

Identification of a CDI system that reduces biofilm formation and host cell adhesion of *Acinetobacter baumannii* DSM30011 strain

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Supplementary data

Supplementary Figure legends

Supplementary Figure S1. Effect of CdiA₂-CT production on *E. coli* cell viability and filamentation. (A) The cell viability of *E. coli* DH5α strain producing CdiA₂-CT with or without CdiI₂ was estimated in colony forming unit (CFU) /ml. Means from three biological replicates are plotted. (B) Cell length repartition of *E. coli* DH5α (*recA*⁺) and RecA-GFP producing MG1655 (*recA*⁺) 3 h after induction of CdiA₂-CT toxin. n indicate the number of analysed cells.

Supplementary Figure S2. The CdiA₁-CT producing *E. coli* strain are viable cells. (A) HU-mCherry producing *E. coli* MG1655 growth monitored by measuring the optical density at 600 nm (dotted lines) and colony forming unit (CFU) /ml (bars) after induction of CdiA₁-CT in presence or in absence of CdiI₁. Means from three biological replicates are plotted. (B) Live and dead assay performed with RecA-GFP *E. coli* MG1655 strain 3 h after induction of CdiA₁-CT in presence or absence of CdiI₁. Before staining, the bacteria were treated with or without (w/o) ethanol. Left panel: representative fluorescence microscopy of CdiA₁-CT producing cells

after staining with SYTO9 and Propidium iodide (PI). Scale bar corresponds to 1 μ m; right panel: percentage of dead cells. n indicate the number of analysed cells.

Supplementary Figure S3. Growth analysis of *A. baumannii* DSM30011 wild-type and mutant strains. The growth was monitored in LB medium by measurement of optical density at 600 nm (OD₆₀₀) (solid lines) and colony forming unit (CFU) /ml (bars).

Supplementary Figure S4. Confocal microscopy images correspond to 3D projections of the maximum intensities obtained from z-stacks of biofilms formed by the wild-type and $\Delta cdiBAI_1$ strains on glass-bottom slides at 3, 5 and 24h. Bacteria were labelled with DAPI. Scale bar 10 μ m.

Supplementary Figure S5. Z-stack analysis of *A. baumannii* adhesion to lung epithelial cells. Confocal microscopy of A549 cells infected with *A. baumannii* DSM30011 wild-type and $\Delta cdiBAI_1$ strains during 2 hours. The corresponding axis are shown. Bacteria were labeled with an anti-*Acinetobacter* antibody (green), the actin cytoskeleton with phalloidin (grey) and nuclei with DAPI (blue).

Supplementary Figure S6. The CdiA₁^{AbSDF} is produced in *A. baumannii* SDF strain. (A) Schematic of the *cdi*₁^{AbSDF} and *cdi*₂^{AbSDF} loci. Genes encoding putative CdiB transporters, CdiA exoproteins and CdiI immunity proteins are colored respectively in brown, green and orange. *cdiC* genes shown in red encode proteins that share high homologies with HlyC acyltransferases that activate the α -hemolysin HlyA in *E. coli* through fatty acylation using acyl-acyl carrier protein (acyl-ACP) as the fatty acid donor (Ogier et al., 2016). *cdi*₁^{AbSDF} locus refers to ABSDF_RS16855, ABSDF_RS16850, ABSDF_RS16845, ABSDF_RS16840 genes and *cdi*₂^{AbSDF} to ABSDF_RS16500, ABSDF_RS16505, ABSDF_RS16510*, ABSDF_RS16515. * indicates the truncated version of the annotated gene on the NCBI database; the 3'-end of the gene was sequenced and the result is shown in Supplementary Figure S7. (B) Cell extract (CE) of wild-type strain grown overnight was analyzed by blue Coomassie staining. Arrow indicates

the CdiA₁^{AbSDF} protein identified by mass spectrometry. Molecular weight marker (kDa) is indicated on the left.

Supplementary Figure S7. Sequence of complete *ABSDF_RS16510* gene in *A. baumannii* SDF strain. Bold sequence shows the annotated gene on the NCBI database, italic the intergenic sequence and the start codon of the potential *cdiI₂^{AbSDF}* (*ABSDF_RS16515*) immunity gene is underlined.

Movie 1. Time-lapse fluorescence microscopy of the recombinant HU-mCherry protein produced by *E. coli* MG1655 strain after induction of the CdiA₂-CT.

Movie 2. Time-lapse fluorescence microscopy of the recombinant HU-mCherry protein produced by *E. coli* MG1655 strain after induction of the CdiA₂-CT in presence of its cognate CdiI₂ immunity protein.

Movie 3. Time-lapse fluorescence microscopy of the recombinant RecA-GFP protein produced by *E. coli* MG1655 strain after induction of the CdiA₂-CT.

Movie 4. Time-lapse fluorescence microscopy of the recombinant RecA-GFP protein produced by *E. coli* MG1655 strain after induction of the CdiA₂-CT in presence of its cognate CdiI₂ immunity protein.

Movie 5. Time-lapse fluorescence microscopy of the recombinant HU-mCherry protein produced by *E. coli* MG1655 strain after induction of the CdiA₁-CT.

Movie 6. Time-lapse fluorescence microscopy of the recombinant RecA-GFP protein produced by *E. coli* MG1655 strain after induction of the CdiA₁-CT.

Movie 7. Time-lapse fluorescence microscopy of the recombinant HU-mCherry protein produced by *E. coli* MG1655 strain after induction of the CdiA₁-CT in presence of its cognate CdiI₁ immunity protein.

Movie 8. Time-lapse fluorescence microscopy of the recombinant RecA-GFP protein produced by *E. coli* MG1655 strain after induction of the CdiA₁-CT in presence of its cognate CdiI₁ immunity protein.

Supplementary Table S1. Plasmids and strains used in this study

Plasmids	Description	Source
pKD3	Carries <i>FRT-Cm-FRT</i> used for λ red integration ; Cm ^R	(Datsenko and Wanner, 2000)
pKD3-AS- <i>recA-Cm</i>	Carries <i>recA</i> gene under its own promoter used for LY653 construction; Cm ^R	This study
pKD4	Carries <i>FRT-kan-FRT</i> used for RecET integration; Kn ^R	(Datsenko and Wanner, 2000)
pCP20	Site-specific excision vector, Flp recombinase expression; Ap ^R	(Datsenko and Wanner, 2000)
pAT02	Expresses the Rec _{Ab} recombinase; Ap ^R	(Tucker et al., 2014)
pFLP2	Site-specific excision vector, Flp recombinase expression; Ap ^R	(Hoang et al., 2000)
pMHL2-2	Apramycin cassette, Apra ^R	(Godeux et al., 2018)
pUA66	<i>gfpmut2</i> gene under the control of <i>rrnB</i> promoter; Kn ^R	(Zaslaver et al., 2006)
pWH1266	Expression plasmid, <i>E. coli-Acinetobacter</i> shuttle plasmid; Ap ^R	(Hunger et al., 1990)
pWH1266-Pempty-gfp	pWH1266 containing promoterless <i>gfp</i> gene; Ap ^R	This study
pWH1266-P <i>cdiB1</i> -gfp	pWH1266 containing 500 bp upstream <i>cdiB1</i> gene from <i>A. baumannii</i> DSM30011 fused to <i>gfp</i> gene; Ap ^R	This study
pWH1266-P <i>cdiA1</i> -gfp	pWH1266 containing 500 bp upstream <i>cdiA1</i> gene from <i>A. baumannii</i> DSM30011 fused to <i>gfp</i> gene; Ap ^R	This study
pWH1266-P <i>cdiI1</i> -gfp	pWH1266 containing 500 bp upstream <i>cdiI1</i> gene from <i>A. baumannii</i> DSM30011 fused to <i>gfp</i> gene; Ap ^R	This study
pWH1266-P <i>cdiB2</i> -gfp	pWH1266 containing 500 bp upstream <i>cdiB2</i> gene from <i>A. baumannii</i> DSM30011 fused to <i>gfp</i> gene; Ap ^R	This study
pWH1266-P <i>cdiA2</i> -gfp	pWH1266 containing 500 bp upstream <i>cdiA2</i> gene from <i>A. baumannii</i> DSM30011 fused to <i>gfp</i> gene; Ap ^R	This study
pWH1266-P <i>cdiI2</i> -gfp	pWH1266 containing 500 bp upstream <i>cdiI2</i> gene from <i>A. baumannii</i> DSM30011 fused to <i>gfp</i> gene; Ap ^R	This study
pUC18T-mini-Tn7T-Gm	mini-Tn7T based vector; mobilizable; Ap ^R Gm ^R	(Choi and Schweizer, 2006)
pUC18T-mini-Tn7T-Ap	Gm ^R removed from pUC18T-mini-Tn7T-Gm by <i>Xba</i> I digestion and self-ligation; Ap ^R	This study
pUC18T-Δ <i>cdiA1</i>	2 kb with homology to the flanking regions of <i>cdiA1</i>	This study
pUC18T-Δ <i>cdiA1</i> -Apra	Apramycin cassette flanked by 2kb upstream and downstream of <i>cdiA1</i> ; Ap ^R , Apra ^R	This study
pUC18T-Δ <i>cdiB1</i>	2 kb with homology to the flanking regions of <i>cdiB1</i>	This study
pUC18T-Δ <i>cdiB1</i> -Apra	Apramycin cassette flanked by 2kb upstream and downstream of <i>cdiB1</i> ; Ap ^R , Apra ^R	This study
pBAD33	Arabinose inducible expression plasmid, Cm ^R	(Guzman et al., 1995)
pBAD33-CdiA ₁ -CT	pBAD33 containing <i>cdiA-CT</i> ₁ ^{Ab30011} toxin (residues Val ⁴⁷⁴⁶ - Arg ⁵¹⁰⁴ of CdiA ₁ ^{Ab30011}); Cm ^R	This study
pBAD33-CdiA ₂ -CT	Expresses <i>cdiA-CT</i> ₂ ^{Ab30011} toxin (residues Ala ¹⁷¹⁵ - Tyr ¹⁹⁵⁰ of CdiA ₂ ^{Ab30011}); Cm ^R	This study
pTrc99a	IPTG-inducible expression plasmid; Ap ^R	GE healthcare
pTrc99a-CdiI ₁ -His ₆	IPTG-inducible expression of CdiI ₁ ^{Ab30011} -His ₆ immunity gene; Ap ^R	This study

pTrc99a-CdiI ₂ -His ₆	IPTG-inducible expression of CdiI ₂ ^{Ab30011} -His ₆ immunity gene; Ap ^R	This study
Strains	Description	Source
<i>E. coli</i>		
Top10	General cloning <i>F-</i> <i>mcrA</i> Δ(<i>mrr-hsdRMS-mcrBC</i>) φ80lacZΔM15 Δ <i>lacX74</i> <i>nupG recA1 araD139</i> Δ(<i>ara-leu</i>)7697 <i>gatE15 galK16 rpsL</i> (<i>Str^R</i>) <i>endA1 λ</i>	Invitrogen
DH5α	General cloning and toxicity assay <i>F-</i> <i>endA1 glnV44 thi-1 recA1 relA1 gyrA96 deoR nupG purB20</i> φ80dlacZΔM15 Δ(<i>lacZYA-argF</i>)U169, <i>hsdR17(rK⁻mK⁺)</i> , <i>λ</i>	Lab collection
BW25141	Δ(<i>araD-araB</i>)567, Δ <i>lacZ4787</i> (::rrnB-3), Δ(<i>phoB-phoR</i>)580, <i>λ</i> , <i>galU95</i> , Δ <i>uidA3::pir⁺</i> , <i>recA1 endA9</i> (del-ins)::FRT, <i>rph-1</i> , Δ(<i>rhaD-rhaB</i>)568, <i>hsdR514</i>	Lab collection
DY330	W3110 Δ <i>lacU169</i> , <i>gal490</i> , λ <i>cI857</i> , Δ(<i>cro-bioA</i>)	(Yu et al., 2000)
MG1655	λ, <i>rph-1</i>	Coli Genetic Stock Center (CGSC) #6300
MS388	MG1655 <i>rpsL</i> ⁺ , <i>Str^R</i>	Gift from F. Cornet
OX468	W1485 <i>F-</i> <i>leu thyA thi deoB</i> or <i>C supE rpsL, hupA-mcherry-FRT-kn-FRT, Str^R, Kn^R</i>	Gift from F. Cornet
LY119	MS388 <i>hupA-mcherry-FRT-kn-FRT, Str^R, Kn^R</i>	This study; MS388 x P1.OX468
LY248	MS388 <i>hupA-mcherry, Str^R</i>	This study; derivative of LY119, <i>kn</i> removed via pCP20
LY653	DY330 <i>fhuB::recA-FRT-Cm-FRT</i>	This study; λred <i>recA</i> at the <i>fhuB</i> chromosomal locus
SS3085	<i>recA4155-gfp-Kn(4155,4136)</i> , <i>Kn^R</i>	(Renzette et al., 2005)
MS388 <i>recA4155-gfp-Kn</i>	MS388 <i>recA4155-gfp-Kn(4155,4136)</i> , <i>Kn^R</i>	This study; MS388 x P1.SS3085
LY769	MS388 <i>recA4155-gfp-Kn, fhuB::recA-FRT-Cm-FRT, Str^R, Kn^R, Cm^R</i>	This study; MS388 <i>recA4155-gfp-Kn</i> x P1.LY653
LY844	MS388 <i>recA4155-gfp-Kn, fhuB::recA-FRT, Str^R, Kn^R</i>	This study; derivative of LY769, <i>Cm</i> removed via pCP20
<i>A. baumannii</i>		
DSM30011	Wild-type	(Repizo et al., 2017)
Δ <i>cdiA1</i>	DSM30011 Δ <i>cdiA1::apra</i> , <i>Apra^R</i>	This study
Δ <i>cdiB1</i>	DSM30011 Δ <i>cdiB1::apra</i> , <i>Apra^R</i>	This study
Δ <i>cdiBA1::kn</i>	DSM30011 Δ <i>cdiA1::FRT-kn-FRT, Kn^R</i>	This study
Δ <i>cdiBA1</i>	DSM30011 Δ <i>cdiA1::FRT</i>	This study; derivative of Δ <i>cdiBA1::kn</i> , <i>kn</i> removed by pFLP2

Ap^R, Cm^R, Kn^R, Str^R, Apra^R, Gm^R resistance to ampicillin, chloramphenicol, kanamycin, streptomycin, apramycin and gentamycin respectively.

Supplementary Table S2. Primers used for plasmid and strain constructions

Name	sequence	construct
Ol359	GGAATAGGAACTAAGGAGGAGAATGCTTCAGCGGCAC	pKD3-AS- <i>recA-Cm</i> construct using Gibson Assembly; PCR on MS388 (Ol359/Ol360) and pKD3 (Ol361/Ol362)
Ol360	AGCCATGGTCATATGAATAACGCAGATGCGACCCTGTG	
Ol361	TCCTCCTTAGTTCCTATTCC	
Ol362	TATTCATATGGACCATGGCT	
P363	CGCCAGCGTCGCATCGGGCATCCGTTGTCGCGTAAACGCTCTTGAGC GATTGTGTAGGC	LY653 strain construct using λred; PCR on pKD3- <i>recA-FRT-Cm-FRT</i>
P364	TTCCATATAACGGCCCTGTACGCCCTGGACGGATAAACGCAGATGCG ACCCTTGTG	pBAD33-CdiA ₁ -CT plasmid construct using SLIC; PCR on DSM30011
Big3	GGGCTAGCGAATTGAGCTCAGGAGGAATTCACCATGGTTGAGAAC AATTATTTGG	pBAD33-CdiA ₂ -CT plasmid construct using SLIC; PCR on DSM30011.
Big4	TTGCATGCCCTGCAGGTGCACTTAACGACGTTTAATGGG	
Big5	GGGCTAGCGAATTGAGCTCAGGAGGAATTCACCATGGCAATCCAG AATAACTCTT	pBAD33-CdiA ₂ -CT plasmid construct using SLIC; PCR on DSM30011.
Big6	TTGCATGCCCTGCAGGTGACCTAATATGGGTGGAGTGTCT	
Big7	ATTCACACAGGAAACAGACCATGATAGATGTTAGCCCAGA	pTrc99a-CdiI ₁ -His ₆ plasmid construct using SLIC; PCR on DSM30011
Big8	GGTCGACTCTAGAGGATCCTAGTGGTGGTGGTGGTGGTGA CCTTATTTG	pTrc99a-CdiI ₂ -His ₆ plasmid construct using SLIC; PCR on DSM30011
Big9	ATTCACACAGGAAACAGACCATGAAGTTAACGCCCTAT	
Big10	GGTCGACTCTAGAGGATCCTAGTGGTGGTGGTGGTGGTGA CTTCCTTAAACGAGTTAA	pUC18T-Δ <i>cdiA1</i> plasmid construct using SLIC. PCR on DSM30011
Big44	AATTCGATCATGCATGAGCTCCGGGTGCTAGCTGATTTTATAA	
Big45	CCGCGGTAGTCCATTGACCATGGCATTCTAAAATCTCACTAAA	Overlapping PCR on DSM30011 (Big44/Big45 and Big46/Big47)
Big46	CCATGGTCAATGGACTACCGCGCGTCGTTAAGGAAAATAGGT	
Big47	TTCCTGCAGCCCCGGGGATCCTGTATAGACCAATTATCAG	
Big84	TGAGATTTAGAATGCCATGGAATCAAGGCCGATCCTTGG	pUC18T-Δ <i>cdiA1</i> -Apra plasmid construct using ligation
Big85	TTTCCTTAACGACGCCCGCGGCCAGGGTTTCCCAGTCA	pUC18T-Δ <i>cdiB1</i> plasmid construct using SLIC; Overlapping PCR on DSM30011 (Big88/Big89 and Big90/Big91). Δ <i>cdiB1</i> strain construct; PCR (Big88/Big91) on pUC18T-Δ <i>cdiB1</i> -Apra plasmid
Big88	AATTCGATCATGCATGAGCTCCGGGCCCTCGAATTATGAATCGA	
Big89	CCGCGGTAGTCCATTGACCATGGCATATAAAATTAAACCGTGA	Δ <i>cdiA1</i> strain construct. PCR on pUC18T-Δ <i>cdiA1</i> -Apra plasmid
Big90	CCATGGTCAATGGACTACCGCGGTAATTCAAGTTAAAGGTCC	
Big91	TTCCTGCAGCCCCGGGGATCCTGAATACACAACACCTGACT	Δ <i>cdiBA1::kn</i> strain construct using Rec _{Ab} ; PCR on pKD4
Big11	CGGACTATAGACTATACTAGTAAGTGCCTTTAAAGTGGGT	
Big14	CCCCTGCAGGTGACGGATCCATAATTCAAGGCATATGAG	Δ <i>cdiBA1::kn</i> strain construct using Rec _{Ab} ; PCR on pKD4
Big229	TTTTACATACAGACAAATCTGTATAAAAAATGCTTCCACTTGTAA GGTTTGTAAAATAAAATTAAAGTTGATATTACCGGTTAATT ATatgTGTAGGCTGGAGCTGCTCG	
Big230	TAAATAATGAAAAGGTATCATTATTCCAAAATCATTATTATAAT TTTATTCCAGTTATTATGTAAAAAAAGGTAGAAATTCTGAGTAA CAATTACCTCCTAGTTCCTATTCCG	

Supplementary Reference

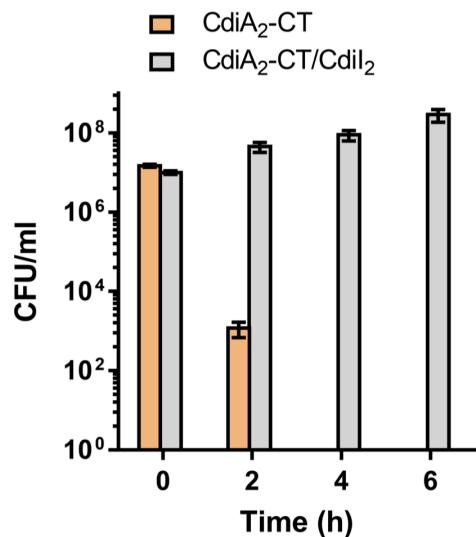
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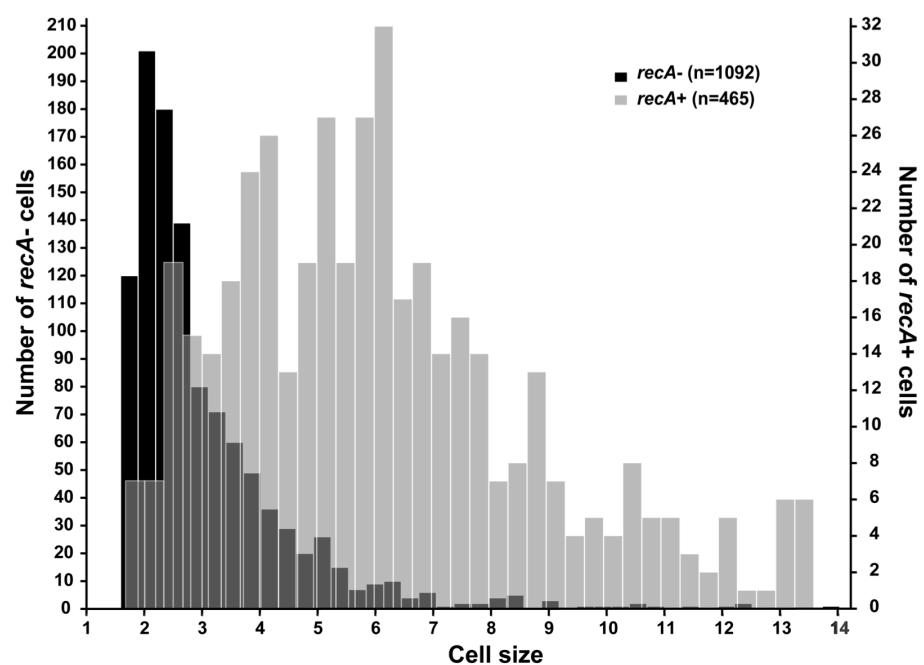
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Supplementary Figure S1

A.

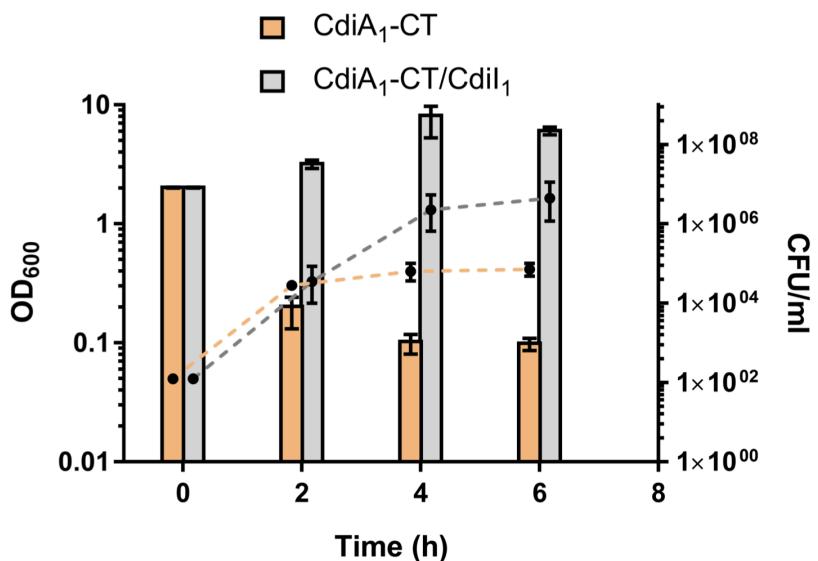


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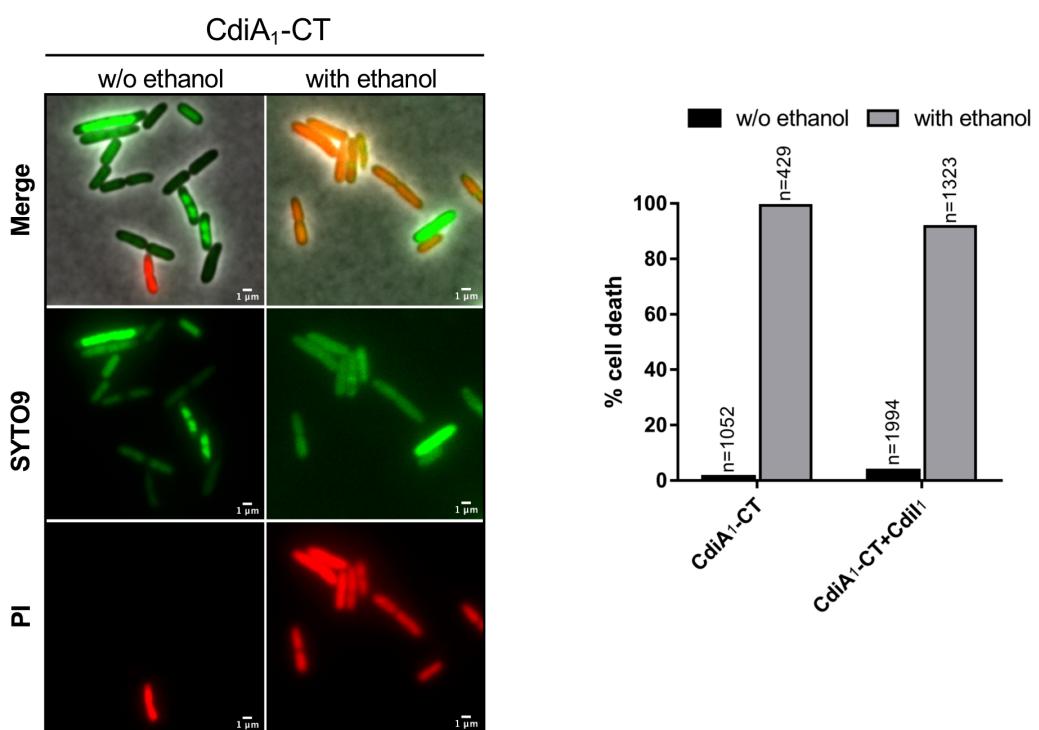


Supplementary Figure S2

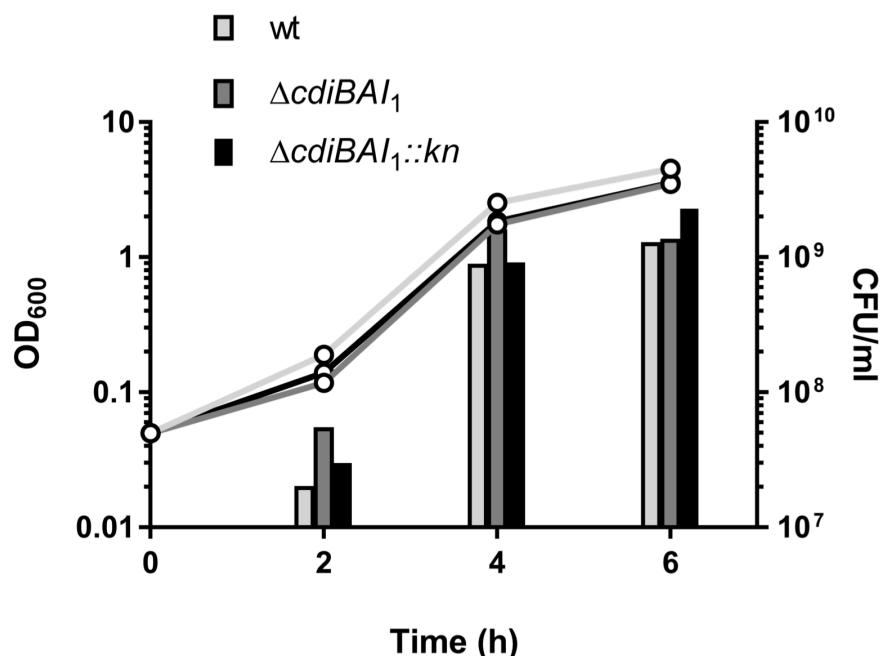
A.



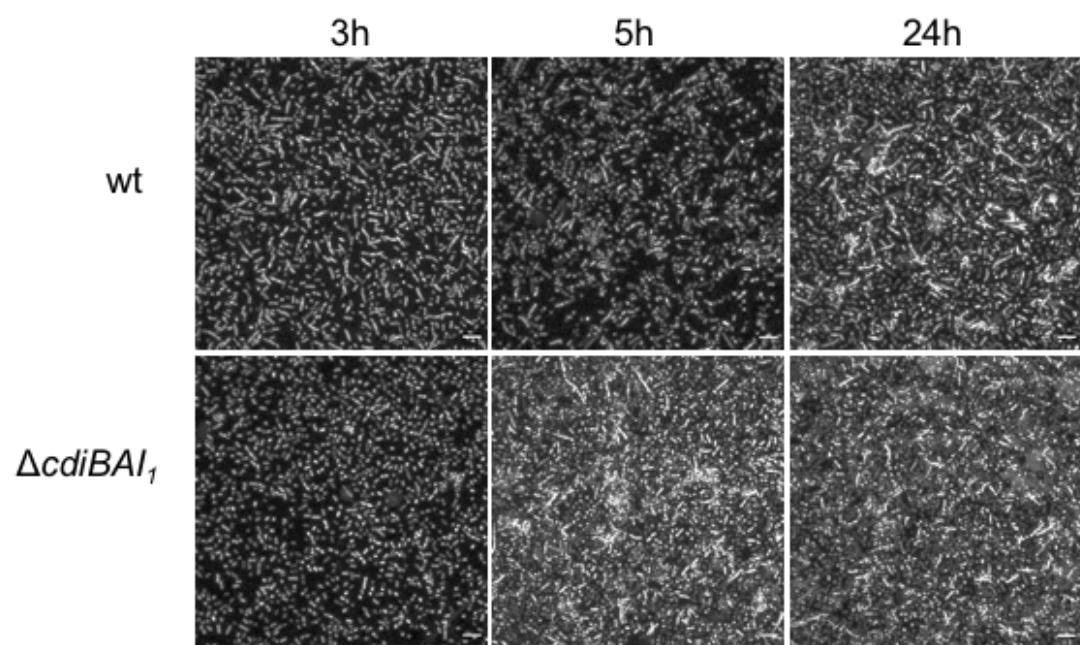
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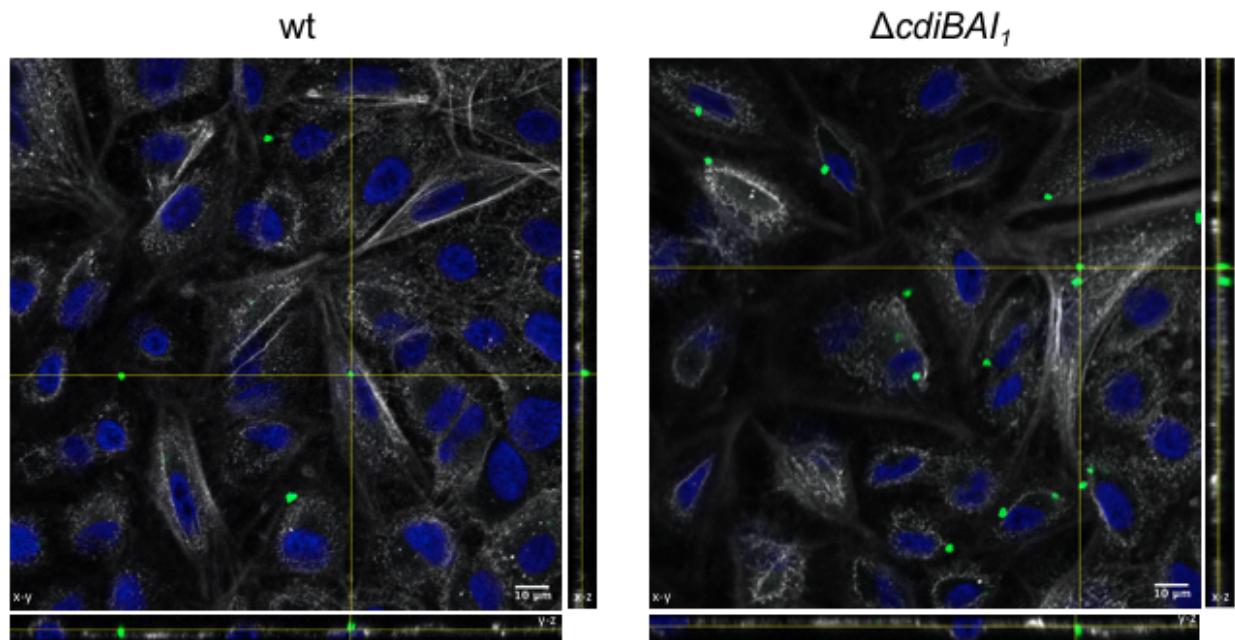
Supplementary Figure S3



Supplementary Figure S4

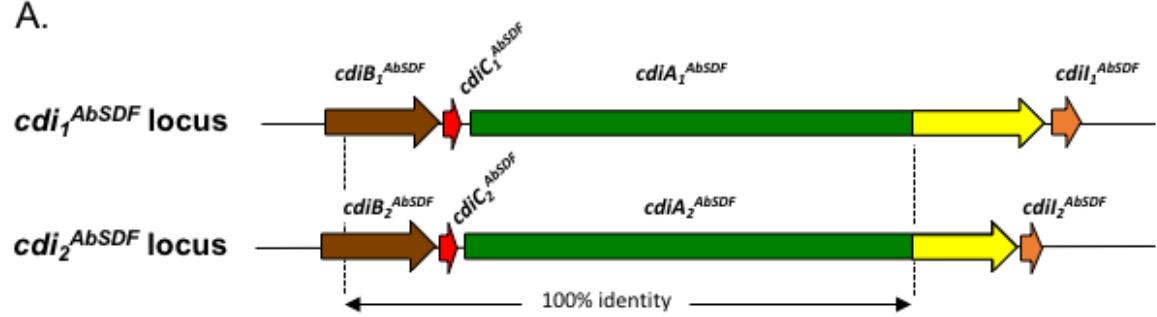


Supplementary Figure S5

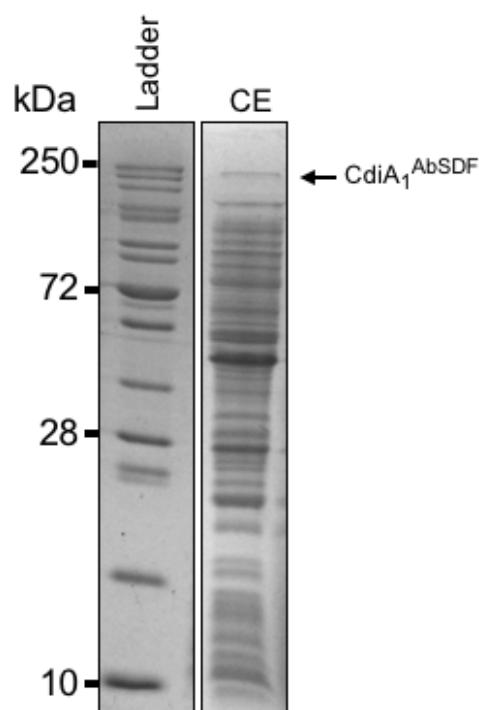


Supplementary Figure S6

A.



B.



Supplementary Figure S7

cdiA2AbSDF (*ABSDF_RS16510*) gene region

AGCATCGTGAATGGAAAAAATTCACAAACCTGATTGCGTGTGAAGCCAATCCGACCACAGACATAAGCTGAAC
AAAACAATATTGATCGCGTTGAGGCCAGAATTAAACCAAGTGAATAACACTGCTTCAGCAGTCGATTGCGACCGATATTACAGTTCGTAC
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