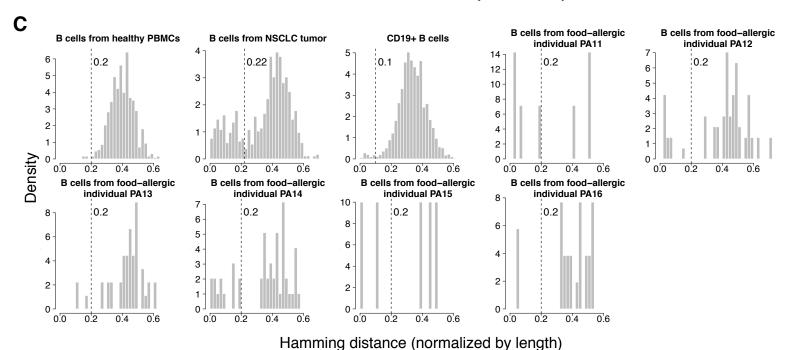
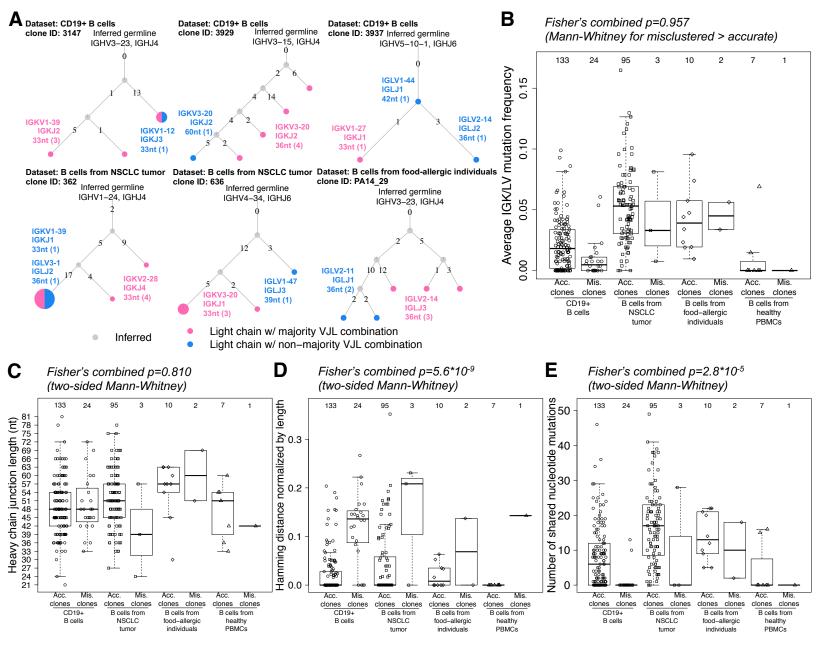
			Number of Cells				
Dataset Description	Platform	Source	Total	BCR contig(s) available	After filtering BCR contigs	After filtering for cells w/ 1 VH and ≥1 VL	After filtering for cells w/ transcriptomic profile of a B cell
PBMCs	V(D)J +	AllCells					
(unsorted) of a	5' Gene	(Catalog no.	7726	1321	1255	1196	1115
healthy donor	Expression	PB001)					
Cells (unsorted)							
from a squamous	V(D)J +	A fresh					
non-small cell	5' Gene	surgical	7802	2953	1817	1530	1388
lung carcinoma	Expression	resection					
(NSCLC) tumor							
CD19+ B cells		AllCells					
isolated from	Direct Ig	(Catalog no.	9465	9465	9219	8454	_
PBMCs of a	Enrichment	PB010-0)	7.05	7.00	,21,	0.15.1	
healthy donor							
B-lymphoblastoid	Direct Ig	Coriell					
cell line	Enrichment	(Catalog no.	854	854	819	799	-
GM12878	Linicillicit	GM12878)					

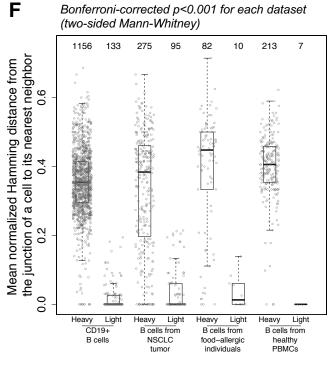
B Cell counts for Croote et al. 2018 dataset

	Number of B cells					
Subject	As in Croote et al., 2018	After filtering BCR contigs	After filtering for cells w/ 1 VH and ≥1 VL			
PA11	59	59	59			
PA12	267	265	265			
PA13	202	200	200			
PA14	212	211	211			
PA15	110	110	110			
PA16	123	123	123			
Total	971	968	968			

Supplemental Figure 1. A. Summary of datasets from 10x Genomics. B. Summary of B cells from foodallergic individuals. C. Distance-to-nearest neighbor plot to guide choosing a fixed distance threshold for inferring clones via hierarchical clustering. Cells were first partitioned into groups sharing the same IGHV gene, IGHJ gene, and heavy chain junction length. Within each partition, for each heavy chain junction sequence, the smallest non-zero nucleotide Hamming distance to other heavy chain junction sequences in the same partition was calculated. A threshold, indicated by a dashed line, was chosen via manual inspection to separate the bimodal distribution representing cells that were likely to be clonally related from unrelated ones.







Supplemental Figure 2. A. Maximum-parsimony trees based on IMGTnumbered nucleotides 1-312 of IGHV for misclustered clones with >2 cells. Clonal members are shown as nodes and colored according to whether their light chain carries the clonal majority VJL combination. In case of a tie, a majority is designated arbitrarily (such as in Clone 3937). Cells with identical IGHV positions 1-312 share the same node, with node size proportionate to the number of cells at that node. Edge labels represent the number of mutations accumulated from one node to the next. B. Average IGK/LV mutation frequency across cells in accurate clones compared with that across cells expressing clonal non-majority VJL combination in misclustered clones. C. Heavy chain junction lengths of accurate clones versus misclustered clones. D. The maximum pairwise distance between heavy chain junction sequences in accurate clones, versus the minimum pairwise distance between cells expressing inconsistent light chains in misclustered clones. E. The minimum pairwise shared IGHV mutations in accurate clones, versus the maximum pairwise shared IGHV mutations between cells expressing inconsistent light chains in misclustered clones. F. Means of the distributions of the smallest normalized Hamming distance between heavy chain junctions in partitions of cells with the same heavy chain VJL combinations, and between light chain junctions in accurate clones inferred via spectral clustering. A distribution was calculated for each heavy chain-based partition or clone. The mean of each distribution is visualized. Acc. = Accurate. Mis. = Misclustered.