

SUPPLEMENTAL MATERIALS

Table S1. Characteristics of the clinical isolates used in this study

Strain	MLST ^a	Enzyme type ^b	Year of isolation	Source ^c	ΦNJS1 sensitivity ^d
A2312NM	ST-11	KPC-2	2014	Feces	+++
A1678	ND	KPC-2	2013	LRS	-
A1679	ST-11	KPC-2	2013	LRS	-
A1705	ST-449	KPC-2, NDM-1	2013	Urine	-
A1706	ST-449	KPC-2, NDM-1	2013	LRS	-
A1763	ST-11	KPC-2	2013	Feces	-
A1806	ST-1493	NDM-1	2013	Feces	+++
A1831	ST-11	KPC-2	2014	Feces	-
A1836	ST-11	KPC-2	2014	Feces	++
A2314	ST-11	KPC-2	2014	LRS	+++
A2402	NEW	KPC-2	2014	Feces	+++
A2440	ND	NDM-1	2014	Blood	-
A1876	ST-1318	NDM-1	2014	Feces	-
A1838	ST-17	IMP	2014	Feces	+++
A1682	ST-17	ND	2013	Blood	++
A2359	ST-307	IMP	2014	LRS	-
A2368	ST-307	IMP	2014	LRS	-
A1871	ST-37	ND	2014	Urine	++
A2612	ST-395	KPC	2015	Feces	+++
A1851	ST-395	ND	2014	Feces	-
A1824	ST-65	KPC-2	2014	Feces	+++
A1749	ST-709	ND	2013	Feces	-
A1860	ST-846	IMP	2014	Blood	++
A1683	ST-86	ND	2013	Blood	-
A2366	ST-875	ND	2014	Feces	-
A2369	NEW	IMP	2014	AF	-
A2373	NEW	KPC	2014	LRS	+++
A2371	NEW	KPC	2014	LRS	-
KP1513	ND	ND	2015	LRS	+++
ATCC BAA-1899		KPC, NDM	unknown	Human	++
ATCC BAA1705		ND	2007	Urine	-
ATCC BAA1706		ND	2007	Urine	+++

^aMLST type: NEW, new ST types; ND, not determined; ^bCarbapenem antibiotic enzyme type: KPC, carbapenemases; NDM, New Delhi metallo-β-lactamase; IMP, imipenemase metallo-β-lactamase; ND, not determined. ^cClinical specimens source: AF, ascitic fluid; LRS, lower respiratory secretions; ^dSensitivity to ΦNJS1: “+++”, EOP (Efficiency of Plaques) >0.01, “++”, EOP>0.0001, “-” EOP<10⁻⁵.

Table S2. *Kp* phages isolated in this study

phage	Genbank number	Genome (bps)	GC content	Host range ^a
ΦNJS1	MH445453	49,292	50.6%	46.88%
ΦNJS2	MH633485	50,132	50.8%	25%
ΦNJS3	MH633486	49,387	51.1%	21.88%
ΦNJR15	MH633487	49,468	51.0%	21.88%
ΦTAH8	MH633484	49,344	51.1%	31.25%

^a. 32 *Kp* clinical isolates were tested. EOP >10⁻⁵ indicates susceptibility to phage.

Table S3. Primers used in this study

Primer	Sequence
mcr1_IS_LR ed_u1	5' GTAACCGTCTCATTAAACCGTCTTTCGCCTCCCTTTCCTGTTT CCGATACCGTTGCACTTGGTTTGACAATTC 3'
mcr1_IS_LR ed_l1	5' CTTGAAAATACATGGTATTGCGTAAAGGGCGGTAAAAGTCTA TCCTGGTGTGTGCGGTGGGTTTGGAAAAAATAC 3'
mcr1_LRed _u1	5' GCAGTATAATTGCCGTAATTATCCCACCGTTTATTTTTTGAGTAG TTTCTCGTGTAGGCTGGAGCTGCTTC 3'
mcr1_LRed _l1	5' GACAAGAGCGATACTCATCTCAGCAAGTAGGCGTTTATTTGATA AATACGCATATGAATATCCTCCTTAG 3'
oriR6kseqpri m1	5' GACACAGGAACACTTAACGGC 3'
psyn-u1	5' CCTGACGGATGGCCTTTTTG 3'

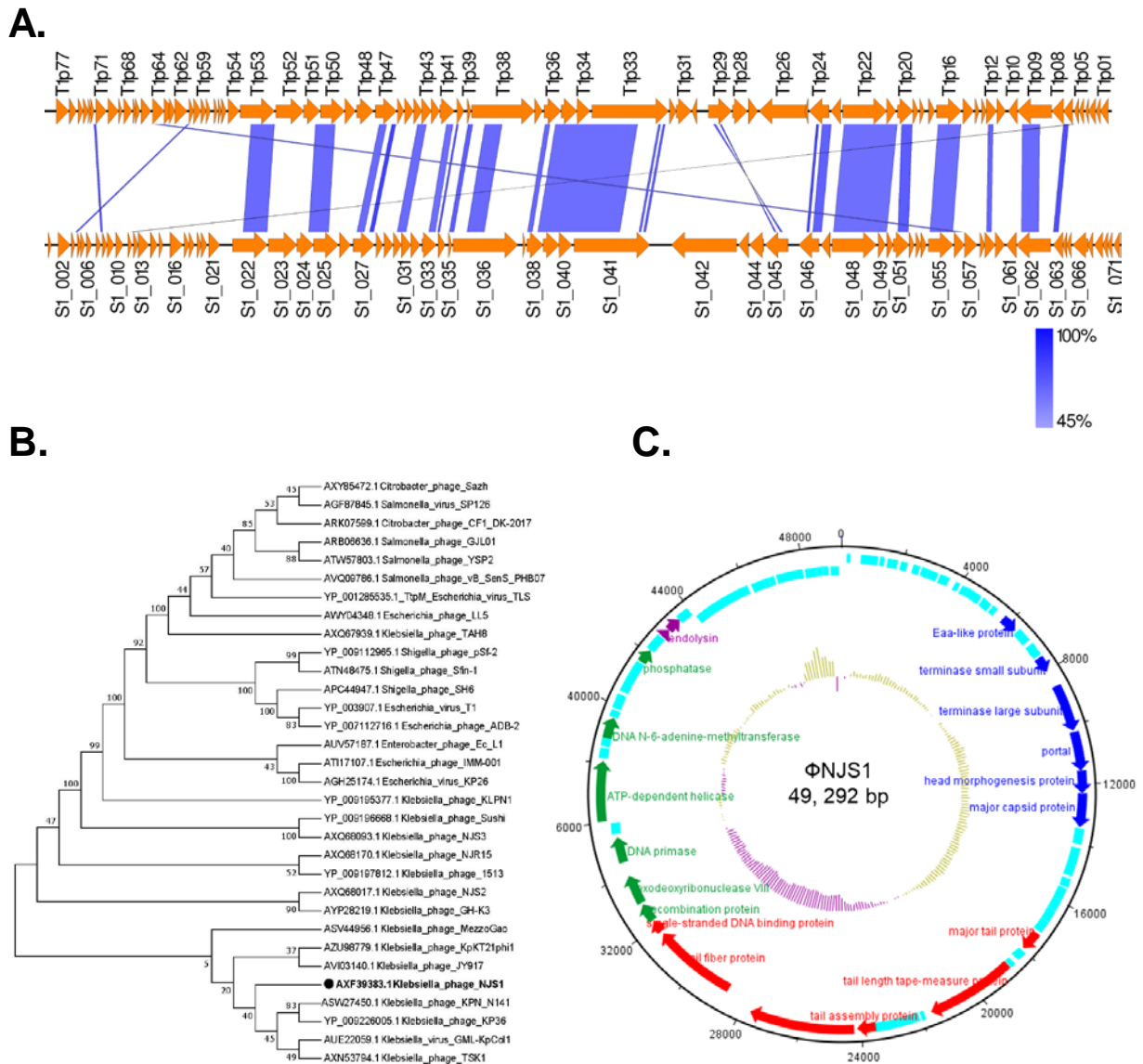


Fig. S1. Genome map and comparison of Φ NJS1 with other phages. **A.** Direct comparison of the genomic structures of NJS1 (accession no. MH445453) and close-related Enterobacteria phage T1 genome (NC_005833) by GATU software, RAST and PHASTER programs. Shaded boxes represent the sequence identity levels between these two genomes. **B.** Phylogenetic tree based on the tape measure proteins (TMP) by using the Neighbor-Joining method. **C.** Genome map. The inner circle indicates the GC content. The color of each gene refers to the functional category: phage structure (blue and red), replication/recombination/repair (green), lysis (purple). The scale units are base pairs.

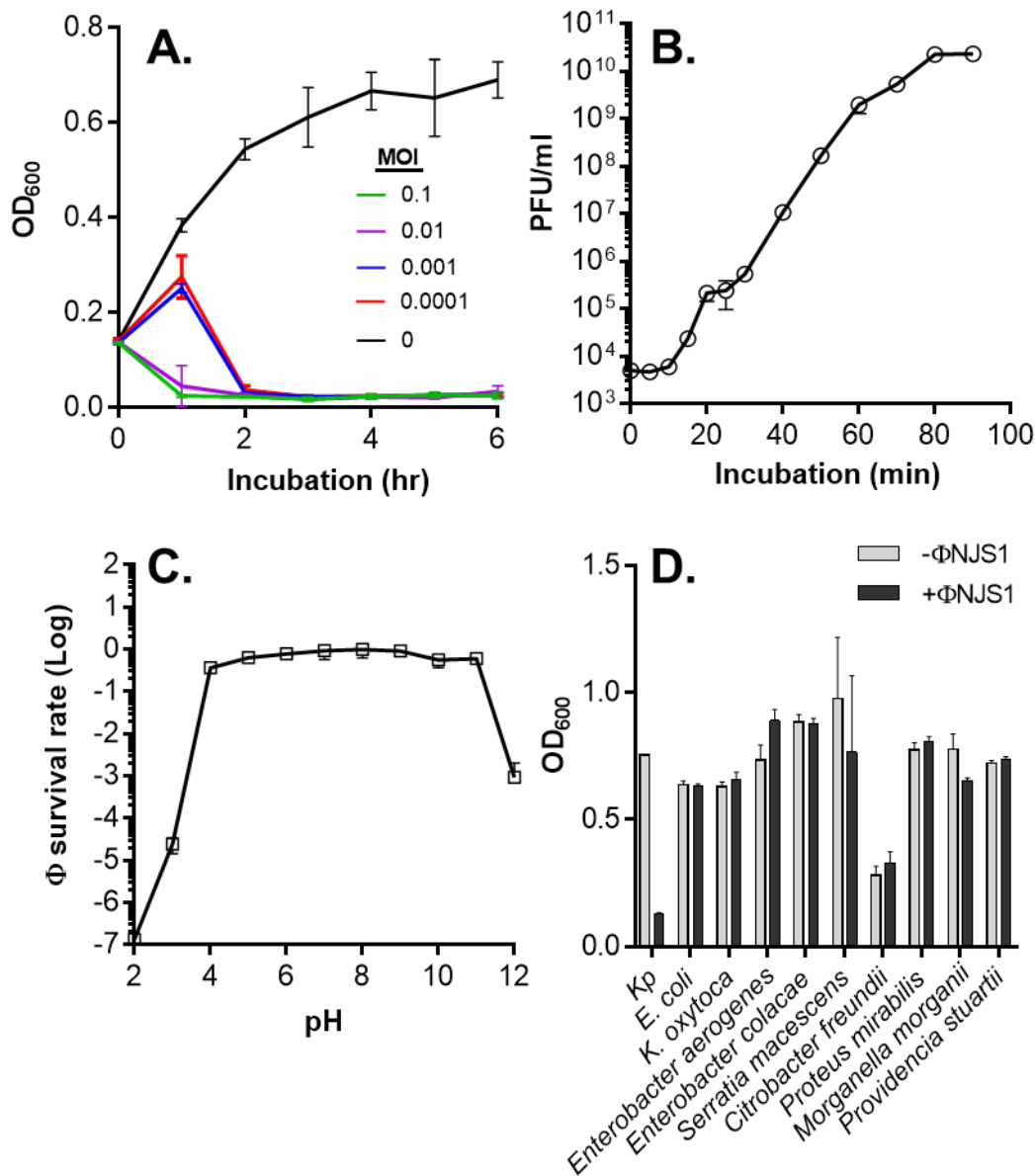


Fig. S2. The characteristics of Φ NJS1. **A.** Infectious dose. Approximately 10^8 CFU/ml mid-log *Kp* cells were incubated with the phage at a multiplicity of infection (MOI) of 0.0001, 0.001, 0.01, or 0.1. Phage infection dynamics (OD₆₀₀) was measured at 1 hr intervals for 6 hr standing incubation at 37°C. The mean of 3 independent assays is shown and error bars represent the standard deviation. **B.** One-step growth curve of the Φ NJS1 with *Kp*. Approximately 3×10^8 CFU/ml mid-log *Kp* cells were incubated with the phage at a MOI of 0.0001. After 10 mins of phage adsorption, unadsorbed phages were removed by centrifugation and bacterial cells were resuspended in fresh LB and incubated at 37°C under constant shaking. Samples were taken at 5-min intervals for titration immediately. The mean of 3 independent assays is shown, and error bars represent the standard deviation. **C.** Phage stability in different pHs. 10^8 PFU/mL phage

particles were incubated in 0.8% NaCl buffered to different pH for 24 hr at 37°C. The survival rate of the phage was compared to that of the control samples (4°C, pH 7). The mean of 3 independent assays is shown and error bars represent the standard deviation. D. Host range specificity. All *Enterobacteriaceae* strains were cocultured without or with phage at MOI=1 under shaking condition at 37°C. At the 6-hr time point, OD₆₀₀ was measured.

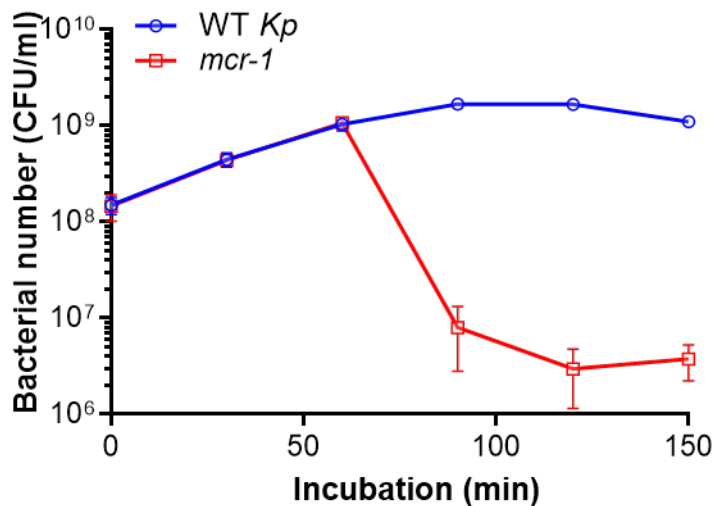


Fig. S3. *Kp* CFU during phage infection. 2×10^8 CFU/ml WT *Kp* and *Kp* (*mcr-1*) were mixed with Φ NJS1 at MOI=10⁻⁴ and incubated at 37°C. Viable bacteria were then counted by serial dilution and plating on LB agar. The mean of three independent assays is shown and error bars represent the standard deviation.

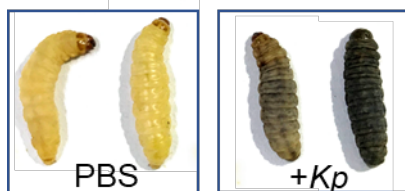


Fig. S4. *G. mellonella* responses to *Kp* infection. Larvae were injected with 10 μ l of *Kp* cells (10^6 CFU) or PBS in parallel. Infected larva became pigmented progressively darker over the course of the infection. Photography was taken after 24 hrs of infection.