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Supplementary Materials for

Low-complexity domain of U1-70K modulates phase separation and aggregation through distinctive basic-acidic motifs

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Predictions of disordered domains

Disorder of human full-length U1-70K (437 amino acids) was predicted using the following services (results shown in Fig. S1):

- 1. SEG (http://mendel.imp.ac.at/METHODS/seg.server.html)
- 2. PONDR (http://www.pondr.com/)
- 3. DISOPRED (bioinf.cs.ucl.ac.uk/disopred/)



Fig. S1. Disordered motif of U1-70K predicted by different models. (**A**) Disordered motif predicted by SEG. (**B**) Disordered motif predicted by PONDR with VL-XT predictor. (**C**) Disordered motif predicted by DISOPRED3 method.



Fig. S2. Statistics of average sizes of droplets of LC1 and LC2. (A) under various of pH (B) with different amino acids added. Ordinary one-way ANOVA test, n>15, depending on the densities of droplets, p<0.1234, p<0.0332, p<0.0001.



Fig. S3. FRAP of the irregular droplets. (A) 200 μ M LC2 under pH 9. (B) 60 μ M of LC1 was mixed with 300 μ M Asp. (C) 60 μ M of LC1 was mixed with 300 μ M Arg.



Fig. S4. Images showing effects of high salt (500 mM NaCl) to LC2 LLPS. The result was compared with LC2 with Asp added.



Fig. S5. Characterization of aggregation and secondary structure of RD19. (A) TEM of $30 \mu M$ RD19 incubated at 37 °C for 3 days. (B) CD spectrum of RD19 incubated for 0 and 24 h.



Fig. S6. The effects of RNA to the LLPS of LC1 and LC2. (**A**) The turbidity changes when adding different ratio of polyU to EGFP-LC1 or -LC2. (**B**) Images of droplets formation with or without 3 equivalents of polyU. (n=3)



Fig. S7. TEM images of aggregates formed by LC1 and LC2 in solution and in droplets. (A) 10 μ M LC1, (B) 60 μ M LC2, (C) 60 μ M EGFFP was incubated at 37 °C for 3 days. The phase separation samples of (D) EGFP-LC1 and (E) EGFP-LC2 were incubated at 37 °C for 12 h, showing aggregates growing inside of droplets.

Droplet count per mm ²	LC1	LC2	
рН 5	2625	3000	
pH 7	4750	3750	
pH 9	9875	1875	
Control	6125	2625	
Arg	1500	2875	
Ala	4500	1875	
Asp	5750	500	

Table S1. Densities of droplets of LC1 and LC2 under various pH values or with different amino acids added.

Table S2. Primers used for gene construction.

Primer name	Primer sequence
U1-BamH231f	5' CGGGATCCAGGGACCGGGACCGGGACCGTGAGCGGGAG 3'
U1-Xho308rstop	5' CCGCTCGAGTCACTCCTCCTTGCGCTCCCGCTCCCGCCGGGC 3'
U1-BamH317f	5' CGGGATCCGCGGAGCCCTCCGAGGCGGGTGACGCGCCC 3'
U1-Xho407rstop	5' CCGCTCGAGTCAGCCCAGACCCTCCAGCCCGTTGTCCTGGCC 3'
U1-Nhe231f	5'CTAGCTAGCATGAGGGACCGGGACCGGGACCGTGAGCGGGAGC
	GCAGA 3'
U1-Sac308rnostop	5' TCCCCGCGGCTCCTCCTTGCGCTCCCGCCGCGG 3'
U1-Nhe317f	5' CTAGCTAGCATGGCGGAGCCCTCCGAGGCGGGTGACGCG 3'
U1-Sac407rnostop	5' TCCCCGCGGGCCCAGACCCTCCAGCCCGTTGTCCTGGCC 3'
U1-Nhe1f	CTAGCTAGCATGACCCAGTTCCTGCCGCCCAACCTTCTG
U1-Sac1314rnostop	GGACCGCGGCTCCGGCGCGCAGCCTCCATCAAATACCC

Table S3. Parameters of LC1, LC2, and LC predicted by CIDAR.

ID	Length	κ	FCR	NCPR	Hydropathy	Disorder promoting
LC1	78	0.086	0.859	0.269	0.896	1.000
LC2	92	0.095	0.543	-0.043	2.127	0.946
LC	178	0.095	0.669	0.096	1.672	0.961

Note: κ - charge patterning parameter, discussed in the help section; FCR- fraction of charged residues; NCPR- net charge per residue