

## Membrane-bound Gaussia luciferase as a tool to track shedding of membrane proteins from the surface of extracellular vesicles

Mikołaj Piotr Zaborowski<sup>1,2,3,\*</sup>, Pike See Cheah<sup>1,2,4</sup>, Xuan Zhang<sup>1,2</sup>, Isabella Bushko<sup>1,2</sup>, Kyunghoon Lee<sup>5</sup>, Alessandro Sammarco<sup>1,2,6</sup>, Valentina Zappulli<sup>1,2,6</sup>, Sybren Lein Nikola Maas<sup>1,2,7</sup>, Ryan M. Allen<sup>8</sup>, Purva Rumde<sup>1,2</sup>, Bence György<sup>1,2,9</sup>, Massimo Aufiero<sup>1,2</sup>, Markus W. Schweiger<sup>1,2</sup>, Charles Pin-Kuang Lai<sup>1,2,10</sup>, Ralph Weissleder<sup>5,11,12</sup>, Hakho Lee<sup>5,11</sup>, Kasey C. Vickers<sup>8</sup>, Bakhos A. Tannous<sup>1,2,11</sup>, Xandra O. Breakefield<sup>1,2,11,\*</sup>

<sup>1</sup>Department of Neurology, Massachusetts General Hospital, Charlestown, MA 02129 USA

<sup>2</sup>Program in Neuroscience, Harvard Medical School, Boston, MA 02115 USA

<sup>3</sup>Department of Gynecology, Obstetrics and Gynecologic Oncology, Division of Gynecologic Oncology, Poznan University of Medical Sciences, 60-535 Poznań, Poland

<sup>4</sup>Department of Human Anatomy, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, 43400, Serdang, Selangor, Malaysia

<sup>5</sup>Center for Systems Biology, Massachusetts General Hospital, Boston, MA 02114 USA

<sup>6</sup>Department of Comparative Biomedicine and Food Science, University of Padua, Padua, Italy

<sup>7</sup>Department of Neurosurgery, UMC Utrecht Brain Center, University Medical Center, Utrecht University, 3584 CX, Utrecht, The Netherlands

<sup>8</sup>Department of Medicine, Vanderbilt University Medical Center, Nashville, TN 37232 USA

<sup>9</sup>Institute of Molecular and Clinical Ophthalmology Basel, 4031 Basel, Switzerland

<sup>10</sup>Present address: Institute of Atomic and Molecular Sciences, Academia Sinica, Taipei 10617, Taiwan

<sup>11</sup>Department of Radiology, Massachusetts General Hospital, Boston, MA 02114 USA

<sup>12</sup>Department of Systems Biology, Harvard Medical School, Boston, MA 02115 USA

\*Corresponding authors: [breakefield@hms.harvard.edu](mailto:breakefield@hms.harvard.edu), [mikolaj.zaborowski@gmail.com](mailto:mikolaj.zaborowski@gmail.com) (ORCID: 0000-0002-4400-6688)

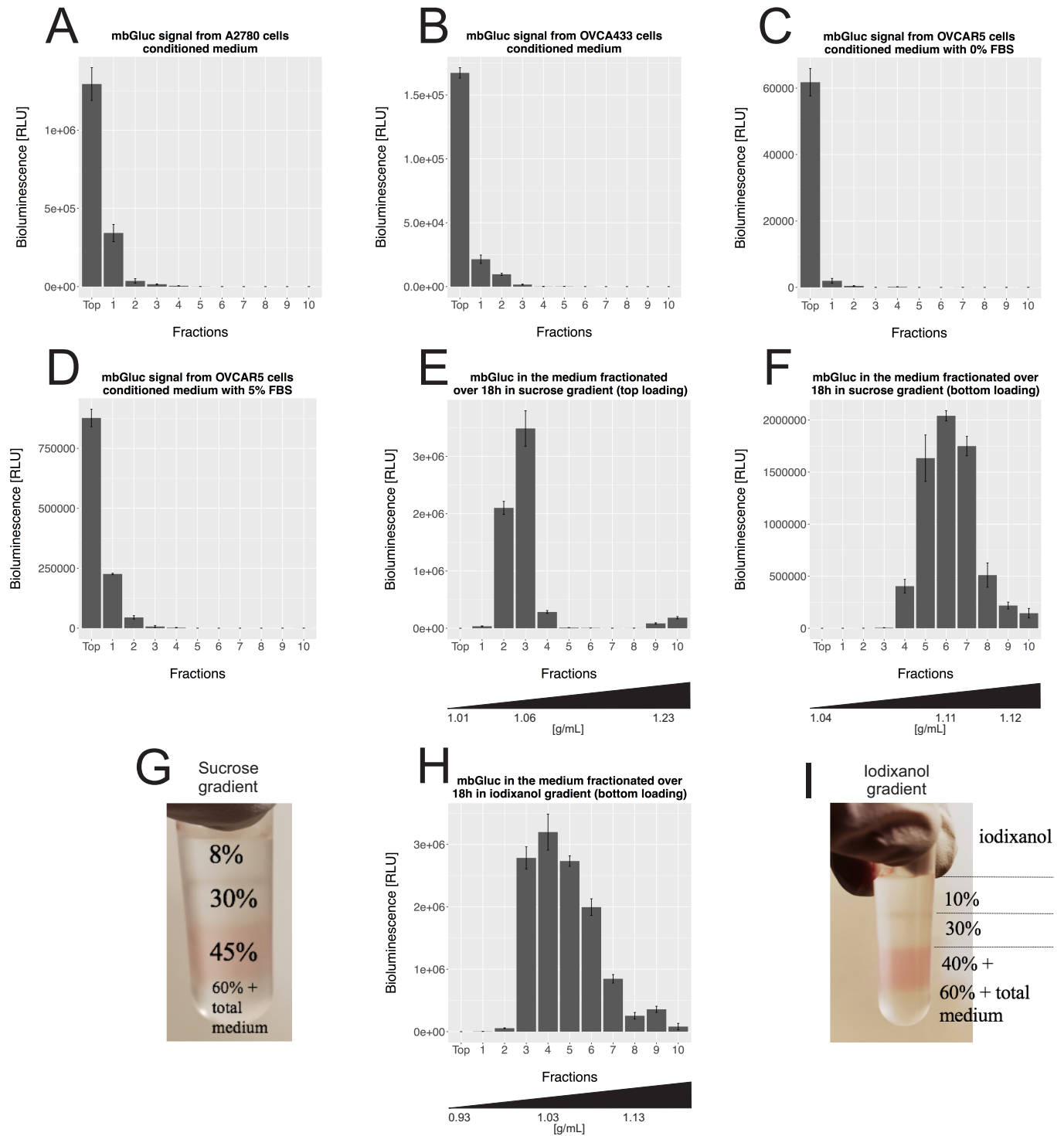
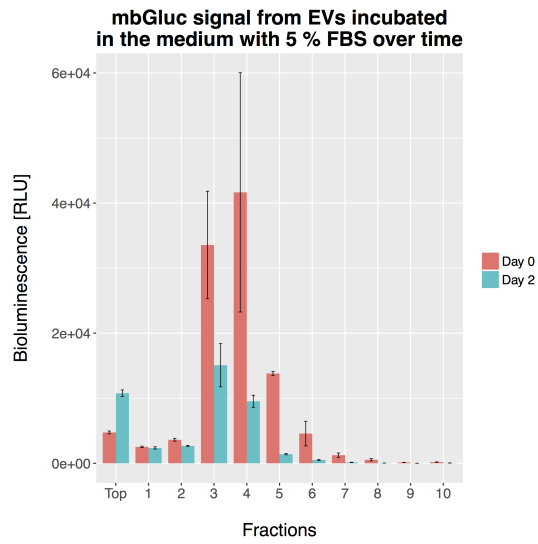


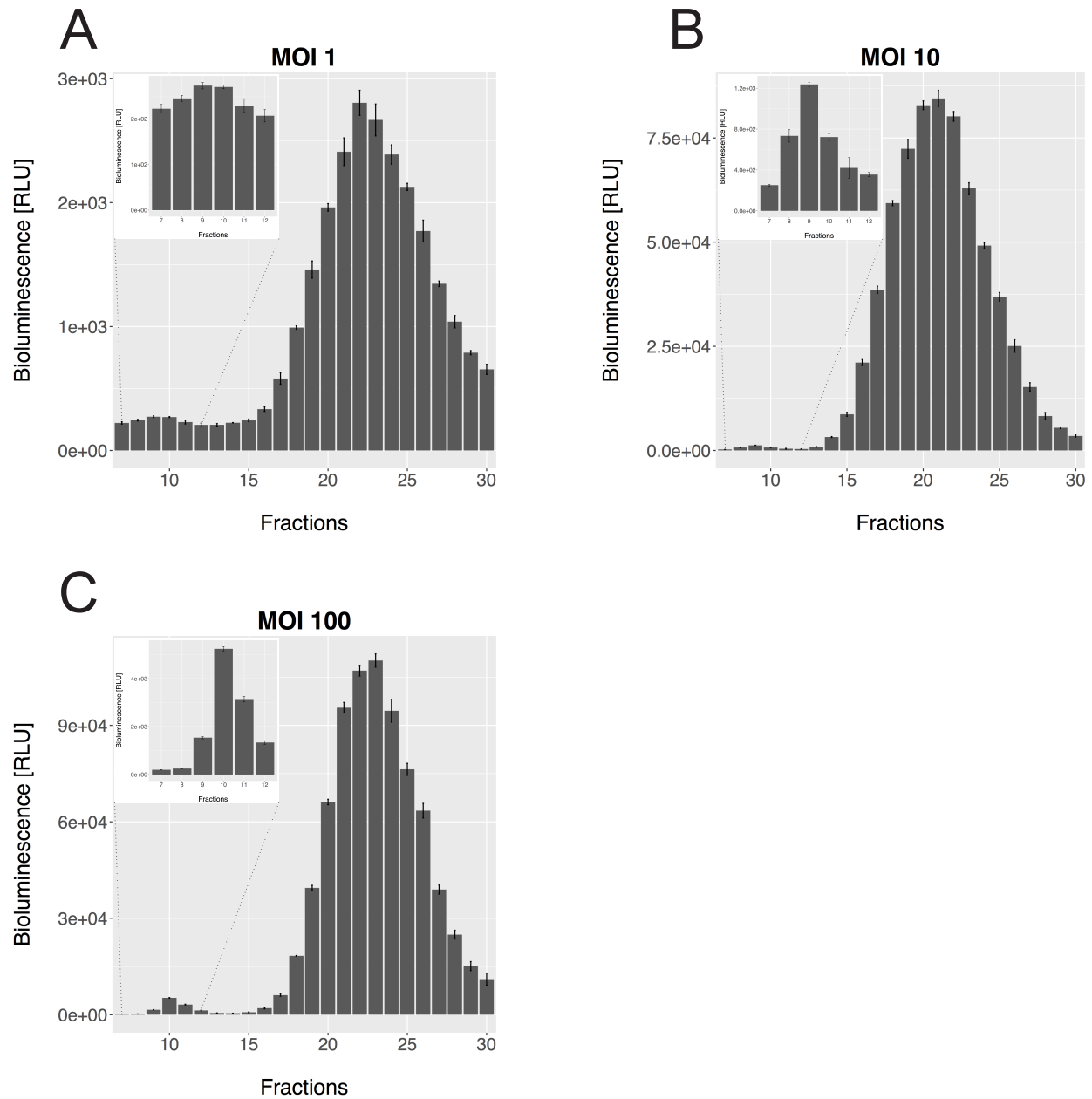
Figure S1.

**Figure S1. mbGluc Is Abundant in Low Density Fractions of Step-Gradient. Related to Figure 2.**

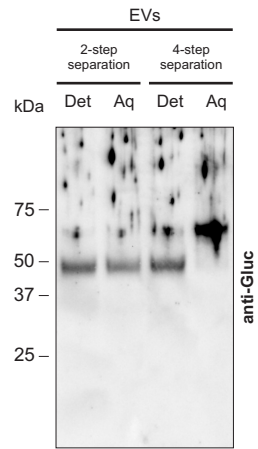
(A) mbGluc activity after fractionation of A2780 cells conditioned medium in sucrose density gradient (procedure in Fig. 2A, sample loaded on top of the gradient, 38 minutes spin at  $200K \times g$ , culture medium supplemented with 5% EV-depleted FBS). (B) mbGluc activity after fractionation of OVCA433 cells conditioned medium in sucrose density gradient (procedure in Fig. 2A, sample loaded on top of the gradient, 38 minutes spin at  $200K \times g$ , culture medium supplemented with 5% EV-depleted FBS). (C) mbGluc activity after fractionation of OVCAR5 cells conditioned medium in sucrose density gradient (procedure in Fig. 2A, sample loaded on top of the gradient, 38 minutes spin at  $200K \times g$ , culture medium supplemented with 0% EV-depleted FBS). (D) mbGluc activity after fractionation of OVCAR5 cells conditioned medium in sucrose density gradient (procedure in Fig. 2A, sample loaded on top of the gradient, 38 minutes spin at  $200K \times g$ , culture medium supplemented with 5% EV-depleted FBS). (E) mbGluc activity after fractionation of OVCAR5 cells conditioned medium in sucrose density gradient (sample loaded on top of the gradient, centrifugation for 18 hours at  $200K \times g$ , culture medium supplemented with 5% EV-depleted FBS). (F) mbGluc activity after fractionation of OVCAR5 cells conditioned medium in sucrose density gradient (sample loaded at the bottom of the gradient, centrifugation for 18 hours at  $200K \times g$ , culture medium supplemented with 5% EV-depleted FBS). (G) OVCAR5 conditioned medium loaded at the bottom of the sucrose density gradient. Medium was mixed with 60% sucrose solution. Loading of 45% solution on that mixture resulted in the move of medium dye to the top of 45%+60% solution (picture before centrifugation). (H) mbGluc activity after fractionation of OVCAR5 cells conditioned medium in iodixanol density gradient (sample loaded at the bottom of the gradient, centrifugation for 18 hours at  $200K \times g$ , culture medium supplemented with 5% EV-depleted FBS). (I) OVCAR5 conditioned medium loaded at the bottom of the iodixanol density gradient. Medium was mixed with 60% iodixanol solution. Loading of 40% solution on that mixture resulted in the move of the pink medium dye to the top of 40%+60% mixed solution (picture before centrifugation).



**Figure S2. mbGluc as a Measure of Release of EV-bound Membrane Proteins. Related to Figure 3.** mbGluc activity measured after fractionation of EVs derived from OVCAR5 mbGluc-positive cells in sucrose density gradient following incubation at 37°C in medium supplemented with 5% EV-depleted medium for 0 and 2 days.

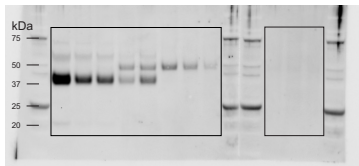
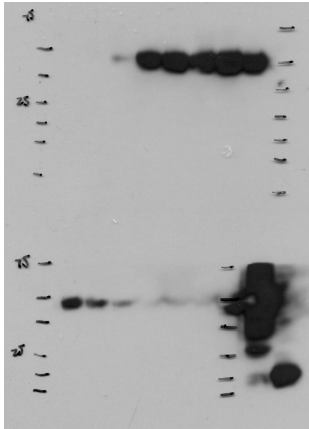
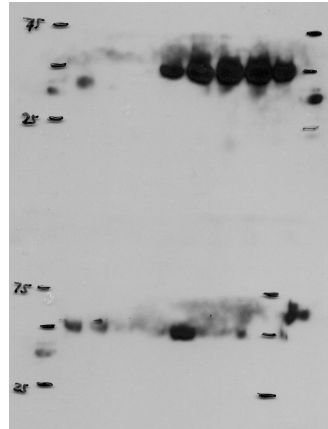
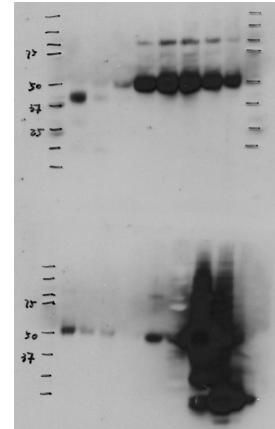
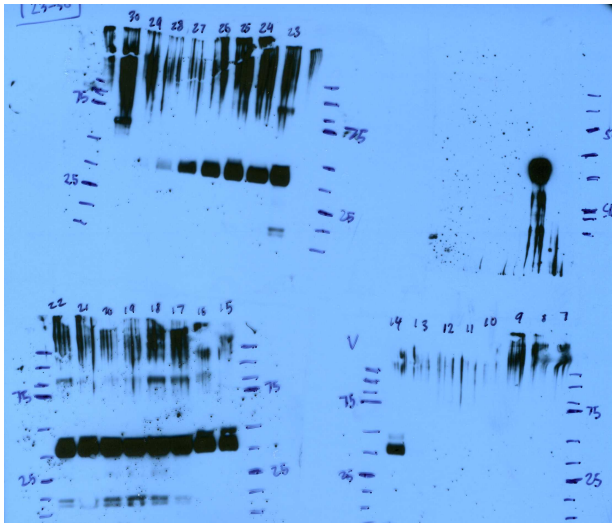
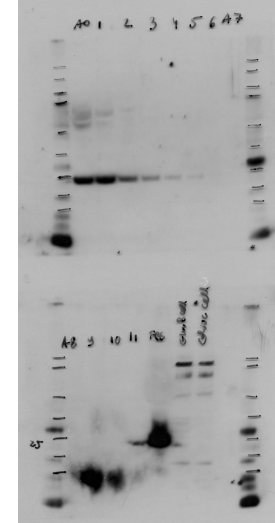
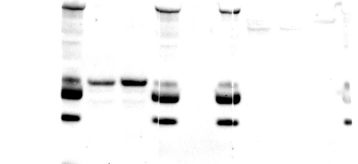


**Figure S3. Level of Expression Does Not Affect Mode of Release of mbGluc. Related to Figure 4.** (A) mbGluc activity in SEC fractions (qEV column) after fractionation of OVCAR5 medium following transduction with a lentivirus at MOI 1 (Procedure in Figure 4A). (B) mbGluc activity in SEC fractions of OVCAR5 medium following transduction with a lentivirus at MOI 10. (C) mbGluc activity in SEC fractions of OVCAR5 medium following transduction with a lentivirus at MOI 100. The insert represents fractions 7-12 with a modified scale on Y axis.



**Figure S4. Increased number of centrifugation steps in TX-114 procedure improves separation of EV-bound mbGluc. Related to Figure 6.**

Western blot of mbGluc protein isolated from OVCAR5-mbGluc+ EVs isolated by UC subjected to TX-114 procedure that included either two or four separation cycles.

**A****B****C****D****E****G****F**

**Figure S5. Full-length gels.**

(A) Full picture of the gel presented in Fig. 2C. (B) Full picture of the gel presented in Fig. 3C (upper panel). (C) Full picture of the gel presented in Fig. 3C (middle panel). (D) Full picture of the gel presented in Fig. 3C (lower panel). (E) Full picture of the gel presented in Fig. 4C. (F) Full picture of the gel presented in Fig. 4G. (G) Full picture of the gel presented in Fig. 5C.