## **Supplementary Information**

#### **Renal Clearable Nanochelator for Iron Overload Therapy**

Homan Kang,<sup>1#</sup> Murui Han,<sup>2#</sup> Jie Xue,<sup>2</sup> Yoonji Baek,<sup>1</sup> JuOae Chang,<sup>2</sup> Shuang Hu,<sup>1</sup> HaYoung Nam,<sup>2</sup> Min Joo Jo,<sup>1</sup> Georges El Fakhri,<sup>1</sup> Michael P. Hutchens,<sup>3,4</sup> Hak Soo Choi,<sup>1,\*</sup> and Jonghan Kim<sup>2,\*</sup>

<sup>1</sup>Gordon Center for Medical Imaging, Department of Radiology, Massachusetts General Hospital and Harvard Medical School, Boston, MA 02114, USA

<sup>2</sup>Department of Pharmaceutical Sciences, Northeastern University, Boston, MA 02115, USA <sup>3</sup>Anesthesiology and Perioperative Medicine, Oregon Health & Science University, Portland, OR 97239, USA <sup>4</sup>Portland Veterans Affairs Medical Center, Portland Oregon, 97239, USA

# These authors contributed equally to this work

\*Correspondence to: HSC at hchoi12@mgh.harvard.edu or JK at j.kim@neu.edu

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# Supplementary Tables

	Deferoxamine (Desferal)	Deferasirox (Exjade)	Deferiprone (Ferriprox, L1)	
Dose (mg kg <sup><math>-1</math></sup> day <sup><math>-1</math></sup> )	30-60	20-30	75-100	
Stoichiometry (chelator:iron)	Hexadentate (1:1)	Tridentate (2:1)	Bidentate (3:1)	
Dose route	IV, SC	Oral	Oral	
Bioavailability	Negligible	70%	40%	
Half-life in humans	20 min	8-16 h	2-4 h	
Elimination route	Urine, Fecal	Fecal (Urine <8%)	Urine	
Adverse effects	Sensorineural hearing loss, visual electroretinographic disturbances, impaired growth, bone development	Gastrointestinal bleeding, renal toxicity, hepatic dysfunction, thrombocytopenia	Agranulocytosis, neutropenia, infection, arthralgia, elevated liver enzymes	

## Supplementary Table 1 General properties of FDA-approved iron chelators

Туре	HD (nm)	Urinary excretion (%ID)	Hepatobiliary excretion (%ID)	Iron excretion in urine <sup>a</sup>	Iron excretion in feces <sup>a</sup>	Ref.
Small size EPL	5.7	82.04 at 4h	<0.2 at 24h	Y	Ν	<b>T</b>
Small size EPL	6.5	58.10 at 4h	<0.2 at 24h	Y	Ν	This study
Degradable nanogel	240	N/P	N/P	Y	Y	1
Nanogel	36	N/P	N/P	Y	Y	2
	101	N/P	N/P	Y	Y	2
Small size melanin	7.0	N/P	N/P	Y	Y	3
	5.4	8.92 at 144 h	43.92 at 144 h	N/P	N/P	
Dendrimer	9.8	4.76 at 144 h	6.45 at 144 h	N/P	N/P	4
	15.4	8.77 at 144 h	8.18 at 144 h	Y	N/P	
Degradable macromolecule	3.5	N/P	N/P	Y	Y	5
Degradable nanogel	136	N/P	N/P	N/P	N/P	6
	9.8	8.77 at 144 h	8.18 at 144 h	N/P	N/P	
Dendrimer	11.2	N/P	N/P	Y	Y	7
	15.4	4.76 at 144 h	6.45 at 144 h	N/P	N/P	
Degradable macromolecule	N/P	N/P	N/P	N/P	N/P	8

**Supplementary Table 2** Summary of polymeric nanoparticle-based iron chelators and their excretion routes

<sup>a</sup>N/P: not provided; Y: significance; N: no significance

	Convert to mg kg <sup>-1</sup>			Conversion
	$0.3 \ \mu mol \ kg^{-1}$	$1 \ \mu mol \ kg^{-1}$	$2 \ \mu mol \ kg^{-1}$	factor
Blank NP	$2.4 \text{ mg kg}^{-1}$	$8 \text{ mg kg}^{-1}$	$16 \text{ mg kg}^{-1}$	8
DFO <sub>2</sub> -NP	$2.73 \text{ mg kg}^{-1}$	9.1 mg kg <sup>-1</sup>	$18.2 \text{ mg kg}^{-1}$	9.1
DFO <sub>4</sub> -NP	$3.06 \text{ mg kg}^{-1}$	$10.2 \text{ mg kg}^{-1}$	$20.4 \text{ mg kg}^{-1}$	10.2
DFO <sub>8</sub> -NP	$3.39 \text{ mg kg}^{-1}$	$11.3 \text{ mg kg}^{-1}$	$22.6~\mathrm{mg}~\mathrm{kg}^{-1}$	11.3

Supplementary Table 3 Conversion of DFO-NP concentration to  $mg kg^{-1}$ 

**Supplementary Table 4** The amount of iron and DFO-NP excretion and molar ratio of iron/DFO-NP at 4 h post-injection (SC and IV) (n=5, mean ± SEM)

Injection route	Excretion	DFO <sub>4</sub> -NP	DFO <sub>8</sub> -NP
IV injection	Urinary excreted Fe (nmol)	4.92±0.42	6.07±1.27
	Urinary excreted DFO-NP (nmol)	14.66±3.72	12.18±2.86
	Fe/NP ratio	$0.43 \pm 0.14$	$0.51 \pm 0.08$
SC injection	Urinary excreted Fe (nmol)	$8.98 \pm 0.2.44$	8.786±1.108
	Urinary excreted DFO-NP (nmol)	3.24±0.843	$4.87 \pm 0.42$
	Fe/NP ratio	2.75±0.06	1.81±0.18

## **Supplementary Figures**



**Supplementary Figure 1.** Synthetic routes of ZW800-1 conjugation on EPL, followed by succinylation.



**Supplementary Figure 2. Ninhydrin test of ZW-EPL samples:** (a) absorbance and fluorescence spectra of DFO-NPs in water (20  $\mu$ M). (b) Photos of each sample. Green color (ZW-EPL<sup>-</sup>) represents the existence of ZW800-1, while purple color (ZW-EPL<sup>+</sup>) indicates the formation of Ruhemann's purple produced by free amines reaction with ninhydrin.



**Supplementary Figure 3. Retention time profiles by HPLC analyses:** (a) Blank NP, (b) DFO<sub>2</sub>-NP, (c) DFO<sub>4</sub>-NP, and (d) DFO<sub>8</sub>-NP.



Supplementary Figure 4. Physical stoichiometry between DFO and EPL: (a) UV absorption spectra of standard DFO solutions with concentrations ranged from 2.5 to 20  $\mu$ M. (b) Standard curve fitting with the linear regression of concentration vs. absorbance of DFO standards. (c) UV absorption spectra of DFO-NPs. (d) Physical stoichiometry calculated by using the linear regression plot shown in **b**.



**Supplementary Figure 5.** <sup>1</sup>**H-NMR spectra of DFO, blank NP, and DFO-NPs in D<sub>2</sub>O.** DFO to NP ratio was calculated by integrating of hydrogen atoms of DFO (positions 6,13 at 2.7 ppm, asterisk) and succinic acid (2.4 ppm, sky blue bar). The calculated values of DFO per chain were based on the assumption of 30 lysine units per entire blank NP.



Supplementary Figure 6. UV-vis absorption spectra of Fe(III)-titrated DFO-NPs (red lines in left panel) and titration curves of absorbance changes (A-A<sub>0</sub>) versus equivalent of iron to each DFO-NP (right panel): (a) DFO<sub>2</sub>-NPs, (b) DFO<sub>4</sub>-NPs, and (c) DFO<sub>8</sub>-NPs. Green lines indicate blank DFO-NP solutions before adding iron solution. Insets: Photos of DFO-NP solutions before and after titration with iron solution. Dotted arrows indicate inflection points where nearly saturated concentrations of iron binding each DFO-NP (20  $\mu$ M). (d) Absorbance spectra of iron in DFO-NP after removing background iron absorptions.



**Supplementary Figure 7. GFC profiles of DFO-NPs to estimate hydrodynamic diameter** (**HD**): Arrows and numbers indicate retention time maxima and corresponding HD of standard proteins, ribonuclease, 3.2 nm; ovalbumin, 6.1 nm; aldolase, 9.6 nm; ferritin, 12.2 nm.



Supplementary Figure 8. UV absorbance curves of competitive iron binding assay as function of ferrozine concentration (0 to 50  $\mu$ M). (n=3 mean ± SEM).



Supplementary Figure 9. Representative absorbance and fluorescence spectra of DFO-NPs in water.



Supplementary Figure 10. Color and NIR fluorescence images of dissected organs of DFO-NPs (0.3  $\mu$ mol kg<sup>-1</sup>) treated mice at 4 h post-injection. He, heart; Lu, lung; Li, liver; Pa/Sp, pancreas/spleen; Ki, kidney; Du, duodenum; In, intestine; Mu, muscle. Scale bar: 1 cm.



**Supplementary Figure 11. Representative NIR images of (a) serum and (b) urine samples** from DFO-NP treated mice and standard sample of DFO-NP with a serial dilution to quantify the amount of DFO-NPs in each sample.



Supplementary Figure 12. Evaluation of excretion route of DFO-NPs. NIR images of abdominal cavity of CD-1 mice injected with (a) DFO4-NP (2  $\mu$ mol kg<sup>-1</sup>) and (b) DFO8-NP (2  $\mu$ mol kg<sup>-1</sup>). He, heart; Lu, lung; Li, liver; Pa/Sp, pancreas/spleen; Ki, kidney; Du, duodenum; In, intestine; Mu, muscle. (c, d) Quantitative analysis of fecal sample collected at 4 h and 24 h post injection. (n=3; mean ± SEM) (e) H&E staining and NIR fluorescence microscope images of kidney, liver, spleen, and heart. NIR fluorescence images indicate that DFO-NP can be excreted only through kidney filtration. Scale bar: 100 µm.



Supplementary Figure 13. Dose dependency on biodistribution PK of DFO-NPs: Color and NIR fluorescence images of mouse abdominal cavity and dissected organs of (a) DFO<sub>4</sub>-NP and (b) DFO<sub>8</sub>-NP. SBR of each organ against Mu. He, heart; Lu, lung; Li, liver; Pa/Sp, pancreas/spleen; Ki, kidney; Du, duodenum; In, intestine; Mu, muscle. Blood clearance curves of DFO-NPs (right, bottom). (n=3; mean  $\pm$  SEM)



**Supplementary Figure 14. Purification and analyses of collected urine samples:** (a) Color and NIR image of collected urine from DFO<sub>8</sub>-NP treated mice at 4 h post-injection, first flow-through, 1<sup>st</sup> FT; after washing with water 3 times, and retentive, RT after filtration with ultrafiltration unit (3,000 Da cutoff). (b) Retention time profile of DFO<sub>8</sub>-NP by HPLC analysis. (c) Mass spectra of reference DFO sample in urine and (d) 1<sup>st</sup> FT of urine from DFO<sub>8</sub>-NP treated mice.



Supplementary Figure 15. PK and biodistribution of DFO<sub>2</sub>-NP 4 h post-SC injection: (a) NIR image of a mouse in prone position after injection. (b) Intraoperative images of abdomen and dissected organs. Urinary excretion value was  $10.72\pm3.81$  %ID. (c) Biodistribution of DFO-NP in major resected organs. SBR of each organ against muscle (Mu) (n = 3 per group, mean ± SEM). He, heart; Lu, lung; Li, liver; Pa/Sp, pancreas/spleen; Ki, kidney; Du, duodenum; In, intestine; Mu, muscle. (d) Blood clearance curve of DFO-NP.



Supplementary Figure 16. Serum ferritin, spleen iron, and ferritin levels of iron overload mice and rats after DFO-NPs treatments. (a, b) CD-1 mice fed iron overload diet and (c) Belgrade (b/b) rats were subcutaneously injected with saline, native DFO and DFO-NPs daily for 5 d. (n = 3-8 per group, mean ± SEM.)







Supplementary Figure 18. Color and NIR images of kidney sections from CD-1 mice administered with native DFO alone (8  $\mu$ mol kg<sup>-1</sup>), blank NP (1  $\mu$ mol kg<sup>-1</sup>), DFO<sub>8</sub>-NP (1  $\mu$ mol kg<sup>-1</sup> as EPL) daily SC injection for 5 days. Another CD-1 mouse was injected a single dose of DFO-NP (1  $\mu$ mol kg<sup>-1</sup>) on the 4th day.



**Supplementary Figure 19. H&E-stained sections of liver, spleen, and heart** following 5 days of daily treatments with saline, DFO, and DFO-NPs as shown in Figure 3d. Scale bar: 100 µm.



Supplementary Figure 20. Biochemical analysis of DFO-NPs in iron-overload animals. (a) Creatinine and (b) blood urea nitrogen (BUN) concentration from iron-loaded CD-1 mice treated with various concentrations of native DFO (8, 16, 80, and 160  $\mu$ mol kg<sup>-1</sup>) and DFO-NPs (2  $\mu$ mol kg<sup>-1</sup>). (n= 5; mean  $\pm$  SEM)



Supplementary Figure 21. Single dose acute toxicity test; biochemical analysis and tissue histology. (a) mouse body weight for 14 days after IV injection. (b) serum LDH, AST, and ALT levels in saline, DFO, and DFO<sub>4</sub>-NP treatment groups. (n=4 to 5 per each group; mean  $\pm$  SEM) (e) H&E staining of heart, liver, spleen, lung and kidney in each treatment group. Scale bar: 100 µm.

#### **Supplementary References**

- 1. Liu, Z., Qiao, J., Nagy, T. & Xiong, M.P. ROS-triggered degradable iron-chelating nanogels: Safely improving iron elimination in vivo. *J. Control. Release* **283**, 84-93 (2018).
- 2. Wang, Y., Liu, Z., Lin, T.M., Chanana, S. & Xiong, M.P. Nanogel-DFO conjugates as a model to investigate pharmacokinetics, biodistribution, and iron chelation in vivo. *Int. J. Pharm.* **538**, 79-86 (2018).
- 3. Yan, J. et al. Melanin nanoparticles as an endogenous agent for efficient iron overload therapy. *J. Mater. Chem. B* **4**, 7233-7240 (2016).
- 4. Hamilton, J.L. et al. In vivo efficacy, toxicity and biodistribution of ultra-long circulating desferrioxamine based polymeric iron chelator. *Biomaterials* **102**, 58-71 (2016).
- 5. Liu, Z., Lin, T.M., Purro, M. & Xiong, M.P. Enzymatically Biodegradable Polyrotaxane-Deferoxamine Conjugates for Iron Chelation. *ACS Appl. Mater. Interfaces* **8**, 25788-25797 (2016).
- 6. Liu, Z., Wang, Y., Purro, M. & Xiong, M.P. Oxidation-Induced Degradable Nanogels for Iron Chelation. *Sci. Rep.* **6**, 20923 (2016).
- 7. Imran ul-haq, M. et al. Design of Long Circulating Nontoxic Dendritic Polymers for the Removal of Iron in Vivo. *ACS Nano* **7**, 10704-10716 (2013).
- 8. Rossi, N.A. et al. In vitro chelating, cytotoxicity, and blood compatibility of degradable poly(ethylene glycol)-based macromolecular iron chelators. *Biomaterials* **30**, 638-648 (2009).