Supporting Information

Atomic structure of the Human Herpesvirus 6B Capsid and Capsid-Associated Tegument Complexes

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Supplementary Figure 1. CryoEM images of HHV-6B particles, sub-particle reconstructions, and resolution assessment. (**a-b**) Representative images recorded on K2 camera indicate a low concentration of HHV-6B sample and show the HHV-6B virion (green) and non-infectious enveloped particle (NIEP, red). (**c-f**) The icosahedral reconstruction in Fig 1b is reshown (c) with one triangular facet in color; the regions (referred to as "sub-particles") surrounding a 5-fold, 3-fold, and 2-fold axis are circled. A total of 77,316 5-fold sub-particles, 128,860 3fold sub-particles, and 196,290 2-fold sub-particles were boxed out from original particle images and refined, yielding improved resolutions for the 5-fold sub-particle (d), 3-fold sub-particle (e), and 2-fold sub-particle (f). The enlarged views from inside these sub-particle reconstructions (right panels) show α-helices and β-strands with well-resolved side chain densities. (g) Gold-standard Fourier shell correlation (FSC) curves of the subparticle reconstructions, indicating the resolutions of sub-particle reconstructions at the 5-fold (red), 3-fold (blue), and 2-fold (green) axes are 3.82 Å, 3.77 Å, and 3.77 Å, respectively, based on the 0.143 FSC criterion.

a

Slices Through ResMap Results

Supplementary Figure 2. Local resolution assessment for the 2-fold sub-particle reconstruction.

Representative slices (**a**) and surface views (**b, c**) of the sub-particle reconstruction showing local resolution heat maps generated by *ResMap¹*. The side view (c) shows the density slab demarked by the two horizontal lines in the top view (b). Color scheme for local resolutions is shown in the color bar. Hexons C, E, P and triplexes Tb, Td are labeled.

Supplementary Figure 3. Local resolution assessment for the 3-fold sub-particle reconstruction.

Representative slices (**a**) and surface views (**b, c**) of the sub-particle reconstruction showing local resolution heat maps generated by *ResMap*1. The side view (c) shows the density slab demarked by the two horizontal lines in the top view (b). Color scheme for local resolutions is shown in the color bar. Hexons C, E and triplexes Te are labeled.

Supplementary Figure 4. Local resolution assessment for the 5-fold sub-particle reconstruction.

Representative slices (**a**) and surface views (**b, c**) of the sub-particle reconstruction, showing local resolution heat maps generated by *ResMap*1. The side view (c) shows the density slab demarked by the two horizontal lines in the top view (b). Color scheme for local resolutions is shown in the color bar. Penton, hexons P, and triplexes Ta, Tc are labeled.

Supplementary Figure 5. Density map and atomic model of a Penton MCP.

The density map (gray) of a penton MCP (segmented out from the 5-fold axis sub-particle reconstruction) at 3.82 Å resolution is superposed with its atomic model (ribbon). Boxed regions are enlarged, with density shown as gray mesh and atomic models as ribbon/sticks, in boxes with corresponding color edges.

Supplementary Figure 6. Density map and atomic model of a hexon MCP and a SCP monomer. (**a-b**) The density map (gray) of a hexon MCP (*e.g.*, MCP E2) (a) and a SCP (*e.g.*, SCP C6) (b) (segmented out from the 3-fold axis sub-particle reconstruction) at 3.77 Å resolution is superposed with the atomic model (ribbon). Boxed regions are enlarged, with density shown as gray mesh and atomic models as ribbon/sticks, in boxes with corresponding color edges.

Supplementary Figure 7. Density maps and atomic models of a Tri2A and a Tri2B monomer.

(**a-b**) The density maps (gray) of a Tri2A (a) and a Tri2B (b) (segmented out from triplex Tb of the 3-fold axis subparticle reconstruction) at 3.77 Å are superposed with their atomic models (ribbon). Boxed regions are enlarged, with density shown as gray mesh and atomic models as ribbon/sticks, in boxes with corresponding color edges.

Supplementary Figure 8. Density maps and atomic models of a Tri1 and a pU11nt monomer.

(**a-b**) The density maps (gray) of a Tri1 (a) and a pU11nt (*e.g.*, pU11-a-2) (b) (segmented out from triplex Tb of the 3-fold axis sub-particle reconstruction) at 3.77 Å are superposed with their atomic models (ribbon). Boxed regions are enlarged, with density shown as gray mesh and atomic models as ribbon/sticks, in boxes with corresponding color edges.

Supplementary Figure 9. Structural differences between corresponding HHV-6B and HCMV subunits of MCP, Tri1, Tri2A, and Tri2B.

(**a**) Schematics of MCP domain organization in HHV-6B and HCMV. (**b**) Superposition of the structures of the C6 subunit of MCP from HHV-6B and HCMV colored as in (a) by domains. Structural differences are located mainly in their upper (left inset) and buttress (right inset) domains, which are shown together with the HHV-6B cryoEM map (semi-transparent gray) in the insets. (**c**) Schematics of Tri1 domain organization in HHV-6B and HCMV. (**d**) Superposition of the structures of Tri1 subunit of triplex Td from HHV-6B and HCMV. Structural differences are located in their N-anchor domains, which is shown together with the HHV-6B cryoEM map (semi-transparent gray) in the inset. (**e**) Schematics of Tri2 domain organization in HHV-6B and HCMV. (**f, g**) Superposition of structures of the Tri2A (f) and Tri2B (g) subunits from triplex Td from HHV-6B and HCMV. Structural differences are located mainly in their embracing-arm domains, which are shown together with the HHV-6B cryoEM map (semitransparent gray).

Supplementary Figure 10. Structural differences between corresponding HHV-6B and HCMV SCP subunits, and between HHV-6B pU11nt and HCMV pUL32nt.

(**a**) Schematics of SCP domain organization in HHV-6B and HCMV. (**b**) Superposition of structures of the C6 conformer of SCP from HHV-6B and HCMV showing minor differences between the two structures. (**c**) Schematics of domain organization in HHV-6B pU11 and HCMV pUL32. (**d**) Superposition of the structures of HHV-6B pU11nt (the a-2 conformer bound to triplex Tb) and HCMV pUL32nt (the c conformer bound to triplex Tb). Structural differences are located mainly in the lower domain, which is shown together with the HHV-6B cryoEM map (semi-transparent gray).

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32) HHV-6B (pU14)

HHV-6B (pU14) HHV-6B (pU11) HCMV (pUL32)

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32)

HHV-6B (pU14)

HHV-6B (pU14) HHV-6B (pU11) HCMV (pUL32) MCMV (pM32) HHV-6A (pU11) HHV-7 (pU11) RatCMV (pR32) PHV-2 (pUL32) SCMV (pUL32) MLCMV (pUL32) RhesusCMV (pUL32) SHV-4 (pUL32) AHV-1 (pUL32) CMCMV (pcyUL32) CHV-5 (pUL32) CHV-2 (pGP32)

411

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32) HHV-6B (pU14)

HHV-6B (pU14) HHV-6B (pU11) HCMV (pUL32)

MCMV (pM32)

HHV-6A (pU11) HHV-7 (pU11) RatCMV (pR32) PHV-2 (pUL32) SCMV (pUL32) MLCMV (pUL32) RhesusCMV (pUL32) SHV-4 (pUL32) AHV-1 (pUL32) CMCMV (pcyUL32) CHV-5 (pUL32) CHV-2 (pGP32)

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32) HHV-6B (pU14)

HHV-6B (pU11) HCMV (pUL32) \overline{M} *CMV HHV-6B (pU14)*

HHV-6B (pU14) HHV-6B (pU11) HCMV (pUL32) MCMV (pM32)

417 388 379

378

389 379 454

HHV-6B (pU11)

HCMV (pUL32) MCMV (pM32) HHV-6B (pU14)

HHV-6B (pU14) HHV-6B (pU11)

 MC **HHV-6A (pU11) HHV-7 (pU11)**

 PH **SCMV (pUL32) ML** Rh SH **AHV-1 (pUL32)** $CN²$ **CHV-5 (pUL32) CHV-2 (pGP32)**

474 .DQLSK...NSTNDLQKILERERIKTIK.....QNNEDIFK......LPSEKRRKEI
459 RVFVSDG.......KTV...TLPVALDAAS.PSPASS...T...RPPPERPGTEIGTDDV
510 RGYAPTGPRPSSSRVSNTLVDLTTPGGNA.....AQPQQTSYKFETVPLVTPQPSSI...
379AEKTKGDSMLDLGD

 ${\tt RGSSPKG}$ ${\tt RSSV}$ $\tt E$ $D L$ $VDRL$ $GD L$ PVK ... $GGKKPMFVFEPPSIPMPSTV$ \tT $.$...

χ.

.....AL......DSSGLLDLEKQLEKLDVG...GVPTQAFVFEPPFKPTETVSQ....

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32)

HHV-6B (pU14) **HHV-6B (pU14)** 587 **HHV-6B (pU11)** $\ldots \ldots \ldots \ldots \ldots$ KHAN $\ldots \ldots \ldots \ldots \ldots \ldots$ DIFAGLNKKYARDVSRGGKGNSRD 587 **HCMV (pUL32)** 621 ..NGNSPWAPTAPLPGD.MNPANWPRERAWALKNPHLAYNPFRMPTT..........ST **MCMV (pM32)** 611 **HHV-6A (pU11)** 598 **HHV-7 (pU11)** 545 **RatCMV (pR32)** 539 **PHV-2 (pUL32)** 595 **SCMV (pUL32)** 448 **MLCMV (pUL32)** 484 **RhesusCMV (pUL32)** $\ldots \ldots \ldots \ldots \ldots \text{LFATGSSETIPGLETPLKEY} \ldots \ldots \text{SG} \ldots \ldots$ 444 **SHV-4 (pUL32)** 439 **AHV-1 (pUL32)** 441 **CMCMV (pcyUL32)** 445 **CHV-5 (pUL32)** 448 **CHV-2 (pGP32)** 538

HHV-6B (pU11) HCMV (pUL32) MC *MV*

HHV-6B (pU14)

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32) HHV-6B (pU14)

HHV-6B (pU14) HHV-6B (pU11) HCMV (pUL32) MCMV (pM32) HHV-6A (pU11) HHV-7 (pU11) RatCMV (pR32) PHV-2 (pUL32) SCMV (pUL32) MLCMV (pUL32) RhesusCMV (pUL32) SHV-4 (pUL32) AHV-1 (pUL32) CMCMV (pcyUL32) CHV-5 (pUL32) CHV-2 (pGP32)

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32)

HHV-6B (pU14)

HHV-6B (pU14) HHV-6B (pU11) HCMV (pUL32) MCMV (pM32)

HHV-6A (pU11) HHV-7 (pU11) RatCMV (pR32) PHV-2 (pUL32) SCMV (pUL32) MLCMV (pUL32) RhesusCMV (pUL32) SHV-4 (pUL32) AHV-1 (pUL32) CMCMV (pcyUL32) CHV-5 (pUL32) CHV-2 (pGP32)

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32) HHV-6B (pU14)

HHV-6B (pU14) HHV-6B (pU11) HCMV (pUL32)

MCMV (pM32) HHV-6A (pU11) HHV-7 (pU11) RatCMV (pR32) PHV-2 (pUL32) SCMV (pUL32) MLCMV (pUL32) RhesusCMV (pUL32) SHV-4 (pUL32) AHV-1 (pUL32) CMCMV (pcyUL32) CHV-5 (pUL32) CHV-2 (pGP32)

HHV-6B (pU11) HCMV (pUL32) MCMV (pM32)

HHV-6B (pU14)

Supplementary Figure 11. Secondary structure for pU11nt, pUL32nt, pM32nt, and pU14nt, and pp150 sequence alignment for 15 CMVs.

Schematic representations of the secondary structure alignment for pU11nt, pUL32nt, pM32nt, and pU14nt², and amino acid sequences of pp150s from HHV-6B³, HHV-6A³, HHV-7⁴, Human cytomegalovirus (HCMV)5, Murine cytomegalovirus (MCMV)6, Rat cytomegalovirus Maastricht (RatCMV)7, Chimpanzee cytomegalovirus (PHV-2)8, Simian cytomegalovirus (SCMV)9, Mandrillus leucophaeus cytomegalovirus (MLCMV)¹⁰, Rhesus cytomegalovirus (RhesusCMV)¹¹, Saimiriine βherpesvirus 4 (SHV-4)¹², Aotine ß-herpesvirus 1 (AHV-1)¹², Cynomolgus macaque cytomegalovirus (CMCMV)¹³, Cercopithecine β -herpesvirus 5 (CHV-5)¹², Caviid β -herpesvirus 2 (CHV-2)¹⁴ analyzed and displayed by $ESPrint 3.0^{15}$. Spiral represents α -helix.

Supplementary Figure 12. Structural differences of CATCs in HHV-6B, HCMV, and MCMV.

(**a**) The atomic models of triplex Te (gray) and pU11 in HHV-6B (light blue) and corresponding structures in HCMV (green) are superposed together as rigid bodies to show differences in their Te-associated CATCs. Conformers b-2 of CATC (*i.e.*, pU11nt-b-2 of HHV-6B and pUL32nt-b in HCMV) exhibit major translational and rotational displacements (upper right panel), while conformers a-2 (pU11nt-a-2 and pUL32nt-a, bottom right panel) and b-1 (pU11nt-b-1 and pUL32nt-c, left panel) exhibit relatively minor rotational displacements between HHV-6B and HCMV. Conformer a-1 (pU11nt-a-1) exists only in HHV-6B. (**b**) The atomic models of triplex Te (gray) and pU11 in HHV-6B (blue) and the corresponding structures in MCMV (gold) are aligned together as rigid bodies to show differences in their associated CATCs. The a-2 CATC conformers (pU11nt-a-2 in HHV-6B and pM32nt-a in MCMV, right panel) exhibit major translational and rotational displacements, while conformers b-2 (pU11nt-b-2 and pM32nt-b, left panel) exhibit relatively minor rotational displacements between HHV-6B and MCMV. Conformer a-1 and b-1 do not exist in MCMV.

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bU11nt-b

pU11nt-b-2

pU11nt-a

pU11nt-a-1

 \mathbf{d}

SCP

 p U11nt-b-2

P1 WP2

 $13\AA$

Tc

(**a**) Zoomed-in view of HHV-6B triplex Ta region with labeled subunits. (**b**) The same region as (a) but models of the HHV-6B Tb triplex and four of its pU11nt subunits are docked by fitting the Tb ribbon model as a rigid body into the density map of HHV-6B Ta region. (**c**) Inset shows the clashes between pU11nt-b-1 (orange) and its neighboring SCP (magenta). (**d**) Inset shows a distance of 13 Å between pU11nt-b-2 (orange) and its neighboring SCP (magenta). (**e**) Zoomed-in view of HHV-6B triplex Tc region. (**f**) The same region as (e) but with ribbon models of the HHV-6B Tb triplex and four of its pU11nt subunits docked by fitting the Tb ribbon model as a rigid body into the HHV-6B Tc region density map. (**g**) Inset shows a distance of 11 Å between pU11nt-b-2 (orange) and its neighboring SCP (magenta). (**h**) Zoomed-in view of HHV-6B triplex Tf region. (**i**) The same region as (h) but with ribbon models of the HHV-6B Tb triplex and four of its pU11nt subunits docked by fitting the Tb ribbon model as a rigid body into HHV-6B Tf region density map. (**j**) Inset shows the clashes between pU11nt-b-1 and its neighboring SCP. In (b, f, i), neighboring MCPs are colored individually while Tri1, Tri2A, Tri2B, pU11nt, and SCP are colored as in Figure 1e.

Supplementary Figure 14. pU11 shares a similar fold with pU14.

(**a**) Atomic models of N-terminal domain of pU11 (pU11nt) and pU14 (pU14nt) proteins shown as pipes and planks (blue at the N-terminus to red at the C-terminus) with their α -helixes labeled. (**b**) Superposed pU11nt (dark blue) and pU14nt (light blue) reveal that pU11nt (a.a. 1-284) and pU14nt (a.a. 1-329) share a highly similar fold. Residues 330-455 are shown as transparency for clarity. pU11nt residues within a distance of 4 Å from neighboring capsid proteins are shown in green. (**c**) Secondary structure and sequence alignment for pU11nt and pU14nt. Those pU11nt residues that are within a distance of 4 Å from neighboring capsid proteins are highlighted in green shades.

Supplementary Tables

Supplementary Table 1. Data collection summarization for HHV-6B, HSV-1, HSV-2, HCMV, and KSHV.

Supplementary Table 2. CryoEM data collection, refinement and validation statistics for HHV-6B icosahedral and sub-particle (2-fold, 3-fold, and 5-fold) reconstructions, and atomic models.

Supplementary Table 3. Comparison of capsid, pU11/pUL32, pU14/pUL25 proteins in HHV-6B and in HCMV.

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