curcumin). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.32 (s, 1H), 7.89 (d, *J* = 8.1 Hz, 1H), 7.75 (d, *J* = 7.8 Hz, 1H), 7.42 (d, *J* = 5.3 Hz, 1H), 7.35 (d, *J* = 5.5 Hz, 1H), 1.38 (s, 12H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  142.8, 139.3, 130.9, 129.9, 126.1, 124.3, 122.0, 84.0, 25.1. Spectroscopic data are in accordance with the literature<sup>5</sup>.



6-(4,4,5,5-*Tetramethyl-1,3,2-dioxaborolan-2-yl)naphthalen-2-ol* (**S7**). The title compound was prepared according to general procedure A, using 6-bromo-2-naphthol (341 mg, 1.50 mmol, 1.00 equiv.), B<sub>2</sub>(pin)<sub>2</sub> (589 mg, 2.25 mmol, 1.50 equiv.), Pd(dppf)Cl<sub>2</sub>•DCM (56 mg, 5 mol%) and KOAc (442 mg, 4.50 mmol, 3.00 equiv.). Purification by silica gel chromatography, eluting with acetone in hexanes (20 cm x 20 mm, gradient elution, 0% → 20%, 2% increases, 50 mL runs, 15 mL fractions) yielded **S7** as a white solid (76% yield, 310 mg, 1.15 mmol). **R**<sub>f</sub> = 0.15 (10% Acetone/Hex, UV, KMnO<sub>4</sub>, Curcumin stain). <sup>1</sup>**H NMR (500 MHz, CDCl<sub>3</sub>)** δ 8.30 (s, 1H), 7.64 (d, *J* = 8.3 Hz, 1H), 7.82 – 7.75 (m, 2H), 7.13 (d, *J* = 2.4 Hz, 1H), 7.09 (dd, *J* = 8.8, 2.5 Hz, 1H), 5.42 (s, 1H), 1.39 (s, 12H). <sup>13</sup>**C NMR (126 MHz, CDCl<sub>3</sub>)** δ 154.6, 136.6, 136.3, 131.3, 130.9, 128.5, 125.7, 117.9, 109.5, 84.1, 25.0. **v**<sub>max</sub> (cm<sup>-1</sup>, thin film, ATR): 3288 (br), 2980 (w), 1624 (m), 1485 (s), 1426 (w), 1389 (m), 1377 (m), 1354 (m), 1330 (s), 1298 (m), 818 (w), 705 (w), 691 (s). **HRMS (ESI+/TOF) m/z:** peak not found in HRMS analysis. **mp:** 158.3 – 160.1 °C (acetone/hexanes).



*2-Bromo-6-ethoxynaphthalene* ( $\mathbf{S8}$ ) was prepared according to literature procedure<sup>6</sup>.

2-(6-ethoxynaphthalen-2-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (S9). The title compound was prepared according to general procedure A, using S8 (126 mg, 0.5 mmol, 1.00 equiv.), B<sub>2</sub>(pin)<sub>2</sub> (196 mg, 0.75 mmol, 1.50 equiv.), Pd(dppf)Cl<sub>2</sub>•DCM (19 mg, 5 mol%) and KOAc (147 mg, 1.50 mmol, 3.00 equiv.). Purification by silica gel chromatography, eluting with AcOEt in Hexanes (20 cm x 20 mm, gradient elution, 0%  $\rightarrow$  10%, 1% increases, 50 mL runs, 15 mL fractions) yielded S9 as a white solid (80% yield, 119 mg, 0.04 mmol). **R**<sub>f</sub> = 0.37 (5% AcOEt/Hexanes, Curcumin Stain). <sup>1</sup>H NMR

(500 MHz, CDCl<sub>3</sub>)  $\delta$  8.29 (s, 1H), 7.80 (dd, J = 8.4, 0.9 Hz, 1H), 7.78 (d, J = 8.9 Hz, 1H), 7.70 (d, J = 8.2 Hz, 1H), 7.13 (dd, J = 8.8, 2.4 Hz, 1H), 7.11 (d, J = 2.0 Hz, 1H), 4.16 (q, J = 7.0 Hz, 2H), 1.48 (t, J = 7.0 Hz, 3H), 1.39 (s, 12H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  158.0, 136.6, 136.1, 131.2, 130.3, 128.5, 126.0, 119.1, 106.5, 83.9, 63.6, 25.1, 14.9. **v**<sub>max</sub> (cm<sup>-1</sup>, thin film, ATR): 2979 (w), 2922 (w), 1625 (m), 1477 (m), 1380 (m), 1342 (m), 1293 (w), 1271 (w), 1206 (s), 1168 (w), 1140 (m), 1111 (w), 1079 (m), 962 (w), 939 (w), 910 (w), 861 (m), 834 (w), 824 (m), 709 (w), 694 (m). HRMS (ESI+/TOF) m/z: peak not found in HRMS analysis. mp: 88.8 – 91.2 °C (AcOEt).



*2-Bromo-6-cyclopropoxynaphthalene* (S10) was prepared according to literature procedure<sup>7</sup>.

2-(6-cyclopropoxynaphthalen-2-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (S11). The title compound was prepared according to general procedure A, using S10 (105 mg, 0.4 mmol, 1.00 equiv.), B<sub>2</sub>(pin)<sub>2</sub> (110 mg, 0.42 mmol, 1.05 equiv.), Pd(dppf)Cl<sub>2</sub>•DCM (15 mg, 5 mol%) and KOAc (118 mg, 1.2 mmol, 3.00 equiv.). Purification by silica gel chromatography, eluting with AcOEt in hexanes (10 cm x 20 mm, gradient elution, 0% → 10%, 1% increases, 30 mL runs, 10 mL fractions) yielded S11 as a white solid (73% yield, 91 mg, 0.29 mmol). **R**<sub>f</sub> = 0.28 (5% AcOEt/Hex, UV, Curcumin Stain). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.29 (s, 1H), 7.81 (dd, *J* = 8.2, 0.9 Hz, 1H), 7.77 (d, *J* = 8.9 Hz, 1H), 7.74 (d, *J* = 8.2 Hz, 1H), 7.43 (d, *J* = 2.4 Hz, 1H), 7.13 (dd, *J* = 2.6, 8.8 Hz, 1H), 3.89 – 3.84 (m, 1H), 1.39 (s, 12H), 0.87 – 0.81 (m, 4H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 158.0, 136.5, 136.1, 131.2, 130.3, 128.7, 126.1, 118.7, 108.1, 83.9, 51.1, 25.1, 6.4. Spectroscopic data are in accordance with the literature<sup>7</sup>.



*Tert-butyl (6-bromonaphthalen-2-yl)carbamate (***S12***).* Following a modified literature procedure<sup>8</sup>. In 10 mL round-bottom flask, 3-amino-2-naphthol (680 mg, 3.00

mmol, 1.00 equiv.) and a magnetic stir bar was added under N<sub>2</sub> atmosphere. Anhydrous THF (6 mL, 0.5 M) was added via syringe followed by (Boc)<sub>2</sub>O (0.88 mL, 3.75 mmol, 1.25 equiv). The reaction mixture was then refluxed with stirring and monitored via TLC. After consumption of the starting material, the reaction mixture was cooled and THF was removed under reduced pressure and the residue was directly purified through flash column chromatography (0% -> 15% EtOAc/Hex, 4% increase until 12%, 12% (2x runs), 3% increase until 15%, 12% (2x runs), 100 mL runs). A pinkish solid was obtained which was triturated with hexanes (5-6 x 30 mL) to afford **S12** as a white solid (74% yield, 714 mg, 2.22 mmol). **R**<sub>f</sub> = 0.60 (20% AcOEt/Hex, UV, *p*-ASD). <sup>1</sup>**H NMR (500 MHz, DMSO)**  $\delta$  9.64 (s, 1H), 8.12 (s, 1H), 8.06 (s, 1H), 7.79 (d, *J* = 8.3 Hz, 1H), 7.74 (d, *J* = 8.2 Hz, 1H), 7.60 – 7.47 (m, 2H), 1.50 (s, 9H). <sup>13</sup>**C NMR (126 MHz, DMSO-***d*<sub>6</sub>)  $\delta$  152.8, 137.8, 132.2, 130.3, 129.3, 129.3, 129.2, 127.6, 120.6, 116.8, 113.3, 79.4, 28.1. Spectroscopic data are in accordance with the literature<sup>9</sup>.

*Tert-butyl* (6-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)naphthalen-2yl)carbamate (S13). The title compound was prepared according to general procedure A, using S12 (488 mg, 1.50 mmol, 1.00 equiv.), B<sub>2</sub>(pin)<sub>2</sub> (589 mg, 2.25 mmol, 1.50 equiv.), Pd(dppf)Cl<sub>2</sub>•DCM (56 mg, 5 mol%) and KOAc (442 mg, 4.5 mmol, 3.00 equiv.). Purification by silica gel chromatography, eluting with AcOEt in hexanes (18 cm x 30 mm, gradient elution,  $0\% \rightarrow 20\%$ , 1% increases until 10% then 2% increases until 20%, 100 mL runs, 20 mL fractions) yielded a white solid which was triturated (1x 10 mL) and washed with hexanes (4x 10 mL) to afford S13 as a white solid (83% yield, 461 mg, 1.25 mmol). **R**<sub>f</sub> = 0.32 (10% AcOEt/Hex, UV, Curcumin Stain). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.28 (s, 1H), 7.99 (s, 1H), 7.80 (dd, J = 8.2, 0.9 Hz, 1H), 7.79 (d, J = 8.8 Hz, 1H), 7.73 (d, J = 8.5 Hz, 1H), 7.32 (dd, J = 8.8, 2.1 Hz, 1H), 6.68 (s, 1H), 1.55 (s, 9H), 1.38 (s, 12H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  152.8, 137.0, 136.0, 135.9, 131.2, 129.7, 129.5, 126.6, 119.1, 114.3, 84.0, 80.9, 28.5, 25.1. Spectroscopic data are in accordance with the literature<sup>9</sup>.



*6-Bromo-1,2,3,4-tetrahydronaphthalene (***S14***)*. Following a modified literature procedure<sup>10</sup>. To a stirred solution of the 7-bromotetralone (1150 mg, 5.00 mmol, 1.00 mmol, 1.00 mmol).

equiv.) in trifluoroacetic acid (3.1 mL, 40.0 mmol, 8.00 equiv.) at room temperature was added the Et<sub>3</sub>SiH (1.8 mL, 11.0 mmol, 2.20 equiv) dropwise. After addition of Et<sub>3</sub>SiH, reaction mixture became slightly warm and, after 30 min, the solution turned cloudy. After 1h40, starting material was not totally consumed. Then, Et<sub>3</sub>SiH (0.8 mL, 5 mmol, 1 equiv.) was added and the reaction mixture was left to stir for 12h. After consumption of the starting material, indicated by TLC analysis, the reaction mixture was added dropwise to a cooled sat. NaHCO<sub>3</sub> solution (60 mL). After neutralization, the aqueous layer was extracted with ether (3x 40 mL). The organic layers were combined, washed with brine and dried over MgSO<sub>4</sub>. Purification by silica gel chromatography, eluting with hexanes (22 cm x 30 mm, isocratic elution, 400 mL run, 20 mL fractions) yielded **S14** as a translucent oil (95% yield, 1.00 g, 4.74 mmol). **R**<sub>f</sub> = 0.48 (Hexanes, KMnO<sub>4</sub> stain). <sup>1</sup>**H NMR (250 MHz, CDCl<sub>3</sub>) δ** 7.23 – 7.15 (m, 2H), 6.92 (d, J = 8.2 Hz, 1H), 2.79 – 2.64 (m, 4H), 1.85 – 1.70 (m, 4H). <sup>13</sup>**C NMR (126 MHz, CDCl<sub>3</sub>) δ** 139.5, 136.2, 131.9, 130.9, 128.5, 119.0, 29.3, 29.0, 23.1, 22.9. Spectroscopic data are in accordance with the literature<sup>11</sup>.

4,4,5,5-*Tetramethyl-2-(5,6,7,8-tetrahydronaphthalen-2-yl)-1,3,2-dioxaborolane* (**S15**). The title compound was prepared according to general procedure A, using **S14** (317 mg, 1.05 mmol, 1.00 equiv.), B<sub>2</sub>(pin)<sub>2</sub> (412 mg, 2.25 mmol, 1.50 equiv.), Pd(dppf)Cl<sub>2</sub>•DCM (56 mg, 5 mol%) and KOAc (442 mg, 4.5 mmol, 3.00 equiv.). Purification by silica gel chromatography, eluting with DCM in hexanes (20 cm x 20 mm, isocratic elution, 25% DCM/Hex, 300 mL run, 12 mL fractions) yielded **S15** as a white solid (70% yield, 273 mg, 1.06 mmol). **R**<sub>f</sub> = 0.37 (25% DCM/Hex, UV, Curcumin Stain, KMnO<sub>4</sub> stain). <sup>1</sup>**H NMR (500 MHz, CDCl<sub>3</sub>)** δ 7.55 – 7.51 (m, 2H), 7.08 (d, *J* = 7.4 Hz, 1H), 2.82 – 2.75 (m, 4H), 1.83 – 1.76 (m, 4H) 1.34 (s, 12H). <sup>13</sup>**C NMR (126 MHz, CDCl<sub>3</sub>)** δ 140.9, 136.7, 135.8, 131.9, 128.8, 83.7, 29.8, 29.3, 25.0, 23.4, 23.2. Spectroscopic data are in accordance with the literature<sup>11</sup>



*2-(6-methoxynaphthalen-2-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane* (S16). The title compound was prepared according to general procedure A, using 2-bromo-6methoxynaphthalene (367 mg, 1.50 mmol, 1.00 equiv.), B<sub>2</sub>(pin)<sub>2</sub> (412 mg, 1.58 mmol, 1.05 equiv.), Pd(dppf)Cl<sub>2</sub>•DCM (56 mg, 5 mol%) and KOAc (442 mg, 4.50 mmol, 3.00 equiv.). Purification by silica gel chromatography, eluting with AcOEt in Hexanes (15 cm x 15 mm, isocratic elution, 10% AcOEt/Hex, 250 mL run, 20 mL fractions) yielded **S16** as a white solid (98% yield, 415 mg, 1.46 mmol). **R**<sub>f</sub> = 0.37 (5% AcOEt/Hexanes, Curcumin Stain). Spectroscopic data are in accordance with the literature<sup>12</sup>



#### 4. TR-FRET binding displacement assay

**Figure S5.** Percentage inhibition for compounds 57 and 59 against STK10 and SLK as measured in a TR-FRET binding–displacement assay. Each measurement was measured in duplicate.

#### 5. References

- Gharpure, S. J.; Anuradha, D.; Prasad, J. V. K.; Srinivasa Rao, P. Stereoselective Synthesis of *Cis* -2,6-Disubstituted Morpholines and 1,4-Oxathianes by Intramolecular Reductive Etherification of 1,5-Diketones. *Eur. J. Org. Chem.* **2015**, 2015 (1), 86–90.
- Bruce, I.; Culshaw, A. J.; Devereux, N. J.; Gessier, F.; MC, K. J.; Neef, J.; Oakman, H. E.; McKenna, J. Organic Compounds. US2010035874 (A1), 2010.
- (3) Creary, X.; Sky, A. F.; Phillips, G.; Alonso, D. E. Reaction of Arylhalodiazirines with Thiophenoxide: A Redox Process. *J. Am. Chem. Soc.* **1993**, *115* (17), 7584–7592.
- (4) Sandham, D. A.; Barker, L.; Brown, L.; Brown, Z.; Budd, D.; Charlton, S. J.; Chatterjee, D.; Cox, B.; Dubois, G.; Duggan, N.; et al. Discovery of Fevipiprant (NVP-QAW039), a Potent and Selective DP 2 Receptor Antagonist for Treatment of Asthma. ACS Med. Chem. Lett. 2017, 8 (5), 582–586.
- (5) Taylor, N. J.; Emer, E.; Preshlock, S.; Schedler, M.; Tredwell, M.; Verhoog, S.; Mercier, J.; Genicot, C.; Gouverneur, V. Derisking the Cu-Mediated 18F-Fluorination of Heterocyclic Positron Emission Tomography Radioligands. *J. Am. Chem. Soc.* 2017, 139 (24), 8267–8276.
- (6) Arakawa, Y.; Nakajima, S.; Kang, S.; Shigeta, M.; Konishi, G.; Watanabe, J. Design of an Extremely High Birefringence Nematic Liquid Crystal Based on a Dinaphthyl-

Diacetylene Mesogen. J. Mater. Chem. 2012, 22 (28), 13908.

- (7) Vidadala, R. S. R.; Rivas, K. L.; Ojo, K. K.; Hulverson, M. A.; Zambriski, J. A.; Bruzual, I.; Schultz, T. L.; Huang, W.; Zhang, Z.; Scheele, S.; et al. Development of an Orally Available and Central Nervous System (CNS) Penetrant Toxoplasma Gondii Calcium-Dependent Protein Kinase 1 (TgCDPK1) Inhibitor with Minimal Human Ether-a-Go-Go-Related Gene (HERG) Activity for the Treatment of Toxoplasmosis. *J. Med. Chem.* **2016**, *59* (13), 6531–6546.
- (8) Kumar, S.; Hernandez, D.; Hoa, B.; Lee, Y.; Yang, J. S.; McCurdy, A. Synthesis, Photochromic Properties, and Light-Controlled Metal Complexation of a Naphthopyran Derivative. *Org. Lett.* **2008**, *10* (17), 3761–3764.
- (9) Naumiec, G. R.; Cai, L.; Lu, S.; Pike, V. W. Quinuclidine and DABCO Enhance the Radiofluorination of 5-Substituted 2-Halopyridines. *European J. Org. Chem.* 2017, 2017 (45), 6593–6603.
- West, C. T.; Donnelly, S. J.; Kooistra, D. A.; Doyle, M. P. Silane Reductions in Acidic Media. II. Reductions of Aryl Aldehydes and Ketones by Trialkylsilanes in Trifluoroacetic Acid. Selective Method for Converting the Carbonyl Group to Methylene. *J. Org. Chem.* **1973**, *38* (15), 2675–2681.
- (11) Sakai, N.; Kobayashi, T.; Ogiwara, Y. One-Pot Synthesis of Tetralin Derivatives from 3-Benzoylpropionic Acids: Indium-Catalyzed Hydrosilylation of Ketones and Carboxylic Acids and Intramolecular Cyclization. *Chem. Lett.* **2015**, *44* (11), 1503–1505.
- (12) Liu, C.; Ji, C.-L.; Hong, X.; Szostak, M. Palladium-Catalyzed Decarbonylative Borylation of Carboxylic Acids: Tuning Reaction Selectivity by Computation. *Angew. Chemie Int. Ed.* **2018**, 57 (51), 16721–16726.













Figure S10. <sup>1</sup>H NMR Spectra (DMSO- $d_6$ /D<sub>2</sub>O/TFA, 600 MHz) of 7 S20



















Figure S17.  $^{13}\text{C}\{^{1}\text{H}\}$  NMR Spectra (DMSO-d\_6, 126 MHz) of 10





Figure S19.  $^{13}\text{C}\{^{1}\text{H}\}$  NMR Spectra (DMSO-d\_6, 126 MHz) of 11









Figure S23.  $^{13}\text{C}\{^1\text{H}\}$  NMR Spectra (DMSO-d\_6, 126 MHz) of 13












Figure S28. <sup>1</sup>H NMR Spectra (MeOD-d<sub>4</sub>, 400 MHz) of **16** 



Figure S29.  ${}^{13}C{}^{1}H$  NMR Spectra (MeOD- $d_4$ , 126 MHz) of 16





Figure S31. <sup>13</sup>C{<sup>1</sup>H} NMR Spectra (DMSO-*de*, 126 MHz) of 17

















Figure S38. <sup>1</sup>H NMR Spectra (DMSO-d<sub>6</sub>, 250 MHz) of 21











Figure S43.  ${}^{13}C{}^{1}H$  NMR Spectra (DMSO-d<sub>6</sub>, 126 MHz) of 23







Figure S46. <sup>1</sup>H NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 400 MHz) of 25

190 180 170	c160.0	TZ Z
160	H 159.7 159.3	
150	$- 138.9 \\ - 153.1 \\ - 146.4$	/ NM
140	/133.3 /130.0	e <sub>2</sub>
130	$ \begin{array}{c}                                     $	<b></b>
120		TE TE 2 121
110	E 115.0 115.0 112.7	120.8
100	$\begin{bmatrix} 112.1\\109.5 \end{bmatrix}$	, 119
06	₩	H H H H H H H H H H H H H H H H H H H
80	$\frac{1}{3}$	1 117
70	-129.0 -127.5 -126.3	115.7
60	idd 	115.0
50		1114
40	<u><u>a</u> 39.7 39.5</u>	μ ω μ μ μ μ μ μ μ μ μ μ μ μ μ μ μ μ μ μ
30	39.3 39.1 38.9	112.1
20		1111
10	-39.9 -39.7 -39.5	E
udd 0	yp 39.3 39.1 38.9	udă 607 Linnulu Whythere

Figure S47.  ${}^{13}C{}^{1}H$  NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 101 MHz) of 25



Figure S48. <sup>1</sup>H NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 400 MHz) of 26

858







Figure S51. <sup>13</sup>C{<sup>1</sup>H} NMR Spectra (DMSO-*d*<sub>6</sub>, 126 MHz) of 27



Figure S52. <sup>1</sup>H NMR Spectra (CDCl<sub>3</sub>, 500 MHz) of 28

		TX
		$ \begin{array}{c}                                     $
	CDC13	$<_{76}^{77}$
		-38 -32 -32 -32 -32 -32 -32 -32 -32
26 ppm	26.2 25.9	

Figure S53.  $^{13}C\{^1H\}$  NMR Spectra (CDCl<sub>3</sub>, 126 MHz) of 28



Figure S54. <sup>1</sup>H NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 400 MHz) of 29



Figure S55.  $^{13}C$ { $^{1}H$ } NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 101 MHz) of 29



190 180 170 160		H Me
150		-149.7
140		-137.6
130		133.1 128.8 126.8
120		~124.8
110		
100		
90		
08	DC13	$4^{77.4}$
70		76.9
60		
50		
40		
30		<u> </u>
20		-22.2
1		-13.9
0		
urda 0		

Figure S57. <sup>13</sup>C{<sup>1</sup>H} NMR Spectra (CDCl<sub>3</sub>, 126 MHz) of **30** 





Figure S59.  $^{13}\text{C}\{^1\text{H}\}$  NMR Spectra (DMSO-d\_6/D\_2O/TFA, 101 MHz) of 31



Figure S60. <sup>1</sup>H NMR Spectra (DMSO-d<sub>6</sub>, 500 MHz) of **32** 




190	Ž
180	TZ Z
170	XZ
160	0~0
150	153.9 152.1 149.7
140	
130	
120	 -118.5
110	
100	
06	
80	 
70	
60	
50	43.1 40.0 739.9
40	- 39.7 DMSO
30	- 1 39.2 5
20	30.4
10	
0	
udd	

Figure S63.  $^{13}\text{C}\{^1\text{H}\}$  NMR Spectra (DMSO-d\_6, 126 MHz) of 36



Figure S64. <sup>1</sup>H NMR Spectra (CDCl<sub>3</sub>, 500 MHz) of 43











Figure S69.  $^{13}C{^1H}$  NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 46







283 20/07/08









כס*י*ר אוגווי סאברנומ (הוגוסס-מי*י*/ ה7ס/ וו







Figure S79.  ${}^{13}C{}^{1}H$  NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 51





Figure S81. <sup>13</sup>C(<sup>1</sup>H) NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 52







S6S















Figure S90. <sup>1</sup>H NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 500 MHz) of 57 S100



Figure S91.  $^{13}C(^{1}H)$  NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 57







Figure S94.  $^{13}C$ C<sup>1</sup>H} NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 58
















**ire S102.** <sup>1</sup>H NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 500 MH) S112



Figure S103. <sup>13</sup>C{<sup>1</sup>H} NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 61





Figure S105. <sup>13</sup>C{<sup>1</sup>H} NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 62





Figure S107.  $^{13}C{^1H}$  NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 63





Figure S109. <sup>13</sup>C{<sup>1</sup>H} NMR Spectra (DMSO-d<sub>6</sub>/D<sub>2</sub>O/TFA, 126 MHz) of 64











Figure S114. <sup>1</sup>H NMR Spectra (CDCl<sub>3</sub>, 250 MHz) of 71



Figure S115.  $^{13}\text{C}\{^{1}\text{H}\}$  NMR Spectra (CDCl3, 126 MHz) of 71







	1	ş			
	190				z
<b>Figure S119.</b> <sup>13</sup> C{ <sup>1</sup> H} NMR Spectra (CD S129	180				S
	170				~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~
	160				CN
	150				
	140				145.8 142.2
	) 13(				-132.6
	0 12	1			-126.5 -122.9
	0 11				-118.4
	0 10				-108.4
	9 0(				
	0			0	
Cl <sub>3</sub> , 126	õ.		 	DC13	$\left\{ \begin{array}{c} 77.2 \\ 76.9 \end{array} \right\}$
5 MHz)	70				
of <b>S3</b>	60 l				
	50				
	40				
	30				
	20				-20.6
	10		_		-14.8
	0				
	mdd	1			



	190 180 170 160 1.			Me
<b>Figure S121.</b> <sup>13</sup> C{ <sup>1</sup> H} NMR Spectra (CDCl <sub>3</sub> , 1: S131	50 14			$<^{144.3}_{144.0}$
	0 130			
	120			-125.3
	110			-110.5
	100			
	90		CDC1	
	08		ώ	$-\left\{\begin{array}{c} 77.4\\ 77.2\\ 77.2\\ 76.2\end{array}\right\}$
26 MHz)	70			-76.9
of <b>S4</b>	60 l			
	50			
	40			
	30			
	20			-20.8
	10			
	udd 0			



	200 190 180 170	-191.4 OS
Figure S123. <sup>13</sup> C{ <sup>1</sup> H} NMR Spectra (CDCl <sub>3</sub> , 101 MHz) of S5 S133	160 150 140 130 120 110 1	$ \begin{array}{c}             144.4 \\             142.1 \\             137.6 \\             132.4 \\             130.6 \\             125.3 \end{array} $
	00 90 80 70 60 50	77.5 CD CL
	40 30 2	
	0 10 0 ppm	- T3.0



190	Me
180	a o a
170	×–×
160	s
150	
140	
130	<u> </u>
120	126.1 124.3 122.0
130110	
100	
. 90	
80	
) 7(	←77.2 G
6	
5	
0 4	
0	
30	25.1
20	
10	
udd 0	

Figure S125.  $^{13}\text{C}\{^1\text{H}\}$  NMR Spectra (CDCl<sub>3</sub>, 126 MHz) of S6



	190 180 170		Me Me Me O B
	160 150		)=/ OH
Figure S127. <sup>13</sup> C{ <sup>1</sup> H} NMR S	140 130 120 110 100	$ \begin{array}{c} 136.6 \\ 136.3 \\ 137 \\ 131.3 \\ 130.9 \\ 128.5 \\ 125.7 \\ 117.9 \\ 117.9 \\ 109.5 \\ 134 \end{array} $	136.6  
pectra (CDCl <sub>3</sub> , 126 MHz) of <b>S7</b>	90 80 70 60	133 132 131 130	— 131.3 130.9
	50 40 30 2	 129 128 128 12 128 12	
	0 10 0 ppm	7 126 ppm	125.7

0000



	190 180 1		Me Me Me O B	
	70 160 1		OEt	
Figure S129. <sup>13</sup> C{ <sup>1</sup> H}	50 140 130 120 110			136.6 136.1 131.2 130.3 128.5 126.0 119.1
} NMR Spectra (CDC S139	100 90 8	 		
<sub>3</sub> , 126 MHz) of	0 70 60			$- < 77.2 \\ 77.2 \\ 76.9 \\ - 63.6$
99	50 40			
	30 20			
	10 0 ppm			



190 180 170 1					Me Me Me O B
60 1				0	>
50			]		-136.5
 140		$\angle^{136.5}_{136.1}$	136		-136.1
130		131.2 130.3 128.7	134		
120	<b></b>	-126.1 -118.7	132		121_0
110		-108.1	130		
100			128	<b> </b>	-128.7
06			. 126	<u> </u>	-126.1
08		-83.9	12		
70		76.9 0	4.		
60			1		
50			20 .		-118.7
40			118 .		
ω			116		
	[	25.1	114		
20			112		
10		<del></del> 6.4	110		
0 pm			اللغ ا	<u> </u>	-108.1

Figure S131.  $^{13}\text{C}\{^1\text{H}\}$  NMR Spectra (CDCl\_3, 126 MHz) of S11



Figure S132.  $^1$ H NMR Spectra (DMSO-d\_6, 500 MHz) of S12





- -H NMR Spectra (CDCI3, 500 MH2 S144
|  | 190 180 170 |  | Me Me<br>Me O B                               |
|--|-------------|--|---|
| Figure C135. 13C(1H) NMR Spectra (CDC), 136 MH | 160 1       |  | ⊐z<br>)=0                                     |
|  | 50          | <b>c</b> 137.0                               | $\succ$                                       |
|  | <br>140     | <br>136.0                                    | 11  |
|  | 130         | 131.2<br>129.7<br>129.5                      | <sup>1</sup> <sup>3</sup> <sub>7</sub> −137.0 |
|  | 120         | $\sim$ 126.6<br>$\sim$ 119.1<br>$\sim$ 114.2 |   |
|  | 110         | 114.5  | н III - 135.9                                 |
|  | 100         |  | 35  |
|  | 90          | ×84.0  | 134   |
|  | 08          | ✓ 80.9 ✓ 77.4 G ✓ 77.4 G                     | 133   |
|  | 70          | \_76.9 <sup>™</sup>                          | 1   |
| ) of <b>S1</b> :                               | 60          |  | × 1   |
| U.   | 50          |  |   |
|  | 40          |  |   |
|  | 30          | <br>28.5                                     | -129.5  |
|  | 20          | 25.1   | 2   |
|  | 10          |  | 28  |
|  | 0           |  | 127   |
|  | mdd         |  | rp 126.6                                      |

**Figure 3133.**  $\mathbb{C}^{-1}$  (The NMR spectral (LDCI3, 126 MHz) of **313** 



Figure S136.  $^1$ H NMR Spectra (CDCl<sub>3</sub>, 250 MHz) of S14

\_



Figure S137.  $^{13}\text{C}\{^1\text{H}\}$  NMR Spectra (CDCl3, 126 MHz) of S14



	190 180 170 160		Me Me Me O B
<b>Figure S139.</b> <sup>13</sup> C{ <sup>1</sup> H}	150 140 130 120 110	$ \begin{array}{c} & 140.9 \\ & 136.7 \\ & 135.8 \\ & 131.9 \\ & 128.8 \end{array} $	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$
NMR Spectra (CDCl <sub>3</sub> , 126 MHz) of <b>S15</b>	100 90 80 70 60	- 83.7 - 77.4 GB 77.201 76.9℃	132 — 131.9 132 — 131.9 130 ppm 30 — 128.8 30 — 29.8 — 29.8 — 29.3
	50 40 30 20 10 0 pj	$ \begin{array}{c} 29.8 \\ 29.3 \\ 25.0 \\ 23.4 \\ 23.2 \end{array} $	28 27 26 25.0 25 24 p



190 180 170					Me Me Me O B
160			-158.7		OMe
150 140 130 120	mqq	<130.4 130.4	136.6 136.1 131.3 130.4 128.5 126.0 118.8	137 136 13	
110			-105.8	5 13	
100				°4	
06				133	
08			$\overset{-83.9}{\leftarrow}^{77.4}_{77.2}$	132	
70			∼76.9₩		
60			<u> </u>		$<^{130.4}_{130.4}$
50			55.4	130	100.1
40				129	
30				128	
20			-25.1		
10				т 27	
udd 0				126 ppm	-126.0

Figure S141.  $^{13}\text{C}\{^1\text{H}\}$  NMR Spectra (CDCl3, 151 MHz) of S16

S151

י ר גר י