

HAMLET, a protein complex from human milk kills and enhances the activity of antibiotics against pathogenic *Streptococci*

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Supplemental Material

Table S1. Pneumococcal strains used in the study

| Strain | Serotype | Antibiotic Resistance (<i>gene</i>)* | Source** |
|-------------------------------------|----------|---|------------|
| <i>S. pneumoniae</i> | | | |
| D39 | 2 | None | (1) |
| D39 Δ <i>dldh</i> (C0832) | 2 | Erythromycin (<i>ermB</i>) | (2) |
| D39 Δ <i>stkP</i> | 2 | Kanamycin (<i>aphA3</i>) | (3) |
| D39 Δ <i>pspC</i> | 2 | Tetracycline (<i>tetM</i>) | (4) |
| D39 Δ <i>pitB</i> | 2 | Chloramphenicol (<i>cat</i>) | (5) |
| D39-Sm | 2 | Streptomycin (<i>rpsL</i>) | This study |
| SP670 | 6A | Penicillin G | (6) |
| ATCC 49619 | 19F | None | (7) EUCAST |
| 3974 | 14 | Erythromycin (<i>ermB</i>) | EUCAST |
| 7545 | 14 | | EUCAST |
| 12627 | 6B | Erythromycin (<i>mef</i> + <i>msrD</i>) | EUCAST |
| 13331 | 14 | | EUCAST |
| 17476 | 23A | | EUCAST |
| 1947 | 19F | Trimethoprim-Sulfoxide (R) | EUCAST |
| 16467 | 6A | | EUCAST |
| 17446 | 16A/F | Trimethoprim-Sulfoxide (I) | EUCAST |
| 18091 | 23F | | EUCAST |
| 4269 | 3 | Fluoroquinolone | EUCAST |
| 13985 | 19F | Tetracycline | EUCAST |
| 17144 | 19F | | EUCAST |
| 6701 | 4 | Penicillin (I) | EUCAST |
| 16998 | 6B | | EUCAST |
| 12116 | 35B | Penicillin (I), Cephalosporine / Ampicillin (R) | EUCAST |
| 16031 | 35A | | EUCAST |
| 19000 | 35B | Penicillin, Cephalosporine | EUCAST |
| 18120 | 6A | | EUCAST |
| 4393 | 23F | | EUCAST |

* I = Intermediate resistance; R = Resistant

** EUCAST; The European Committee on Antimicrobial Susceptibility Testing

Table S2. Primers used for Erm-resistance determination.

| Gene name | Primer name | Primer sequence (5' to 3') | Reference |
|-------------------|--------------|-----------------------------------|------------|
| <i>ermB</i> | ErmB-F | GGTACCATGAACA AAAATATAAAAATATTCTC | This study |
| | ErmB-R | GGATCCTTATTTCTCCCGTTAAATAATAG | This study |
| | ErmB-F2 | GAAAAGGTA CTCAACCAAATA | (8) |
| | ErmB-R2 | AGTAACGGTACTTAAATTGTTTAC | (8) |
| <i>ermTR</i> | ErmTR-F | GGGTCAGGAAAAGGACATT | (9) |
| | ErmTR-R | CATTCGCATGCTTCAGC | This study |
| <i>ermT</i> | ErmT-F | CCGCCATTGAAATAGATCCT | (10) |
| | ErmT-R | GCTTGATAAAAATTGGTTTTTTGGA | (10) |
| <i>mefA/mefE</i> | MefAE-F | AGGGCAAGCAGTATCATTAAATCA | (10) |
| | MefAE-R | CTGCAAAGACTGACTATAGCCT | (10) |
| <i>mefA</i> (GBS) | MefA-F | AGTATCATTAAATCACTAGTGC | (8) |
| | MefA-R | TTCTTCTGGTACTAAAAGTGG | (8) |
| <i>linB</i> | LinB-F | CCTACCTATTGTTTGTGGAA | (11) |
| | LinB-R | ATAACGTTACTCTCCTATTC | (11) |
| <i>msrD</i> | MsrD-F | TTGGACGAAGTAACTCTG | (12) |
| | MsrD-R (GAS) | GCTTGTCTCTTACGTTC | This study |
| | MsrD-R (SPN) | GCTTGGCTCTTACGTTC | (12) |

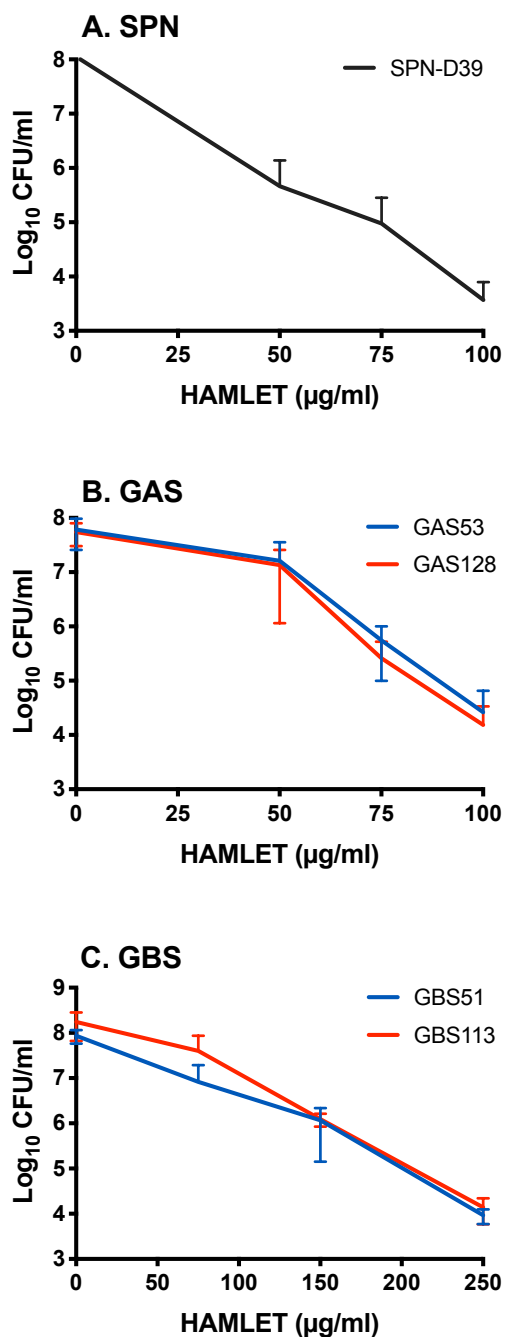


Figure S1. Bactericidal activity of HAMLET against *S. pneumoniae*, GAS and GBS. Bacteria were grown in THY, washed and resuspended in PBS with 25 mM glucose to keep the bacteria energized. The bacterial suspensions were prepared to obtain a starting concentration of approximately 1×10^8 CFU/ml. Then, increasing concentrations of HAMLET were added to wells of each bacterial strain and the bacteria were allowed to incubate for 1 hour at 37°C. Bacteria were then serially diluted and dilutions were plated onto agar that were allowed to grow for 24-48 hours to detect colonies. The viable CFU counts were counted and the concentration as CFU/ml was calculated and depicted in the graphs. The results represent the mean data from at least 3 separate experiments with standard deviations.

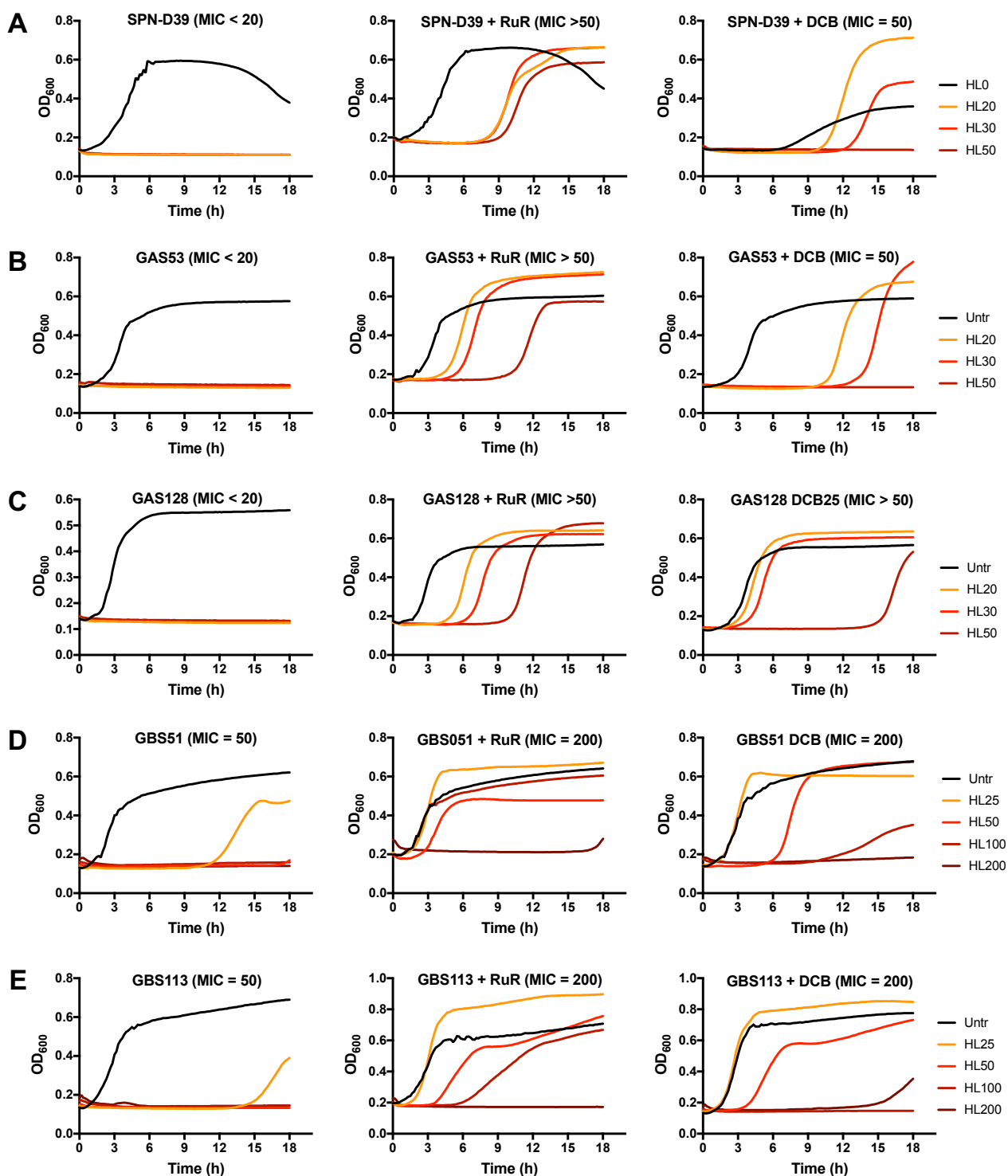


Figure S2. Inhibition of MIC growth curves of pathogenic *Streptococci* with ion transport inhibitors. MIC growth curves are shown for (A) D39, (B and C) GAS clinical isolates 53 and 128, and (D and E) GBS clinical isolates 51 and 113 in the presence of increasing concentrations of HAMLET

(left) and Ruthenium red (30 μ M; middle) or Dichlorobenzamil (25 μ M; right). Bacteria were grown in the presence of HAMLET alone or in the presence of inhibitors for 18 h at 37°C with the absorbance at 600 nm (OD_{600}) recorded every 10 min. The lowest concentration of HAMLET where no growth was detected over 18 h was considered the MIC value. The figure shows a representative growth curve for each strain. However, the MIC values are based on three separate experiments with samples run in duplicate wells.

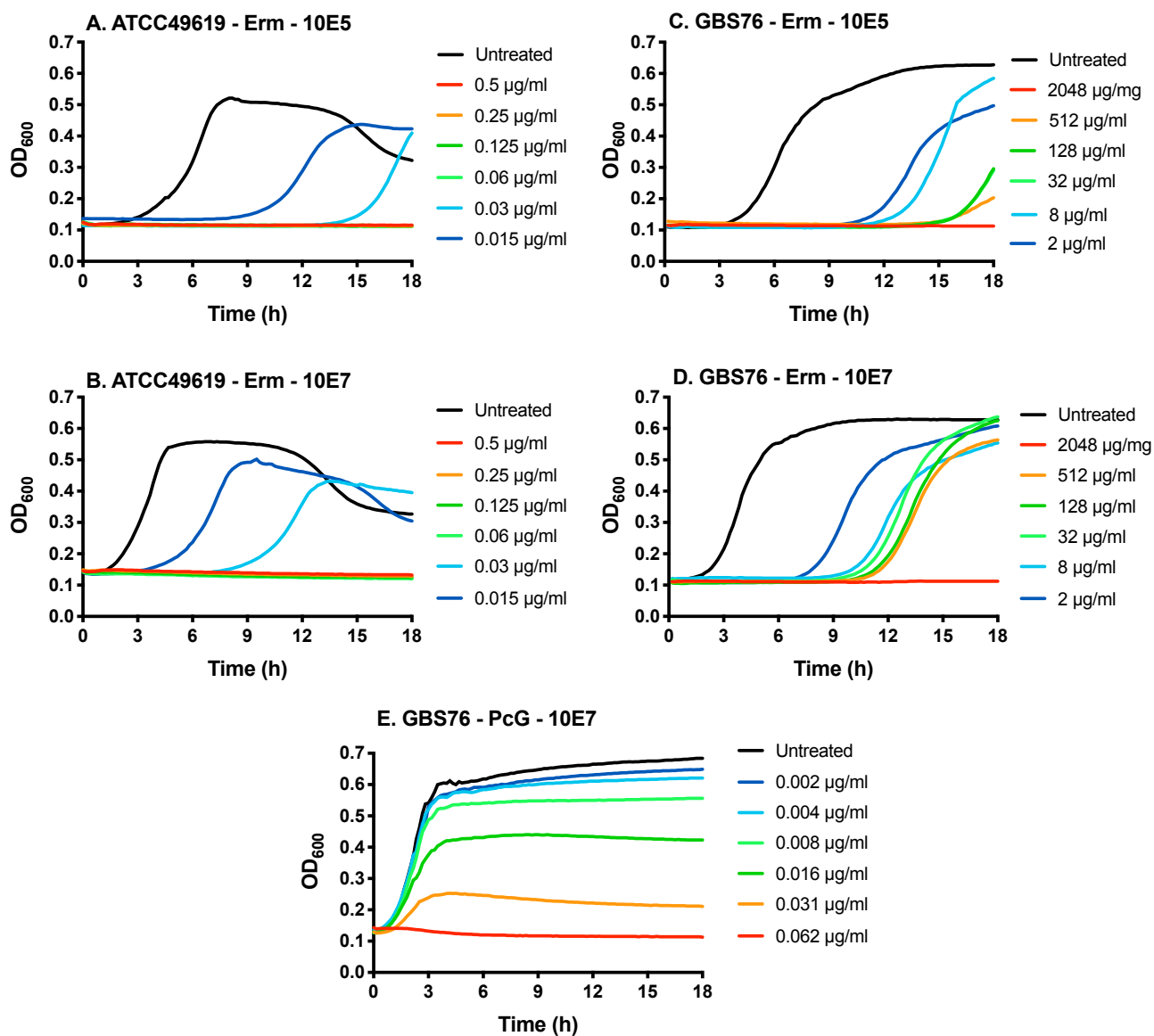


Figure S3. Validation of MIC assays using different inoculum concentrations. MIC growth curves are shown for *S. pneumoniae* ATCC49619 or GBS strain 76 exposed to various erythromycin concentrations with the bacterial inoculum being 10⁵ CFU/ml (A and C, respectively) or 10⁷ CFU/ml (B and D, respectively). The growth curves show a right shift in the presence of sub-MIC concentration of erythromycin that is less substantial at the higher inoculum, without the MIC for erythromycin being affected. Exposure of GAS 76 to penicillin G (E) resulted in a concentration-dependent decrease in maximal optical density, without a right-shift of the growth curve.

References

1. **Avery, O. T., Macleod, C. M., and McCarty, M.** 1944. Studies on the chemical nature of the substance inducing transformation of pneumococcal types: induction of transformation by a deoxyribonucleic acid fraction isolated from pneumococcus type III. *J Exp Med* **79**:137-158.
2. **Tyx, R. E., Roche-Hakansson, H., and Hakansson, A. P.** 2011. Role of Dihydrolipoamide Dehydrogenase in Regulation of Raffinose Transport in *Streptococcus pneumoniae*. *J Bacteriol* **193**:3512-3524.
3. **Echenique, J., Kadioglu, A., Romao, S., Andrew, P. W., and Trombe, M. C.** 2004. Protein serine/threonine kinase StkP positively controls virulence and competence in *Streptococcus pneumoniae*. *Infect Immun* **72**:2434-2437.
4. **Balachandran, P., Brooks-Walter, A., Virolainen-Julkunen, A., Hollingshead, S. K., and Briles, D. E.** 2002. Role of pneumococcal surface protein C in nasopharyngeal carriage and pneumonia and its ability to elicit protection against carriage of *Streptococcus pneumoniae*. *Infect Immun* **70**:2526-2534.
5. **Brown, J. S., Gilliland, S. M., Ruiz-Albert, J., and Holden, D. W.** 2002. Characterization of pit, a *Streptococcus pneumoniae* iron uptake ABC transporter. *Infect Immun* **70**:4389-4398.
6. **Hakenbeck, R., Briese, T., Chalkley, L., Ellerbrok, H., Kalliokoski, R., Latorre, C., Leinonen, M., and Martin, C.** 1991. Variability of penicillin-binding proteins from penicillin-sensitive *Streptococcus pneumoniae*. *J Infect Dis* **164**:307-312.
7. **Jorgensen, J. H., Doern, G. V., Ferraro, M. J., Knapp, C. C., Swenson, J. M., and Washington, J. A.** 1992. Multicenter evaluation of the use of Haemophilus test medium for broth microdilution antimicrobial susceptibility testing of *Streptococcus pneumoniae* and development of quality control limits. *J Clin Microbiol* **30**:961-966.
8. **Bolukaoto, J. Y., Monyama, C. M., Chukwu, M. O., Lekala, S. M., Nchabeleng, M., Maloba, M. R., Mavenyengwa, R. T., Lebelo, S. L., Monokoane, S. T., Tshepuwane, C., and Moyo, S.**

- R. 2015. Antibiotic resistance of *Streptococcus agalactiae* isolated from pregnant women in Garankuwa, South Africa. *BMC Res Notes* **8**:364.
9. **Seppälä, H., Skurnik, M., Soini, H., Roberts, M. C., and Huovinen, P.** 1998. A novel erythromycin resistance methylase gene (*ermTR*) in *Streptococcus pyogenes*. *Antimicrob Agents Chemother* **42**:257-262.
10. **Woodbury, R. L., Klammer, K. A., Xiong, Y., Bailiff, T., Glennen, A., Bartkus, J. M., Lynfield, R., Van Beneden, C., Beall, B. W., and Active, B. C. S. T.** 2008. Plasmid-Borne *erm(T)* from invasive, macrolide-resistant *Streptococcus pyogenes* strains. *Antimicrob Agents Chemother* **52**:1140-1143.
11. **Gygax, S. E., Schuyler, J. A., Kimmel, L. E., Trama, J. P., Mordechai, E., and Adelson, M. E.** 2006. Erythromycin and clindamycin resistance in group B streptococcal clinical isolates. *Antimicrob Agents Chemother* **50**:1875-1877.
12. **Daly, M. M., Doktor, S., Flamm, R., and Shortridge, D.** 2004. Characterization and prevalence of *MefA*, *MefE*, and the associated *msr(D)* gene in *Streptococcus pneumoniae* clinical isolates. *J Clin Microbiol* **42**:3570-3574.