

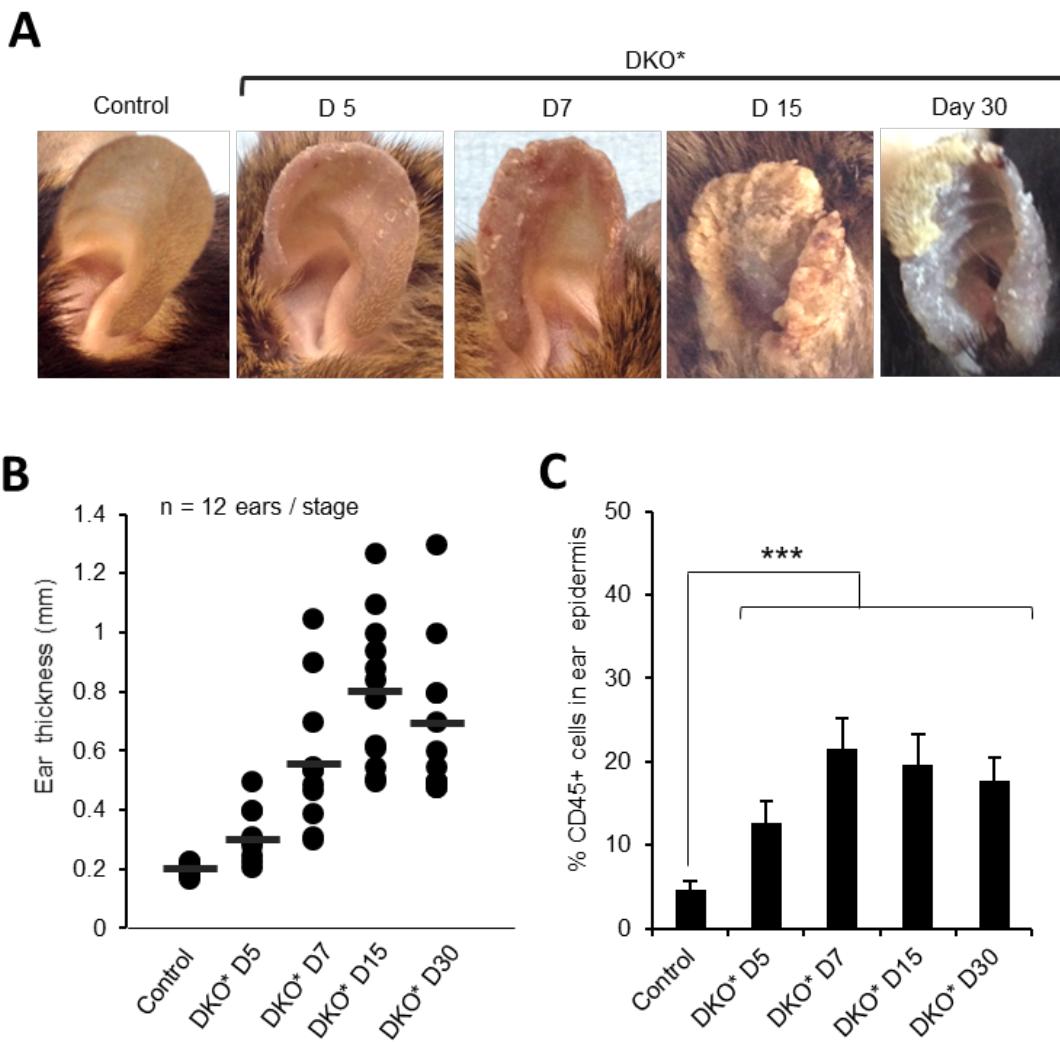
# **Role of bulge epidermal stem cells and TSLP signaling in psoriasis**

**Nuria Gago-Lopez et al.**

## **APPENDIX FIGURES AND LEGENDS**

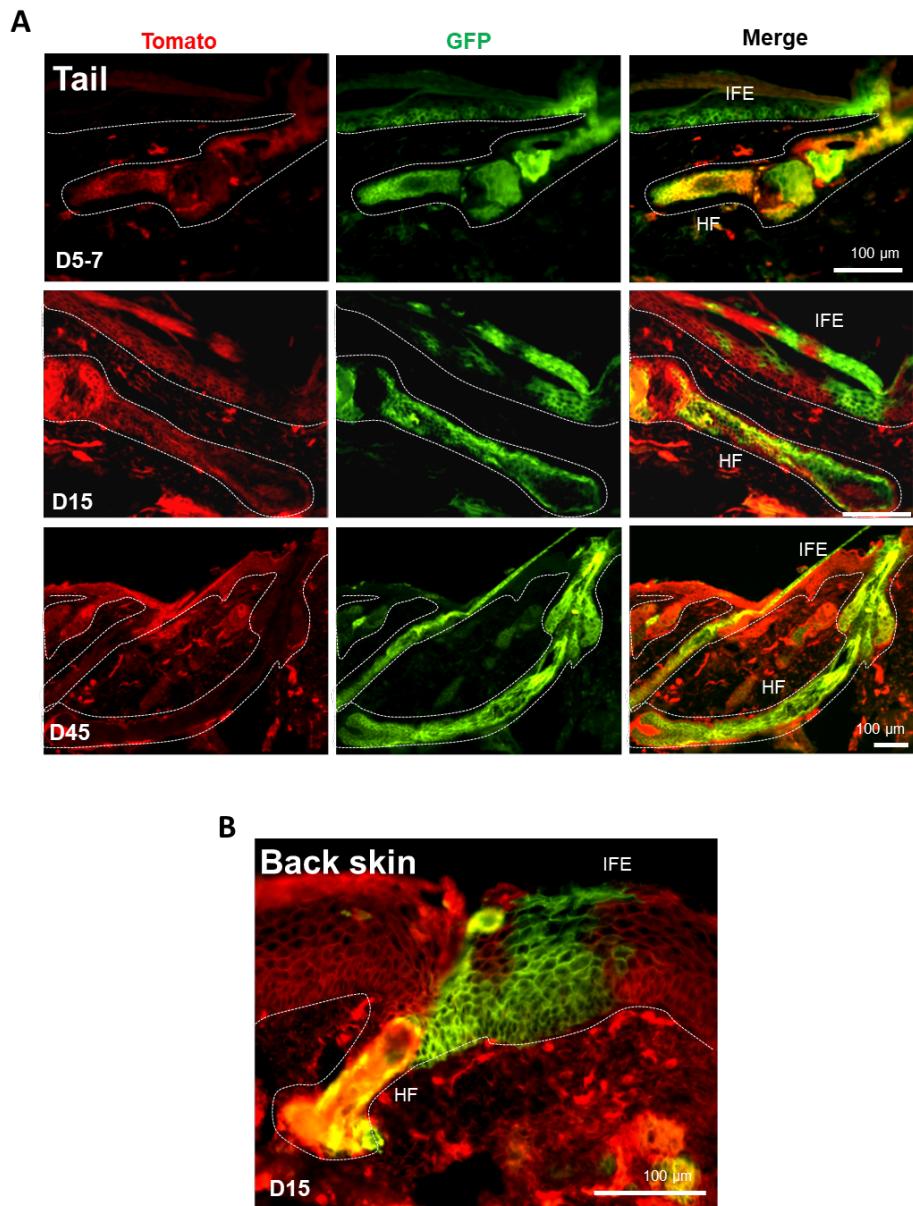
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**Appendix Fig S1. Characterization of DKO\*-mT/mG mice during psoriasis-like progression.**

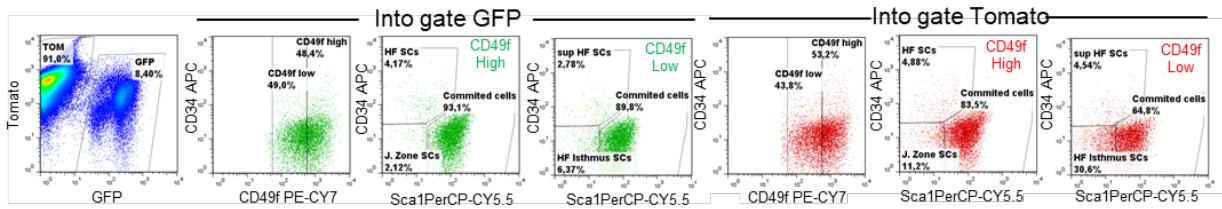
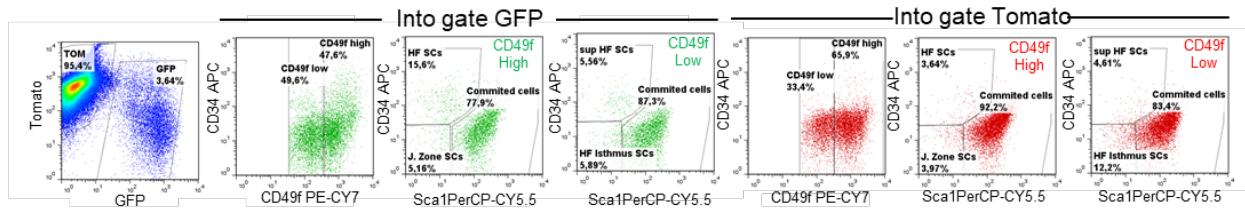
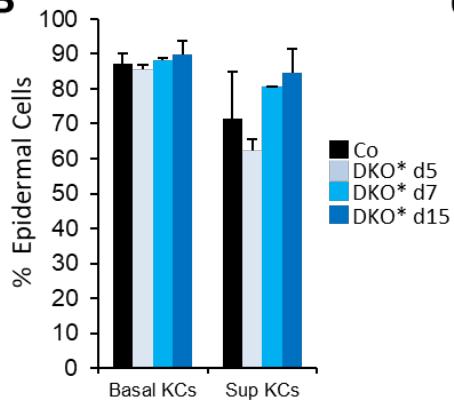
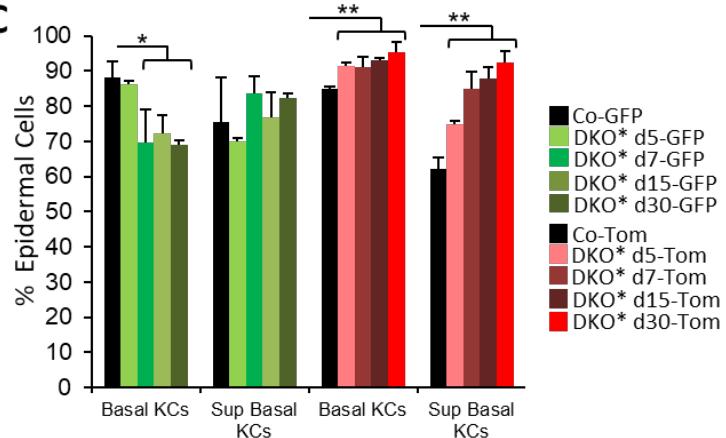
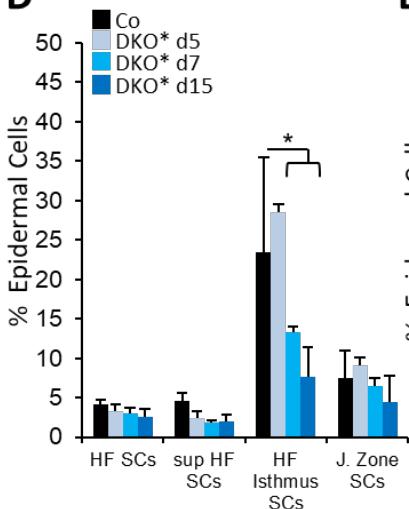
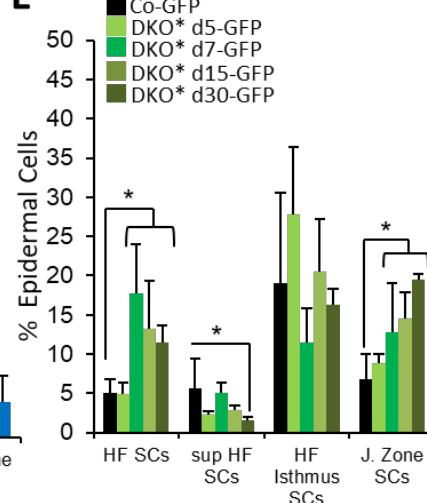
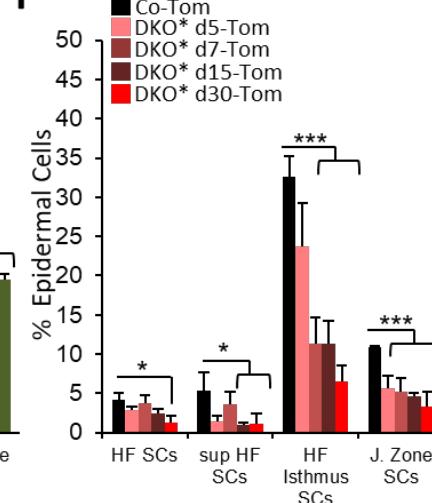
- A.** Images of ear skin at different time points during psoriasis-like development. Same control ear image was included in Fig. 3B.
- B.** Ear thickness measurement at different time points during psoriasis-like development.
- C.** FACS quantification of CD45<sup>+</sup> cells from ear skin of control and DKO\* mice at different time points during psoriasis-like development. n = 5-10 per time point. Statistical significance \*\*\*p<0.0001 (t-student two tailed-test relative to control group). See Supplemental Appendix Table 2 for exact p-Value.



**Appendix Fig S2. Dynamic pattern of GFP/Tomato epidermal cells in tail and back skin of DKO\*-mT/mG mice during psoriasis-like progression.**

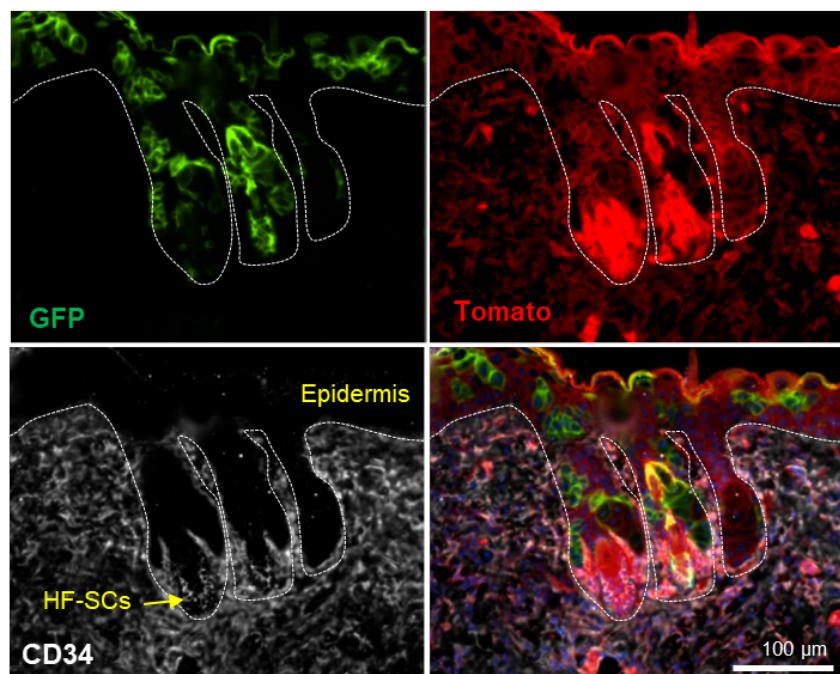
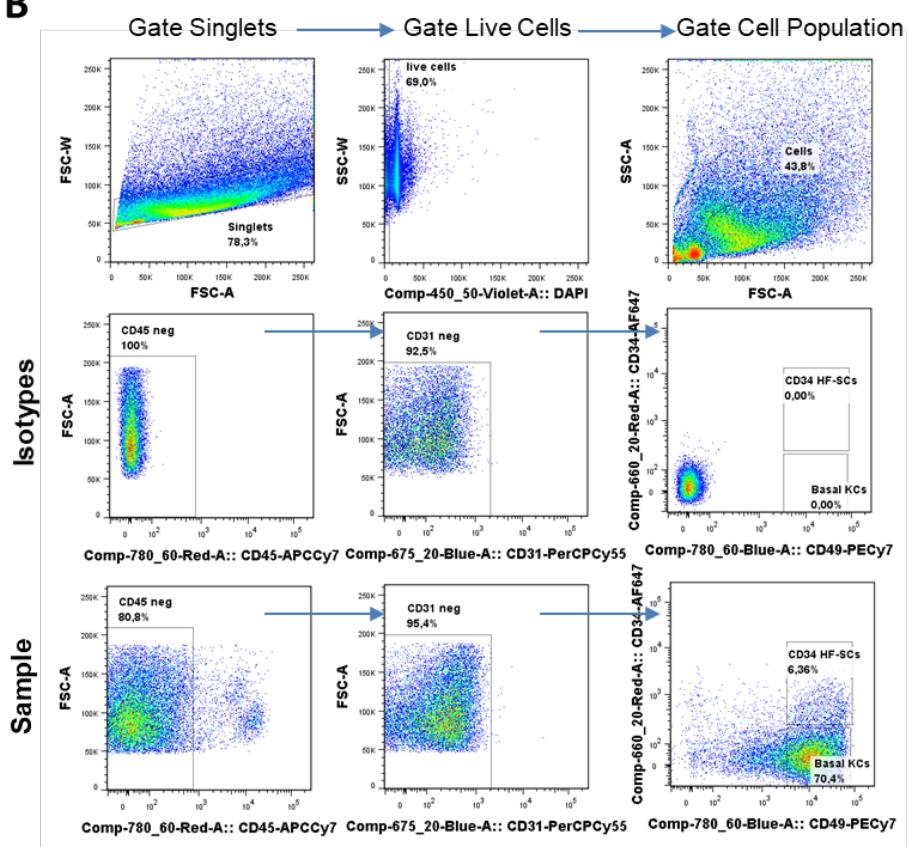
**A.** Fluorescence imaging of DKO\*-mT/mG tail skin at different time points during psoriasis-like progression. White dotted line separates epidermis and dermis.

**B.** Fluorescence imaging of DKO\*-mT/mG back skin at day 15 after tamoxifen treatment. White dotted line separates epidermis and dermis.

**A****Control-mT/mG : Into CD45-CD49f+ population****DKO\*-mT/mG: Into CD45-CD49f+ population****B****C****D****E****F**

**Appendix Fig S3. FACS analyses of different epidermal stem cell populations in ear skin of Co-mT/mG and DKO\*-mT/mG mice during psoriasis-like progression.**

- A.** Strategy for FACS analysis of different epidermal stem cell populations in Co-mT/mG and DKO\*-mT/mG mice.
- B.** FACS quantification of basal and suprabasal keratinocytes from IFE at different time points during psoriasis-like development. n=3 per time point. Statistical significance \*p<0.05 (t-student two tailed-test relative to control groups). See Supplemental Appendix Table 2 for exact p-Value.
- C.** FACS quantification of basal and suprabasal keratinocytes from IFE into GFP<sup>+</sup> and Tomato<sup>+</sup> gating at different time points during psoriasis-like development. Reduction in mutant<sup>GFP</sup> IFE basal keratinocytes and enrichment of non-mutant<sup>Tom</sup> IFE populations. Statistical significance \*p<0.05 (t-student two tailed-test relative to control groups). See Supplemental Appendix Table 2 for exact p-Value.
- D.** FACS quantification of HF-SC subpopulations at different time points during psoriasis-like development. n=3 per time point. Statistical significance \*p<0.05 (t-student two tailed-test relative to control groups). See Supplemental Appendix Table 2 for exact p-Value.
- E.** FACS quantification of HF-SC subpopulations into GFP<sup>+</sup> gating at different time points during psoriasis-like development. Enrichment of bulge HF-SCs and junctional zone SCs. Statistical significance \*p<0.05 (t-student two tailed-test relative to control groups). See Supplemental Appendix Table 2 for exact p-Value.
- F.** FACS quantification of HF-SC subpopulations into Tomato<sup>+</sup> gating at different time points during psoriasis-like development. Reduction in all HF-SC subpopulations along psoriasis-like progression. Statistical significance \*p<0.05 (t-student two tailed-test relative to control groups). See Supplemental Appendix Table 2 for exact p-Value.

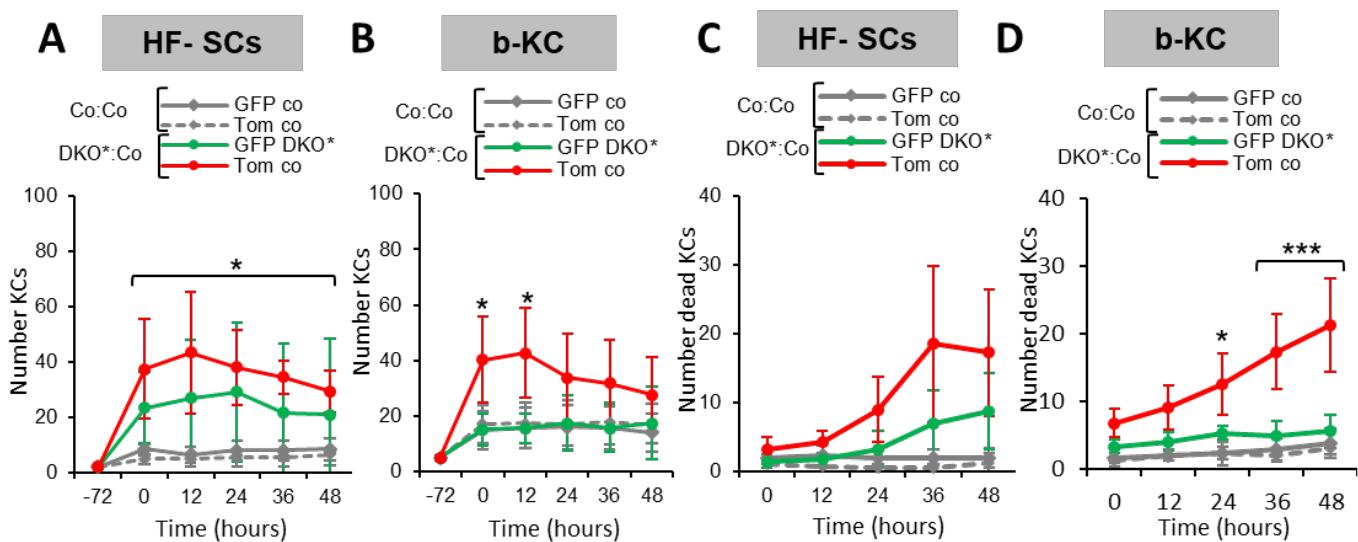
**A****B**

**Appendix Fig S4. CD34 expression in bulge HF-SCs of DKO\*-mT/mG mice by immunofluorescence and FACS strategy for sorting.**

**A.** Immunofluorescence image of CD34 staining in ear skin of psoriatic DKO\*-mT/mG mice.

White dotted line separates epidermis and dermis.

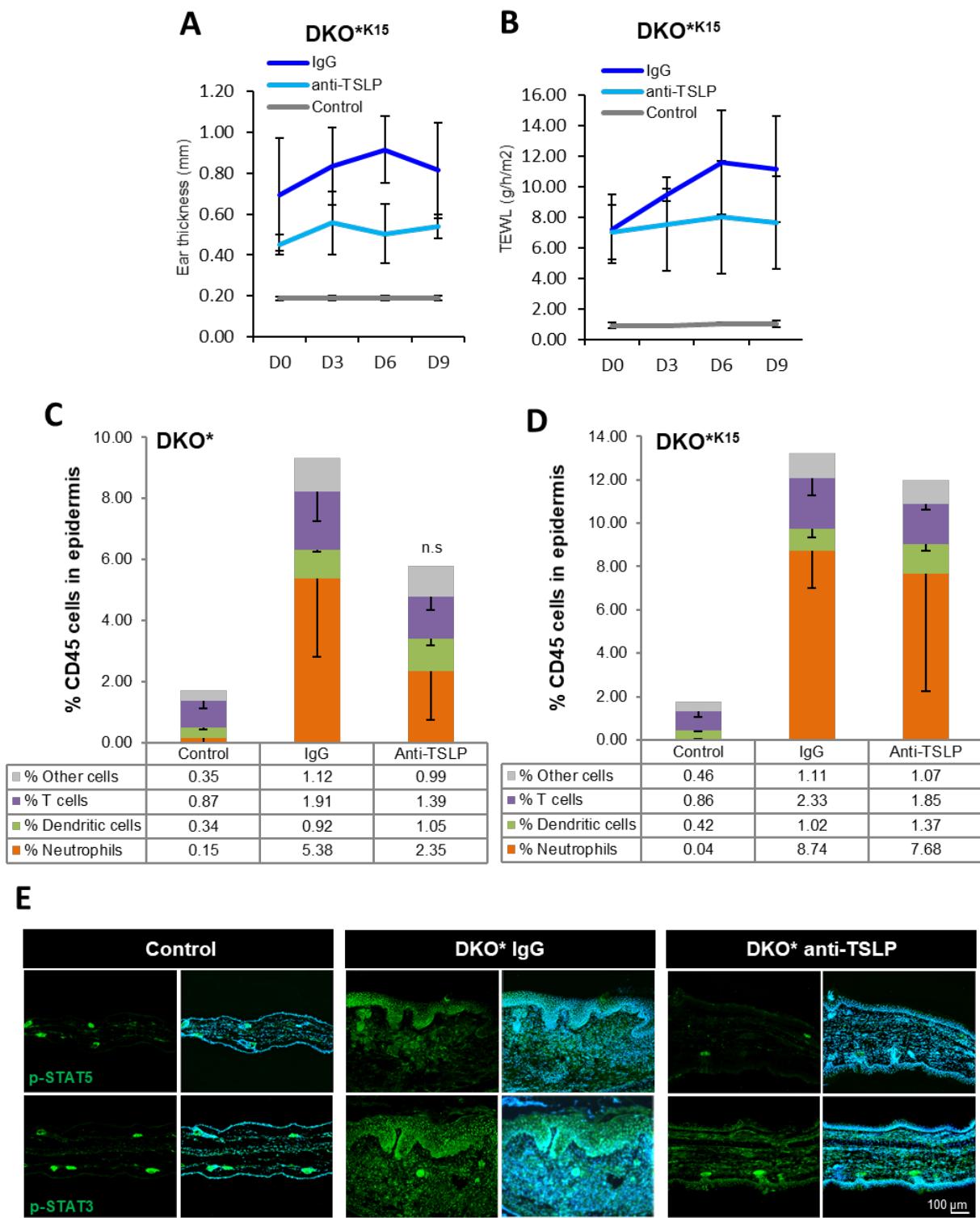
**B.** Strategy for FACS sorting of CD34 bulge HF-SCs and CD49f basal KCs. Sorting procedures were done excluding dead cells and doublets. Cell population positive for CD45 and CD31 were discarded and cells positive for  $CD34^+/CD49f^{high}$  and  $CD34^-/CD49f^{high}$  were sorted for further experiments.



**Appendix Fig S5. Mutant<sup>GFP</sup> keratinocytes induce hyper-proliferation in control<sup>Tom</sup> keratinocytes.**

**A, B.** Average of total number of live and dead keratinocytes per field from co-cultures of mutant or control GFP<sup>+</sup> HF-SCs with control Tomato<sup>+</sup> HF-SCs at different time points of time lapse capture during 48 hours. n = 10-12 fields analyzed in total from two independent experiments. Statistical significance \*p<0.05 (Two-way ANOVA and Bonferroni post test).

**C, D.** Average of total number of live or dead keratinocytes per field from co-cultures of mutant or control GFP<sup>+</sup> basal keratinocytes with control Tomato<sup>+</sup> basal keratinocytes at different time points of time lapse capture during 48 hours. n = 4 fields from two independent experiments. Statistical significance \*p<0.05, \*\*\*p<0.001 (Two-way ANOVA and Bonferroni post test).



**Appendix Fig S6. Characterization of immune cell infiltration and TSLP signaling in DKO\* and DKO\*<sup>15</sup> mice treated with anti-TSLP.**

**A, B.** Longitudinal analysis of ear thickness (I) and Transepidermal water loss (TEWL) (J) in DKO\*<sup>15</sup> mice at different time points during psoriasis-like disease progression (Two-way ANOVA and Bonferroni post test, n=3, n.s = non statistical significance).

**C, D.** FACS quantification of immune cell subtypes (T cells, dendritic cells and neutrophils) on ear skin of DKO\* and DKO\*<sup>15</sup> mice after anti-TSLP or IgG treatment by FACS analysis. After anti-TSLP treatment, neutrophils are reduced in DKO\* mice. DKO\* n=4, DKO\*<sup>15</sup> n=3. n.s = non statistical significance (t-student two tailed-test relative to control groups).

**E.** Representative immune-fluorescence images of ear sections from DKO\* mice treated with IgG (left panel) or anti-TSLP (right panel) stained for p-STAT5 and p-STAT3. p-STAT5 expression was inhibited in mice treated with anti-TSLP, whereas p-STAT3 expression was maintained in both groups, IgG and anti-TSLP (n =3).

**Table S1. Primers for qPCR**

Name	Sequence
Tslp-F	GAGGACTGTGAGAGCAAGCCAG
Tslp-R	GGCAGTGGTCATTGAGGGCTT
Tslpr-F	CGGGAGAGCAATGACGATG
Tslpr-R	CCGAACCCGGAAGTCATAGC
IL-7ra-F	TGTGAGTTCAATCCGAAAGT
IL-7ra-R	CTGGCTGTGCAGGAAGATCA
Vegfa-F	CAGTCCGAGCCGGAGAGGGAGC
Vegfa-R	CGGACGGCAGTAGCTTCGCTGG
Foxc1a-F	CACTCGGTGCGGGAAATGT
Foxc1a-R	GTGCGGTACAGAGACTGACTG
Nfatc1-F	GGCGGGAAAGAAGATGGTCTGTC
Nfatc1-R	TGGTTGCGGAAAGGTGGTATCTCA
Ptgs2-F	AAGCCGAGCACCTTGAG
Ptgs2-R	ATTGATGGTGGCTTTGGTAG
Tnfa-F	CCCCAAAGGGATGAGAAGTT
Tnfa-R	CACTTGGTGGTTGCTACGA
II-23a-F	AGCAGGACATATGAATCTACTAAGAGA
II-23a-R	GTCCTAGTAGGGAGGTGTGAAGTTG
II-1 $\alpha$ -F	GGCTCACTCATGAGACTTGC
II-1 $\alpha$ -R	AGGTGTAAGGTGCTGATCTGG
II-6-F	CCGGAGAGGAGACTCACAG
II-6-R	CAGAATTGCCATTGCACAAC
II-1 $\beta$ -F	GGGCTGGACTGTTCTAATGCCTT
II-1 $\beta$ -R	CCATCAGAGGCAAGGAGGAAACA
p65-F	CCAAAAAGACGTGCCTCCTG
p65-R	CCCCCAGCCCATAAGGAAAC
G-CSF-F	CTGCCACCATCCCTGCCTCT
G-CSF-R	CCATCTGCTGCCAGATGGTGGT
IFN- $\gamma$ -F	GAAAATCCTGCAGAGCCAGATT
IFN- $\gamma$ -R	TGATGCCCTGATTGTCTTCAA
S100a9-F	GTGGTGTCCAGGTCTCC
S100a9-R	CACCTTCTCAGATGGAGCG

**Table S2. List of p-Values**

Figure	sample number	exact p-Value
1	I J	n = 2-6 n = 2-6
		Co vs Lesional: *=0.021; Non Lesional vs Lesional: **=0.0044 Co vs Lesional: **=0.01; Non Lesional vs Lesional: ***=0.0007
2	D	n = 6
		IFE Thickness. Co vs DKO* D5: n.s.=0.055; Co vs DKO* D7: **=0.0015; Co vs DKO* D15: ***=0.0001; Co vs DKO* D30: ***=0.0002
2	E	n = 3
		cCas3. DKO*GFP vs DKO*Tom D5: n.s.=0.13; DKO*GFP vs DKO*Tom D7: *=0.044; DKO*GFP vs DKO*Tom D15: n.s.=0.79 Ki67. DKO*GFP vs DKO*Tom D5: n.s.=0.87; DKO*GFP vs DKO*Tom D7: ***=0.0007; DKO*GFP vs DKO*Tom D15: *=0.05
3	D	n = 5-12
		Co vs DKO* D5: *=-0.014; Co vs DKO* D7: **=0.0065; Co vs DKO* D15: ***=0.0005; Co vs DKO* D30: **=0.0027; Co vs DKO15* D5: **=0.0032; Co vs DKO15* D7: ***=0.0001; Co vs DKO15* D15: **=0.0014; Co vs DKO15* D30: ***=0.0001
3	E	n = 4-6
		Co vs DKO* D5, D15, D30: ***=0.0001; Co vs DKO15* D5, D15, D30: ***<0.0001
3	F	n = 6-10
		Co vs DKO*: ***=0.0003; Co vs DKO15*: ***<0.0001
3	G	n = 4-5
		Co vs DKO*: *=0.039; Co vs DKO15*: *=0.035
3	H	n = 4-5
		Co vs DKO*: ***=0.0096; Co vs DKO15*: ***=0.0038
3	I	n = 3-6
		GFP. Co vs DKO* D5: n.s.=0.73; Co vs DKO* D15: **=0.0028; Co vs DKO* D30: **=0.0027; Co vs DKO15* D5: **=0.0026; Co vs DKO15* D15: **=0.0036; Co vs DKO15* D30: **=0.003
3	K	n = 3-5
		cCas3. DKO15*GFP vs DKO15*Tom D5: n.s.=0.2; DKO15*GFP vs DKO15*Tom D7: n.s.=0.119; DKO15*GFP vs DKO15*Tom D15: **=0.0027
3	K	n = 3-5
		Ki67. DKO15*GFP vs DKO15*Tom D5: n.s.=0.06; DKO15*GFP vs DKO15*Tom D7: *=0.012; DKO15*GFP vs DKO15*Tom D15: ***=0.0001
5	D	n = 3
		Co CD34 vs CD34GFP D7: **=0.0047; Co CD34 vs CD34GFP D30: *=-0.05; Co CD34 vs CD34Tom D7: ***=0.0031; Co CD34 vs CD34Tom D30: **=0.0025; Co CD49f vs CD49fGFP D7: *=0.02; Co CD49f vs CD49fGFP D30: *=-0.05
5	E	n = 2
		Co CD34 vs CD34GFP D7: *=-0.017; Co CD34 vs CD34GFP D30: *=-0.05; Co CD34 vs CD34Tom D7: *=0.016; Co CD34 vs CD34Tom D30: *=-0.011; Co CD49f vs CD49fGFP D7: *=0.011; Co CD49f vs CD49fGFP D30: *=-0.012
5	F	n = 4
		Co vs DKO*: **=0.0018; Co vs DKO15*: *=-0.04
6	B	n = 3
		Ad-Empty vs Ad-Cre: ***=<0.0001
6	D	n = 3
		IgG vs AbTSLP Ad-Empty: *=-0.02; IgG vs AbTSLP Ad-Cre: *=0.05
6	E	n = 3
		IgG vs AbTSLP Ad-Cre: *=-0.05
6	F	n = 3
		IgG vs AbTSLP Ad-Empty: **=0.007; IgG vs AbTSLP Ad-Cre: *=0.011
6	K	n = 3
		IgG vs AbTSLP DKO*: *=0.03; IgG vs AbTSLP DKO15*: *=0.015
7	B	n = 3
		Control or non-Lesional vs Lesional: ***=0.001
7	C	n = 3
		Control or non-Lesional vs Lesional: *=-0.001
EV3	B	n = 3
		Co vs DKO* D15: ***=0.0001
EV3	C	n = 3
		Co vs DKO* D15: ***=0.0001
EV3	D	n = 3
		Co CD34 vs CD34GFP D7: **=0.007
EV4	F	n = 3-4
		Co CD34 vs CD34GFP: *=-0.015; Co CD34 vs CD34Tom: n.s.=0.089
EV4	G	n = 3-4
		Co CD34 vs CD34GFP: *=0.021
EV4	H	n = 3
		Co CD34 vs CD34GFP: *=-0.017; Co CD34 vs CD34Tom: *=0.016
EV4	I	n = 3
		Co CD34 vs CD34GFP: *=0.035
EV4	J	n = 3
		Co CD34 vs CD34Tom: *=-0.024; Co CD49f vs CD49fTom: n.s.=0.18
EV4	K	n = 3
		Co CD49f vs CD49fTom: **=0.0077
EV4	L	n = 3
		All cell subpopulations in relation with the control: **<0.0011
EV5	E	n = 2
		Ad-cre IgG GFP vs Ad-cre AbTSLP GFP: *=0.047
EV5	F	n = 2
		Ad-cre IgG GFP vs Ad-cre AbTSLP GFP: *=0.05
EV5	G	n = 2
		Ad-cre IgG GFP vs Ad-cre AbTSLP GFP: n.s.=0.2; Ad-cre IgG Tom vs Ad-cre AbTSLPTom: *=0.05
EV5	H	n = 2
		Ad-cre IgG Tom vs Ad-cre AbTSLPTom: n.s.=0.08
EV5	I	n = 2
		Ad-cre IgG GFP vs Ad-cre AbTSLP GFP: *=-0.05
EV5	J	n = 2
		Ad-cre IgG GFP vs Ad-cre AbTSLP GFP: *=0.05
EV5	L	n = 2
		Ad-cre IgG GFP vs Ad-cre AbTSLP GFP: *=0.029
Appendix S1	C	n = 5-10
		Co vs DKO* D5, D15, D30: ***<0.0001
Appendix S3	C	n = 3
		b-KCs Co vs b-KCs GFP D7-30: *=0.042; b-KCs Co vs b-KCs Tom D7-30: **=0.003; Supb-KCs Co vs Supb-KCs Tom D7-30: **=0.0012
Appendix S3	D	n = 3
		HF Isthmus SCs Co vs HF Isthmus SCs D7-15: *=0.05
Appendix S3	E	n = 3
		HF-SCs Co vs HF-SCs GFP D7-30: *=0.03; Sup HF-SCs Co vs Sup HF-SCs GFP D30: *=-0.05; J. Zone SCs Co vs J. Zone SCs GFP D30: *=-0.012
Appendix S3	F	n = 3
		HF-SCs Co vs HF-SCs Tom D30: *=-0.04; Sup HF-SCs Co vs Sup HF-SCs Tom D15-30: *=-0.05; HF Isthmus SCs Co vs HF Isthmus SCs Tom D7-30: ***=0.0006; J. Zone SCs Co vs J. Zone SCs Tom D30: ***=0.0004