Supplementary Information

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Supplementary Movie Legends

Movie S1. Swarming assays in which *P. aeruginosa* is spotted (A) without phage or (B) with phage. (C) Pairwise swarming assay in which *P. aeruginosa* is spotted (left) without phage or (right) with phage. Time-lapse images are acquired over the course of 23 hours at 37 $^{\circ}$ C

Movie S2. Six-way swarming assays in which *P. aeruginosa* is spotted at the center. The same strain is spotted (A) without phage or (B) with phage at the satellite positions. Time-lapse images are acquired over the course of 17 hours at 37 °C.

Movie S3. Swarming assays of *P. aeruginosa* in the center and (A) rhamnolipids, (B) PQS, and (C) HHQ at the satellite positions for concentrations indicated in millimolar units. Time-lapse images are acquired over the course of 17 hours at 37 °C.

Movie S4. Swarming assays of uninfected wild-type *P. aeruginosa* in the center. The $\Delta rhlAB$ strain is spotted (A) without phage or (B) with phage at the satellite positions. The $\Delta rhlAB$ $\Delta pqsA$ strain is spotted (C) without phage or (D) with phage at the satellite positions. Time-lapse images are acquired over the course of 17 hours at 37 °C.

Movie S5. Swarming assays of wild-type *P. aeruginosa* in the center and the (A) wild-type, (B) $\Delta rhlAB$, or (C) $\Delta rhlAB \Delta pqsA$ strain mixed with gentamycin and spotted at satellite positions. Time-lapse images are acquired over the course of 18 hours at 37 °C.





Center:WTSatellites: $\Delta CRISPR \Delta cas$

Figure S1. Swarming of wild-type *P. aeruginosa* at the center and (A) phage only at satellite positions, imaged after 16 hours of growth or (B) the $\Delta rhlAB$ strain at the satellite positions, imaged after 14 hours of growth. The experiment in (B) is identical to that in Fig. 3B except that image was acquired at an earlier time point. Swarming assays of wild-type *P. aeruginosa* at the center and (C) $\Delta lasR$, (D) $\Delta rhlR$, and (E) $\Delta lasR \Delta rhlR$ spotted at the satellite positions (left images) without phage or (right images) with phage. (F) Swarming assays of wild-type *P. aeruginosa* at the satellite positions.



Satellites: $\Delta pqsA$

WT $\Delta pqsA + phage$



Center: WT **Satellites:** $\Delta rhlAB$ filtered sup.

WT $\Delta rhlAB$ sonicated & filtered sup.





Center: WT **Satellites:** $\Delta pqsH + Kan$



WТ $\Delta rhlAB \Delta pqsH + Kan$

> Figure S3. Swarming assay of wild-type P. aeruginosa at the center and 6 µL of (A) 25 mg / mL kanamycin or (B) 40 mg / mL fosfomycin spotted alone or with wild-type, $\Delta rhlAB$, $\Delta pqsA$, or $\Delta rhlAB \Delta pqsA$ *P. aeruginosa* at the satellite positions on antibiotic-free plates. (C) Swarming assay with 6 μ L of 25 mg / mL kanamycin spotted with the $\Delta pqsH$ or $\Delta rhlAB \Delta pqsH$ strains at the satellite positions and (D) the corresponding repulsion radii at the satellite positions. (E) Swarming assay and (F) corresponding repulsion radii in which 6 μ L of the initial culture of $\Delta rhlAB$ is spotted with gentamycin (concentrations in $\mu g / mL$) at the satellite positions. White dots on the satellite positions indicate the precise centers of positions where the cultures or antibiotics were spotted. The dashed lines indicate the boundaries of the initial spots. Bars are the average of at least 6 positions and error bars indicate standard deviation.

Center:

WT

Satellites: $\Delta rhlAB$ + Gent



Center: PA14 Satellites: MPAO1

PA14 MPAO1 + Kan



Center: PA14 Satellites: P2m



Figure S4. Swarming assay of wild-type *P. aeruginosa* PA14 at the center and 6 μ L at the satellite positions on antibiotic-free plates of (A) *P. aeruginosa* MPAO1 alone or with 25 mg / mL of kanamycin or (B) *P. aeruginosa* P2m. (C) Swarming assay with carbenicillin, cefsulodin, ciprofloxacin, or tetracycline (concentrations in mg / mL) spotted alone or with wild-type *P. aeruginosa* PA14 at the satellite positions on antibiotic-free plates. White dots at the satellite positions indicate the precise centers of positions where the cultures or antibiotics were spotted. The dashed lines indicate the boundaries of the initial spots.



Figure S5. (A) LC-MS analysis of agar extracts for PA14 $\Delta rhlAB$ or PA14 $\Delta rhlAB \Delta pqsA$ in which cultures are mixed with 40 mg / mL of fosfomycin. 6 µL of the mixture is spotted onto antibiotic-free swarming medium, incubated for 16-18 hours, and the area surrounding the colony is extracted. Pure 10 µM PQS, which is not spotted onto agar, is provided as a reference. (B) LC-MS analysis of agar extracts surrounding the LESB58 or PA14 $\Delta rhlAB \Delta pqsA$ strains. (C) Schematic indicating the arc (dashed line) along which the radius of repulsion is measured (yellow). The blue dot indicates the center of the satellite colony.

Supplementary Tables

SI Strams an	Jusinius		
Strain or plasmid	Lab strain name	Genotype	Reference / source
Strains			
WT	AFS27E.1	PA14 attTn7::[PA1/04/03-mCherry] aacC1::FRT	This study
ΔCRISPR Δcas	NMHK326	PA14 \triangle CRISPR \triangle cas	(1)
AFS27E	AFS27E	PA14 attTn7::[PA1/04/03-mCherry] aacC1 ⁺	(2)
$\Delta lasR$	AFS20.1	PA14 $\Delta(lasR)$::FRT	(3)
rhlR	SM32	PA14 $\Delta rhlR$	(4)
$\Delta lasR \ rhlR$	AFS49H	PA14 Δ(lasR)::FRT rhlR::MAR2xT7	(3)
$\Delta rhlAB$	BR04.1	PA14 Δ (<i>rhlAB</i>)::FRT	This study
$\Delta pqsA$	AFS79.1	AFS27E.1 $\Delta(pqsA)$::FRT	This study
$\Delta rhlAB \Delta pqsA$	AFS82.1	PA14 Δ (<i>rhlAB</i>)::FRT Δ (<i>pqsA</i>)::FRT	This study
pqsH	AFS77	AFS27E.1 pqsH::Mar2xT7	This study
$\Delta rhlAB pqsH$	BR07	PA14 Δ(<i>rhlAB</i>)::FRT <i>pqsH</i> ::Mar2xT7	This study
LESB58		Hyper-virulent P. aeruginosa clinical isolate	(5)
P2m		Mucoid <i>P. aeruginosa</i> cystic fibrosis isolate PAmFLR02	(6)
MPAO1		P. aeruginosa strain MPAO1	(7), from Gitai lab
Plasmids			
pFLP2		Plasmid expressing Flp to recombine FRT sites	(8)
pAS03		Plasmid containing FRT-aacC1-FRT template	(9)
pUCP18-RedS		λ Red recombineering vector	(10)
Phage			
DMS3vir	DMS3vir		(11)

Table S2. Primers

Primer name	Sequence
Lambda red primers	
rhlA-lred-u1	CCCTCGAGTTCTCCAATACC
rhlA-lred-l1	CATCTCACACCTCCCAAAAATTTTC
rhlA-lred-u2	GCTGTTTGCCTGTTCGAAAATTTTTGGGAGGTGTGAGATGattccggggatccgtcgacc
rhlB-lred-l2	TCTGTTATGCCAGCACCGTTCAGGACGCAGCCTTCAGCCAtgtaggctggagctgcttcg
rhlB-lred-u3	TGGCTGAAGGCTGCGTCCTGA
rhlB-lred-13	TCGAAGCTGGAGATGTTCTG
pqsA-lred-u1	CGACCGGCGATTCCATTTTC
pqsA-lred-l1	CATGACAGAACGTTCCCTCT
pqsA-lred-u2	ATCCGGATGCATATCGCTGAAGAGGGAACGTTCTGTCATGattccggggatccgtcgacc
pqsA-lred-12	TTCACCCCCACAGCCTGAATCAACATGCCCGTTCCTCCGGtgtaggctggagctgcttcg
pqsA-lred-u3	CCGGAGGAACGGGCATGTTG
pqsA-lred-13	GTTGAGGTGTCCCTTGACGT
pqsH-u1	CCTGGATGATCGTCGTGGGC
pqsH-11	CCAACTCCTCGAGGTCGTTGT
qPCR primers	
pqsA_qPCR_u2	AGGCGGTTCTGGTTCCTAC
pqsA_qPCR_l2	GCATGTGCGAGGGAATCTG
pqsB_qPCR_u1	CTCTACGTGGTCGATACCCTCG
pqsB_qPCR_11	CAGACTCGCTGTCCACTTCCAA
pqsH_qPCR_u1	TTCCAGTCTGGGTGGGAGTTC
pqsH_qPCR_11	TCGAGGATCTGCACGATGGAG
rhlA_qPCR_u1	GCGCTTCGAGGTCAATCAC
rhlA_qPCR_11	AGCATCGCCTGGTTCAGTC
rhlB_qPCR_u1	CGGCGACGTATTTCCCTTCATC
rhlB_qPCR_11	TAGGTCAGTTCGTCGCTCAG
5S_qPCR_u1	GAACCACCTGATCCCTTCCC
5S gPCR 11	TAGGAGCTTGACGATGACCT

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