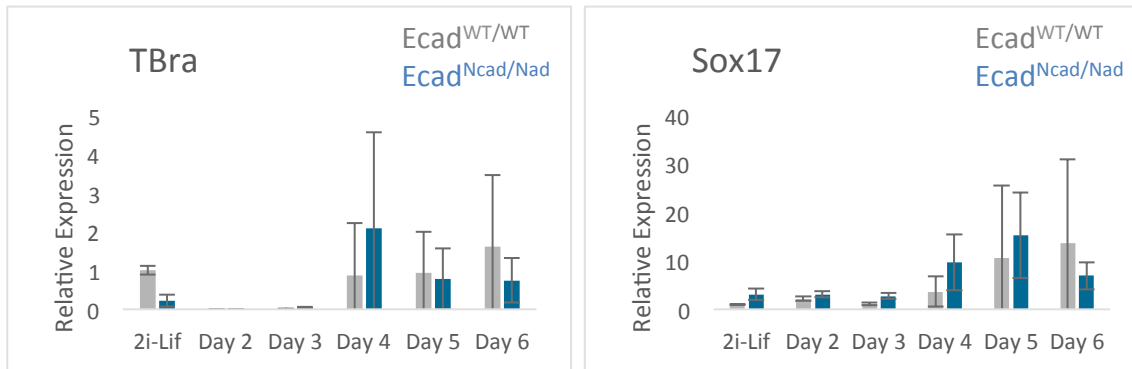


Figure S1: Single-cell views of protein co-expression in EpiSCs and during neural differentiation. White arrowheads point out individual cells of the given protein expression profile. Percentages below images indicate the proportion of cells of a given identity out of all cells analysed (manual quantification). Scale bars=25µm. **A.** EpiSCs stained for E-cadherin (green), N-cadherin (red), the pluripotency marker Oct4 (white) and nuclear envelope marker LaminB1 (blue). N=2596 cells from three biological replicates **B.** Cells on day four of neural differentiation from a 2i-Lif starting population stained for E-cadherin (green), N-cadherin (red), Sox1-GFP (shown in white), and nuclear envelope marker LaminB1 (blue). N=2275 cells from three biological replicates.

A: Ecadh^{Ncadh/Ncadh} cells



B: Dox-inducible N-Cadherin

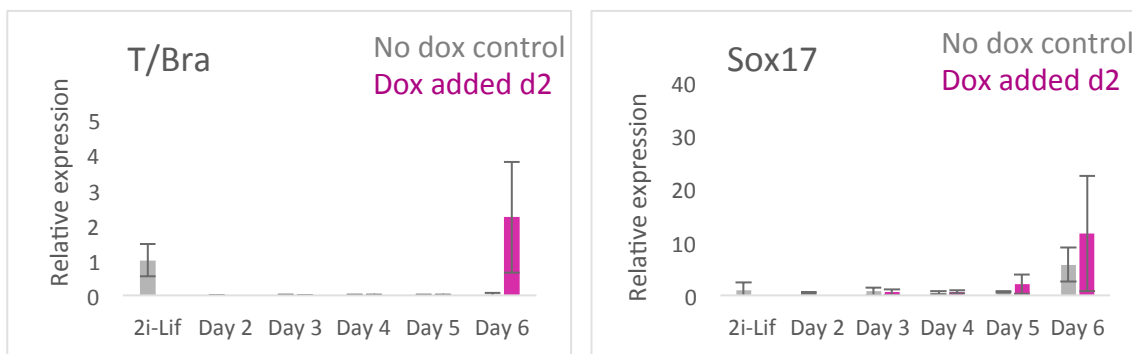


Figure S2: Cadherin switching has no clear effect on mesoderm or endoderm markers in the context of neural differentiation

A: qPCR analysis of *Ecadh*^{Ncadh/Ncadh} cells during successive days of neural differentiation. N=3. Values normalised to control cell line on D0.

B: qPCR analysis of inducible N-cadherin overexpressing cells during neural differentiation; Dox added on day 2. N=9 (three biological replicates of three independent clones). Values normalised to control cell line on D0

Error bars=SD.

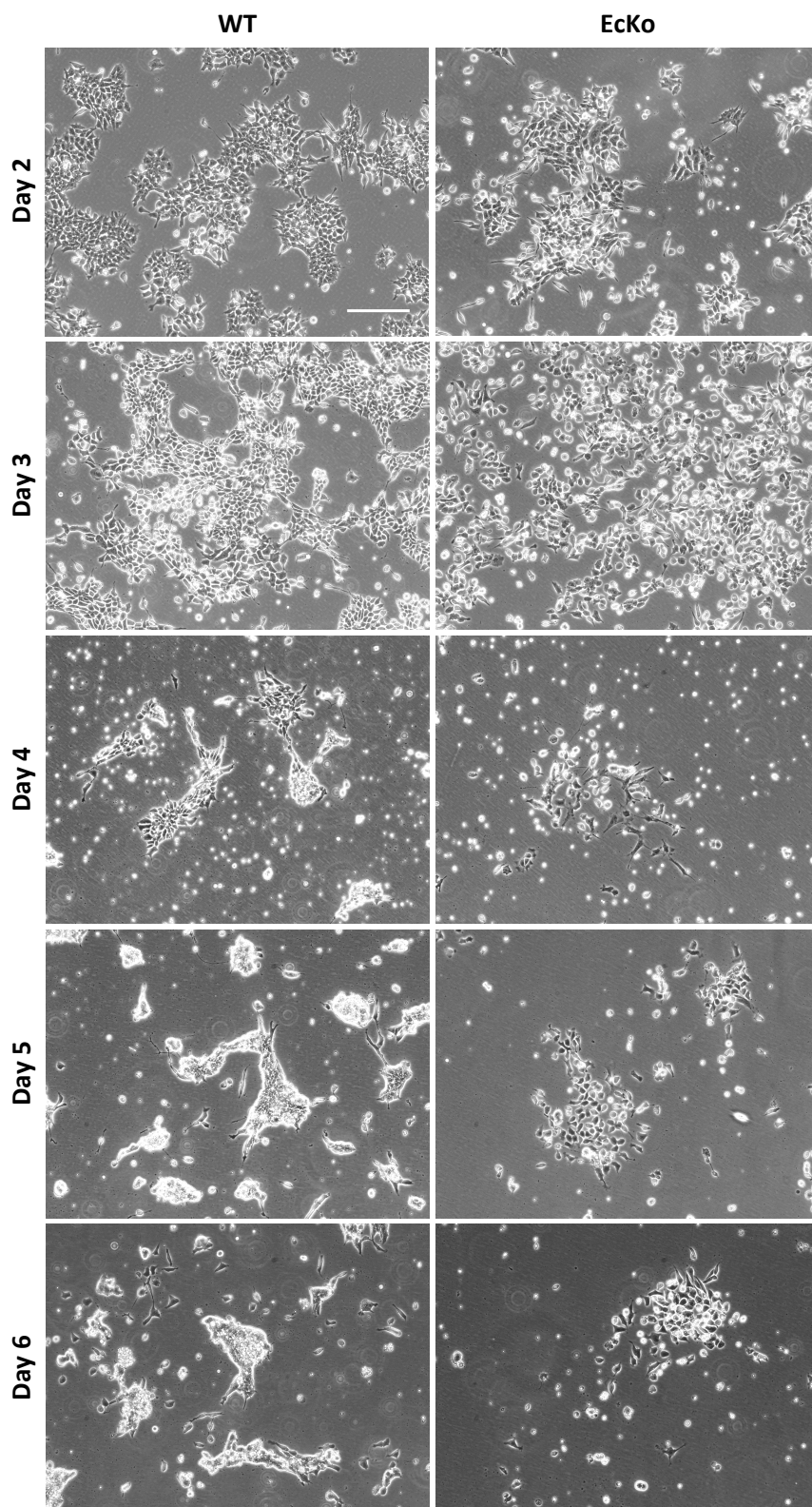


Figure S3: $Ecad^{-/-}$ cells die after prolonged culture in N2B27. Phase contrast images of $Ecad^{Flx/Flx}$ and $Ecad^{-/-}$ cells on days 2-6 of neural differentiation from a 2i-Lif starting population. Scalebar= 100 μ m.

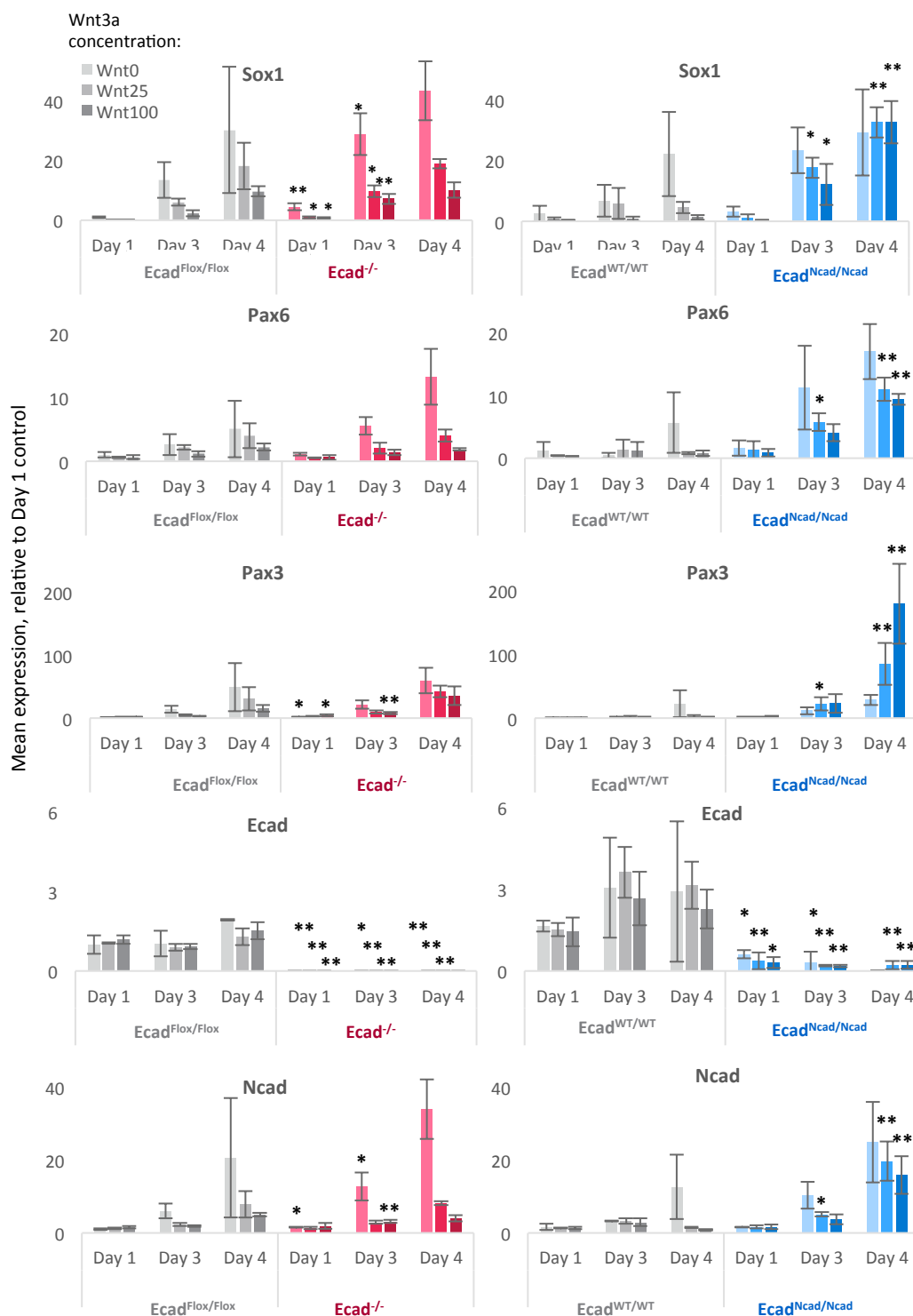


Figure S4: Effects of cadherin switching on β -catenin and WNT signalling. qPCR analysis of Ecad^{Flox/Flox} and Ecad^{Ncad/Ncad} cells during neural differentiation in increasing concentrations of Wnt3a; bars denote mean expression relative to the Day 1 condition of the relevant control cell line (grey). Asterisks denote significant difference compared to the paired control cell line in the same condition. N=3 biological replicates. Error bars=SD, *p<0.05, **p<0.01, paired T-test.

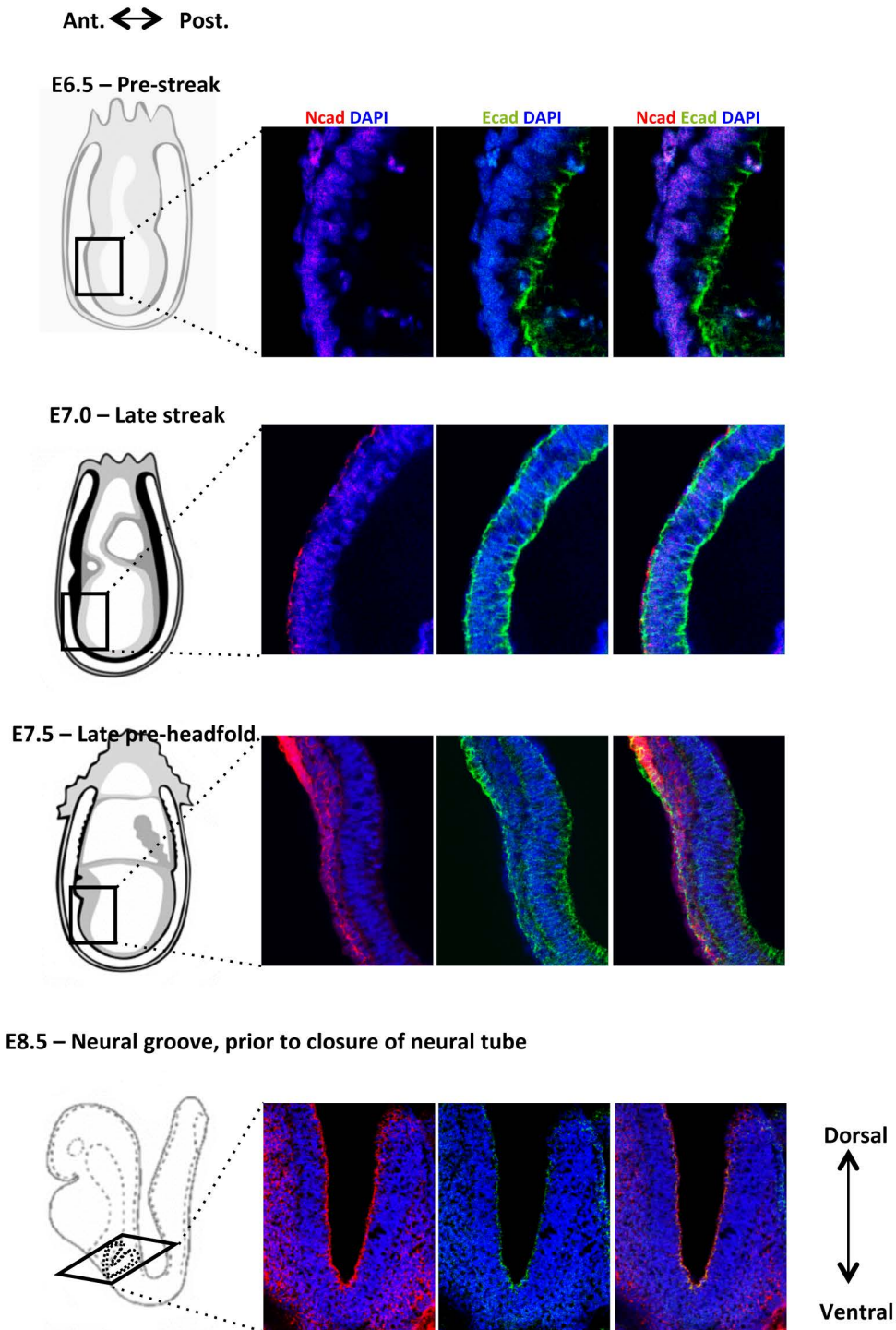


Figure S5: Cadherin switching in vivo. E-Cadherin (green) and N-Cadherin (red) immunostaining at various stages of mouse development as indicated. Embryo drawings are adapted from the EMAP eMouse Atlas Project (<http://www.emouseatlas.org>).

Table S1: qPCR primer sequences.

Primers used with the Universal Probe Library qPCR system from Roche.

Target gene	Forward sequence	Reverse sequence	UPL probe number
<i>Ap2a</i>	CCGGGTATTAACATCCCAGAT	CCGAAGAGGTTGTCCTTGTTA	94
<i>Axin2</i>	TGGGGAGCAGTTTTGTGC	CGGCTGACTCGTTCTCCT	96
<i>Cdh1</i>	ATCCTCGCCCTGCTGATT	ACCACCGTTCTCCTCCGTA	18
<i>Cdh2</i>	GCCATCATCGCTATCCTTCT	CCGTTTCATCCATACCACAAA	18
<i>Dusp4</i>	GCCTGGCCTACCTGATGAT	GCTGCTTGACGAACTCAAAA	25
<i>Etv4</i>	GGGTACCTTGGTGAGCACAG	CCCTGAGGAGATGTGAAGGA	66
<i>Nanog</i>	CCTCCAGCAGATGCAAGAA	GCTTGCACTTCATCCTTTGG	25
<i>Pax3</i>	AAAAGGCTAAACACAGCATCG	CAATATCGGAGCCTTCATCTG	110
<i>Pax6</i>	GTTCCCTGTCCTGTGGACTC	ACCGCCCTTGGTTAAAGTCT	78
<i>Pou3f1</i>	CTCAAGCCGCTGCTCAAC	CGCGATCTTGCCAGGTT	25
<i>Pou5f1</i>	GTTGGAGAAGGTGGAACCAA	CTCCTTCTGCAGGGCTTTC	95
<i>SDHA</i>	CAGTTCCACCCCACAGGTA	TCTCCACGACACCCTTCTGT	71
<i>Sox1</i>	GTGACATCTGCCCCATC	GAGGCCAGTCTGGTGTCAG	60
<i>Sox17</i>	CACAACGCAGAGCTAAGCAA	CGCTTCTCTGCCAAGGTC	97
<i>T</i>	ACTGGTCTAGCCTCGGAGTG	TTGCTCACAGACCAGAGACTG	27
<i>TBP</i>	GGGGAGCTGTGATGTGAAGT	CCAGGAAATAATTCTGGCTCA	97
<i>Ywhaz</i>	TTACTTGGCCGAGGTTGCT	TGCTGTGACTGGTCCACAAT	9

Table S2: RPPA antibodies

All antibodies were raised in rabbit

Epitope	Supplier	Catalogue no.
Akt	Cell Signaling Technologies	9272
Akt P Ser473	Cell Signaling Technologies	4060
Akt P Thr308	Cell Signaling Technologies	2965
beta-actin	Cell Signaling Technologies	4970
beta-Catenin	Cell Signaling Technologies	9562
beta-Catenin P Ser33,Ser37,Thr41	Cell Signaling Technologies	9561
beta-Catenin P Thr41,Ser45	Cell Signaling Technologies	9565
Caspase 3	Cell Signaling Technologies	9662
Caspase 3 cleaved	Cell Signaling Technologies	9664
c-Jun N-term	Epitomics	1254-1
c-Jun P Ser73	Cell Signaling Technologies	9164
E-Cadherin	Cell Signaling Technologies	3195
GSK-3-alpha/beta P Ser21/Ser9	Cell Signaling Technologies	9331
GSK-3-beta	Cell Signaling Technologies	9315
GSK-3-beta P Ser9	Cell Signaling Technologies	9336
IGF-1R beta	Cell Signaling Technologies	3027
ILK1 (4G9)	Cell Signaling Technologies	3856
Integrin alpha 4	Cell Signaling Technologies	4600
Integrin Beta 1 [EP1041Y]	Abcam	ab52971
Integrin beta3	Cell Signaling Technologies	4702
Integrin beta4	Cell Signaling Technologies	4707
IRS-1	Cell Signaling Technologies	2382
JAK1	Cell Signaling Technologies	3332
JAK1 P Tyr1022,Thr1023	Invitrogen (Biosource)	44-422G
MAPKAPK-2	Epitomics	1497-1
MAPKAPK-2 P Thr334	Cell Signaling Technologies	3041
MEK1/2	Cell Signaling Technologies	9122
MEK1/2 P Ser217/221	Cell Signaling Technologies	9154
MEK6 [EP558Y]	Abcam	ab52937
mTOR	Cell Signaling Technologies	2972
mTOR P Ser2448	Cell Signaling Technologies	2971
NFkB p105/p50	Calbiochem	GTX110585
NFkB p65 Ser536	Cell Signaling Technologies	3033
p38 MAPK	Cell Signaling Technologies	9212
p38 MAPK PThr180,Tyr182	Cell Signaling Technologies	9211
p44/42 MAPK (ERK1/2)	Cell Signaling Technologies	9102
p44/42 MAPK (ERK1/2) P Thr202/Thr185,Tyr204/Tyr187	Cell Signaling Technologies	4370
p90 S6 kinase (Rsk1-3)	Santa Cruz	sc-231
PDK-1	Cell Signaling Technologies	3062
PDK-1 P Ser241	Cell Signaling Technologies	3061
PKA	Abcam	ab26322
Prohibitin	Santa Cruz	sc-28259
Raf P Ser338	Cell Signaling Technologies	9427
Rsk2 Pser 227	Cell Signaling Technologies	3556
Slug (C19G7	Cell Signaling Technologies	9585
Smad1/5 P Ser463/Ser465	Cell Signaling Technologies	9516
Smad2 P Ser465,Ser467	Cell Signaling Technologies	3108
Smad2/3 P Ser465/Ser423,Ser467/Ser425	Cell Signaling Technologies	8828
Smad3 P Ser423,Ser425	Cell Signaling Technologies	9520
Stat3	Cell Signaling Technologies	12640
Stat3 P Y705	Cell Signaling Technologies	9131
Stat5	Invitrogen (Biosource)	44-368G
Stat5 P Tyr694	Cell Signaling Technologies	9351
Stat6	Cell Signaling Technologies	9362
Stat6 P Tyr641	Cell Signaling Technologies	9361
Tsc-2 (Tuberin)	Cell Signaling Technologies	3612
Tsc-2 (Tuberin) P Thr1462	Cell Signaling Technologies	3617
YAP P Ser127	Cell Signaling Technologies	4911
YAP1 [EP1674Y]	Abcam	ab52771
Zyxin	Cell Signaling Technologies	3553

Table S3: Immunocytochemistry and flow cytometry antibodies

All antibodies were diluted to the specified concentration in blocking buffer.

Epitope recognised	Clone	Host species	Dilution factor	Supplier	Cat. number
β -catenin (active), dephosphorylated on Ser37 or Thr41	8E7	Mouse	1:1000	Millipore	05-665
E-cadherin	DECMA-1	Rat	1:200	Sigma	U3254
E-cadherin, eFluor660-conjugated	DECMA-1	Rat	1:300	eBioscience	50-3249-82
GFP	Polyclonal	Chicken	1:1000	Abcam	13970
HA	HA-7	Mouse	1:1000	Sigma	H3663
Lamin β 1	Polyclonal	Chicken	1:1000	Abcam	Ab90169
Lamin β 1	Polyclonal	Rabbit	1:1000	Abcam	Ab16048
N-cadherin	32	Mouse	1:200	BD	610920
N-cadherin, AlexaFluor488-conjugated	Polyclonal	Sheep	1:50	R&D	FAB6426G
Oct4	N-19	Goat	1:200	Santa Cruz	SC-8628
Sox1	N23-844	Mouse	1:200	Pharmigen	560749

Table S4. Significant changes in gene expression 48 h after N-cadherin overexpression during neural differentiation. Cells were cultured for 48 h in neural differentiation conditions when N-cadherin overexpression was induced by addition of Dox. RNA samples were collected for Nanostring gene expression analysis 48 h later. The analysis included 770 genes involved in cellular signalling pathways. Values show mean enrichment compared to un-induced controls. $N=3$ biological replicates.

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