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Supplemental Information

Lgr5 and Col22a1 Mark Progenitor Cells in the Lineage toward Juvenile

Articular Chondrocytes

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Figure S1. Quantitative PCR showed expression of Gdf5 and Lgr5 in digit interzone mRNA was extracted from the digit interzone (IZ) and surrounding non-interzone regions of a E14.5 embryo to generate cDNA for QPCR. Expression of *Gapdh* was normalized to 1.0, values higher than 1.0 indicates these genes are upregulated, vice versa. Among all the selected stem cell/progenitor markers, known marker of joint interzone *Gdf5* and novel marker identified in this paper *Lgr5* were upregulated in the interzone region.



Figure S2. Lgr5-GFP mutant developed normal knee and digit joints

Representative histology sections stained with Alcian blue and nuclear fast red, showing no observable difference between the developing knee (A) and digit joints (B) of Lgr5^{+/+}, Lgr5^{GFP/4} and Lgr5^{GFP/GFP} E18.5 embryos (n=3 per genotype). (C) The articular cartilages appeared to have formed normally with the expected expression and localization of COLXXII in the superficial layer. F=femur, T=tibia, CI=cruciate ligament, M=metacarpal, P1= proximal phalange, P2=middle phalange, P3=distal phalange. Scale bar, 200µm



Figure S3. Bulk transcriptomic analysis of cell populations in joint development.

(A) Three population of cells isolated. One from Sox9 expressing cells in E13.5 digit of Sox9-eGFP mice, and two populations from the E14.5 interzone region of the developing digits that contains the Lgr5+ interzone (red arrows) and surrounding non-GFP region (yellow arrows) from Lgr5-GFP mice (B). Scalebars, 1mm. (C) Diagrammatic illustration of the protocol with all cell isolated were sorted for the appropriate population of cells for subsequent for RNA-seq for the generating the three datasets. (D) Workflow for bulk RNAseq data processing. (E) Venn diagram showing the number of genes identified, and overlaps between datasets. Genes with higher expression in Lgr5+ population are shown on the right. (F) Heat maps depicting comparative gene expression levels of the top genes involved in specified molecular pathways. + Fold change (FC) indicates an increase and -FC for decrease in Lgr5+ cells. "+ ∞ " are given to indicate genes that are expressed only in Lgr5+ cells. Related to figure 3.



Figure S4. Expression of Lgr5 and COLXXII in phalangeal and knee joints.

Co-localization of Lgr5 (Green) and COLXXII (Red) in E18.5 (A) and newborn (P0) (B) digit joints. (C) Expression in M/P1, Ta/M and Knee joints at P10. M, metatarsal. P1, proximal phalange. P2, middle phalange. P3, distal phalange. Ta/M, tarsometatarsal joint. (D) Cells embedded in the COLXXII layer are flat compare to the round chondrocytes in the deeper zones (yellow arrow) of P6 mouse. Focal adhesions (β 1-integrin) are found at both sides of the cells where they are in direct contact with the COLXXII matrix (white arrows). Scale bars in (A-C), 100µm and in (D), 10µm. Related to figure 3.



Figure S5 Additional information for single cell analysis

(A) Volcano plot showing *Gdf5* (red circle) is the only differential expressing gene (DEG) between *Lgr5+/Gdf5-* and *Lgr5+/Gdf5+* subpopulations, and these cells are essential identical. (B) Volcano plot shows DEGs between Lgr5+ cells from cluster 1 and cluster 5, also show they are the same cells, differ only as proliferating cells in cluster 5. (C) GO term analysis of the DEGs from panel (B) (circled). (d-g) Principle component analysis (PCA) for the 5,460 E14.5 knee joint cells. (D) A contour plot showing the density of cells, two major peaks are found. (E) Peak 1 corresponds to interzone cells expressing *Gdf5* and *Lgr5*, and peak 2 as non-interzone expressing mature chondrocyte markers, *Comp* and *Epyc.* (F) t-SNE clusters 1, 2, 5, and 6 are located in peak 1, whereas clusters 3 and 4 are in peak 2, consistent with the 2D t-SNE finding. (G) Contour plot of the cell cycle genes showing the existence of proliferating subpopulation of cell in the interzone (cluster 5) and non-interzone (cluster 4) clusters. (H) Violin plots showing the expression profiles of representative cartilage, ligament and cell cycle genes in each cluster supporting their identities/signatures. Related to figure 4.



Puncture site

Puncture site

Figure S6. Lgr5+ interzone tissue transplantation for repairing of a cartilage lesion

(A) No tdTomato+ cells was detected in the puncture site without interzone tissue insert. Bright-field image of the same region was shown on the right. (B) Analysis of two additional animals receiving interzone tissue transplantation. Similar positive outcomes among all three operated animals, with differentiation of the interzones to chondrocytes expressing COL II, reestablishment of the superficial layer (CILP1 expressing), and suppressing COL I expression and fibrotic tissue. Scalebars, 50µm. **Related to figure 6.**

	Expressed in Sox9+ only (EPKM>5)			Expressed in Lgr5- only (FPKM>5)			Expressed in Lgr5+ only (FPKM>5)					
	Cene	e13.5	e14.5	e14.5	Gene	e13.5	e14.5	e14.5	Cene	e13.5	e14.5	e14.5
	Gene	Sox9+	Lgr5-	Lgr5+		Sox9+	Lgr5-	Lgr5+	Gul	Sox9+	Lgr5-	Lgr5+
1	Epyc	48.82	0.63	1.2	Alas2	0.86	25.41	1.16		0.2	0.75	58.03
2	Smpd3	30.66	0.29	0.19	Foxf2	2.52	20.2	2.85	GlisI	4.33	3.54	52.57
3	Panx3	27.67	1.16	1.14	Strab	4.58	18.26	4.13	Adck3	0.99	2.62	35.46
4	Epha2	20.86	4.24	4.68	Cpa3	0.81	15.64	2.79	Cdh13	2.91	2.97	35.39
5	Frzb	20.82	1.77	2.12	Sst	0.23	15.56	4.07	Hhip	2.97	4.94	30.22
6	Foxd1	19.47	3.43	0.71	Cd74	1.82	12.69	4.52	Atp1a2	3.4	1.89	27.54
7	Cxcl10	18.67	1.73	3.35	Zfhx4	4.26	12.46	4.66	Lgr5	3.48	1.36	26.75
8	Pltp	17.52	3.89	2.93	Cmal	0.39	12.41	2.26	Chst5	4.06	3.01	26.54
9	Hoxc10	17.02	0.21	0.37	Ablim1	3.74	12.18	4.9	Susd4	3.6	4.89	22.94
10	Tnfaip3	16.98	4.13	4.71	S100a8	0	11.5	0.86	Msc	2.18	1.28	22.79
11	Peg10	14.87	3.71	2.4	Grp	1.24	11.19	4.11	Barx2	1.27	0.25	20.41
12	Spp1	14.72	2.5	2.13	Snca	0.69	9.95	1.52	Pknox2	3.74	1.42	19.37
13	Sgms2	14.54	3.67	3.36	Fgf2	4.52	9.61	4.19	Btc	0.46	0.56	19.22
14	Chac1	14.27	0.64	1.38	Angpt2	0.78	9.45	1.97	Gfra1	2.9	3.67	18.25
15	Ndufa4l2	14.12	2.13	3.35	Кср	4.12	9.03	4.84	Akr1b10	3.87	1.78	18.06
16	Chst11	14.06	4.4	4.75	Arhgdig	4.34	9.01	4.21	Klc3	3.59	4.05	17.07
17	Ptgs2	13.6	1.04	1.41	Axin2	2.12	8.94	3.13	Tubb4a	4.29	4.63	16.12
18	Mpzl2	13.06	3.79	4.67	Cd52	0	8.86	2.87	Gcgr	3.2	1.77	15.97
19	Edil3	12.86	3.73	4.66	Kcnk2	3.13	8.79	4.11	Col22a1	1.5	1.93	15.82
20	S100b	12.1	2.56	2.62	Stmn2	2.71	8.7	1.51	Svopl	1.11	1	15.72
21	Foxo6	12.07	4.31	3.36	Dbx2	1.4	8.68	0.74	Vaultrc5	4.63	2.57	15.48
22	Sytl2	12.03	2.32	4.18	Alox5ap	1.35	8.64	2.61	Pde2a	2.11	2.81	15.23
23	C1galt1	11.68	3.47	4.69	Nxnl2	2.17	8.62	3.73	Ppp1r1b	2	2.62	14.44
24	Cgrefl	11.62	4.64	4.25	Srgn	0.45	8.58	2.88	Sntb1	1.77	2.9	13.36
25	Pmaip1	11.61	2.98	2.64	Cmpk2	3.86	8.56	4.93	Thbs4	2.09	2.94	12.9
26	Tcf15	11.41	4.13	3.25	F11r	3.94	8.27	3.13	Krt5	4.61	1.24	12.62
27	Saa1	11.33	2.08	4.06	Kif26a	3.06	8.07	1.13	Lrrn3	2.5	1.52	12.38
28	Hck	10.72	4.42	1.37	Gstm6	2.7	8.06	3.74	Tnnc1	1.83	1.07	12.27
29	Xylt1	10.7	4.44	3.26	Heph	2.99	7.92	3.35	Ephx1	1.74	2.72	12.15
30	Entpd6	10.53	4.55	4.95	H2-T22	2.59	7.91	4.31	Stc1	3.61	4.5	11.55
31	Col15a1	10.4	3.93	2.15	Il17rd	2.63	7.89	2.76	Cerl	2.53	0.03	11.04
32	Magel2	10.39	4.97	3.5	Gpr124	4.32	7.8	2.74	Ptpro	0.68	0.76	10.97
33	Ifitm5	10.38	2.22	1.74	Fbxl19	4.86	7.78	4.63	Matk	2.46	2.32	10.56
34	Bmp5	10.25	4.5	2.69	Tmtc2	2.81	7.77	2.37	Acta2	2.64	3.18	10.21
35	<i>E2f</i> 7	9.81	4.32	3.38	Lgals7	2.52	7.75	3.95	Ppplrla	3.75	2.33	9.98
36	Foxa3	9.6	2.47	3.36	Raver1-fdx11	3.61	7.36	3.59	Kcnma1	4.52	2.85	9.71
37	Ssfa2	9.58	4.32	4.96	Adamts9	3.79	7.33	3.19	Sh2b2	3.44	4.96	9.6
38	Clec11a	9.43	2.23	1.37	Nek8	4.51	7.31	4.84	Glrb	2.27	4.98	9.5
39	Eif3j	9.34	0.33	0.66	Hdc	0.85	7.31	0.71	Des	1.17	1.61	9.33
40	Serinc5	9.13	2.57	2.94	Ngfr	0.67	7.28	3.72	Camklg	3.22	4.51	9.32
41	Ncoa7	9.03	2.27	2.52	Scara5	1.44	7.15	3.18	Pitpnm1	4.26	4.74	9.26
42	Hoxa5	8.96	2.36	1.97	Pdxk	4.82	7.12	4.5	Gdf1	3.37	3.67	9.1
43	Dlc1	8.89	3.62	3.41	Kctd1	4.39	7.12	3.9	Krtdap	0.62	0.19	9.02
44	E2f8	8.67	2.56	3.61	Cav1	2.19	6.92	3.6	Kcnj15	0.4	0.47	9.02
45	Otor	8.65	3.95	2.44	Tubb3	2.06	6.9	1.24	Ccl4	1.41	4.75	8.88
46	Stil	8.53	3.47	3.27	Arhgap39	4.5	6.89	3.82	Sfn	3.54	1.38	8.86
47	Avprla	8.45	1.25	0.8	Lypd6	2.57	6.71	3.31	Sema3e	3.21	3.2	8.8
48	Fndc3c1	8.39	2.26	2.93	Lbh	4.96	6.7	2.52	Myod1	0.46	1.69	8.77

 Table S1. Genes expressed by only one population (top 100 ranked by level) (Digit bulk)

49	Frat1	8.29	2.48	4.36	Slc25a14	3.08	6.7	4.78	Smtnl2	3.1	4.07	8.64
50	Armcx5	8.25	3.75	4.53	Ophn1	4.37	6.68	4.33	Pf4	1.69	3.34	8.57
51	Nol12	8.23	4.59	4.21	Adamts17	3.42	6.67	3.66	Snap91	2.29	1.93	8.43
52	Ptger4	8.16	2.88	2.04	Mterfd3	3.9	6.62	4.78	Dcx	2.93	2.18	8.41
53	Ttll3	8.13	4.77	4.21	Rev3l	4.38	6.58	4.67	Fam159a	3.68	3.42	8.39
54	Dclre1c	8.02	2.77	4.02	Acsf3	3.84	6.54	3.97	Accs	2.66	3.37	8.32
55	Stk17b	8	2.3	1.7	Stom	2.8	6.54	4.95	Gata3	3.49	0.84	8.3
56	Hist1h2bn	8	3.32	3.49	Ank2	3.39	6.48	4.86	Myl1	1.83	2.24	8.28
57	Omd	7.96	3.38	3.72	Bcl2l11	3.94	6.42	4.94	Slc25a23	2.77	4.17	8.13
58	Pcdh1	7.9	0.67	2.57	Ankrd44	2.78	6.42	3.49	Tcea3	1.68	4.53	8.08
59	Npat	7.88	4.84	4.74	Nkain l	3.17	6.4	3.93	Doc2g	4.83	4.62	7.9
60	Cbr2	7.86	4.12	3.25	Ptges	3.93	6.39	3.44	Spon2	0.21	2.12	7.89
61	Hspg2	7.84	3.74	2.28	Kit	2.35	6.38	2.5	Abcg4	3.87	4.93	7.88
62	Pygl	7.81	3.93	3.2	Mgat3	3.77	6.35	3.81	Adamts14	3.42	2.32	7.81
63	Krtcap3	7.75	2.34	4.88	Fzd10	1.6	6.33	1.17	Krt15	2.78	1.78	7.81
64	Shroom3	7.72	4.68	4.46	Slitrk5	3.45	6.31	3.4	Dleu2	1.11	3.16	7.77
65	Hist1h3g	7.67	4.3	2.68	Ddx23	4.78	6.29	4.63	Cntfr	2.19	1.1	7.71
66	Scd1	7.62	3.42	1.66	Camk2g	4.35	6.26	4.27	Phactr1	1.35	1.55	7.69
67	Ung	7.59	3.32	2.78	Fap	1.5	6.26	3.85	Fbp2	4.11	1.92	7.65
68	Nppc	7.59	3.69	3.88	Rhobtb2	4.82	6.25	4.68	Anol	2.91	2.32	7.62
69	Baiap211	7.58	2.65	2.78	Tgfbr3	4.87	6.22	3.44	Ehbp111	3.16	3.34	7.59
70	Tgfa	7.57	2.66	1.99	Atxn7l1	4.59	6.19	3.89	Uap111	3.42	4.25	7.53
71	Adamts3	7.53	4.01	2.9	Wbp7	4.39	6.12	4.97	Wdr67	4.62	4.69	7.41
72	Prkg2	7.48	1.73	1.22	Iqgap2	4.99	6.1	4.34	Tmem117	1.95	1.4	7.39
73	Orc1	7.4	3.33	3.57	Sdpr	2.35	6.09	3.2	Dynlt3	4.2	4.23	7.38
74	Aldoc	7.38	3.34	4.4	Zfp322a	4.84	6.06	4.9	Tle6	3.26	4.42	7.36
75	Foxfla	7.37	0.58	1.26	Slc2a4	3.46	6.06	2.19	Ramp1	0.56	0.62	7.33
76	Cenpk	7.36	4.04	4.36	Pear1	4.79	6.04	3.57	Rxrg	4.78	2.31	7.22
77	Fam180a	7.35	1.15	2.41	Mtss1	3.04	6.03	4.92	St7l	4.32	4.72	7.18
78	Tgm2	7.34	3.85	3.84	Traf5	4.46	6.01	4.74	Wnt9a	3.13	3.77	7.17
79	Esco2	7.34	4.01	4.35	Fgf10	2.45	6	2.21	Tesk2	4.7	4.25	7.16
80	Hiplr	7.33	3.6	4.68	Gdf11	3.4	5.98	4.88	Coq10a	4.14	4.78	7.16
81	Slc19a2	7.32	4.28	4.71	Txnrd2	3.51	5.95	3.92	Fam20c	3.93	3.24	7.16
82	Cenpf	7.32	3.48	4.93	Tspan11	1.58	5.95	3.57	Rarres2	3.09	3.66	7.16
83	Pdlim1	7.29	3.41	2.27	Kpna6	4.89	5.94	4.26	Pip4k2a	3.77	3.38	7.12
84	Per2	7.28	2.93	4.73	Fert2	4.55	5.9	4.66	Dmrt2	1.31	2.73	7.06
85	Ascc3	7.27	4.03	4.96	Zfp397	4.26	5.9	4.14	Nt5e	2.7	1.32	7.02
86	St6galnac2	7.25	4.83	3.19	Smcr8	4.69	5.89	3.4	Tagln	3.33	2.95	7.01
87	Scn1b	7.22	2.19	2.08	Gng8	3.13	5.88	3.46	Col13a1	0.75	4.35	6.98
88	Grem1	7.16	1.11	2.71	Sall1	2.8	5.88	2.77	Aard	3.16	3.38	6.97
89	Nptx2	7.12	1.28	0.73	Nlrx1	2.62	5.87	4.8	Tesc	1.57	0.78	6.95
90	Usp2	6.93	4.18	3.64	Prmt10	4.21	5.86	4.73	Shq1	4.9	4.87	6.91
91	Gpd2	6.93	4.65	4.22	Tcf7	2.91	5.86	3.27	Ell3	3.03	2.34	6.91
92	Snhg4	6.91	3.3	2.85	Slc22a5	3.73	5.84	4.4	Clqa	0.31	0.86	6.91
93	Isl1	6.86	0.14	0.02	Ptch2	2.46	5.83	1.86	Tmem175	4.76	4.96	6.87
94	Lingol	6.78	3.68	2.82	Prr5l	1.38	5.83	1.75	Dner	0.57	1.67	6.83
95	Pcp4	6.75	2.28	1.82	Tyk2	4.38	5.79	4.23	Apba2	3.22	3.57	6.81
96	Fam84b	6.75	2.9	3.26	Cdc42bpa	4.25	5.75	4.38	Ppp1r3g	3.54	1.43	6.79
97	Dlx3	6.69	3.11	1.26	Srd5a1	4.44	5.73	4.28	Dcaf6	3.38	3.82	6.78
98	Rfx1	6.68	4.95	3.69	Ccdc28a	3.3	5.73	3.59	Igsfl 1	2.7	1.46	6.77
99	Arl4c	6.67	2.97	3.03	Pcnx	4.42	5.72	4.81	Krt14	4.43	0.54	6.76
100	Foxn2	6.64	3.72	4.97	Hnrnpul2	4.27	5.69	3.96	Fbp1	1.5	1.46	6.76

Gene	e13.5 Sox9+	e14.5 Lgr5- (FPKM)	e14.5 Lgr5+
Transcription factors ex	spressed in Lgr5+ only		
Glis1	4.33	3.54	52.57
Msc	2.18	1.28	22.79
Barx2	1.27	0.25	20.41
Pknox2	3.74	1.42	19.37
Gata3	3.49	0.84	8.3
Tcea3	1.68	4.53	8.08
Rxrg	4.78	2.31	7.22
Dmrt2	1.31	2.73	7.06
Rorc	1.03	0.62	6.02
Pitx3	0.79	1.01	5.71
Transcription factors ex	pressed in Lgr5- only		
Foxf2	2.52	20.2	2.85
Dbx2	1.4	8.68	0.74
Lbh	4.96	6.7	2.52
Ptges	3.93	6.39	3.44
Wbp7	4.39	6.12	4.97
Rfx5	4.76	5.54	4
Tcf20	3.21	5.48	3.03
Transcription factors ex	xpressed in Sox9+ only		
Foxd1	19.47	3.43	0.71
Hoxc10	17.02	0.21	0.37
Tnfaip3	16.98	4.13	4.71
Foxo6	12.07	4.31	3.36
Tcf15	11.41	4.13	3.25
Foxa3	9.6	2.47	3.36
Hoxa5	8.96	2.36	1.97
Foxfla	7.37	0.58	1.26
Pdlim1	7.29	3.41	2.27
Per2	7.28	2.93	4.73
Rfx1	6.68	4.95	3.69
Foxn2	6.64	3.72	4.97
Pou3f3	6.58	1.83	2.45
Ziml	6.44	1.13	1.16
Perl	6.41	3.16	2.64
Hoxc9	6.32	0.06	0.34
Meoxl	6.03	1.63	3.26
Nkx6-2	5.41	4.07	4

 Table S2. Transcription factors expressed in specific cell population (Bulk transcriptome of digit joint)

Gene	e13.5 Sox9+	e14.5 Lgr5- (FPKM)	e14.5 Lgr5+	Fold change Lgr5+/Lgr5-	Fold change Lgr5+/Sox9+
Irx2	9.97	5.84	57.21	+9.80	+5.74
Six2	50.34	53.86	277.21	+5.15	+5.51
Six1	16.24	9.49	37.33	+3.93	+2.30
Barx1	55.41	23.81	92.04	+3.87	+1.66
Irx1	15.6	11.76	43.98	+3.74	+2.82
Sox5	21.32	10.02	35.08	+3.50	+1.65
Trps1	15.71	12.68	33.33	+2.63	+2.12
Erg	10.34	9.99	25.21	+2.52	+2.44
Scx	42.52	80.56	194.54	+2.41	+4.58
Sox4	416.76	318.43	701.26	+2.20	+1.68
Cuxl	47.5	33.43	73.42	+2.20	+1.55
Tle4	14.27	11.65	25.58	+2.20	+1.79
Hes1	298.41	284.69	604.86	+2.12	+2.03
Nfatc 1	52.48	39.2	80.79	+2.06	+1.54
Creb5	19.1	25.8	49.93	+1.94	+2.61
Gli2	13	19.73	38.16	+1.93	+2.94
Gli3	22.69	32.62	60.75	+1.86	+2.68
Cited1	10.79	11.28	19.14	+1.70	+1.77
Gabpb1	30.91	29.95	50.67	+1.69	+1.64
Ebf4	7.67	11.14	18.09	+1.62	+2.36

Table S3. Transcription factors expressed in all three populations AND Lgr5+ expressed 1.5-fold higher than the other two populations (Bulk transcriptome of digit joint)

	E14.5 Lgr5-	E14.5 Lgr5+	Fold 		E14.5 Lgr5-	E14.5 Lgr5+	Fold change
Gene	(FP	PKM)	Lgr5-	Gene	(FP	KM)	Lgr5-
IHH/PTHrP pathway	7			Cell surface	markers and rec	eptors	
Glis1	3.54	52.57	$+\infty$	Cd9	105	296.73	+2.83
Hhip	4.94	30.22	$\infty + \infty$	Cd200	6.66	14.9	+2.24
Hhipl1	0.92	6.36	$\infty + \infty$	Cd83	22.25	45.54	+2.05
Pthlh	11	50.11	+4.56	Cd47	22.63	37.49	+1.66
Disp1	19.78	74.28	+3.76	Cd52	8.86	2.87	-00
Cdon	10.93	28.96	+2.65	Cd74	12.69	4.52	-∞
Gli2	19.73	38.16	+1.93	Cd248	46.41	21.55	-2.15
Gli3	32.62	60.75	+1.86	Cd82	10.93	6.35	-1.72
Ihh	45.36	23.54	-1.93	Cdh13	2.97	35.39	$\infty + \infty$
<i>Pth1r</i>	131.09	72.3	-1.81	Itgal 1	16.04	81.87	+5.10
Gli1	26.77	16.03	-1.67	Itea5	29.83	52.72	+1.77
				Iteav	9.39	5.39	+1.37
BMP/TGF B pathway				Itga9	8.32	11.38	+1.05
Chrdl1	3 52	8 64	$+\infty$	Itohl	92.38	97.13	+1.05
Chrdl?	0.85	14 32	$+\infty$	Itoh5	70.15	73.66	-2.02
Inhha	6.76	74.18	+10.97	Cdh3	33.83	16 78	-1.62
Gdf5	71 72	614.83	+8 57	Cdh2	21 30	13.23	-1.76
Tafhi	136.76	863 5	+6.31	Ddr2	15.06	8 55	-1.70
Tafh?	27 44	120.3	+4.38	Itaa6	5 78	5.44	-1.74
1gj02 I thn4	16 72	26.21	+4.38	nguo	5.78	5.44	-1.00
Li0p4 Lthp2	61.22	118 27	+2.17 ±1.02	Enzymos for	FCM dogradati	0 n	
Chud	5.60	110.57	+1.93	LIZYINES IOI	2 22	7.01	1.00
Chra Smadl	22.44	11.43	+2.01	Adamis14	2.32	/.01	± 0
Smaa1	32.44	39.33	+1.65	Addmisi2 Time 2	0.99	14.62	± 2.12
Twsg1	45.51	73.92	+1./1	Timps	49.55	85.52	+1.73
Gajio	86.28	32.02	-2.69	Cisj	24.52	36.84	+1.50
Sostac1	36.56	13.65	-2.68	ChstIO	10.7	6.78	-1.58
Bmp/	52.52	21.81	-2.41	Chst15	/.46	3.1/	-00
Bmper	10.35	5.08	-2.04	Mmp11	124.76	//.61	-1.61
Bmp4	18.37	10.07	-1.82	ECM			
Tgjbr2	33.03	19.82	-1./0	ECM compo	nents	59.02	
WNT nother				Cup	0.75	38.03	
wini patnway	1.20	26.75			1.95	13.82	+∞
Lgro	1.36	26.75	$+\infty$	Inbs4	2.94	12.9	∞^+
Lgr4	28.27	/6.06	+2.69	Spon2	2.12	/.89	$+\infty$
Wnt4	3.52	8.09	$+\infty$	Igfbi	136.76	863.5	+6.31
Wnt9a	3.77	/.1/	+∞	Fmod	24.45	134.56	+5.50
Wnt11	23.2	43.28	+1.87	Col8a1	6.86	22.27	+3.25
Dkk3	37.72	164.84	+4.37	Dcn	35.98	115.28	+3.20
Sfrp2	203.06	450.71	+2.22	Efempl	12.14	34.69	+2.86
Sfrp1	28.5	44.32	+1.56	Col9a2	216.81	479.99	+2.21
Fzd4	20.58	36.73	+1.78	Ltbp4	16.72	36.21	+2.17
Fzd8	21.71	53.42	+2.46	Col9a3	206.32	440.71	+2.14
Wispl	5.11	8.68	+1.70	Col6a1	82.63	167.27	+2.02
Tax1bp3	92.74	151.9	+1.64	Ltbp3	61.32	118.37	+1.93
Tle4	11.65	25.58	+2.20	Vcan	31.28	56.73	+1.81
Ebf2	27.59	8.27	-3.34	Wisp1	5.11	8.68	+1.70
Lefl	36.27	12.73	-2.85	Col9a1	184.09	302.35	+1.64
Sostdc1	36.56	13.65	-2.68	CollIal	104.96	169.86	+1.62
Dkk2	30.03	11.99	-2.50	Col2a1	1400.84	2253.09	+1.61
Nkd2	32.47	14.43	-2.25	Postn	52.87	81.99	+1.55
Nkd1	13.91	6.74	-2.06	Emilin1	159.14	244.21	+1.53
Hicl	78.08	36.77	-2.12	Fbln2	159.5	243.39	+1.53
Nxn	16.76	9.19	-1.82	Prg4	2.36	1.14	not exp. in both
Hmga2	23.57	13.6	-1.73	Comp	15.75	3.9	-∞
Fzd7	19.6	13.08	-1.50	Emilin2	18.61	6.65	-2.80
Gpc2	15.06	10.06	-1.50	Col23a1	25.86	10.96	-2.36
				Matn1	93.79	41.46	-2.26
				Fbn1	26.53	12.34	-2.15
				Col4a1	50.96	28.98	-1.76
				Fbln5	21.76	13.65	-1.59
				Lox	47.87	31.36	-1.53

Table S4. Genes involved in molecular signaling pathways, cell-matrix contact, ECMdegradation and organisation (Bulk transcriptome of E14.5 digit joint)

 $+\infty$ and $-\infty$ indicate genes that are not expressed in Lgr5- and Lgr5+ respectively, fold change is not applicable.

Table S5. Details and parameters for single cell transcriptome sequencing andanalysis. Related to Figures 4 and 5.

Sequencing and quality sheet				
Sequencing and quanty check	50Chr/101hr rain and			
Total road rains	204 million			
Percentage achieving a quality score of Q20	304 million 729/			
(aquivalent to 1 mistake in 1,000 kmc)	/2%0			
(equivalent to 1 mistake in 1,000 bps)	02.00/			
Mapping rate (to mouse reference mm10)	93.0%			
Percentage of valid barcodes	97.7%			
Sequencing saturation	1.2%			
Fraction of reads in cells and beads				
Cells	90.9%			
Background beads	9.1%			
Transcriptome mapping				
Total number of genes detected in aggregate	18,247			
Total number of UMIs detected	46,239,500			
Number of reads (R2) per UMI detected	6.6 (=304million/46million)			
Total number of cells captured and sequenced	5,649			
Average number of gene detected	2,058 (s.d. 637 genes)			
Number of UMIs per cell	6,601			
Blood cell captured	189			
Average number of genes detected in blood cell	216			
Non-blood cells	5,460 (=5649-189)			
Total number of Lgr5+ cell detected	207			
Express normal Lgr5 allele only	94			
Express Lgr5-CRE-ERT2 allele only	87			
Express both alleles	26			
Data analysis				
DEG stringency	• FDR cutoff of <0.05			
DE a buildene j	expression difference at least 20%			
	(defined as % (turned on in cluster i)			
	(defined as /0{turned on in eluster_1}			
Clustering of dispersed gapes (Heatman)	- Top 50/ dispersed gene (generate			
Clustering of dispersed genes (freatinap)	• Top 5% dispersed gene (genome-			
Descriptions to instance	• at least 50 cells expressed			
Pseudotime trajectory	Negative binomial distribution			
	• based on the genome-wide UMI count			
	data for genes expressing in 10 cells			
	or more			

Table S6. DEGs "Cluster 1 vs non-cluster 1" and "Lgr5+ vs Lgr5-" (Single cell transcriptome of E14.5 knee joint). Related to Figure 4

	Genelist A (n=62)
Genelist B (n=27) DEGs (Lgr5+ (cluster 1) vs Lgr5-)	Osr2, Man1a, Nb11, Angptl2, Dusp1, Egfl6, Gas2, Rbms3, Creb5, Rnd3, Zfp503, Dclk1, F2r, Zfp3611, Igfbp4, 3632451006rik, Epb4113, Svil, Tpm1, Palld, Tuba1a, Hmcn1, Cdh11, Hspa1a, Pdgfra, Cpq, Epha7, Foxp1, Loxl2, Csrp1, Dbn1, Col6a1, Col8a2, Vcan, Ddah2, Col6a2, Cdon, Prrx1, Tagln2, Sulf2, Col5a2, Tax1bp3, Zyx, Nrep, Ltbp3, Nfia, Rcan1, Fb1n1, Igfbp7, Arhgap31, Marcks11, Kctd12, Hoxa10, Abracl, Marcks, Hmgb3, Timp2, Map11c3a, Ifitm3, Aes, Trps1, Barx1
	Genelist B (n=27)
Genelist C (n=48) 62	F2rl1, Fhod3, Adam33, Tagln, Fst, Khdrbs2, Hspb8, Chst5, Inhba, Tnni2, Etl4, Meox2 , Sorcs2, Tlr2, Bmp4, Gfra4, Syt11, Mafb , Ppm11, Pdgfrl, Gpm6b, Fjx1, Tmem119, Pcdh18, Enpp1, Rgcc, Pdgfc
Genelist A (n=62)	Genelist C (n=48)
DEGs (Cluster 1 vs non-cluster 1)	Lrrc17, Pamr1, Htra1, Cdh13, Phactr1, Aqp1, Tm4sf1, Nupr1, Tcea3, Lrrn1, Sfrp2, Ccdc80, Cntfr, Tgfbi, Ssc5d, Gas1, Pax9, Mkx, Pid1, Map1b, Gdf5, Dkk3, Sulf1, Gm26771, Dusp8, Mylk, Shisa2, Adamts11, Prss23, Krt17, Cited2, Wisp1, Loxl1, Sox4, Lmo1, Ackr3, Bcl11a, Scx, Phldb2, Myo1b, Spint2, Fbn2, Postn, Mfap4, Col12a1, Tgfb2, Fn1, Ier3

Bold = transcription factors

Table S7. Primers used in this paper

	Forward primer	Reverse primer							
Primers for get	Primers for genotyping Lgr5-eGFP-IRES-CreER ^{T2} mice								
wildtype mutant	5'-CTGCTCTCTGCTCCCAGTCT-3'	5'-ATACCCCATCCCTTTTGAGC-3' 5'-GAACTTCAGGGTCAGCTTGC-3'							
Primers for get	Primers for genotyping R26R-lacZ mice								
wildtype mutant	5'-AAAGTCGCTCTGAGTTGTTAT-3'	5'-GGAGCGGGAGAAATGGATATG-3' 5'-GCGAAGAGTTTGTCCTCAACC-3'							
Primers for Qu	Primers for Quantitative PCR								
I out	5' CACAACCTCTCTCATCTCCAT 2'	5' TOCTOOTTATTTTTOTOCOTOT 2'							
Lgr4 Lar5	5'-CACAGCCACTGCGGCGACTT_3'	5'-CAATGGGCGTCTGCCGGGTC-3'							
Lgi 5 Larh	5'-CTCCTTGCTGGACAAGCTGA-3'	5'-TCTCAAAGAGGTGCTCGCAG-3'							
Gdf5	5'-AAAGGGAGGTAACAGCAGCG-3'	5'-CCCATCCTTCTCCAAGGCAC-3'							
Cd90	5'-TCTCCTGCTCTCAGTCTTGC-3'	5'-TATTCTCATGGCGGCAGTCC-3'							
Cd105	5'-TAGCACCTTGTCCCAGGAAG-3'	5'-CAGTACAGAGGGCAGGACAA-3'							
Notch I	5'-TGTGAGTCCAACCCTTGTGT-3'	5'-CGTTGATGTTGGTCTGGCAA-3'							
Klf4	5'-AAAAGAACAGCCACCCACACTT-3'	5'-CCCAGTCACAGTGGTAAGGTTTC-3'							
Gapdh	5'-TGCACCACCAACTGCTTAG-3'	5'-GATGCAGGGATGATGTTC-3'							