

Supplementary Figure 1 : Differential DNA denaturation 3DPCR.

A) APOBEC cytidine deaminases deaminate cytidine into uridine in single stranded DNA. **B)** APOBEC activity leads to the accumulation of GC \rightarrow AT mutations. **C)** As GC basepairs with 3 hydrogen bonds and AT with 2 hydrogen bonds, AT rich DNA requires less energy for denaturation allowing PCR amplification at lower denaturation $T_d/^\circ\text{C}$. **D)** PCR amplification with a gradient of denaturation temperatures allows to pick up AT rich APOBEC mutated DNA below the restrictive temperature of non mutated DNA, represented by the yellow dotted line.

A

Kanamycin 3D-PCR at 84.6°C

Mouse					Rabbit					Pig					Cow					Dog				
↶	T	C	G	A	↶	T	C	G	A	↶	T	C	G	A	↶	T	C	G	A	↶	T	C	G	A
T		1	0	0	T		0	0	0	T		0	0	0	T		0	0	0	T		0	0	0
C	59		0	1	C	53		1	0	C	42		0	0	C	93		0	0	C	63		0	0
G	0	0		87	G	0	0		43	G	0	0		35	G	0	0		39	G	0	0		58
A	0	0	0		A	0	0	1		A	1	0	0		A	0	0	0		A	0	0	1	
n=2430 bp					n=2565 bp					n=2430 bp					n=2565 bp					n=2160 bp				

B

Cytochrome C 3D-PCR at 82.3°C

Mouse					Rabbit					Pig					Cow					Dog				
↶	T	C	G	A	↶	T	C	G	A	↶	T	C	G	A	↶	T	C	G	A	↶	T	C	G	A
T		0	1	0	T		0	0	0	T		0	1	0	T		0	0	0	T		1	0	0
C	0		0	0	C	0		0	0	C	33		0	0	C	0		0	0	C	0		0	1
G	0	0		133	G	0	0		107	G	0	0		16	G	0	0		71	G	0	0		117
A	0	0	1		A	0	0	0		A	1	0	0		A	0	0	0		A	0	0	1	
n=1288 bp					n=1656 bp					n=920 bp					n= 1472 bp					n=1288 bp				

C

CMYC 3D-PCR at 89.4°C

Mouse

↶	T	C	G	A
T		0	0	0
C	82		0	0
G	0	0		0
A	0	0	0	

n=1616 bp

D

HIV-1 V1V2 3D-PCR at 81.2°C

Mouse dC

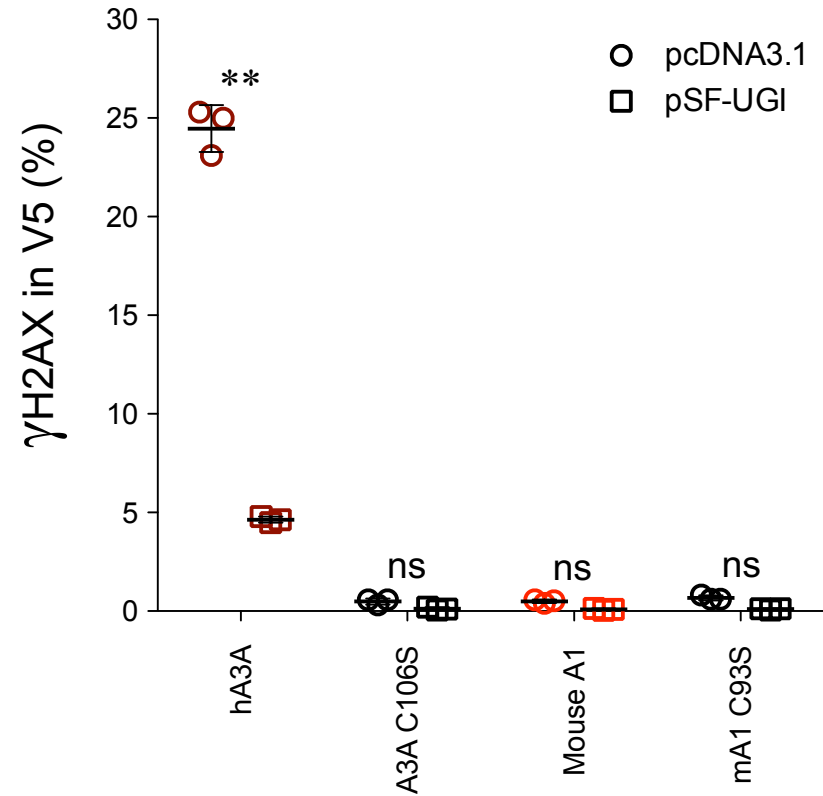
Mouse 5MedC

Mouse dC					Mouse 5MedC				
↶	T	C	G	A	↶	T	C	G	A
T		1	0	1	T		1	0	2
C	5		0	0	C	24		0	1
G	0	0		283	G	0	0		110
A	1	0	1		A	0	0	1	

n=3948 bp

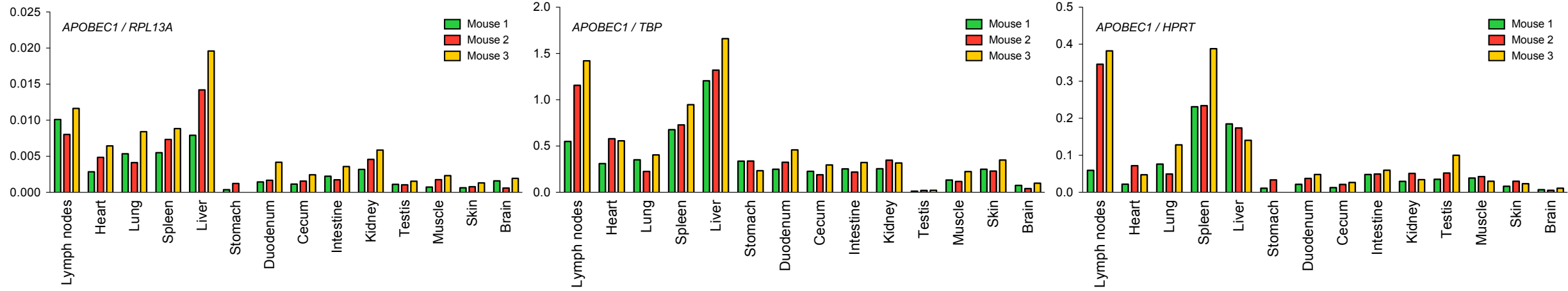
n=2820 bp

Supplementary Figure 2 : Mutation matrices of APOBEC1 mutated sequences.



Supplementary Figure 3 : Double strand breaks formation upon APOBEC transfection requires UNG.

Double strand breaks formation upon A1 transfection in QT6 cells by flow cytometry analysis of γ H2AX staining in V5 transfected cells 48 hours post-transfection. Human APOBEC3A (hA3A) was used as positive control. Circles represent data from γ H2AX staining upon transfection with pcDNA3.1 APOBEC plasmids while squares represent γ H2AX staining upon transfection with a dual promoter vector co-expressing APOBEC sequences along with the UGI UNG inhibitor. Error bars represent the standard deviations from three independent transfections. Differences between pcDNA3.1 and pSF-UGI transfections were calculated using student t test (** p<0.01).



Supplementary Figure 4 : Expression profile of APOBEC1.

APOBEC1 expression in 3 C57/BL6 mice tissues normalized on RPL13A, TBP, and HPRT reference genes.

Plasmid	Matrix	Primers
Human A1	NM_001644	For : 5'- CACC ATG ACTTCTGAGAAAGGTCC Rev : 5'- TCTCCAAGCCACAGAAGGATGTATC
Rabbit A1	NM_001082341	For : 5'- CACC ATG GCTTCGAGAAAGGTC Rev : 5'- TCTCCAAGGCACAGAAGGTTG
Mouse A1	NM_031159	For : 5'- CACC ATG AGTTCCGAGACAGGCC Rev : 5'- TTCAACCCCTGTAGCCCAAAGG
Opossum A1	NM_001032982	For : 5'- CACC ATG AATTCTAAGACAGGTCCA Rev : 5'- TCTCCAGGTCACAAATGGCTGG
Armadillo A1	XM_004455274	For : 5'- CACC ATG ACTTCTGAGACAGGTCCTTC Rev : 5'- TCTCCAAGTCATGGAAGGTTGTATT
Cow A1	XM_002687817	For : 5'- CACC ATG GCTTCTGACAGAGGTCCTCCA Rev : 5'- TCTCCAAGTCATAGGAAGTTGTACC
Pig A1	XM_003126519	For : 5'- CACC ATG GCTTCTGACAGAGGTCCTTC Rev : 5'- TCTGTAAATCACAGGTAGTTGTATC
Macaque A1	XM_005570020	For : 5'- CACC ATG ACTTCTGAGAAAGGTCCTTC Rev : 5'- TCTCCAAGTCACGGAAGGCTGTATC
Marmoset A1	XM_009003530	For : 5'- ATGACTTCTGAGAGAGGTCCTTC Rev : 5'- TCTCCAAGTCACAGAAGGCTGGATC
Hedgehog A1	XM_004717433	For : 5'- CACC ATG ACTGCTGAGAAAGGTCCTTC Rev : 5'- CTGAGTCACAGAAGGTGGTATCAAC
Dog A1	XM_543826	For : 5'- CACC ATG GCTTCTGACAAAGGTCCTTC Rev : 5'- TCTCCAAGTCACAAGAGGTTGTATC
Cat A1	XM_543826	For : 5'- CACC ATG GCTTCTGACAAAGGTCCTTC Rev : 5'- TCTCCAAGTCACAAGAGGTTGTATC

Supplementary table 1 : Compendium of primers used for APOBEC1 amplification and cloning

Plasmid	Matrix	Primers
Mouse A1 C93S	pcDNA3.1 Mouse A1	For : 5'- CTGTCCTGGAGTCCCAGCGGGGAGTGCTCCAGG Rev : 5'- CCTGGAGCACTCCCCGCTGGGACTCCAGGACAG

Supplementary table 2 : Primers used for mutagenesis

Primer	Application	Sequence	Cycling conditions
Kanamycin out fwd	kanamycin PCR	5'- CGACCTGTCCGGTGCCCTGAATGAA	95°C 5 min 37x (95°C 30 sec, 60°C 30 sec, 72°C 1 min), 72°C 10 min
Kanamycin out rev	kanamycin PCR	5'- GACGGCCACAGTCGATGAATCCAGAA	
Kanamycin in fwd	kanamycin 3DPCR	5'- TCATCTCACCTTGCTCCTGCCGAGAA	81-89°C 5 min 42x (81-89°C 1 min, 60°C 45 sec, 72°C 2 min), 72°C 10 min
Kanamycin in rev	kanamycin 3DPCR	5'- CTTGTCAGATCATCCTGATCGACAA	
CytC out fwd	Cytochrome C PCR	5'- CATCCAAAGCAGGTCAACCTATGACAA	95°C 5 min 35x (95°C 30 sec, 60°C 30 sec, 72°C 1 min), 72°C 10 min
CytC out rev	Cytochrome C PCR	5'- TCTTGGTTAAGAGGTGTGGTCTGTAA	
CytC in fwd	Cytochrome C 3DPCR	5'- CCTCTACACAGCGTAGACTTACATCAA	80-86°C 5 min 35x (80-86°C 1 min, 60°C 30 sec, 72°C 2 min, 72°C 10 min)
CytC in rev	Cytochrome C 3DPCR	5'- AAGACCTCGTTTAGCCATTCATACAA	
QT6 cMyc out fwd	cMyc PCR	5'- GATGCCGCTCAGCGCCAGCCTCCCCAGCAA	95°C 5 min 40x (95°C 1 min, 58°C 45 sec, 72°C 2 min), 72°C 10 min
QT6 cMyc out rev	cMyc PCR	5'- GCAGTCTGGATGATGATGGATTTGACGAA	
QT6 cMyc in fwd	cMyc 3DPCR	5'- TTCGAGGAGGAGGAGGAGAACTTCTA	87-95°C 5 min 42x (87-95°C 1 min, 60°C 1 min, 72°C 2 min), 72°C 10 min
QT6 cMyc in rev	cMyc 3DPCR	5'- GATTGCTCGTCCGGTCCGAGATGAA	

Supplementary Table 3 : Compendium of primers and PCR conditions used for nested PCR/3DPCR amplifications

Target	UPL Probe	Primers
Mouse APOBEC1	27	For : 5'-CCCATGAGCGTTGGATTC Rev : 5'-TCAACCACGGGCAGTCTT
Mouse ribosomal protein L13A (RPL13A)	108	For : 5'- CCCTCCACCCTATGACAAGA Rev : 5'- GCCCCAGGTAAGCAAACCTT
Mouse hypoxanthine guanine phosphoribosyl transferase (HPRT)	95	For : 5'- TCCTCCTCAGACCGCTTTT Rev : 5'- CCTGGTTCATCATCGCTAATC
Mouse TATA box binding protein (TBP)	107	For : 5'- GGCGGTTTGGCTAGGTTT Rev : 5'- GGGTTATCTTCACACACCATGA

Supplementary Table 4 : Compendium of primers and UPL probes used for mouse transcriptome analysis.