

Supplementary Materials

Characterization of the Noncanonical Regulatory and Transporter Genes in Atratumycin Biosynthesis and Production in a Heterologous Host

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Table S1. Summary of strains and plasmids used in this study.

Strains/ plasmids	Relevant phenotype	Source/ [Ref]
<i>S. atratus</i> SCSIO ZH16NS-80S	Wild-type (WT) producer of atartumycin	[1]
$\Delta atr1$	<i>S. atratus</i> SCSIO ZH16NS with a 624 bp of <i>atr1</i> substituted by <i>aac(3)IV</i> +OriT	This work
$\Delta atr2$	<i>S. atratus</i> SCSIO ZH16NS with a 315 bp of <i>atr2</i> substituted by <i>aac(3)IV</i> +OriT	This work
$\Delta atr29$	<i>S. atratus</i> SCSIO ZH16NS with a 729 bp of <i>atr29</i> substituted by <i>aac(3)IV</i> +OriT	This work
$\Delta atr30$	<i>S. atratus</i> SCSIO ZH16NS with a 447 bp of <i>atr30</i> substituted by <i>aac(3)IV</i> +OriT	This work
$\Delta atr32$	<i>S. atratus</i> SCSIO ZH16NS with a 612 bp of <i>atr32</i> substituted by <i>aac(3)IV</i> +OriT	This work
<i>E. coli</i>		
Bw25113	K-12 derivative: <i>araBAD</i> , <i>rhaBAD</i>	[2]
ET12567	<i>dam</i> , <i>dcm</i> , <i>hsdM</i> , <i>hsdS</i> , <i>hsdR</i> , <i>catR</i> , <i>tetR</i>	[3]
Plasmids		
pIJ773	P1-FRT-oriT- <i>aac(3)IV</i> -FRT-P2	[4]
pIJ790	λ -RED (<i>gam bet exo</i>) CmlR <i>araCrep101ts</i>	[4]
pUZ8002	<i>tra</i> , <i>neo</i> , RP4	[5]
pL646ATE	<i>Tsr</i> , <i>acc(3)IV</i> , <i>ermE</i> *P	[6]
cosmid-56G	A cosmid which contains partial atratumycin biosynthesis cluster	This work
cosmid-198F	A cosmid which contains partial atratumycin biosynthesis cluster	This work
cosmid-1610C	A cosmid which contains partial atratumycin biosynthesis cluster	This work
PAC-434C	A cosmid which contains complete actinomycin biosynthesis cluster	This work
$p\Delta atr1$	A 624 bp fragment in <i>atr1</i> in cosmid 56G was substituted by the <i>aac(IV)</i> +OriT cassette using the PCR-targeting strategy	This work
$p\Delta atr2$	A 315 bp fragment in <i>atr2</i> in cosmid 56G was substituted by the <i>aac(IV)</i> +OriT cassette using the PCR-targeting strategy	This work
$p\Delta atr29$	A 729 bp fragment in <i>atr29</i> in cosmid 1610C was substituted by the <i>aac(IV)</i> +OriT cassette using the PCR-targeting strategy	This work
$p\Delta atr30$	A 447 bp fragment in <i>atr30</i> in cosmid 1610C was substituted by the <i>aac(IV)</i> +OriT cassette using the PCR-targeting strategy	This work
$p\Delta atr32$	A 612 bp fragment in <i>atr32</i> in cosmid 1610C was substituted by the <i>aac(IV)</i> +OriT cassette using the PCR-targeting strategy	This work
ZH16NS-80S:: <i>atr1</i>	An integrated vector pL646ATE with complete <i>atr1</i> for over-expression	This work

ZH16NS-80S:: <i>atr2</i>	An integrated vector pL646ATE with complete <i>atr2</i> for over-expression	This work
ZH16NS-80S:: <i>atr29</i>	An integrated vector pL646ATE with complete <i>atr29</i> for over-expression	This work
ZH16NS-80S:: <i>atr30</i>	An integrated vector pL646ATE with complete <i>atr30</i> for over-expression	This work
ZH16NS-80S:: <i>atr32</i>	An integrated vector pL646ATE with complete <i>atr32</i> for over-expression	This work

Table S2. Summary of primers used in this study.

Primer Name	Sequence (5'→3')	purpose
DEL- <i>atr1</i> -F	gaaagcgcgttgcttcaggcccgtgcgctgatcgaatcattccggggatccgacgacc	For disrupting <i>atr1</i>
DEL- <i>atr1</i> -R	ctggaaacgggagttggccccgagctcgcgcgatgatctctgtaggctggagctgcttc	
DEL- <i>atr2</i> -F	gaactcggctcctgtgcaccaggcgatgctcgacgacattccggggatccgacgacc	For disrupting <i>atr2</i>
DEL- <i>atr2</i> -R	gacgtaccggcgataggtgcgaccgagacggagagctctgtaggctggagctgcttc	
DEL- <i>atr29</i> -F	gagggaacggtcctcggactgctcgcccccaacggtgacattccggggatccgacgacc	For disrupting <i>atr29</i>
DEL- <i>atr29</i> -R	gcgccgaacggcgaaactccgctatgtcgattccgagttctgtaggctggagctgcttc	
DEL- <i>atr30</i> -F	atgatgtcaacatctcccagacggccctggcgctgctattccggggatccgacgacc	For disrupting <i>atr30</i>
DEL- <i>atr30</i> -R	ccccgtaccgatgccacggtgggattgccgaagagttgtgtaggctggagctgcttc	
DEL- <i>atr32</i> -F	gccaccgacggccccagatctacacctatgtctcccagattccggggatccgacgacc	For disrupting <i>atr32</i>
DEL- <i>atr32</i> -R	catcggggaggacgcccgtccaggcggcggtcccgtgtaggctggagctgcttc	
COM- <i>atr1</i> -F	aaaacatgatgaagacttccacgtctaaac	For cloning complete <i>atr1</i>
COM- <i>atr1</i> -R	aaaaactagttcatgacgtcgtgattctctg	
COM- <i>atr2</i> -F	aaaacatgatgaccgattcctattctgtc	For cloning complete <i>atr2</i>
COM- <i>atr2</i> -R	aaaaactagtctacaggaagctgccagtgc	
COM- <i>atr29</i> -F	aaaacatgatgacgaacgccatcgaggc	For cloning complete <i>atr29</i>
COM- <i>atr29</i> -R	aaaaactagttcacgccgattccctctccc	
COM- <i>atr30</i> -F	aaaacataggtgaccgagacactcagtgc	For cloning complete <i>Atr30</i>
COM- <i>atr30</i> -R	aaaaactagttcagcgagcggtcgccg	
COM- <i>atr32</i> -F	aaaacataggtgacggcttcgacc	For cloning complete <i>Atr32</i>
COM- <i>atr32</i> -R	aaaaactagtctacatcgactccctga	
ID- <i>atr1</i> -F	gccaataccgccgaaggcatc	For verifying mutant $\Delta atr1$
ID- <i>atr1</i> -R	cagacctctccacggctc	
ID- <i>atr2</i> -F	cggaaagcagatcgaacttcgg	For verifying mutant $\Delta atr2$
ID- <i>atr2</i> -R	caggagcgccatgatctccgc	
ID- <i>atr29</i> -F	gccatcgaggccgaaggctc	For verifying mutant $\Delta atr29$
ID- <i>atr29</i> -R	gacgtcgtcagcgaagggcg	

ID- <i>atr30</i> -F	caggagtacatcatggcgggc	For verifying mutant Δ <i>atr30</i>
ID- <i>atr30</i> -R	cagcgacatcacgatgggggtg	
ID- <i>atr32</i> -F	gaagcagcgcaccgtcctcg	For verifying mutant Δ <i>atr32</i>
ID- <i>atr32</i> -R	gacttgccggtacctgcagc	
ID- <i>atr19</i> -F	gcgggtggacctggcgaag	For screening PAC clones containing <i>atr</i> BGC
ID- <i>atr19</i> -R	gcagccgtcgggagcgacg	
ID- <i>orf(-2)</i> -F	cccatacgccgctacgaacgc	For screening PAC clones containing <i>atr</i> BGC
ID- <i>orf(-2)</i> -R	ccgttgcaagtggcacgagttc	
ID- <i>orf(+2)</i> -F	gaaatggtgctggagggtcacg	For screening PAC clones containing <i>atr</i> BGC
ID- <i>orf(+2)</i> -R	ctagatccgccgctccgcacc	
ID-com-F	gatctgaccgacgcggtccac	For verifying complete gene fragments
ID-com-R	gcgcggtcgatccccgatg	

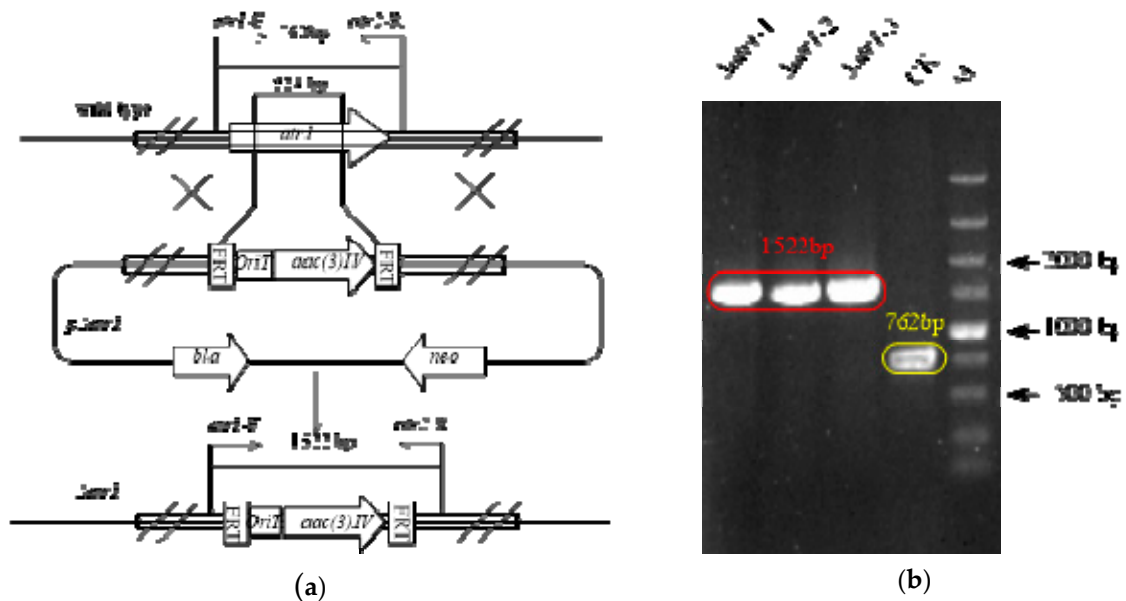


Figure S1. Disruption of *atr1* in *S. atratus* SCSIO ZH16NS-80S via PCR-targeting. **a)** Schematic representation for disruption of *atr1*. **b)** PCR analysis of the control strain (*S. atratus* SCSIO ZH16NS-80S) and the *atr1* mutants carried out using primers listed in **Table S2**. M: DNA molecular ladder; CK: using the genomic DNA of *S. atratus* SCSIO ZH16NS-80S as template, Δ *atr1* (1-3): using the genomic DNA of three different colonies of Δ *atr1* mutant as template.

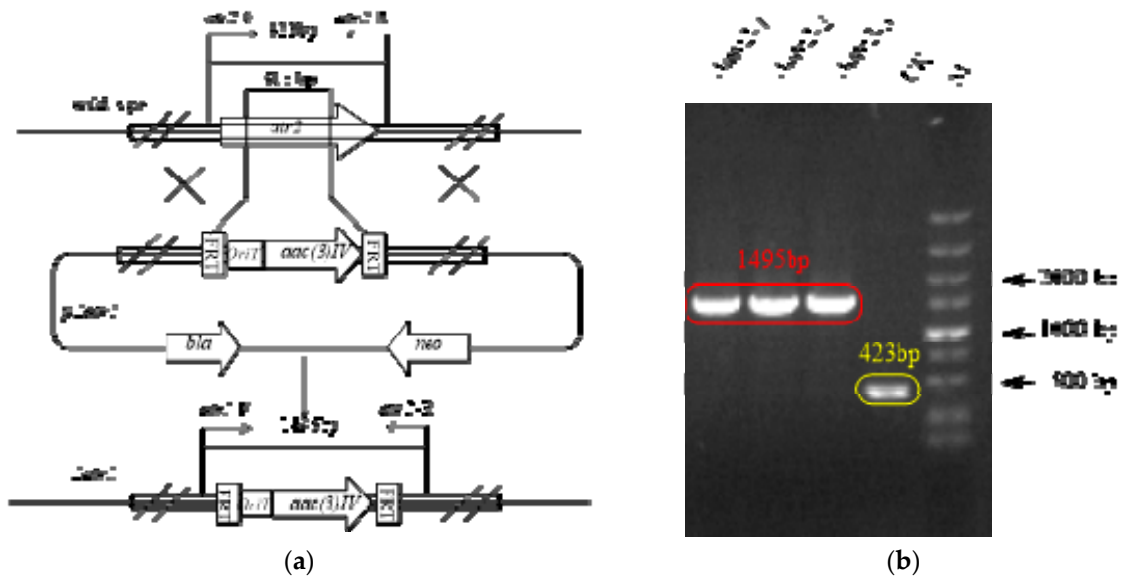


Figure S2. Disruption of *atr2* in *S. atratus* SCSIO ZH16NS-80S via PCR-targeting. **a)** Schematic representation for disruption of *atr2*. **b)** PCR analysis of the control strain (*S. atratus* SCSIO ZH16NS-80S) and the *atr2* mutants carried out using primers listed in **Table S2**. M: DNA molecular ladder; CK: using the genomic DNA of *S. atratus* SCSIO ZH16NS-80S as template, $\Delta atr2$ (1-3): using the genomic DNA of three different colonies of $\Delta atr2$ mutant as template.

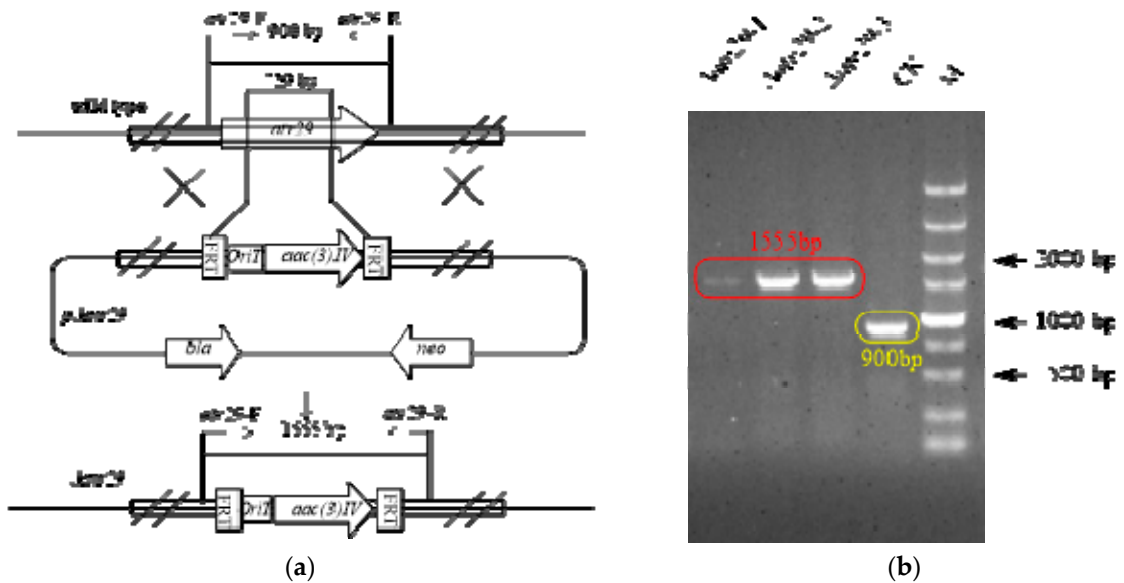


Figure S3. Disruption of *atr29* in *S. atratus* SCSIO ZH16NS-80S via PCR-targeting. **a)** Schematic representation for disruption of *atr29*. **b)** PCR analysis of the control strain (*S. atratus* SCSIO ZH16NS-80S) and the *atr29* mutants carried out using primers listed in **Table S2**. M: DNA molecular ladder; CK: using the genomic DNA of *S. atratus* SCSIO ZH16NS-80S as template, $\Delta atr29$ (1-3): using the genomic DNA of three different colonies of $\Delta atr29$ mutant as template.

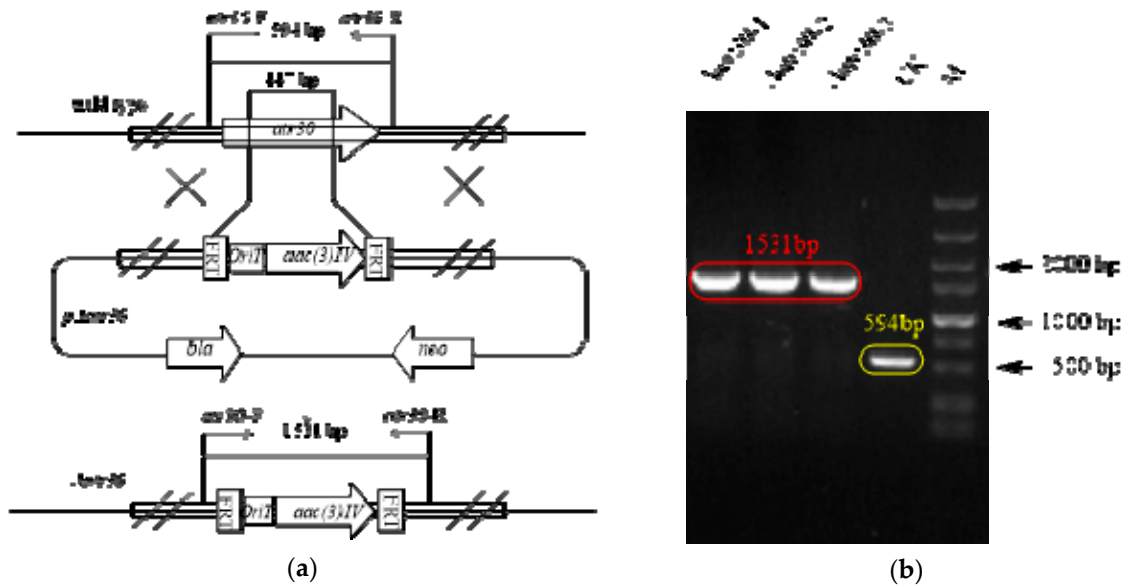


Figure S4. Disruption of *atr30* in *S. atratus* SCSIO ZH16NS-80S via PCR-targeting. **a)** Schematic representation for disruption of *atr30*. **b)** PCR analysis of the control strain (*S. atratus* SCSIO ZH16NS-80S) and the *atr30* mutants carried out with the primers listed in **Table S2**. M: DNA molecular ladder; CK: using the genomic DNA of *S. atratus* SCSIO ZH16NS-80S as template, $\Delta atr30$ (1-3): using the genomic DNA of three different colonies of $\Delta atr30$ mutant as template.

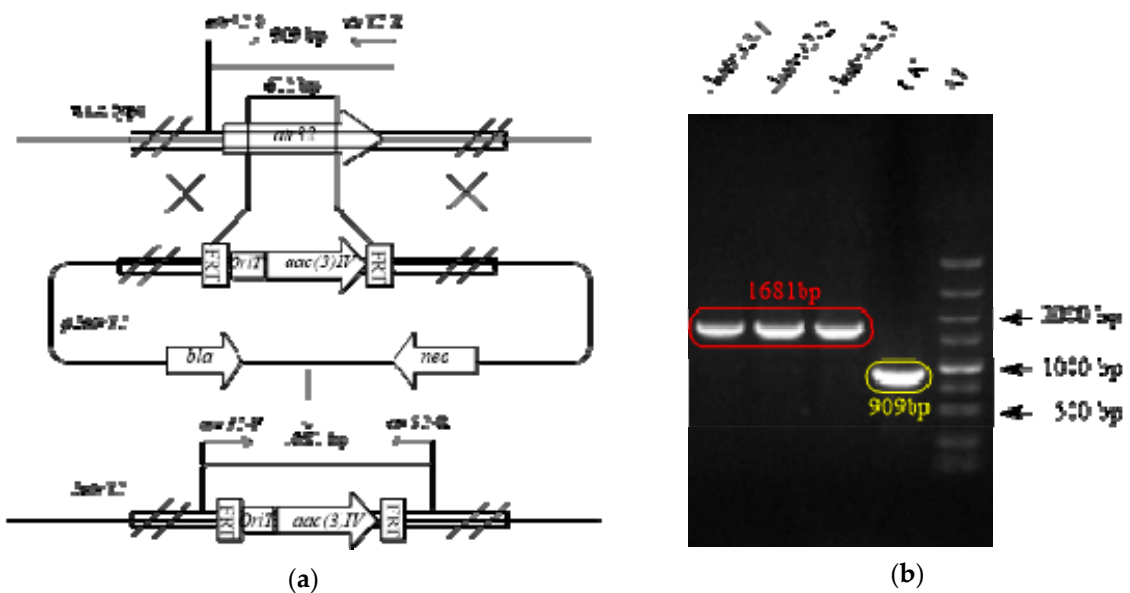


Figure S5. Disruption of *atr32* in *S. atratus* SCSIO ZH16NS-80S via PCR-targeting. **a)** Schematic representation for disruption of *atr32*. **b)** PCR analysis of the control strain (*S. atratus* SCSIO ZH16NS-80S) and the *atr32* mutants carried out using primers listed in **Table S2**. M: DNA molecular ladder; CK: using the genomic DNA of *S. atratus* SCSIO ZH16NS-80S as template, $\Delta atr32$ (1-3): using the genomic DNA of three different colonies of $\Delta atr32$ mutant as template.

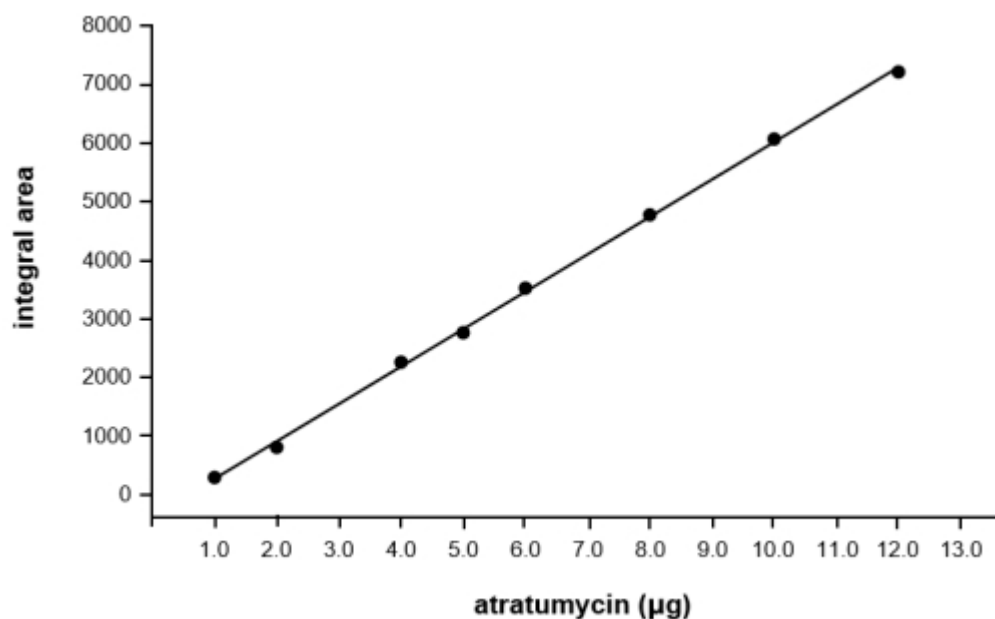


Figure S6. The quantitative HPLC standard curve for atratumycin. The curve was generated via analysis of a concentration gradient of 1.0 μg, 2.0 μg, 4.0 μg, 5.0 μg, 6.0 μg, 8.0 μg, 10.0 μg, 12.0 μg of atratumycin. UV absorption was maintained below 1 A unit to ensure appropriate confidence of the generated standard curve and avoid deviation from linearity.

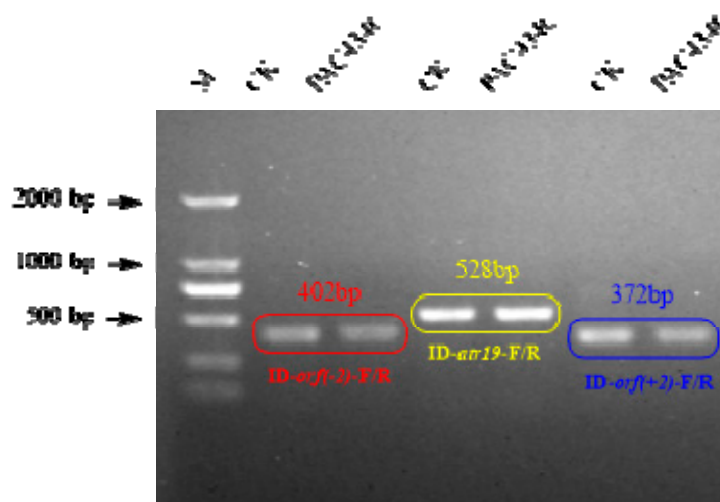


Figure S7. PCR verification of PAC434 containing entire *atr* gene cluster. M: DNA molecular ladder; CK: using the genomic DNA of *S. atratus* SCSIO ZH16NS-80S as template.

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