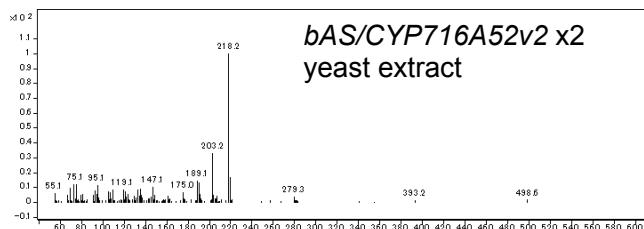
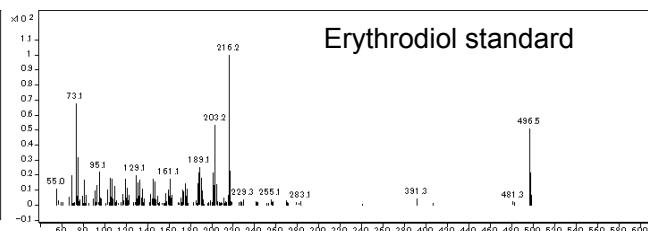
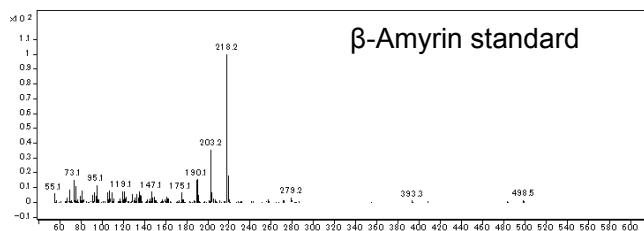
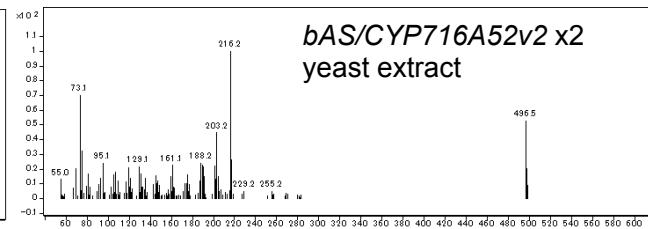
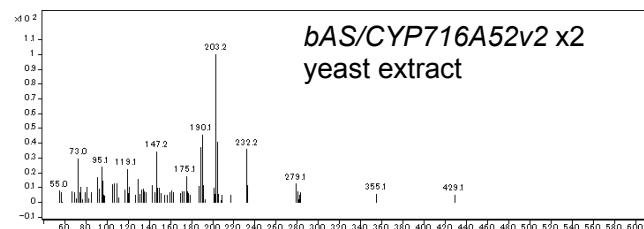
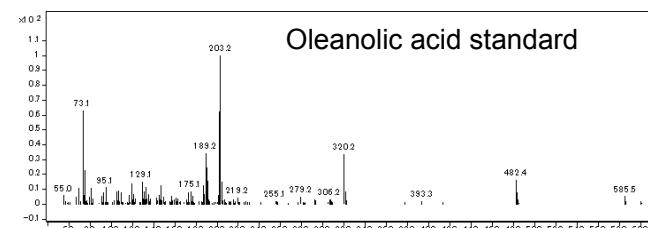
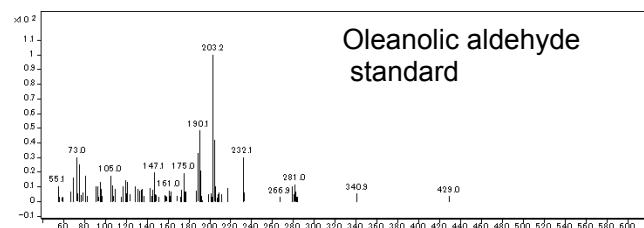
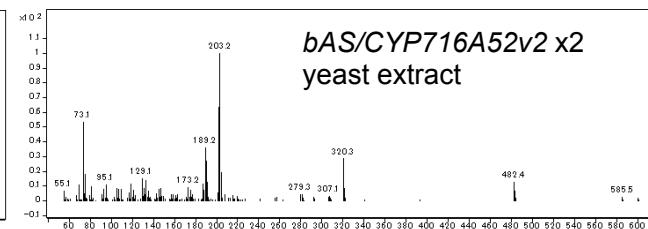
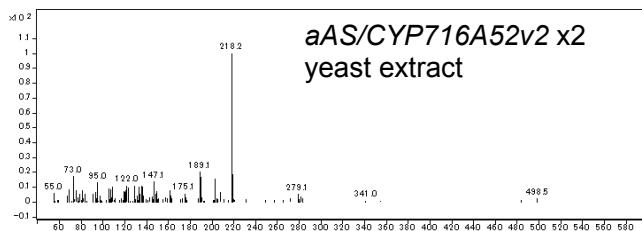
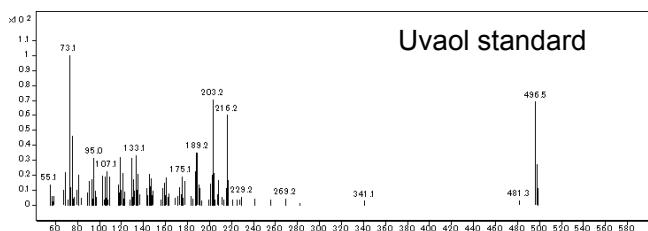
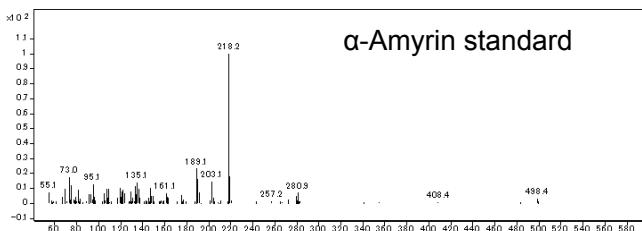
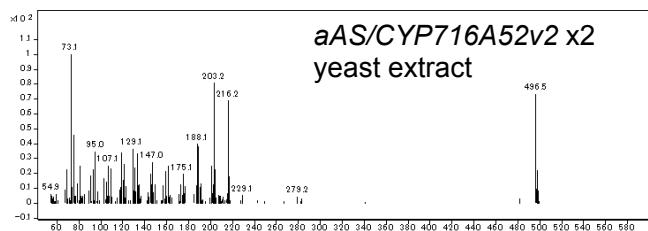
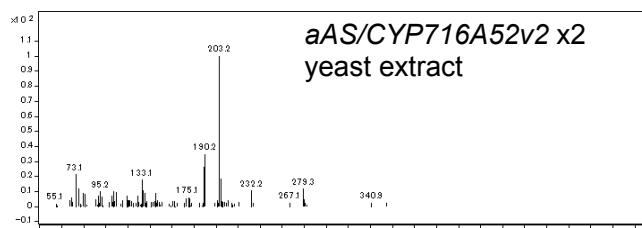
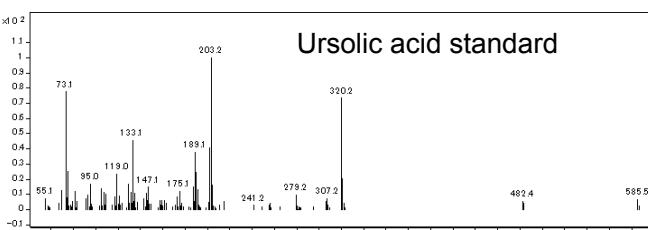
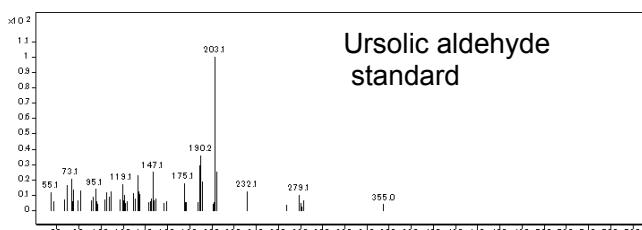
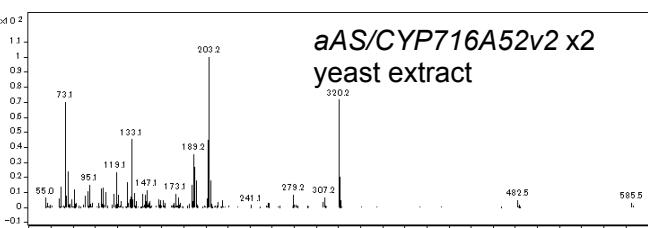
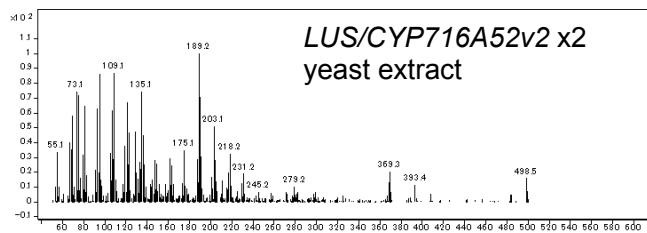
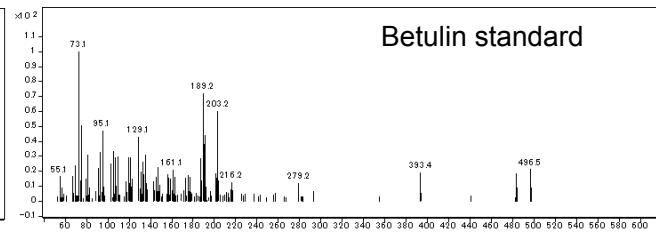
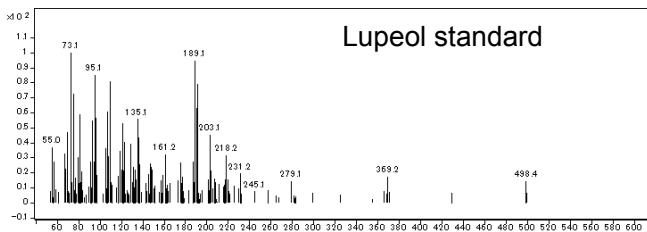
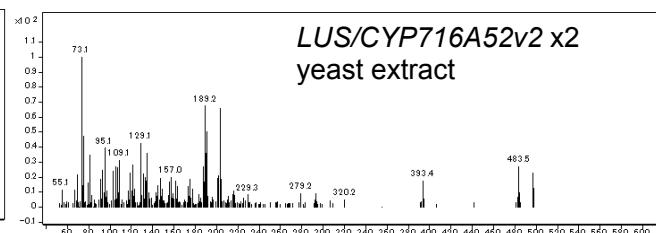
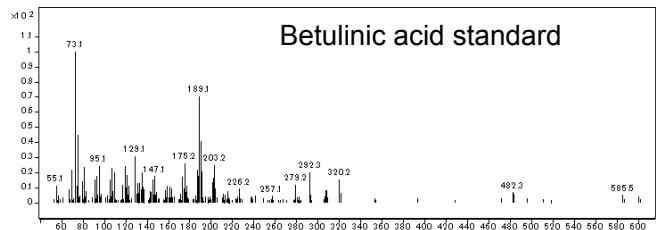
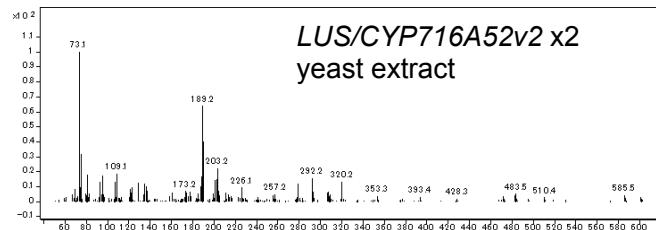
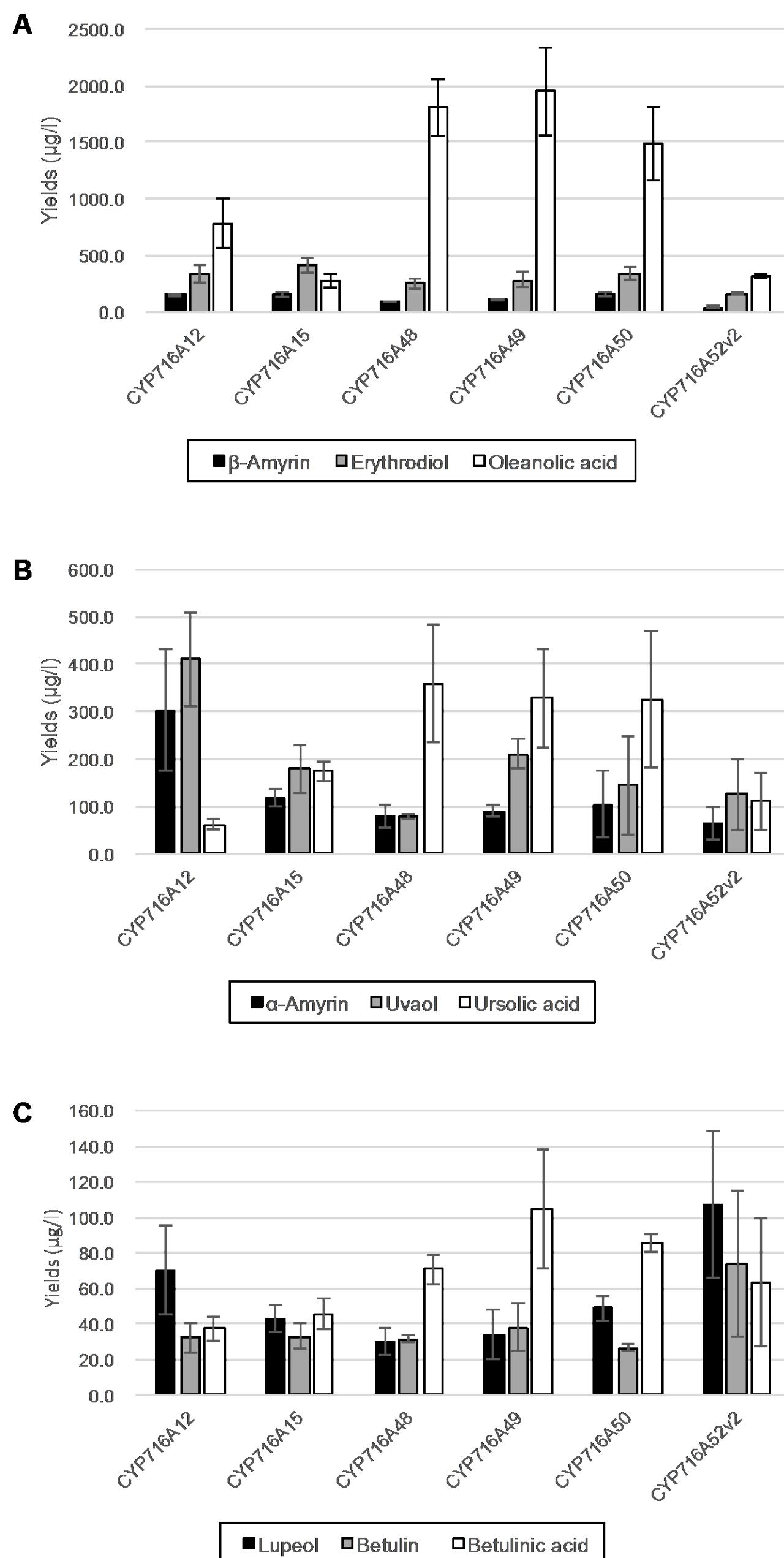


**A****Peak 1 (Rt=29.04 min)****Peak 2 (Rt=34.44 min)****Peak 3 (Rt=35.79 min)****Peak 4 (Rt=36.57 min)**

**Figure S1. Mass spectra of target compounds and authentic standards shown in Figure 2.**  
**A)** Mass spectra of β-amyrin derivatives. **B)** Mass spectra of α-amyrin derivatives. **C)** Mass spectra of lupeol derivatives.

**B****Peak 5 (Rt=29.93 min)****Peak 6 (Rt=35.63 min)****Peak 7 (Rt=37.32 min)****Peak 8 (Rt=38.40 min)****Figure S1. Cont.**

**C****Peak 9 (Rt=30.21 min)****Peak 10 (Rt=36.33 min)****Peak 12 (Rt=37.29 min)****Figure S1. Cont.**



**Supplemental Figure S2. Absolute amounts of detected triterpenoids.**

The amounts of (A)  $\beta$ -amyrin derivatives in *bAS/CPR/CYP716A*-expressing yeast extracts, (B)  $\alpha$ -amyrin derivatives in *aAS/CPR/CYP716A*-expressing yeast extracts, and (C) lupeol derivatives in *LUS/CPR/CYP716A*-expressing yeast extracts are given. Quantitation and error bars correspond to the mean and standard deviation, respectively ( $n = 3$ ).

CYP716A12	1	MEPNTYLSLILFVFSISSLFFYKQKSPLN---LPPGKGMPIIGESLEFLSTGWKGKPEKKIFDRMKYSS-E-LFKTSIVGESTVCCGAASNKFLFSNENKLVT	105
CYP716A15	1	-MEVFLSLLLIIFVLSVSIHLHFLYKIRSHFT-PNLPGKIGIPMVGESLEFLSTGWKGKPEKKIFDRISKYSSE-VFKTSILGEPAAMFGAAACNKFLFSNENKLVI	107
CYP716A48	1	-MEFVYSLCLFVFLISLHFLYKNIKSSFG---QIPPGKTGIVPVIGESLEFLSTGWKGKPEKKIFDRISKYSSE-VFKTSILGEPAAMFGAAACNKFLFSNENKLVI	106
CYP716A49	1	-MEFLFCGLTFL-LSLASLYLNYINIST-KG-YRVPPTGIVPVGESLEFLSTGWKGKPEKKIFDRILSKYAPNQFTKTSILGEPAAMFGAAACNKFLFSNENKLVI	106
CYP716A50	1	-MEFVYSLCLFLLTFLSLHFLYKSRSLALSG---PLPPGKTGIPMVGESLEFLSTGWKGKPEKKIFDRMAYKSSH-VFKTILLGEKAACFGCAPNKFLFSNENKLVI	106
CYP716A52v2	1	-MEFVYVPLLSLFLVLTISLHFYKSKPSSSGFPFLPPGKTGIPITGESYELSTGWKGKPEKKIFDRMTKYSM-VFKTISFEGPAAMFGAAACNKFLFSNENKLVI	108
Proline			
CYP716A12	106	AWPVPSVKVIFPITSLSLNSLKEEKS1KMRKLLPQFPKPEALQRQYGVMDVIAQRFVTHDKNETWPLAKRYTFLLACRLFMSVEDENHVAKEFSDPQLIAQGTLSP	215
CYP716A15	108	AWPVSVDKVIFPSSQTQSS-KEEA1KMRKLLPQFPKPEALQRQYIGIMDHIAQRIFADSWDRDVEIWPLAKRFTFLWFLACRLFMSIEDPAHVAKEFKPFIWLGTLITVP	216
CYP716A48	107	AWPVSVKVIFPSSQTQSS-KEEA1KMRKLLPQFPKPEALQRQYGVMDHIAQRIFSDGWDKNEVWPLAKRFTFLWFLACRLFMSIEDPAHVAKEFKPFIWLGTLITVP	215
CYP716A49	107	AWPVSVDKVIFPSSQTQSS-KEEAKMRKLLPQFPKPEALQRQYITPMIDTAIRMESGWDGDKIVFVPLAKRFTFLWFLACRLFMSIEDPAHVAKEFPNDIAGHTLSP	215
CYP716A50	107	AWPVSVKVIFPSSQTQSS-KEEAKMRKLLPQFPKPEALQRQYIGIMDHIAQRIFSSGGENKDQVWVPLC1VTFD1A2RSLEMSVEEPTAKEAKLLEPFWNLASGLTSP	215
CYP716A52v2	109	AWPVSVKVIFPSSQTQSS-KEEAKMRKLLPQFPKPEALQRQYIGIMDQIANFESQENKEVNWVPLAKS1VTFD1A2KV1FVEEPAQVAILLEPESATASGHTSP	217
Oxygen activation			
CYP716A12	216	IDLPGTPPNKATKASNFIRKEITLIIKORRDLDAEGTAS1TQDI1SHMLLTSDENGKSMELNIAADKILGLLGGHDATASVCTFLVYLCEPITIDKYMQEOMEIAKS	325
CYP716A15	217	IDLPGTPPHRAIKASNFIRKEITLIIKORKDIAEGKAS1QXDI1SHMLLTADEDGCHMENMXTADKILGLLGGHDATASAAITFLIKYMAELPITYKVEQEOMEIAKS	326
CYP716A48	216	IDLPGTPPHRAIKSSNFIRKEITLIIKORKDIAEGKAS1TQDI1SHMLLTSDENGKFMELDIAADKILGLLGGHDATASAACTFVVKYLAELPITYKVEQEOMEIAKS	325
CYP716A49	216	VNLPGTPPNRGK1SSNVWVKEITLIIKORKDIAEGKAS1TQDI1SHMLLTADEDGFMELDIAADKILGLLGGHDATASAACTFVVKYLAELPITYKVEAVCKEQEOMEIAKS	325
CYP716A50	216	IDLPGTPPNRANKASNOIRKMLVALIKORKDIAEGKAS1TQDI1SHMLLTSDENGKFMELDIAADKILGLLGGHDATASAACTFVVKFELAELPITYKVEQEOMEIVKS	325
CYP716A52v2	218	IDLPGTPNSATKSSIVRKLVG1ITLIIKORKDIAEGKAS1TQDI1SHMLLTSDESCKFMCEGDIADKILGLLGGHDATASAACTFVVKFELAELPQTYKVEQEOMEIVKS	327
Heme binding			
CYP716A12	326	KPAGELLNWDD1KLYYSNNVACEVLR1SPPLQGCF-REAIT1DFMFNGFS1PKWGKWLWYSANSTHKNAECFPMPKEDP1TRFEQGSPAPYTFVVFPGGGPRMCPGKEYARL	434
CYP716A15	327	KAPGE--NWDDVOMYRYSNNVACEVLR1APPLQGAF-FREAIT1DFMFNGFS1PKWGKWLWYSANSTHKSPECPOPENFDP1TRFEQGSPAPYTFVVFPGGGPRMCPGKEYARL	434
CYP716A48	326	KAPGEELLNWDD1KLYYSNNVACEVLR1APPLQGCF-REAIT1DFMFNGFS1PKWGKWLWYSANSTHRNPECPOPENFDP1TRFEQGSPAPYTFVVFPGGGPRMCPGKEYARL	434
CYP716A49	326	KAEGLLNEWDD1KLYYSNNVACEVLR1APPLQGAF-REAI1DFMFNGFS1PKWGKWLWYSANSTHRNPECPOPENFDP1TRFEQGSPAPYTFVVFPGGGPRMCPGKEYARL	434
CYP716A50	326	KAPGEELLNWDD1KLYYSNNVACEVLR1APPLQGAF-REALAD1MFNGFS1PKWGKWLWYSANSTHRNPECPOPENFDP1TRFEQGSPAPYTFVVFPGGGPRMCPGKEYARL	434
CYP716A52v2	328	KKAGELLNEWDD1KLYYSNNVACEVLR1APPLQGAF-REALSDFTYNGFS1PKWGKWLWYSANSTHINSEVEPEP1KEDPSRF1GAGP1PES1VVFPGGGPRMCPGKEYARL	436
ExoR			
CYP716A12	435	E1LVEHMNLVVKRFEKVIWPKDEK1I1VDP1PKAKDLP1IRLYPHKA----	479
CYP716A15	435	E1LVMHMNLVVKRFEKVIWPKDEK1I1VDP1PKAKDLP1IRLYPHKA----	479
CYP716A48	435	E1LVEHMHLVVKRFEKVIWPKDEK1I1VDP1PKAKDLP1IRLYPHLA----	479
CYP716A49	435	E1LVEHMHLVVKRFEKVIWPKDEK1I1VDP1PKAKDLP1IRLYPH1PQTVAA	483
CYP716A50	435	E1LVEHMHLVVKRFEKVIWPKDEK1I1VDP1PKAKDLP1IRLYPHKA----	479
CYP716A52v2	437	E1LVEHMHLVVKRFEKVIWPKDEK1I1VDP1PKAKDLP1IRLYPHKA----	481
PERF			

### Supplemental Figure S3. Protein alignment of CYP716A enzymes used in this study.

Full primary sequences of CYP716s were aligned using GENETYX Mac (ver. 12.0).

Common features found in plant P450s are highlighted.

**Supplemental Table 1. List of primers used in this study**

No.	Target seq	Sequence (5' to 3')
1	CYP716A48 F	<u>CACCATGGAGTTCTTCTATGTCTCTCTTC</u>
2	CYP716A48 R	TTAACATTAAGGGGATAAAGAC
3	CYP716A49 F	<u>CACCATGGAGCTCTTCTTCCTT</u>
4	CYP716A49 R	TTAACAGCAACAATTGAGG
5	CYP716A50 F1	CGCTCACAAACAATCTGGAA
6	CYP716A50 F2	<u>CACCATGGAGTTTTCTATGTCTCTTG</u>
7	CYP716A50 R	TTAGGCCTTGTGTGGAAAAAA
8	CYP716A52v2 F1	<u>CACCATGGAACTCTTCTATGTCCCTCT</u>
9	CYP716A52v2 R	TTAGGCTTGTGTGGAAATAGGC

The underlined sequences were added to facilitate the unidirectional cloning of the product into pENTR/D-TOPO (Invitrogen)

**pplemental Table 2. Amino acid identities of the enzymes analyzed in this stu**

	CYP716A12	CYP716A15	CYP716A48	CYP716A49	CYP716A50	CYP716A52v2
CYP716A12						
CYP716A15	74					
CYP716A48	77	79				
CYP716A49	71	74	74			
CYP716A50	71	75	83	73		
CYP716A52v2	73	70	81	73	79	