# Kisspeptin-Activated Autophagy Independently Suppresses Non-Glucose-Stimulated Insulin Secretion from Pancreatic β-Cells

Chien Huang<sup>1, #</sup>, Hao-Yi Wang<sup>1, #</sup>, Mu-En Wang<sup>1, 2</sup>, Meng-Chieh Hsu<sup>1</sup>, Yi-Hsieng Samuel Wu<sup>1</sup>, Yi-Fan Jiang<sup>3</sup>, Leang-Shin Wu<sup>1</sup>, De-Shien Jong<sup>1</sup>, and Chih-Hsien Chiu<sup>1, \*</sup>

<sup>1</sup>Laboratory of Animal Physiology, Department of Animal Science and Technology, National Taiwan University, Taipei 10617, Taiwan

<sup>2</sup>Department of Pathology, Duke University School of Medicine, Duke Cancer Institute, Duke University, Durham, NC 27514, USA.

<sup>3</sup>Graduate Institute of Molecular and Comparative Pathobiology, School of Medicine, National Taiwan University, Taipei 10617, Taiwan

#Equal contribution

#### \*Corresponding Author: Chih-Hsien Chiu

Laboratory of Animal Physiology, Department of Animal Science and Technology National Taiwan University

No. 50, Lane 155, Section 3, Keelung Road, Taipei City 106, Taiwan

Tel.: +886-2-3366-4171; Fax: +886-2-3366-4070; Email: chiuchihhsien@ntu.edu.tw

### **Supplementary Tables**

Table 1. Antibodies used in Western Blotting

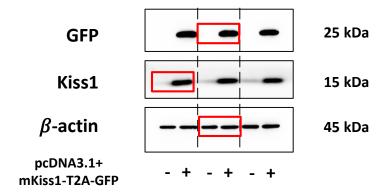
Antibody Name	Company	Product Number	Dilution
Anti-GFP	Santa Cruz Biotechnology	sc-9996	1:1000
Anti-Kiss1	Cloud-Clone Corp	PAC559Mu01	1:400
Anti-β-actin	Santa Cruz Biotechnology	sc-47778	1:2500
Anti-SQSTM1/p62	abcam	ab109012	1:1000
Anti-LC3	Cell Signaling Technology	#2775	1:1000
Anti-insulin	Cell Signaling Technology	#8138	1:1000
Anti-ATG5	Cell Signaling Technology	#12994	1:1000
Anti-GAPDH	Cell Signaling Technology	#2118	1:2500
Anti-PARP	Cell Signaling Technology	#9532	1:1000
Anti-Cleaved	Cell Signaling Technology	#9661	1:250
Caspase 3			

### Table 2. Primers used in qPCR analyses

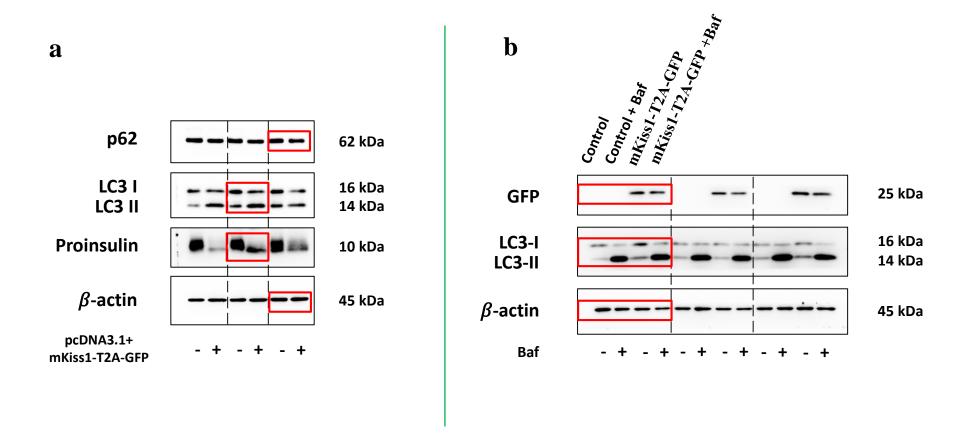
Gene Name	Forward (5' to 3')	Reverse (5' to 3')
Insulin	GCAGAGAGGAGGTACTTTGGA	GGTAGGAAGTGCACCAACAG
RPL19	GCTCTTTCCTTTCGCTGCTGC	CAGTCACAGGCTTGCGGATGAT

#### **Supplementary Figure Legends**

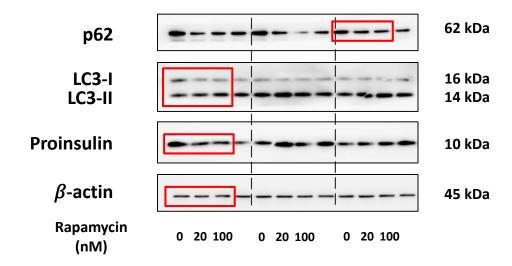
Supplementary Figure S1. The full-length blot of GFP, Kiss1, β-actin presented in Figure 2 of the main text. Supplementary Figure S2. The full-length blot of GFP, p62, LC3, Proinsulin, β-actin presented in Figure 3 of the main text. Supplementary Figure S3. The full-length blot of p62, LC3, Proinsulin, β-actin presented in Figure 4 of the main text. Supplementary Figure S4. The full-length blot of ATG5, p62, LC3, Proinsulin, β-actin presented in Figure 5 of the main text. Supplementary Figure S5. The full-length blot of ATG5, p62, LC3, Proinsulin, β-actin presented in Figure 7 of the main text. Supplementary Figure S6. The dosage response of the glucose-stimulated insulin secretion in NIT-1 cells by measuring luminescent activity of luciferase. After 30-minute collection, the insulin secretion from NIT-1 under 0, 5.5, 11 and 16.5 mM glucose challenge was measured by luciferase activities. Quantifications normalized by total protein in NIT-1 cells are shown as the means ± standard errors of the mean (n = 3). Different letters represent significant difference determined by one-way ANOVA with post-hoc tests. Supplementary Figure S7. The bafilomycin treatment does not trigger apoptosis in NIT-1 cells. Representative blots of apoptosis markers in NIT-1 cells after treating with 0, 1, 5, or 20 nM bafilomycin for 6 h; quantifications normalized by β-actin are shown as the means  $\pm$  standard errors of the mean (n = 3). Indicated markers have no significant differences determined by one-way ANOVA. Supplementary Figure S8. Short-term exposure of kisspeptin decreases (pro)insulin level and activated autophagy in NIT-1 cells. After transfecting pcDNA3.1+mKiss1-T2A-GFP plasmid for 66 h, the growth media for NIT-1 cells were changed to blank media for another 6-hour culture. After 6-hour culture, the conditioned media from control and Kiss1-overexpressing group were collected and then treated to the non-treated NIT-1 cells for 2 minute. Then, the conditioned media-treated cell lysates from two group were collected for further analysis. The treated conditioned media to control group were collected from NIT-1 cells transfected with reagent only. Representative blots of autophagy markers and (pro)insulin in short-term treated NIT-1 cells and quantifications normalized by GAPDH are shown as the means  $\pm$  standard errors of the mean (SEM). \*compared with the control; \*p < 0.05, \*\*p < 0.01.



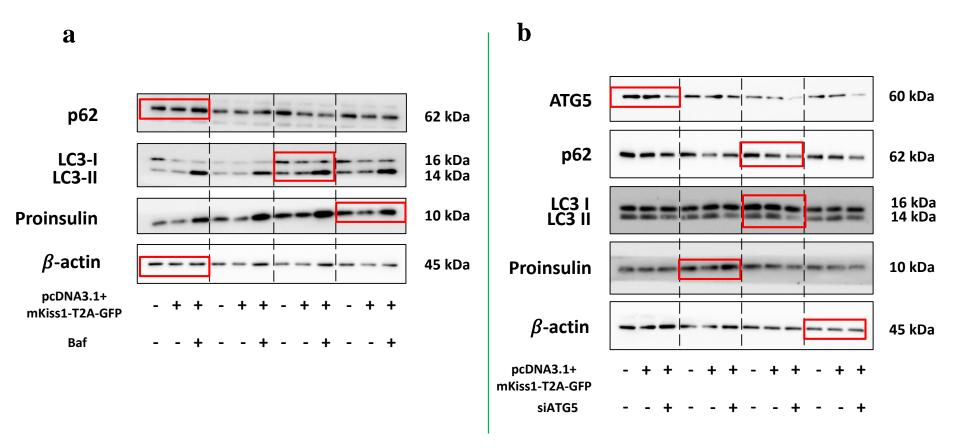
### **Supplementary Figure S1**



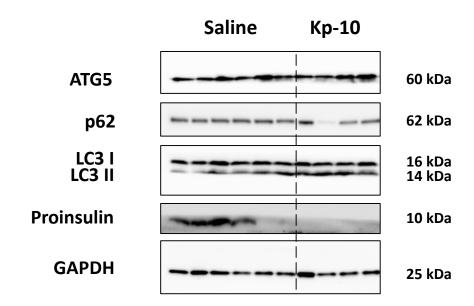
**Supplementary Figure S2** 



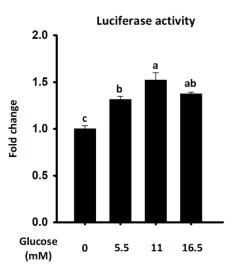
**Supplementary Figure S3** 



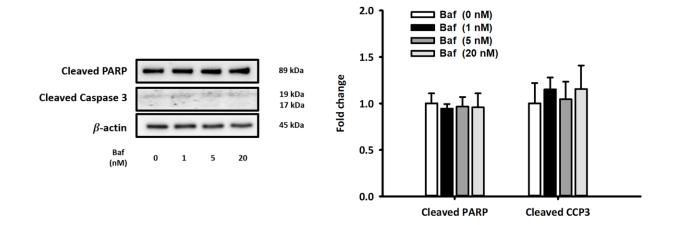
**Supplementary Figure S4** 



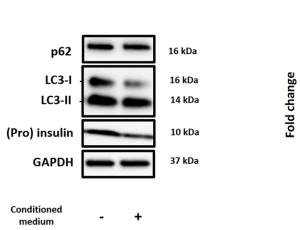
**Supplementary Figure S5** 

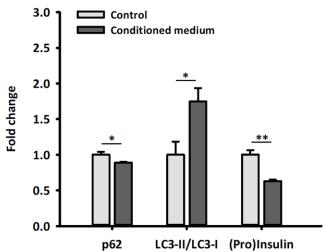


# **Supplementary Figure S6**



**Supplementary Figure S7** 





# **Supplementary Figure S8**