## Extensive Expression Analysis of *Htt* Transcripts in Brain Regions from the zQ175 HD Mouse Model Using a QuantiGene Multiplex Assay

Aikaterini S. Papadopoulou<sup>1</sup>, Casandra Gomez-Paredes<sup>1</sup>, Michael A. Mason<sup>1</sup>,

Bridget A. Taxy<sup>1</sup>, David Howland<sup>2</sup> and Gillian P. Bates<sup>1\*</sup>

<sup>1</sup>Huntington's Disease Centre, Department of Neurodegenerative Disease and UK Dementia

Research Institute at UCL, Queen Square Institute of Neurology, University College London,

London WC1N 3BG, UK.

<sup>2</sup>CHDI Management / CHDI Foundation Inc., New York, NY 10001, USA

\*Corresponding Author: <a href="mailto:gillian.bates@ucl.ac.uk">gillian.bates@ucl.ac.uk</a>

Supplementary Information

Brain region	Dilution of starting material (10 mg/300 μL)	Final input (µg/µL)
Striatum	1:3	11
Cortex	1:4-1:5	6.6 – 8.25
Hippocampus	1:3-1:5	6.6 – 11
Cerebellum	1:4-1:5	6.6 – 8.25
Brainstem	1:3-1:5	6.6 – 11

Supplementary Table S1: Recommended dilution of tissue lysate (10 mg / 300  $\mu$ L) to ensure signals are within the linear range of detection for genes of interest.

**Supplementary Table S2:** qPCR primer / probe set sequences for the measurement of *Htt* transcripts and for the discrimination of endogenous *Htt* and mutant *Htt* in zQ175 mice.

Assay name	ID	Sequence	Assay	
		(5'-3')	Efficiency	
(Htt)-I <sub>1</sub> -pA <sub>1</sub>	FW	TCCTCATCAGGCCTAAGAGCTGG	1.9	
	RV	GAGACCTCCTAAAAGCATTATGTCATC		
	probe	AGTGCAGGACAGCGTGAGAGATGTG		
( <i>Htt</i> )- <i>I</i> ₁- <i>p</i> A₂	FW	TAGCTGCTTGTGACTGGAGA	2	
	RV	CCCAGAGTTGAGAGAAAGGA		
	probe	AGCTGCAAGAGAGCACAGGGC		
(Htt)- I1-3'	FW	GCTTACTGCCTCTGTCCATT	2.1	
	RV	TCAATCAGTCAGCTTGGAGA		
	probe	CCCAAAGGTGCTAGCCTCCA		
Htt-FL	FW	TTAGTGCCATTCATCGTAATTC	2	
	RV	TGAAGTGTTTTCTTCAGAGTGG		
	probe	TCCAGGCAATTCAGTCTCGCTG		
Endogenous (WT) allele	FW	CAGGTCCGGCAGAGGAACC	1.9	
	RV	TTCACACGGTCTTTCTTGGTGG		
	probe	TGCACCGACCAAAGAAGGAACTCT		

FW = forward; RV = Reverse

**Supplementary Table S3**: Design of the 14-plex QuantiGene assay for the detection of all *Htt* transcripts.

Transcript/gene symbol	Transcript/gene name	Accession Number	Probe Set Region
(Htt)-I₁-pA₁	Huntingtin – Intron 1 -	NC000071	521-983
	poly(A)1		
(Htt)-I₁-pA₂	Huntingtin – Intron 1 -	NC000071	1104-1465
	poly(A)₂		
(Htt)- I₁-3'	Huntingtin – Intron 1 - 3'	NC000071	16339-16922
(Htt)-I₃	Huntingtin – Intron 3	NC000071	30195-30846
Htt-FL (Exons 50-53)	Huntingtin coding	NM_010414	6901-7433
(Htt-FL)-s-3'UTR	Huntingtin –short- 3' UTR	NM_010414	9553-9993
(Htt-FL)-m-3'UTR	Huntingtin –mid- 3' UTR	NM_010414	10720-11120
(Htt-FL)-I-3'UTR	Huntingtin –long- 3' UTR	NM_010414	12640-13104
Eif4a2	Eukaryotic Initiation factor	NM_013506	710-1271
	4a2		
Rpl13a	Ribosomal protein L13a	NM_009438	2-467
Ubc	Ubiquitin C	NM_019639	113-676
CanX	Calnexin	NM_007597	1195-1720
Atp5b	ATP synthase subunit beta	NM_016774	22-406
Ppib	Cyclophilin B	NM_011149	207-588



QuantiGene 9-plex QuantiGene 10-plex

**Supplementary Figure S1: Establishment and optimisation of QuantiGene** *Htt* **multiplex assays in striatal tissue in WT and zQ175 heterozygous mice.** The MFI signals corresponding to 2-fold serial dilutions of the QuantiGene striatal lysate are shown for each housekeeping gene probe set. **(a-e)** 9-plex QuantiGene assay and **(f-j)** 10-plex QuantiGene assay. *Eif4a2* and *Ubc* reached saturation at the highest lysate concentrations for both the 9-plex and 10-plex QuantiGene assays. Data were from two technical replicates of equimolar pools of striatal samples (n = 4/genotype). Statistical analysis by linear regression. MFI = mean fluorescence intensity.



Supplementary Figure S2: Establishment and optimisation of QuantiGene *Htt* 10-plex assay in cortical tissue in WT and zQ175 heterozygous mice.

The MFI signals corresponding to 2-fold serial dilutions of the QuantiGene cortical lysate are shown for each transcript probe set. **(a-e)** *Htt* transcripts and **(f-j)** Housekeeping reference genes. **(a, b)** The linear regression slopes of the *Httexon1* transcript ( $I_1$ – $pA_1$  and  $I_1$ – $pA_2$ ) between the two genotypes are significantly different. **(c, d)** The  $I_1$ -3' and  $I_3$  probe sets give comparable signals between WT and zQ175 samples. **(e)** *Htt-FL* probe set signals are shown. **(f-i)** The linear regression slopes for the *Atp5b*, *Canx*, *Eif4a2* and *Rpl13a* housekeeping genes are significantly different between genotypes, but the signals are comparable at the recommended dilutions (Supplementary Table S1). Data were from two technical replicates of equimolar pools of striatal samples (n = 4/genotype). Statistical analysis by linear regression, \*p<0.05, \*\*p<0.01. MFI = mean fluorescence intensity.



## Supplementary Figure S3: Establishment and optimisation of QuantiGene *Htt* 10-plex assay in hippocampal tissue in WT and zQ175 heterozygous mice.

The MFI signals corresponding to 2-fold serial dilutions of the QuantiGene hippocampal lysate are shown for each transcript probe set. (a-e) *Htt* transcripts and (f-j) Housekeeping reference genes. (a, b) The linear regression slopes of the *Httexon1* transcript ( $I_1$ – $pA_1$  and  $I_1$ – $pA_2$ ) between the two genotypes are significantly different. (c, d) The  $I_1$ -3' and  $I_3$  probe sets give comparable signals between WT and zQ175 samples. (e) *Htt-FL* probe set signals are shown. (f, i) The linear regression slopes for the *Atp5b* and *Rpl13a* housekeeping genes are significantly different between genotypes, but the signals are comparable at the recommended dilutions (Supplementary Table S1). Data were from two technical replicates of equimolar pools of striatal samples (n = 4/genotype). Statistical analysis by linear regression, \*p<0.05, \*\*p<0.01. MFI=mean fluorescence intensity.



## Supplementary Figure S4: Establishment and optimisation of QuantiGene *Htt* 10-plex assay in cerebellar tissue in WT and zQ175 heterozygous mice.

The MFI signals corresponding to 2-fold serial dilutions of the QuantiGene cerebellar lysate are shown for each transcript probe set. (a-e) *Htt* transcripts and (f-j) Housekeeping reference genes. (a) There is a trend toward a difference between the linear regression slopes between the two genotypes for the *Httexon1* transcript  $(I_1-pA_1) \# = 0.069$ . (c, d) The  $I_1$ -3' and  $I_3$  probe sets give comparable signals between WT and zQ175 samples. (e) *Htt-FL* probe set signals are shown. (f, i) The linear regression slopes for the *Atp5b* and *Rpl13a* housekeeping genes are significantly different between genotypes, but the signals are comparable at the recommended dilutions (Supplementary Table S1). Data were from two technical replicates of equimolar pools of striatal samples (n = 4/genotype). Statistical analysis by linear regression. MFI = mean fluorescence intensity.



Supplementary Figure S5: Establishment and optimisation of QuantiGene Htt 10-plex assay in brain stem tissue in WT and zQ175 heterozygous mice. The MFI signals corresponding to 2-fold serial dilutions of the QuantiGene brain stem lysate are shown for each transcript probe set. (a-e) Htt transcripts and (f-j) Housekeeping reference genes. (a) The linear regression slope of the Httexon1 transcript with the  $I_1-pA_1$  probe set between the two genotypes is significantly different. (c, d) The  $I_1$ -3' and  $I_3$  probe sets give comparable signals between WT and zQ175 samples. (e) Htt-FL probe set signals are shown. Data were from two technical replicates of equimolar pools of striatal samples (n = 4/genotype). Statistical analysis by linear regression, \*p<0.05. MFI = mean fluorescence intensity.



Supplementary Figure S6: Expression analysis of *Htt* transcripts in five brain regions using the custom-made 14-plex QuantiGene. The expression of intronic *Htt* transcripts is presented relative to that for housekeeping genes. (a-e) The *Httexon1* transcript is present in all zQ175 brain regions, but the difference between zQ175 and WT in the cerebellum did not reach statistical significance. (f-j) The level of the *Htt-FL* transcript is significantly decreased in the striatum, cortex and hippocampus as measured by the E50-53 and all 3'UTR probe sets. # = 0.058. Data were from n = 4/genotype samples with two technical replicates. One sample was excluded from the WT cortex and hippocampus and one from the zQ175 hippocampus, as all of the *Htt-FL* probe sets failed to show a signal above background. Statistical analysis was by multiple *t*-test with Benjamini-Hochberg correction with an alpha of 0.05, \*p<0.05, \*\*p<0.01, \*\*\*p<0.001. Tissue lysate dilutions were 1:3 for striatum and brain stem, 1:4 for cortex and cerebellum and 1:5 for hippocampus.