Supplemental Table S1. Oligonucleotides, plasmids and strains used in this work.

Primers	Sequence (5'to 3')	Purpose	Source
BaF	GCACCAAATGCAGCAACTCA	<i>blp1_{IC1}</i> gene allele detection	This work
BaR	TGCTCGTTACAGAAAGCGGT		
BaF	GCACCAAATGCAGCAACTCA	<i>blp1_{ICII}</i> gene allele detection	This work
Blp2R	TACTCGTTACGGAAGGTTCT		
BaprB	GATCAGTATGGTCAATTAT	Expression of <i>blp1</i> gene	This work
BapRaiR	CAGCGTGATCAACTACAACTAC		
Fjgms	AGGACAGAAATGCCTCGAC	<i>aac(3)-I</i> amplification	[1]
Rjgms	ATCTCGGCTTGAACGAATT		
Bav1	CATTACAATGCTTAAGCTA	<i>blp1</i> upstream region	This work
Bav2	CATATTAAACAATAATTTGCTTCCTTCATCGGTAGAAAC	amplification; additional sequence underlined	
Bav3	GCAAATTATTGTTTAATATG	<i>blp1</i> downstream region	This work
Bav4	<u>GTCGAGGCATTTCTGTC</u> CTGGTTTAGCAATAGAACGGAT	amplification; additional sequence	THIS WORK
Dave	<u>diedAddeAlHeidie</u> elddilladeAllAdaaeddal	underlined	
Bpat	AAGAGACTTTTAATAGGCGAT	<i>blp1</i> gene deletion confirmation	This work
		with Bav4 primer	
BlpKlF	ACTGGAATTCCTATAAGACACTTAACTTATGAG	<i>blp1</i> gene with upstream region	This work
BlpKlR	AGTCGGTACCAATTAACTCCATCAGACTTAGTC	amplification	
Aac3I_seqR	CGAAGTCGAGGCATTTCTGT	Confirmation of transformed	This work
AcORI_seqR	AGGCTGTTGATAACTTTTGGAA	bacteria	
BldBamF	TTCTA <u>GGATCC</u> GAATATTGCTCCAGTAATT	<i>blp1</i> ₂₆₅₂₋₃₃₆₂ cloning into	This work
BlXhR	TTCTA <u>CTCGAG</u> TTAAACAATAATTTGCTGG	expression vector; restrictions	
		sites underlined	

sites under med		
Relevant characteristics	Source	
pUC19 derivative with <i>sacB</i> gene from <i>Bacillus</i> sp.; for the generation of	[1]	
e		
pUC19 derivative with <i>aac(3)-I</i> gentamicin aminoglycoside acetyltransferase	This work	
cassette and ori site from <i>Acinetobacter</i> ; for the complementation experiments		
pUC19_sacB derivative with upstream and downstream regions of A. baumannii	This work	
<i>blp1</i> gene and <i>aac(3)-I</i> gentamicin aminoglycoside acetyltransferase cassette from		
	Relevant characteristicspUC19 derivative with sacB gene from Bacillus sp.; for the generation of markerless gene deletion mutants pUC19 derivative with aac(3)-I gentamicin aminoglycoside acetyltransferase cassette and ori site from Acinetobacter; for the complementation experiments pUC19_sacB derivative with upstream and downstream regions of A. baumannii	

p <i>blp1</i> _{IC1}	clinical <i>A. baumanni</i> strain; for the generation of markerless gene deletion mutants pUC19_gm_AcORI derivative with <i>blp1</i> gene along with upstream region (plausible promoter) from Ab _{IC1} strain; for the complementation experiments	This work
р <i>blp1</i> _{IC II}	pUC19_gm_AcORI derivative with $blp1$ gene along with upstream region (plausible promoter) from Ab _{IC II} strain; for the complementation experiments	This work
pET-28b	Protein expression vector	Novagen
pET-His- Blp1 ₂₆₅₂₋₃₃₆₂	Blp1 ₂₆₅₂₋₃₃₆₂ expression plasmid, His-tag fused N-terminally to protein	This work
Strains	Relevant characteristics	Source
Acinetobacter	Representative IC I clone strain ^a ; MDR strain, gentamicin sensitive	[2]
<i>baumannii</i> Ab _{IC I}		
Acinetobacter	Representative IC II clone strain ^a ; MDR strain, gentamicin sensitive	[2]
<i>baumannii</i> Ab _{IC II}		
Ab _{IC I} ⊿blp1	<i>blp1</i> gene-negative mutant of <i>A. baumannii</i> strain Ab _{IC I} ; markerless	This work
Ab _{IC II} ⊿blp1	<i>blp1</i> gene-negative mutant of <i>A. baumannii</i> strain Ab _{IC II} ; markerless	This work
Ab _{IC I} ⊿ <i>blp1</i> ::p	Ab _{IC I} <i>Ablp1</i> strain with pUC19_gm_AcORI plasmid	This work
Ab _{IC II} ⊿ <i>blp1</i> ::p	Ab _{IC II} <i>dblp1</i> strain with pUC19_gm_AcORI plasmid	This work
Ab _{IC I} ⊿blp1∷pblp1 _{IC I}	Ab _{IC I} $\Delta blp1$ strain complemented with pblp1 IC I	This work
Ab _{IC I} ⊿ <i>blp1∷pblp1</i> _{IC II}	Ab _{IC I} $\Delta blp1$ strain complemented with pblp1 IC II	This work
Ab _{IC II} ⊿blp1∷pblp1 _{ICI}	Ab _{IC II} $\Delta blp1$ strain complemented with pblp1 IC I	This work
Ab _{IC II} ⊿ <i>blp1</i> ∷pblp1 _{IC II}	Ab _{IC II} $\Delta blp1$ strain complemented with pblp1 IC II	This work
Escherichia coli OP50	Wild type, bacterial food source for C. elegans	[3]
<i>E. coli</i> JM107	endA1 glnV44 thi-1 relA1 gyrA96 Δ (lac-proAB) [F_ traD36 proAB+ lacIq lacZ Δ M15] hsdR17(RK- mK+) λ -	[4]
<i>E. coli</i> ArcticExpress (DE3)	B F ⁻ ompT hsdS($r_B^- m_B^-$) dcm ⁺ Tet ^r gal λ (DE3) endA Hte [cpn10 cpn60 Gent ^r]	Thermo Fisher Scientific

3 of Health Sciences Kauno Klinikos Hospital in 2010.

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