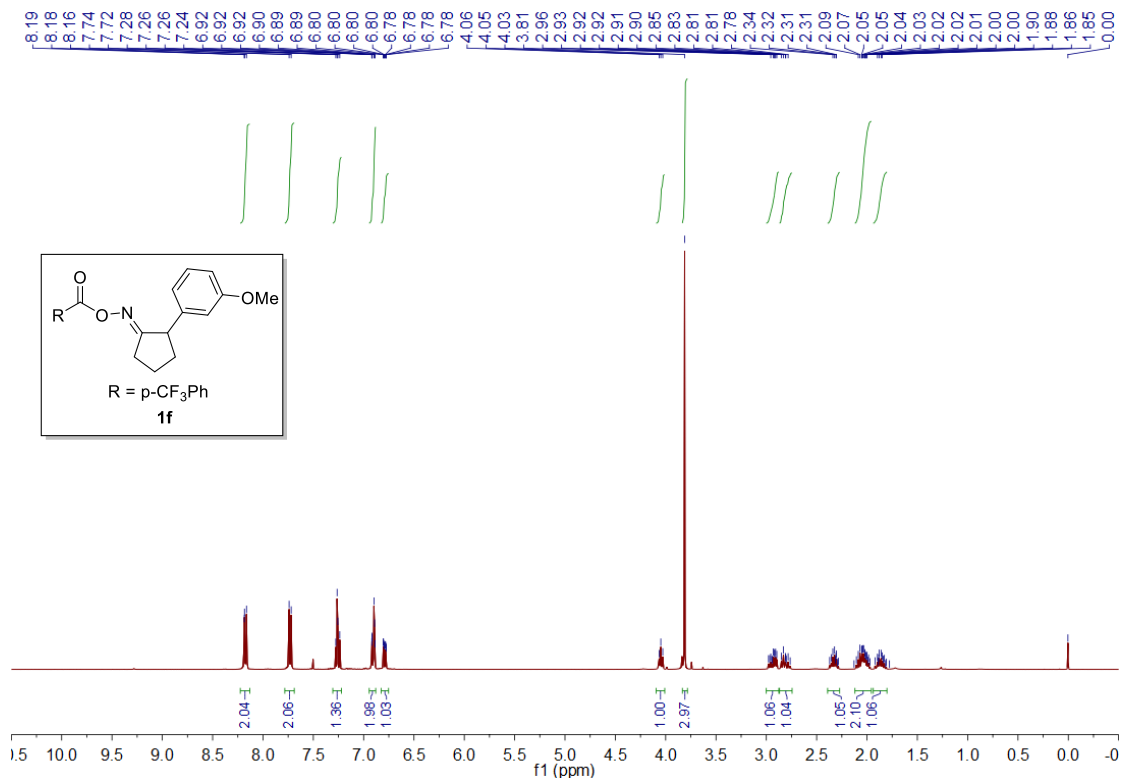


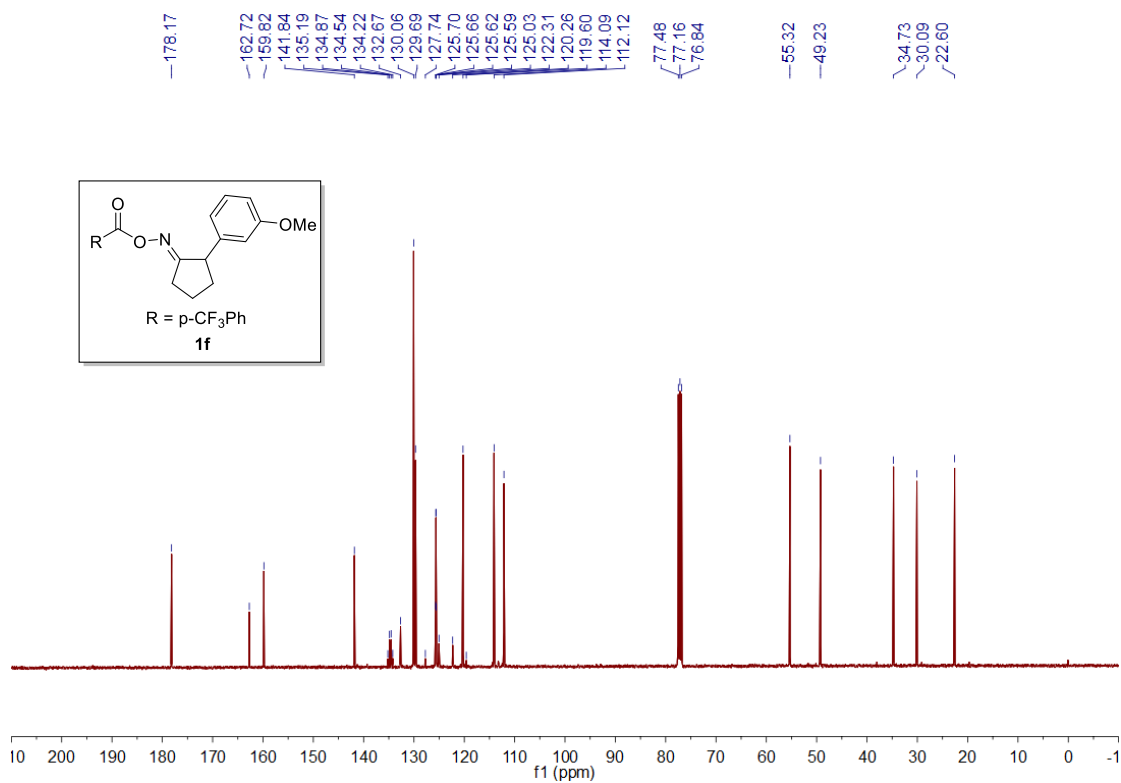
**Enantioselective cyanation via radical-mediated C–C single bond cleavage for
synthesis of chiral dinitriles**

Wang *et al.*

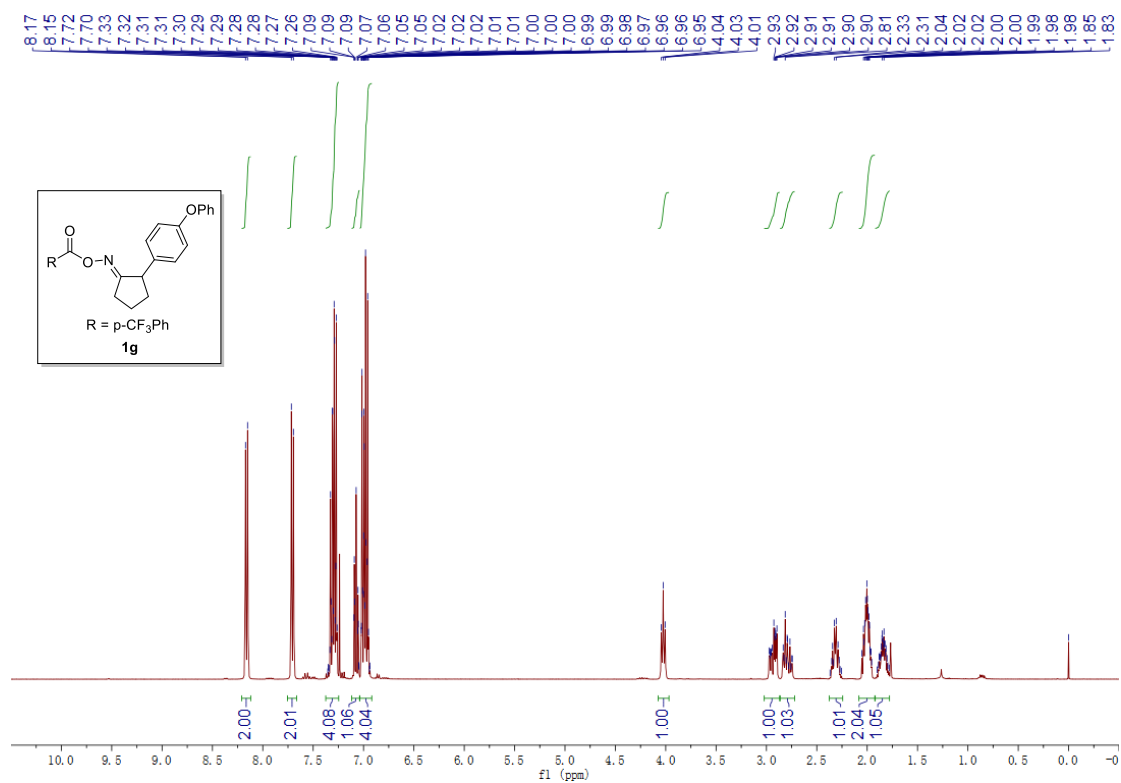
Supplementary Figures



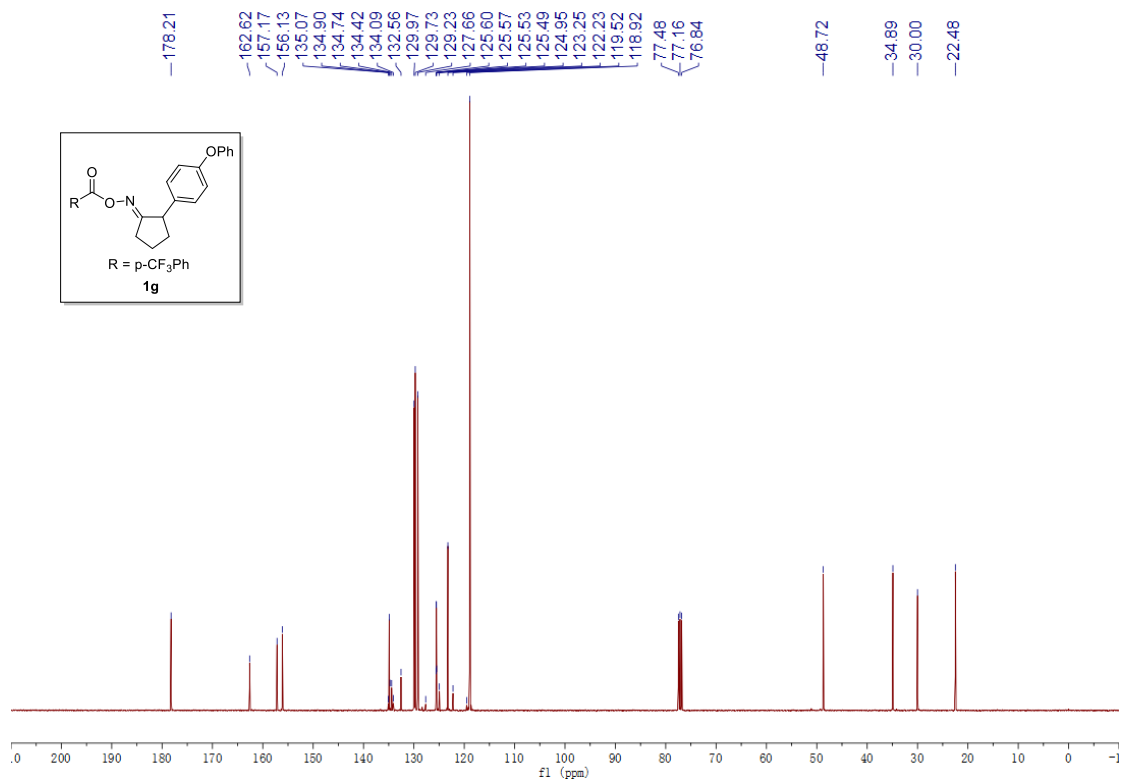
Supplementary Figure 1. ¹H NMR of **1f**



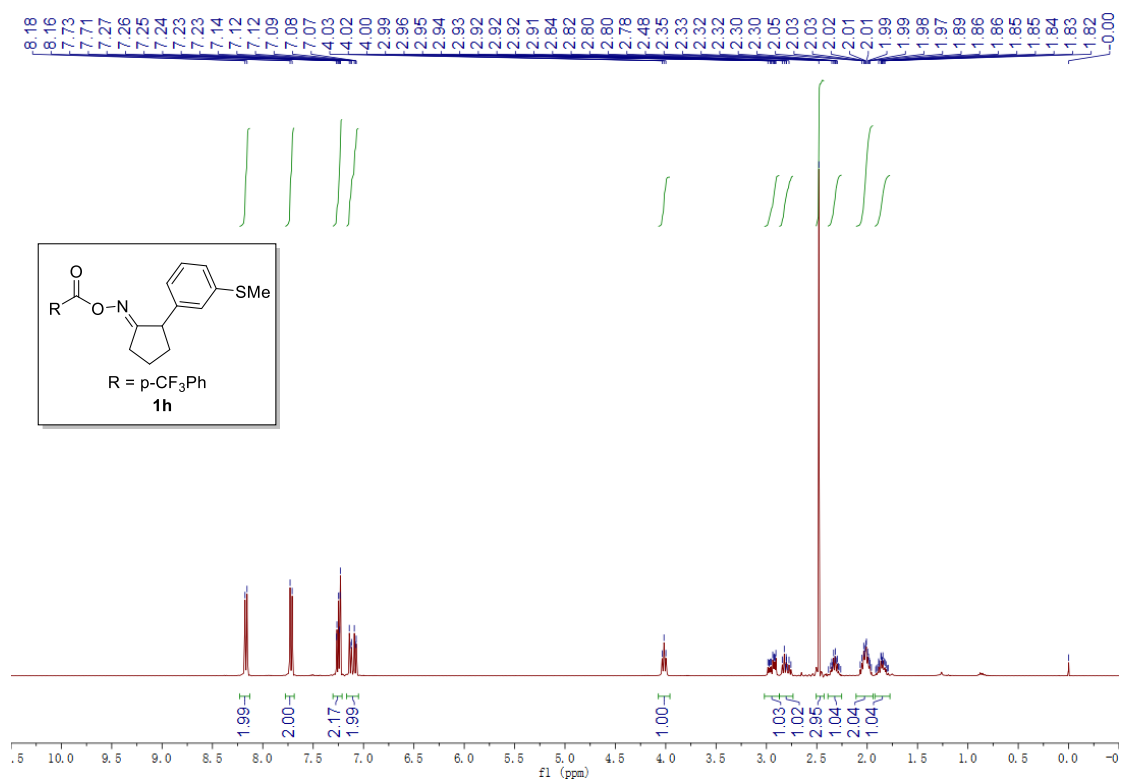
Supplementary Figure 2. ¹³C NMR of **1f**



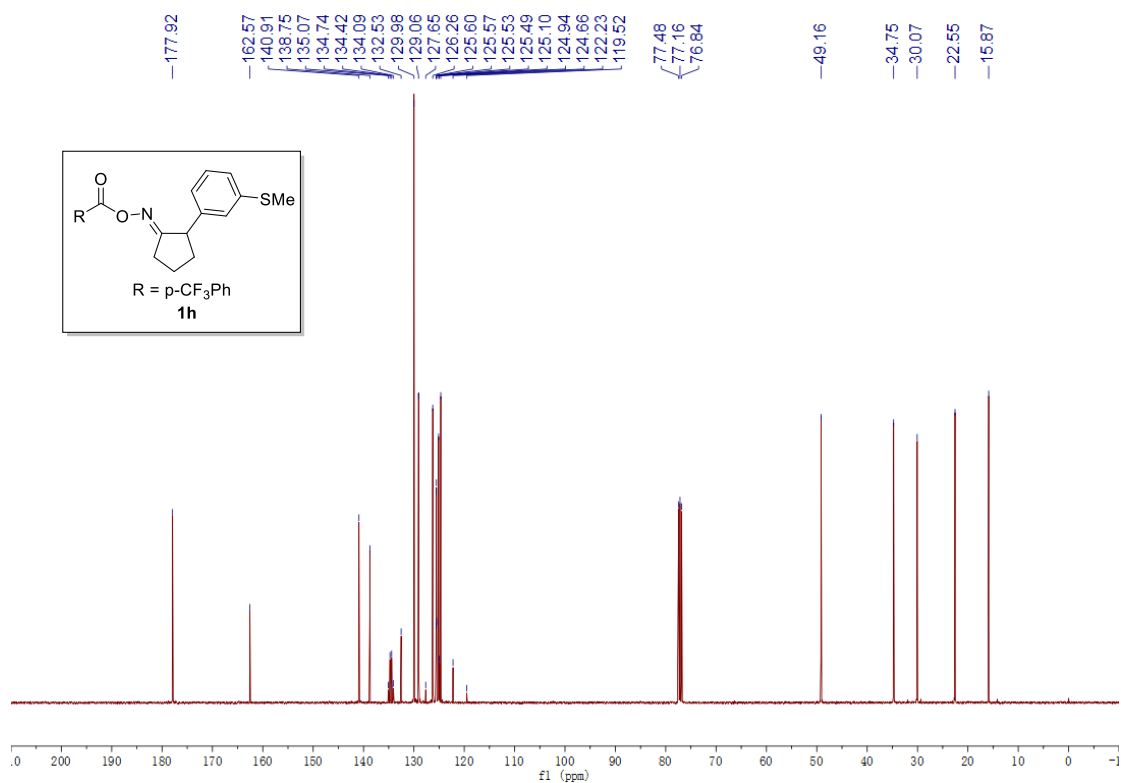
Supplementary Figure 3. ¹H NMR of **1g**



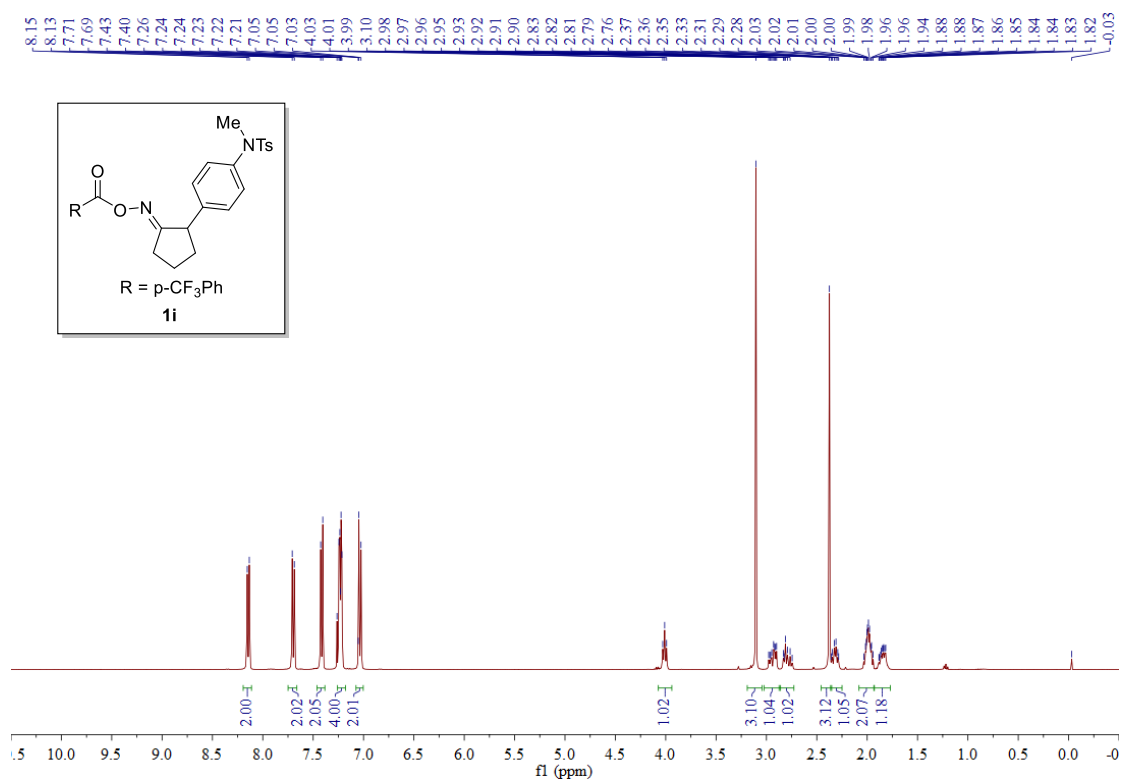
Supplementary Figure 4. ¹³C NMR of **1g**



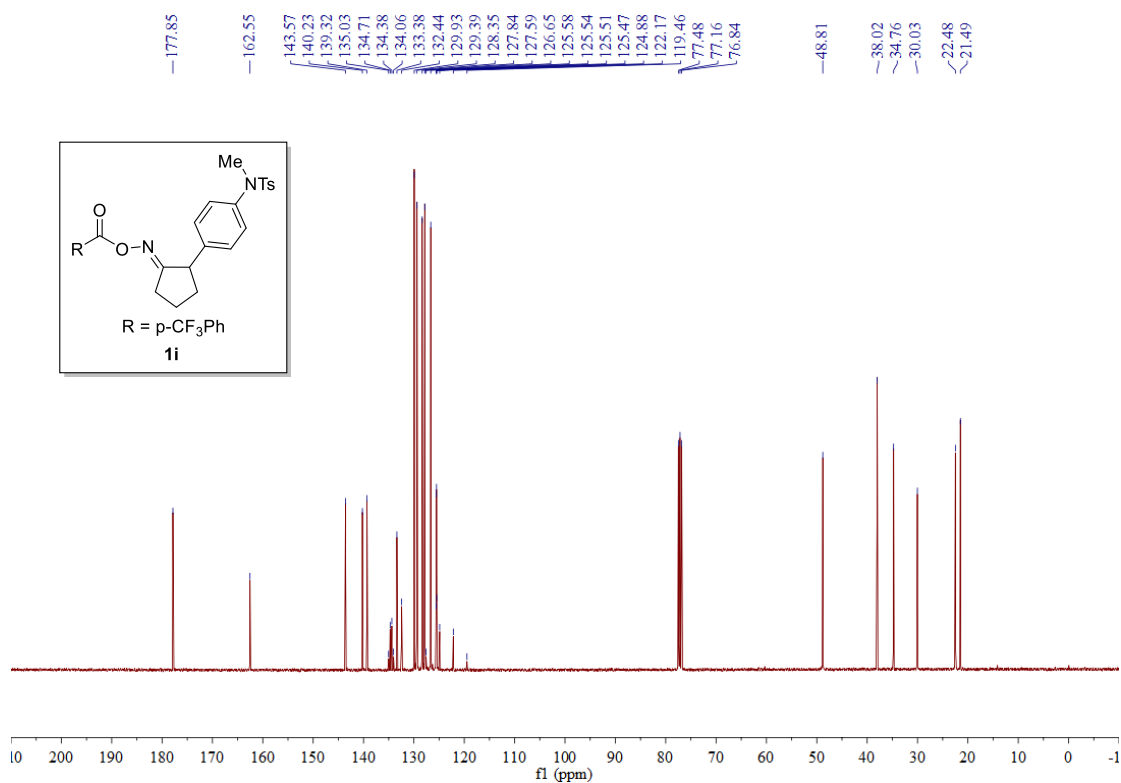
Supplementary Figure 5. ¹H NMR of **1h**



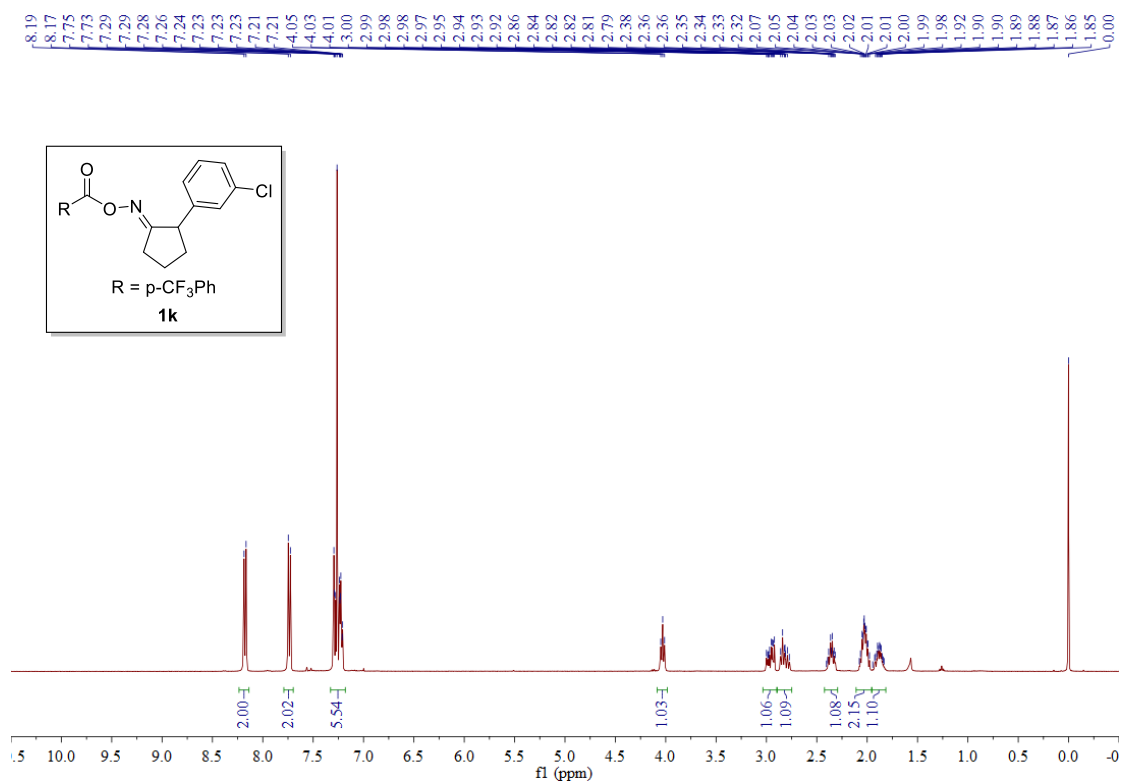
Supplementary Figure 6. ¹³C NMR of **1h**



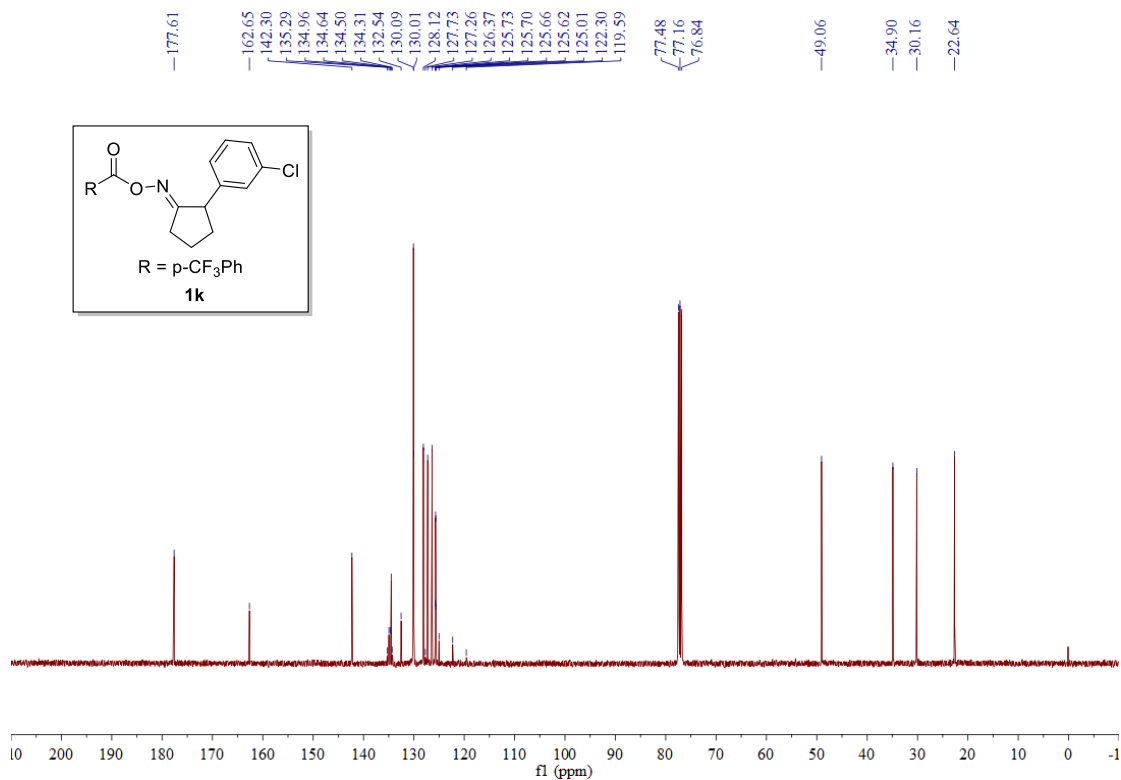
Supplementary Figure 7. ¹H NMR of **1i**



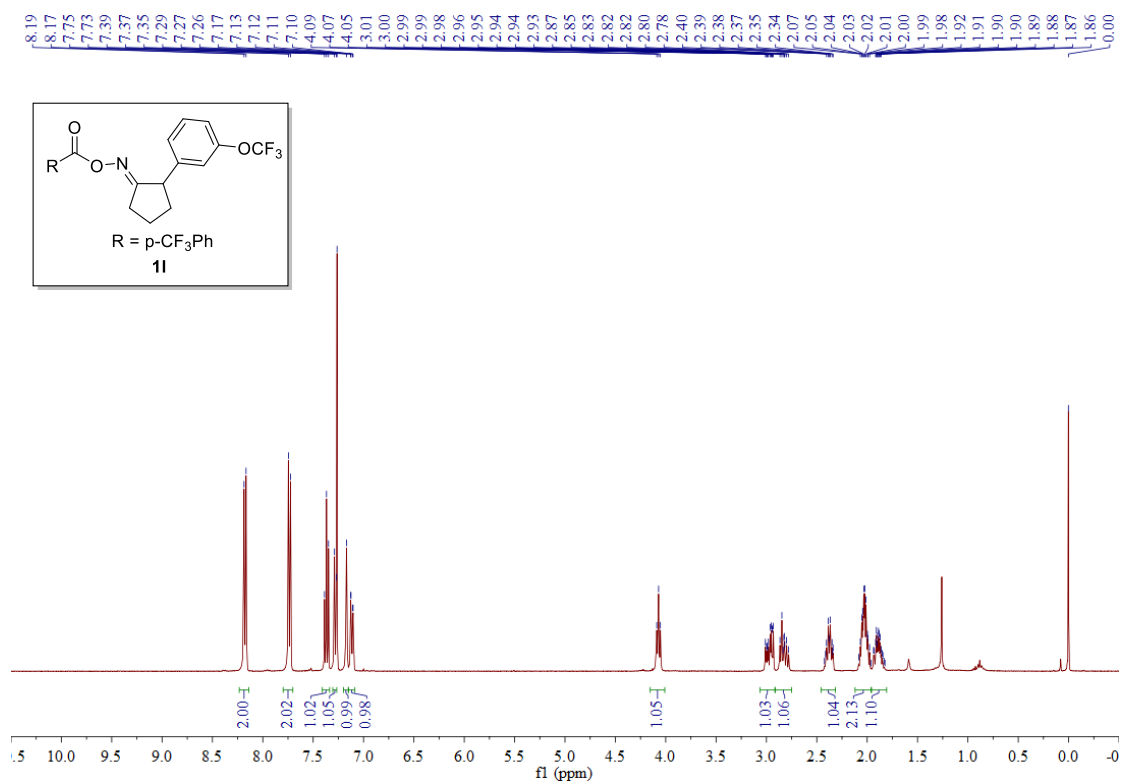
Supplementary Figure 8. ¹³C NMR of **1i**



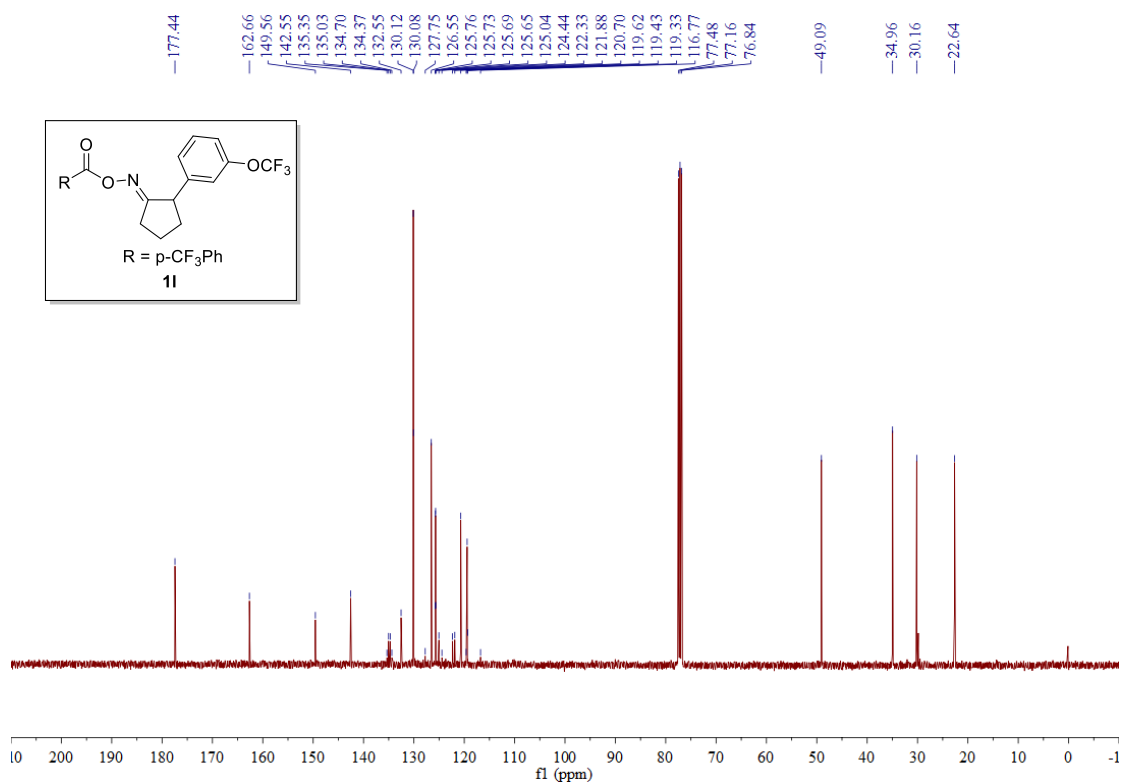
Supplementary Figure 9. ¹H NMR of **1k**



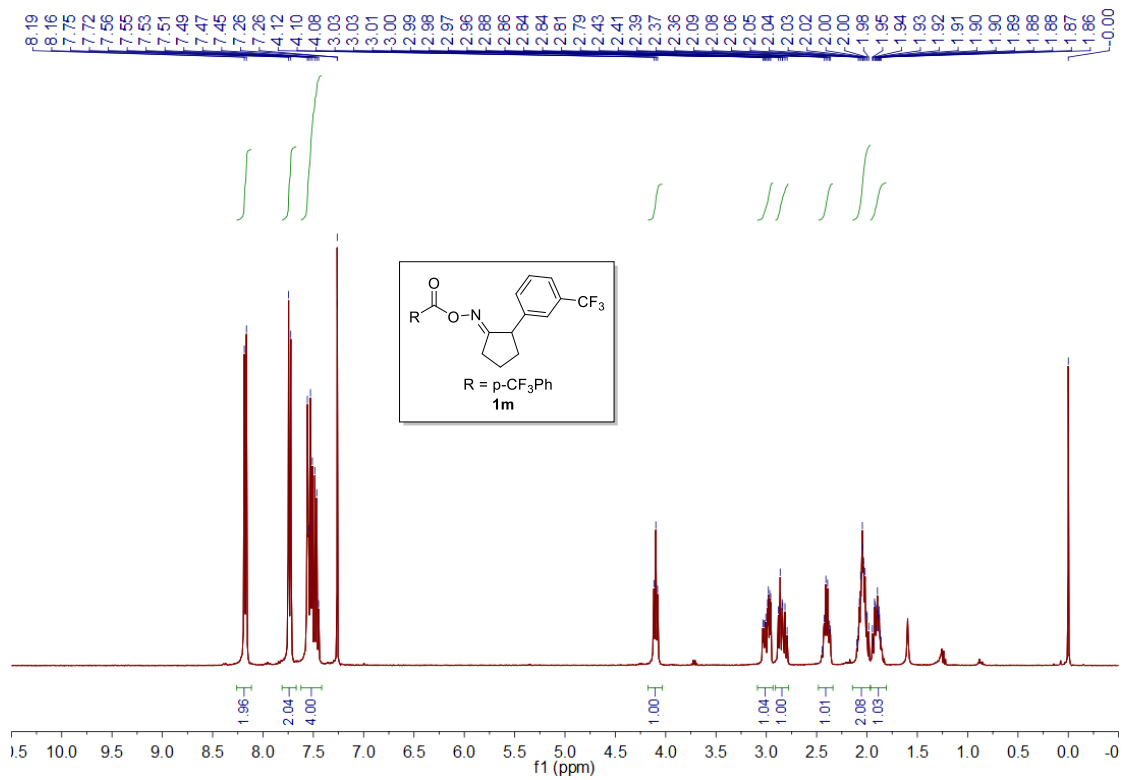
Supplementary Figure 10. ¹³C NMR of **1k**



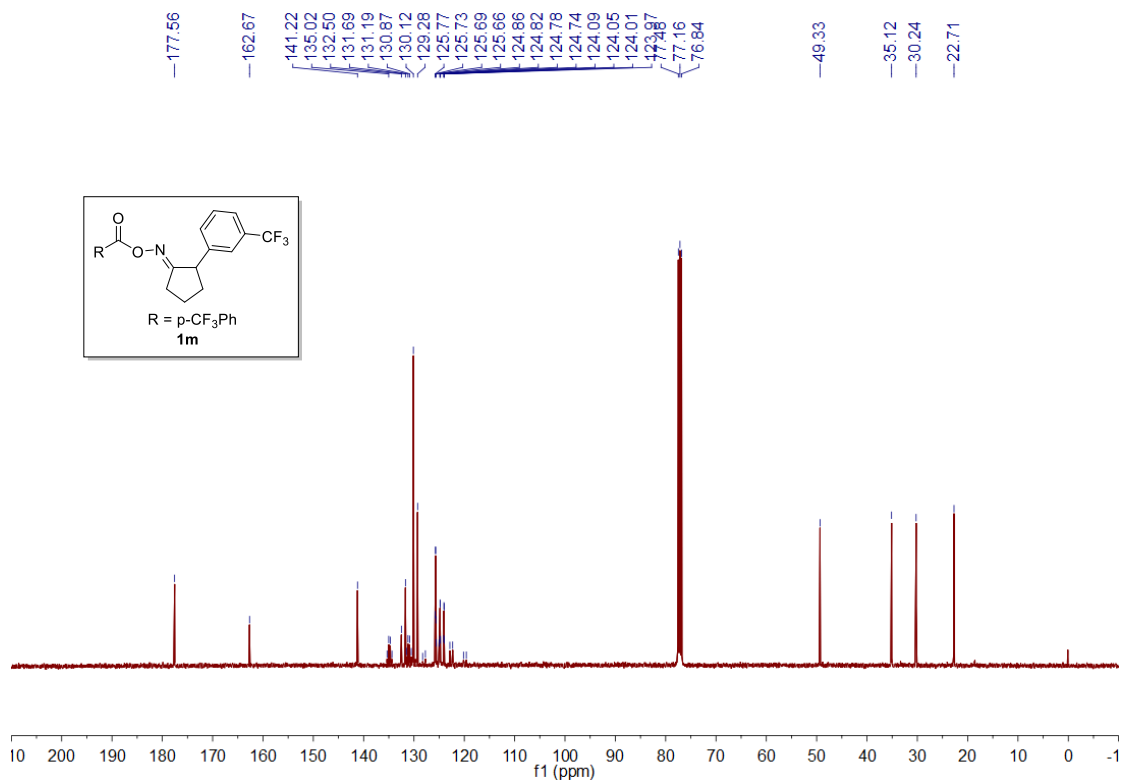
Supplementary Figure 11. ¹H NMR of **11**



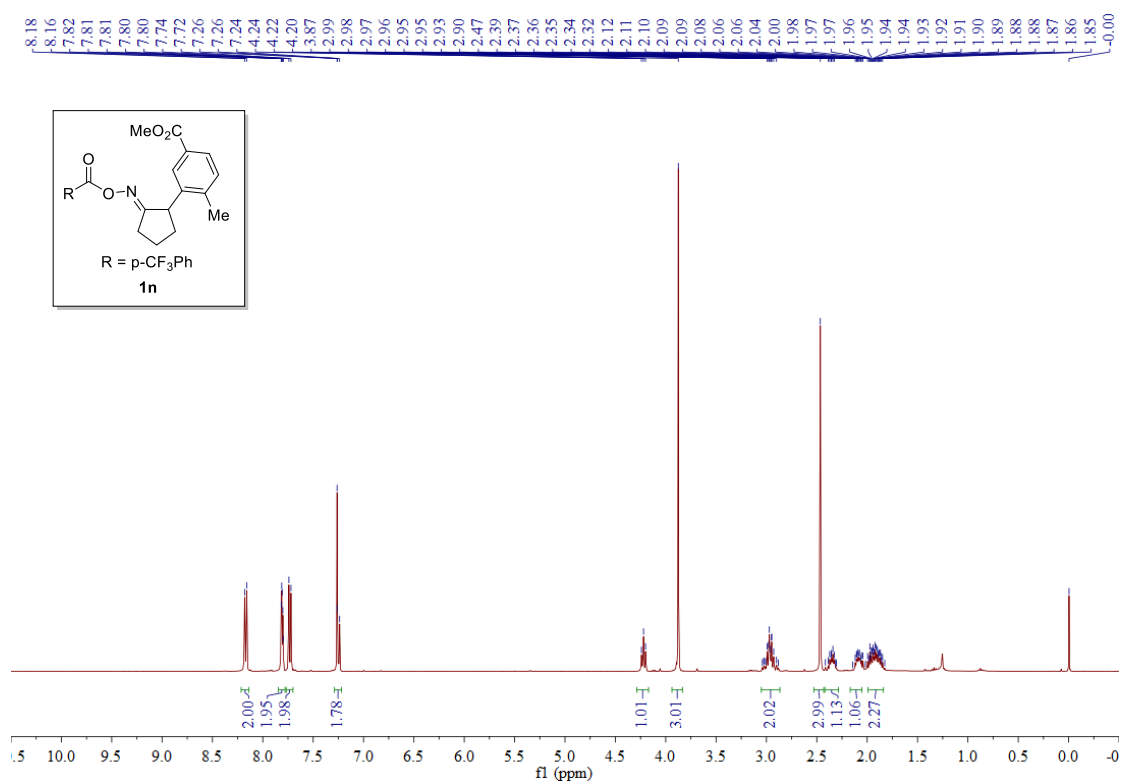
Supplementary Figure 12. ¹³C NMR of **11**



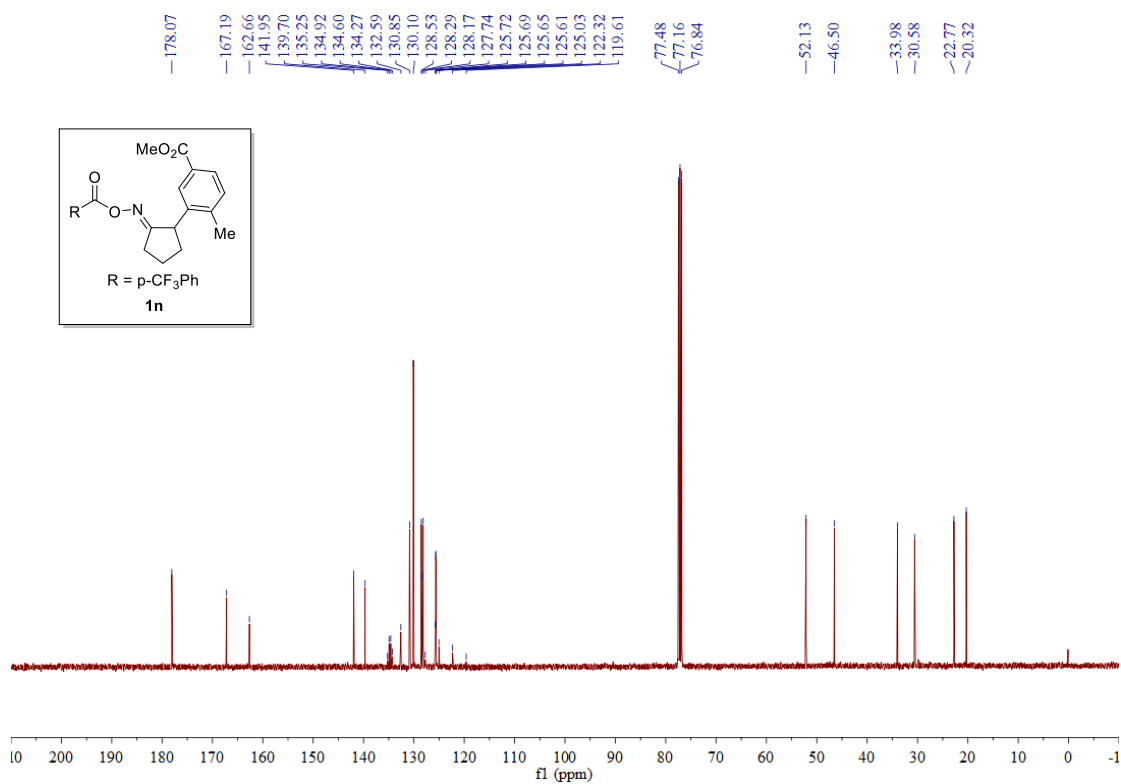
Supplementary Figure 13. ¹H NMR of **1m**



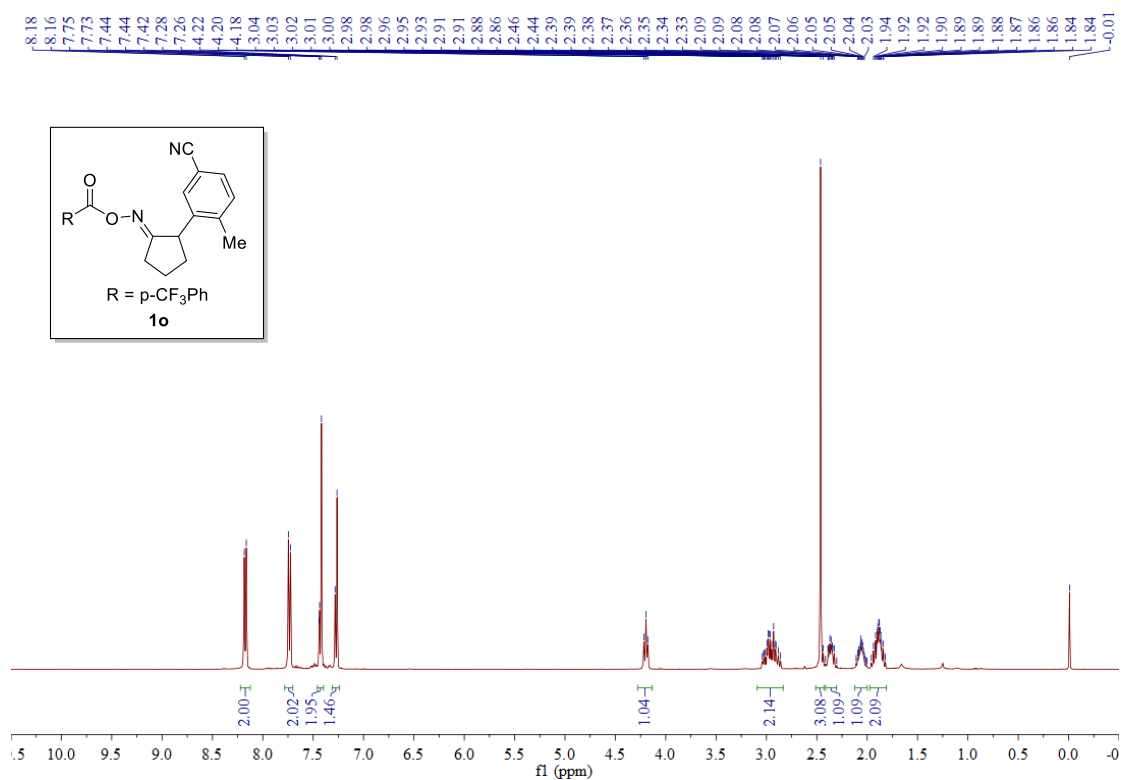
Supplementary Figure 14. ¹³C NMR of **1m**



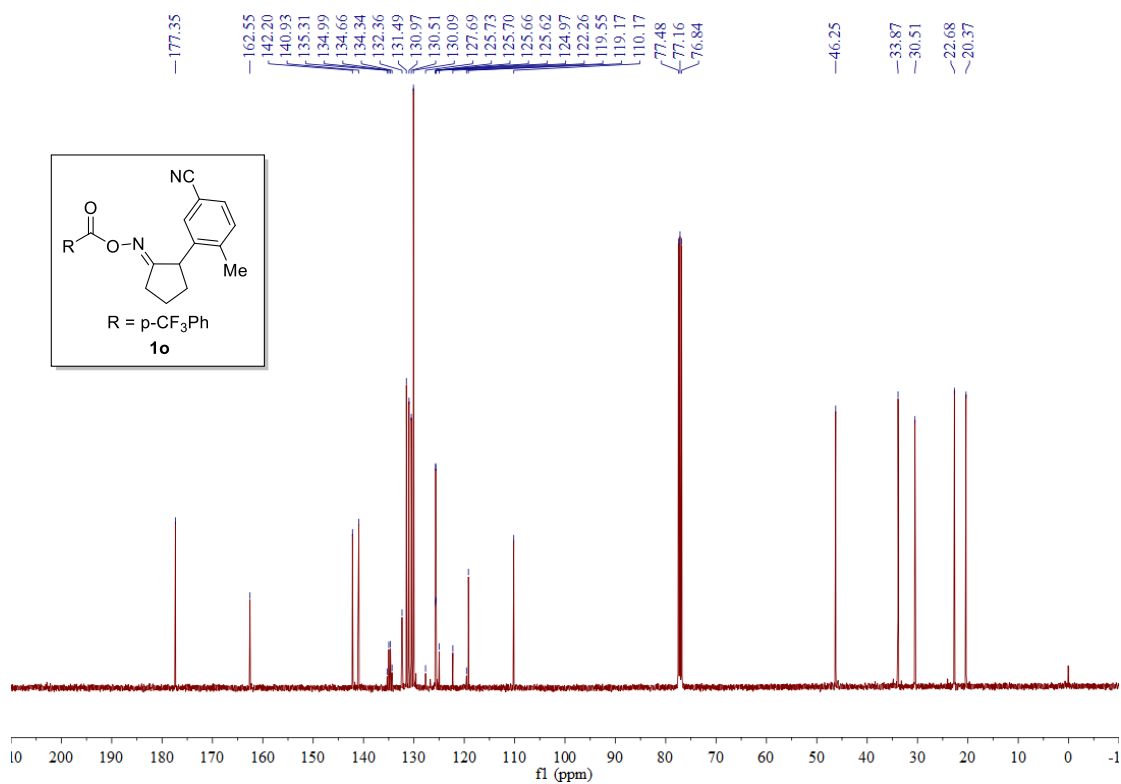
Supplementary Figure 15. ¹H NMR of **1n**



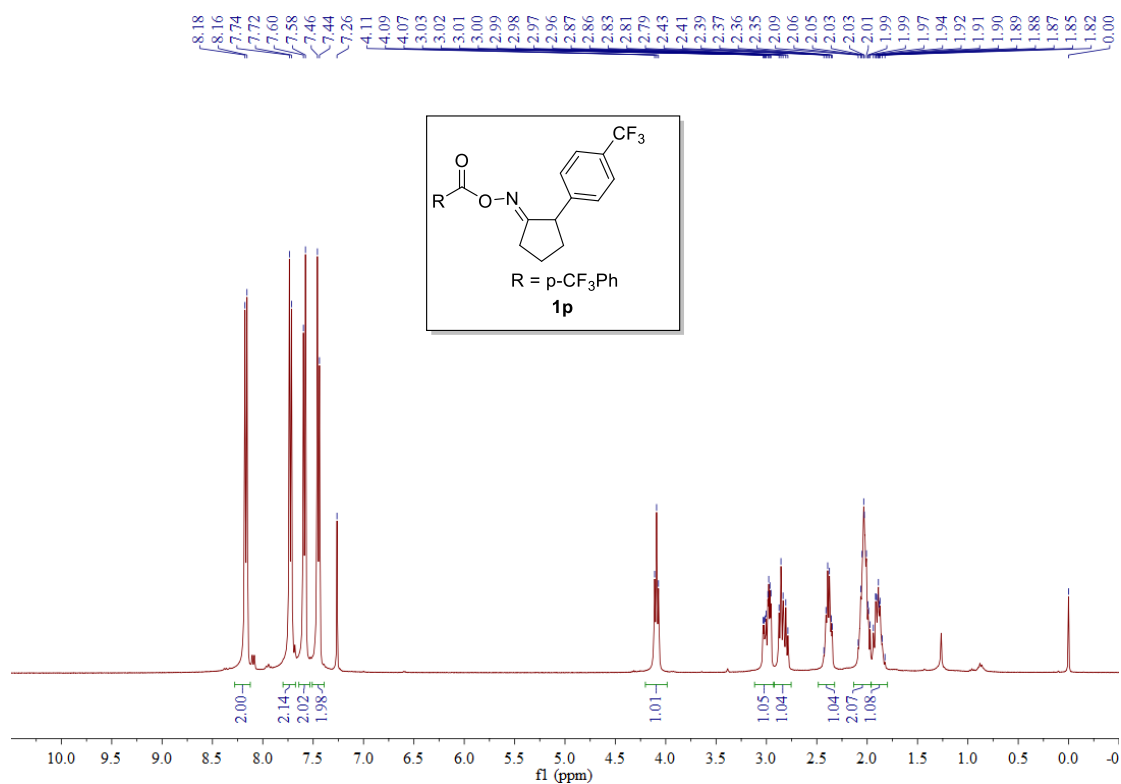
Supplementary Figure 16. ¹³C NMR of **1n**



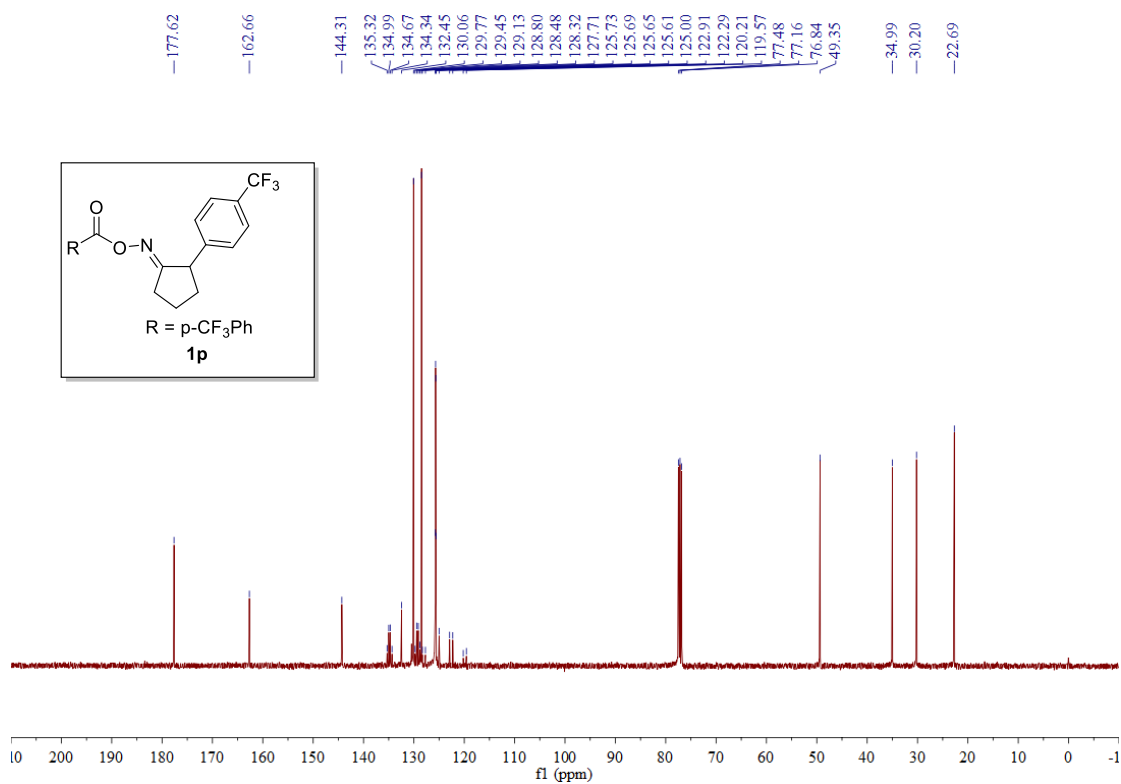
Supplementary Figure 17. ¹H NMR of **1o**



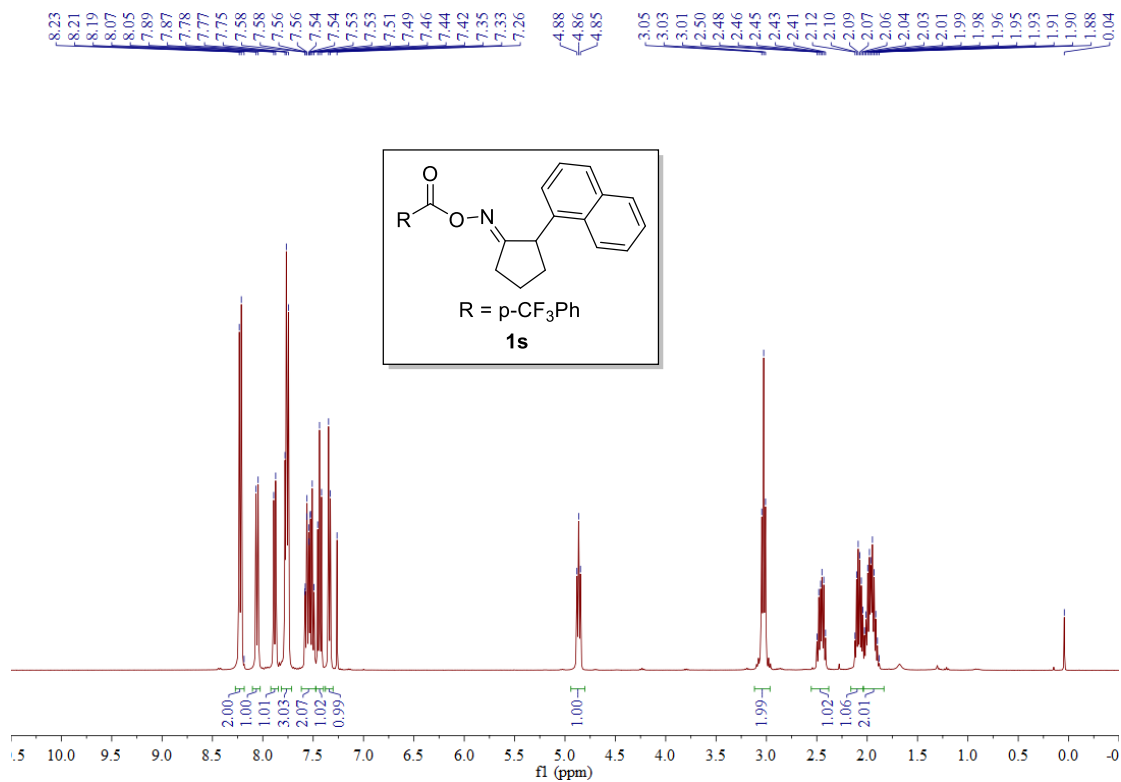
Supplementary Figure 18. ¹³C NMR of **1o**



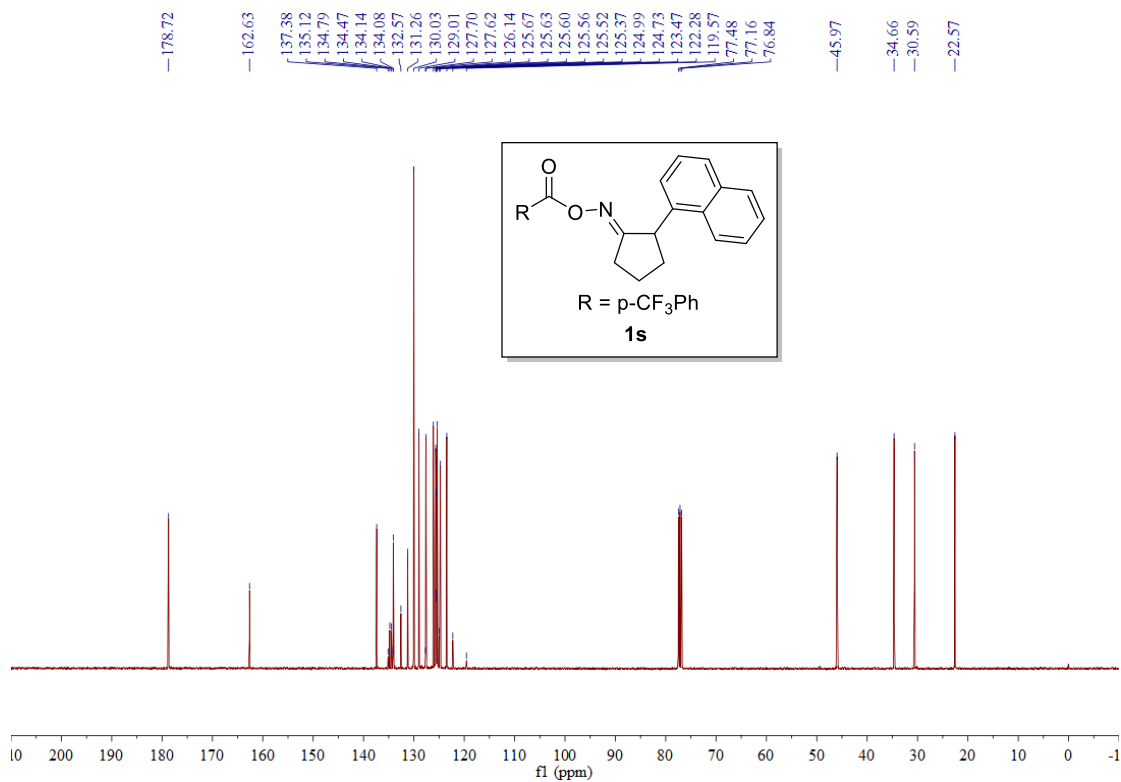
Supplementary Figure 19. ¹H NMR of **1p**



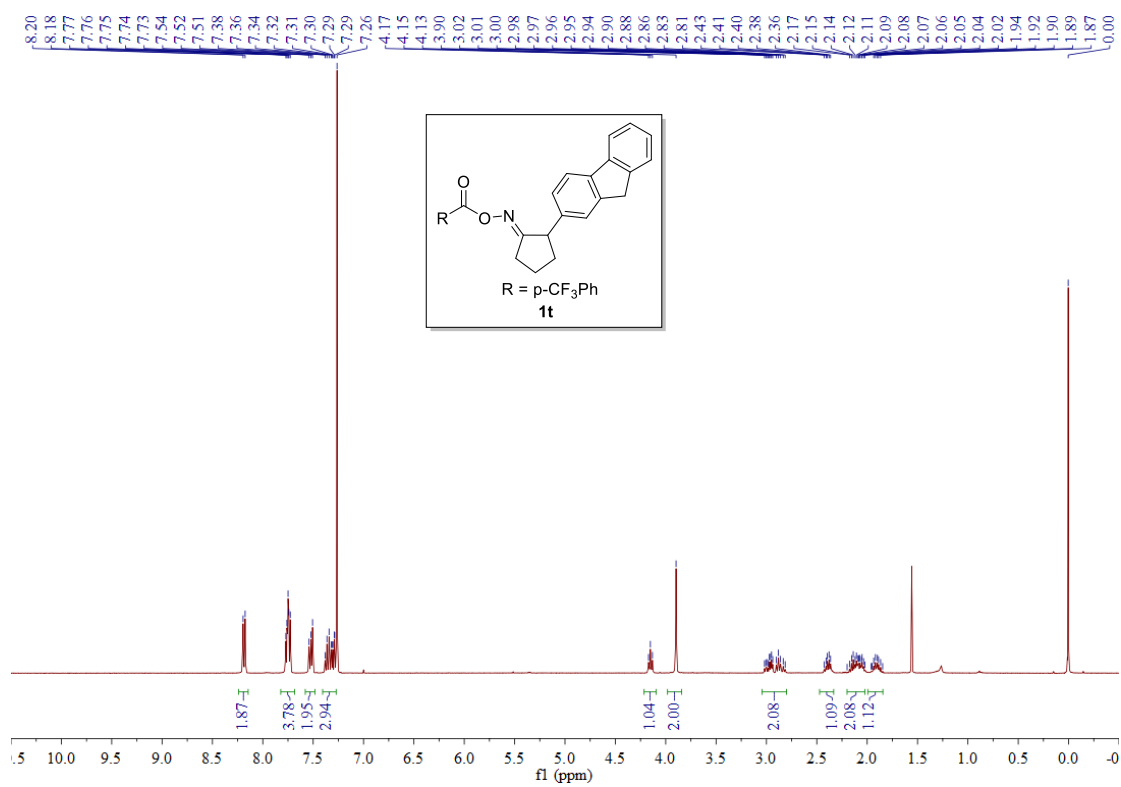
Supplementary Figure 20. ¹³C NMR of **1p**



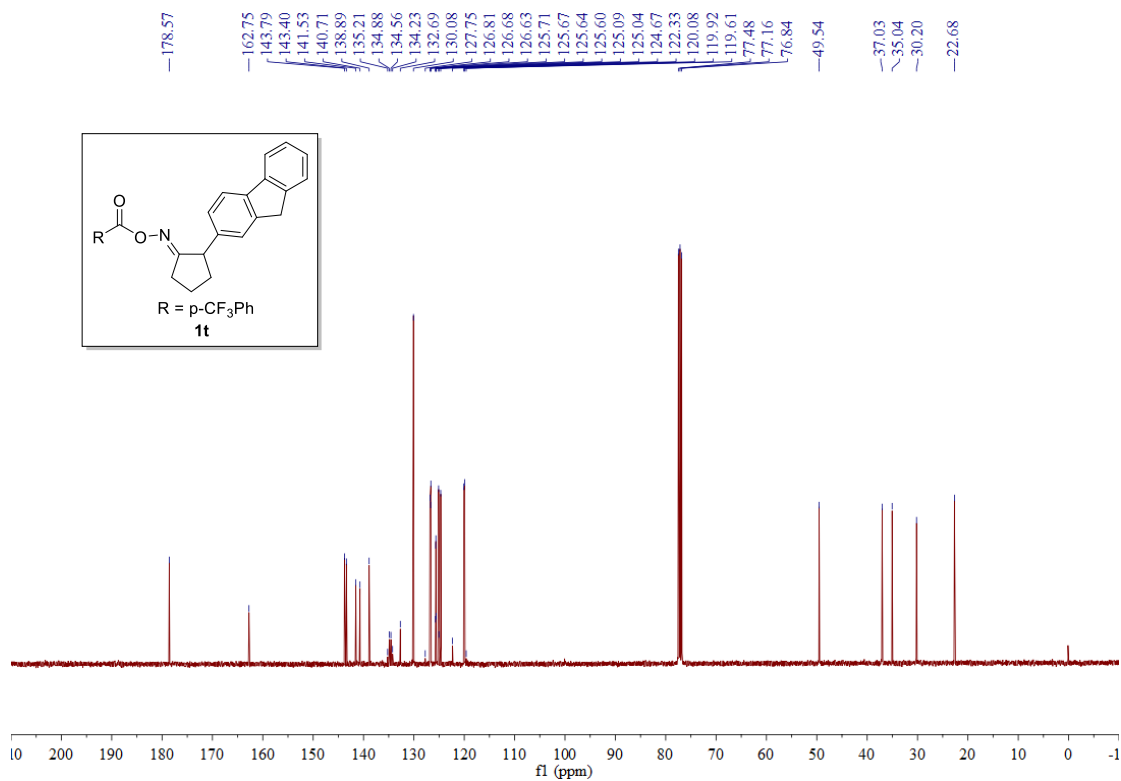
Supplementary Figure 21. ¹H NMR of **1s**



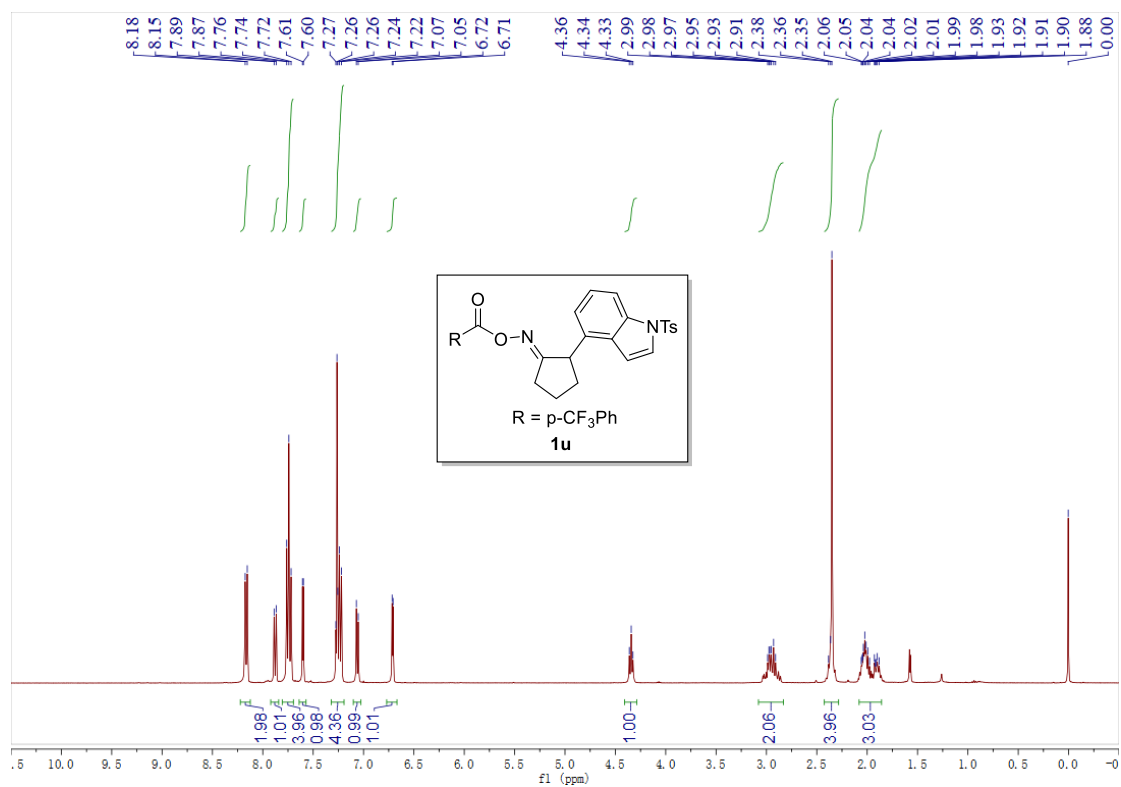
Supplementary Figure 22. ^{13}C NMR of **1s**



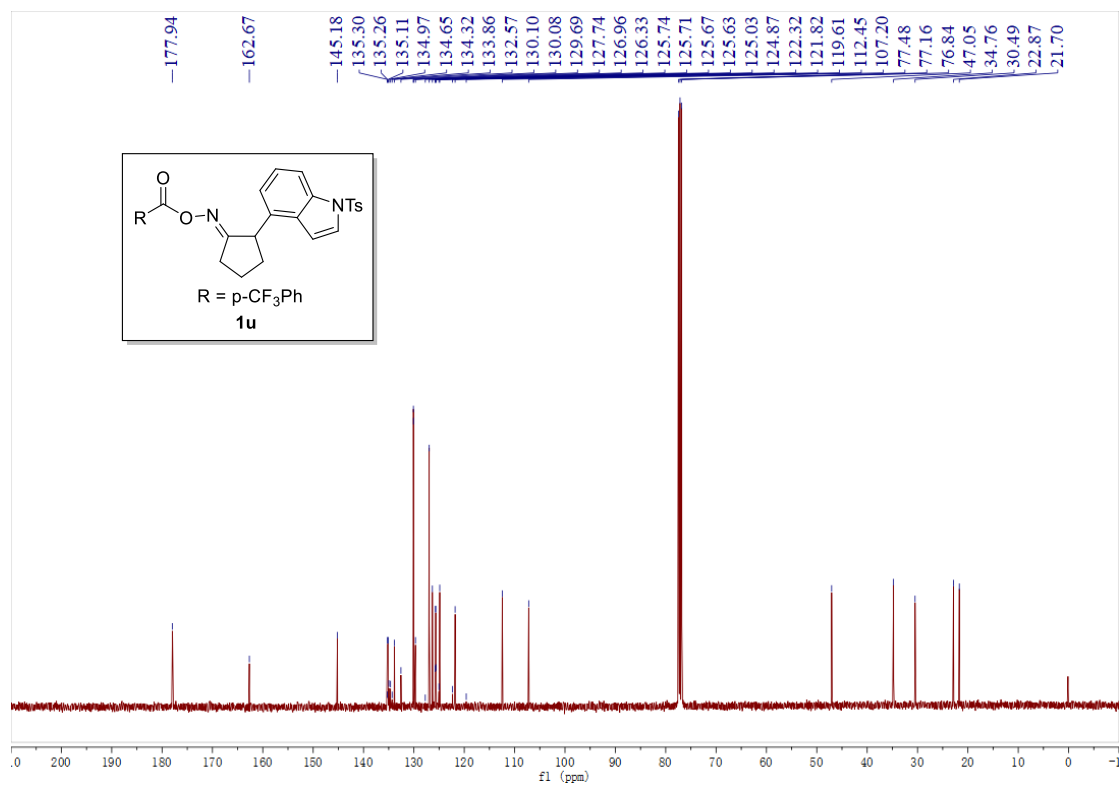
Supplementary Figure 23. ^1H NMR of **1t**



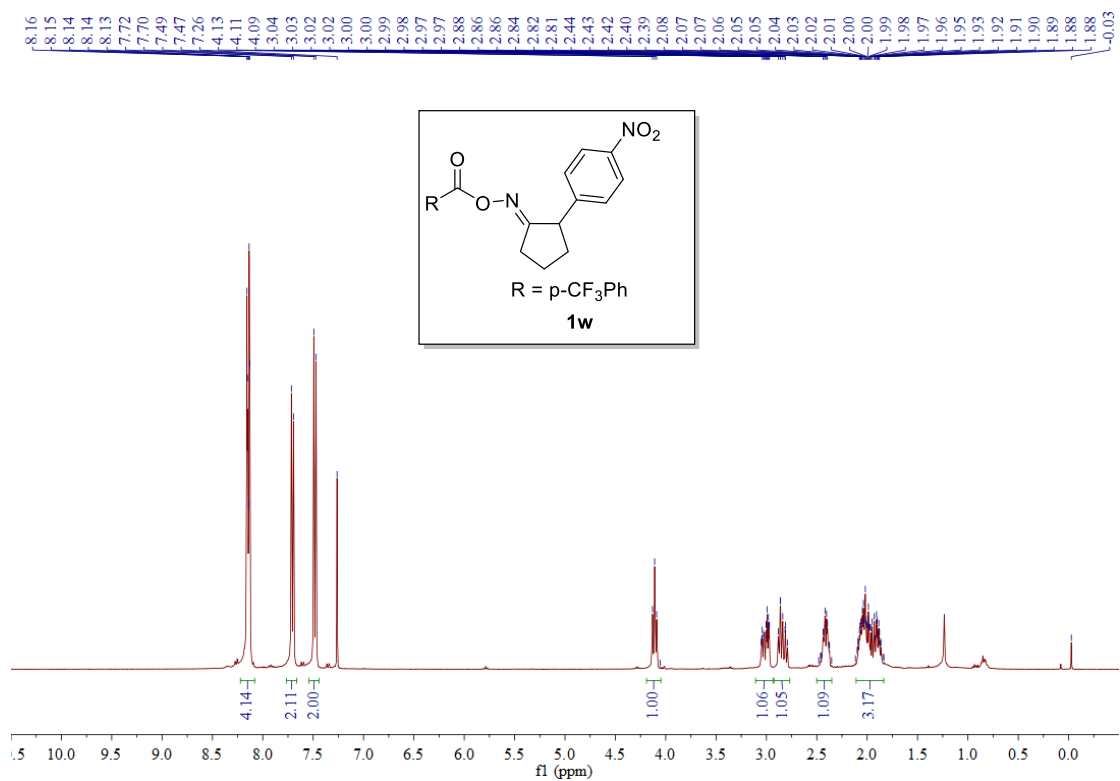
Supplementary Figure 24. ¹³C NMR of **1t**



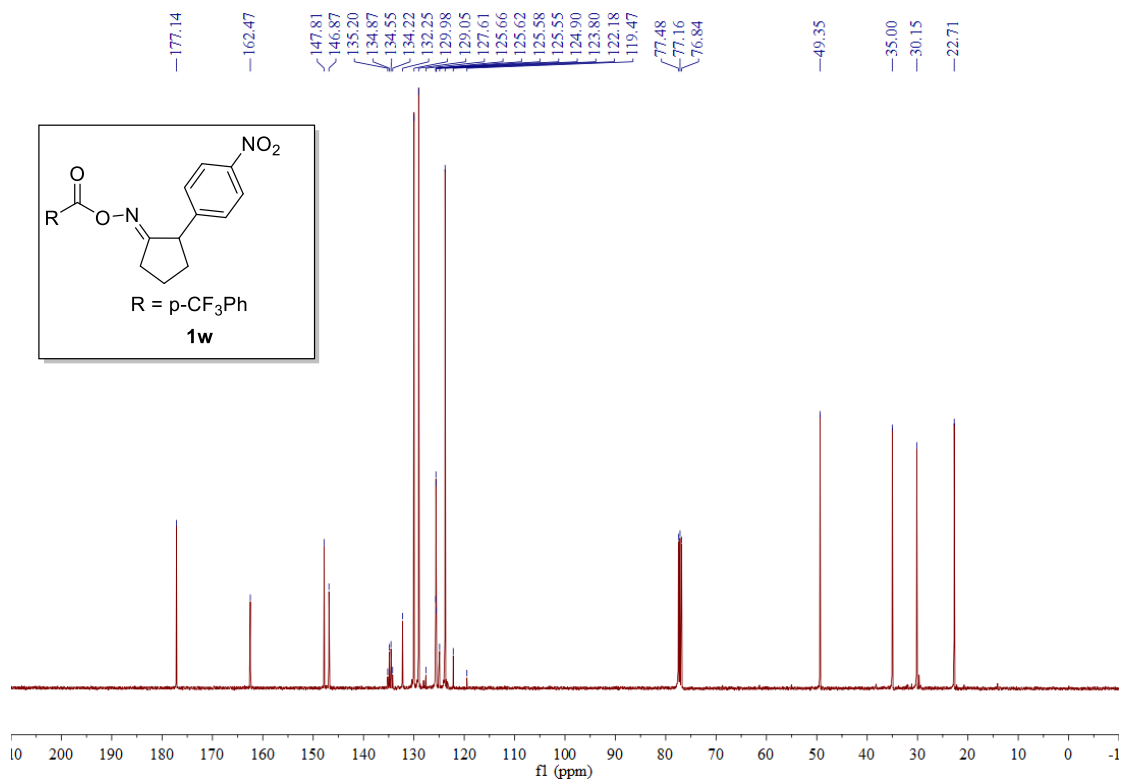
Supplementary Figure 25. ¹H NMR of **1u**



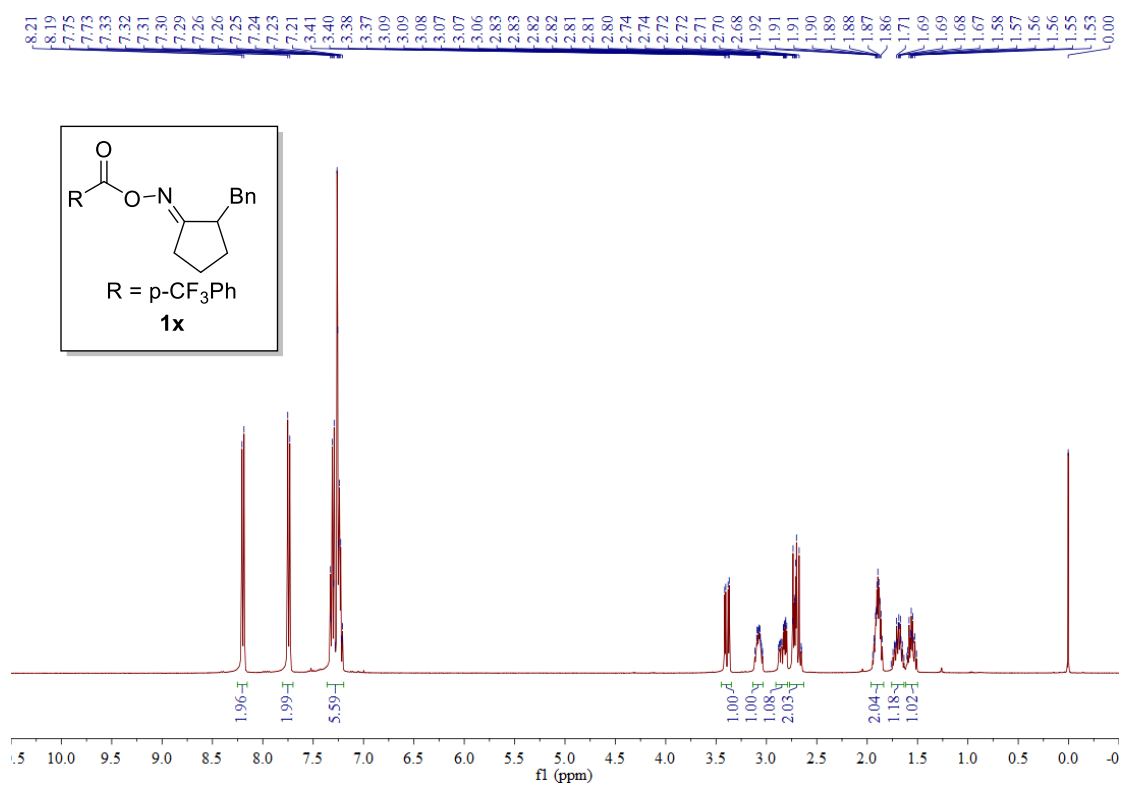
Supplementary Figure 26. ^{13}C NMR of **1u**



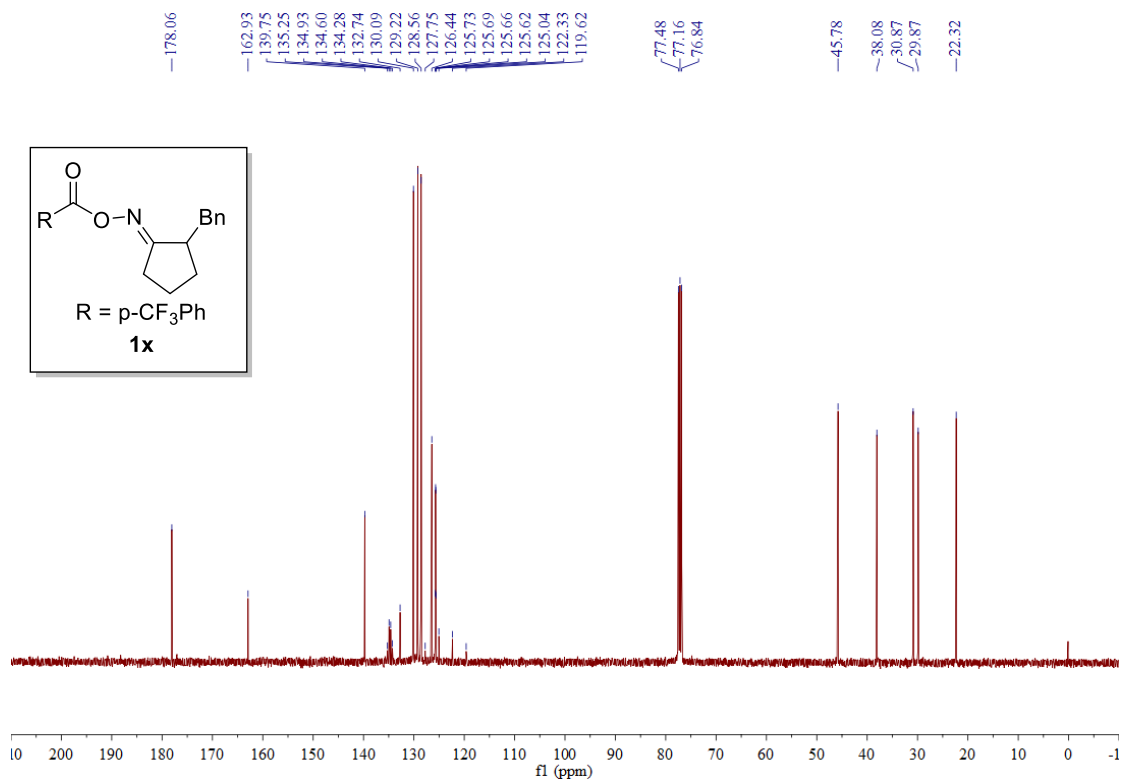
Supplementary Figure 27. ^1H NMR of **1w**



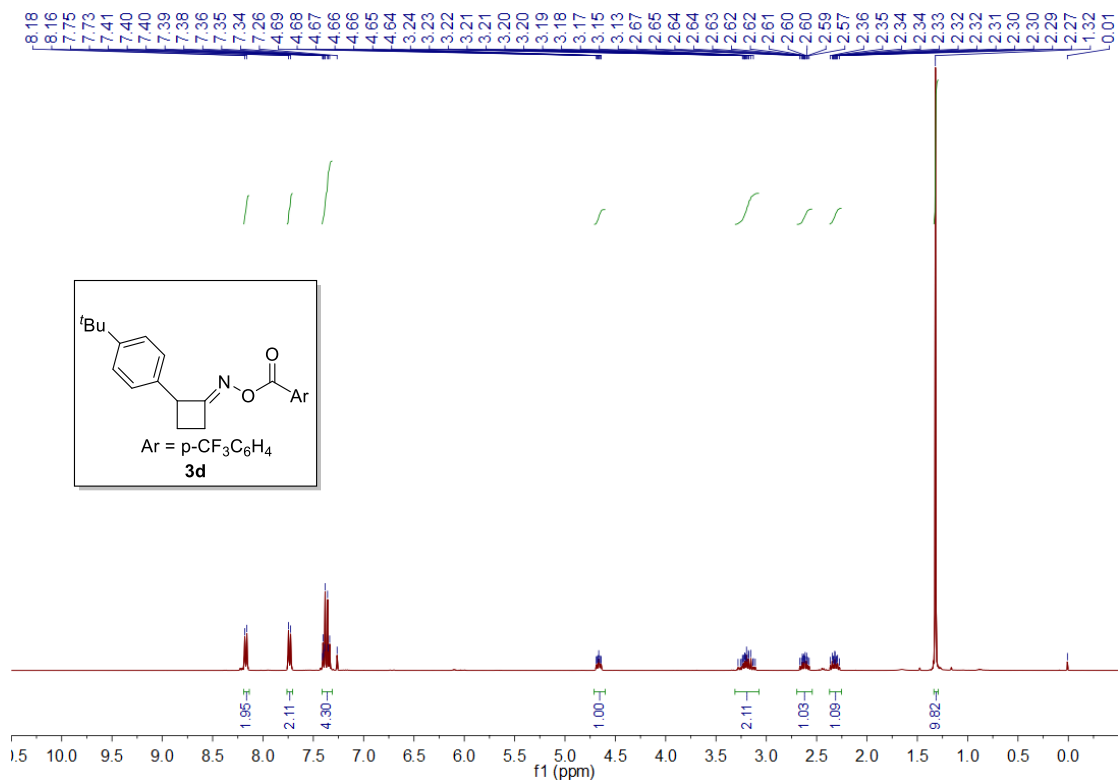
Supplementary Figure 28. ¹³C NMR of **1w**



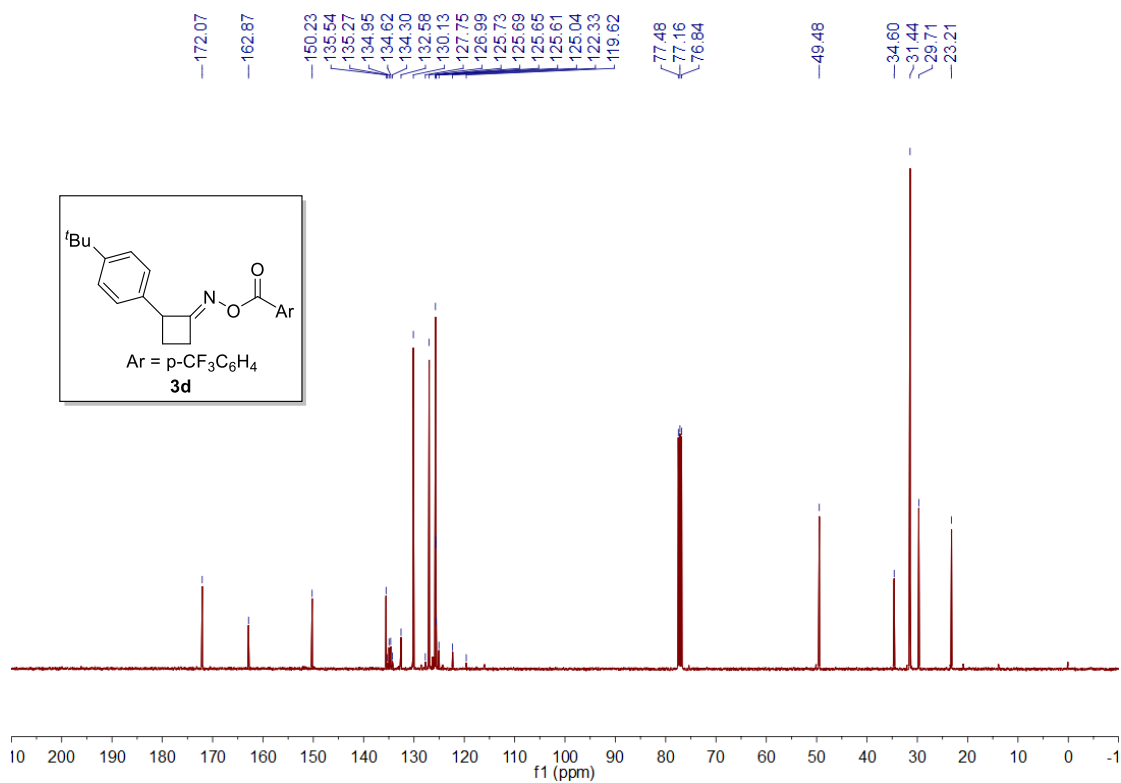
Supplementary Figure 29. ¹H NMR of **1x**



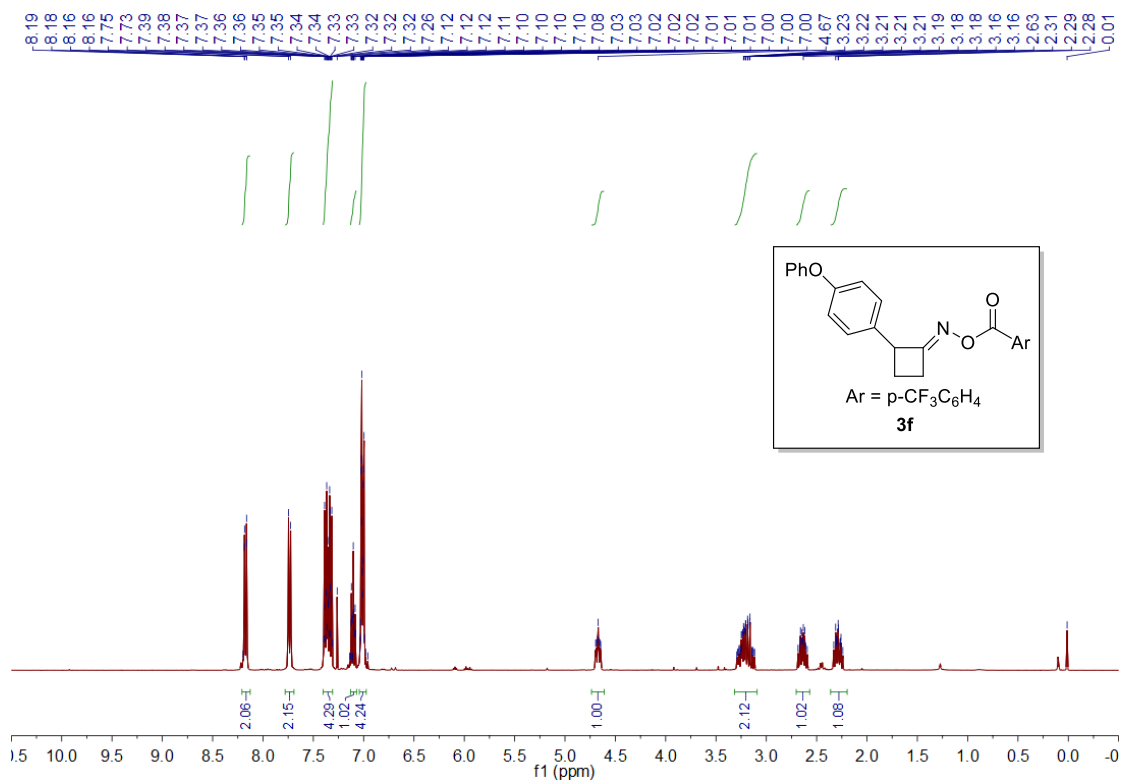
Supplementary Figure 30. ¹³C NMR of **1x**



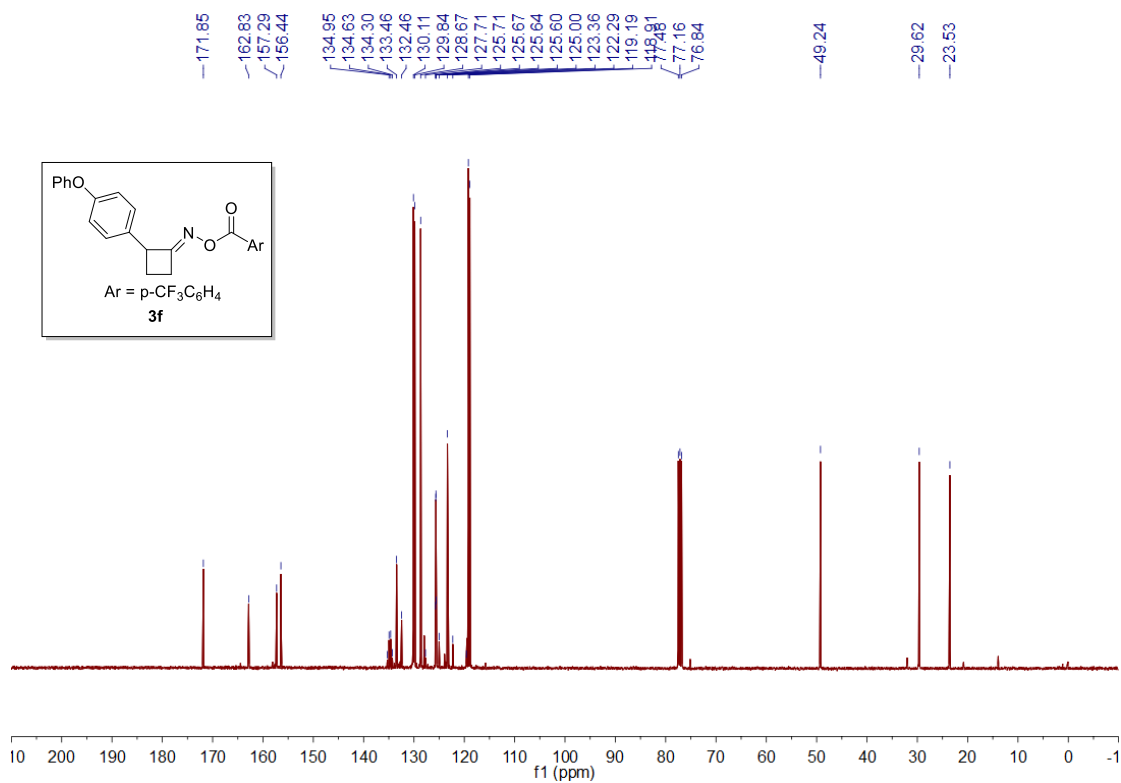
Supplementary Figure 31. ¹H NMR of **3d**



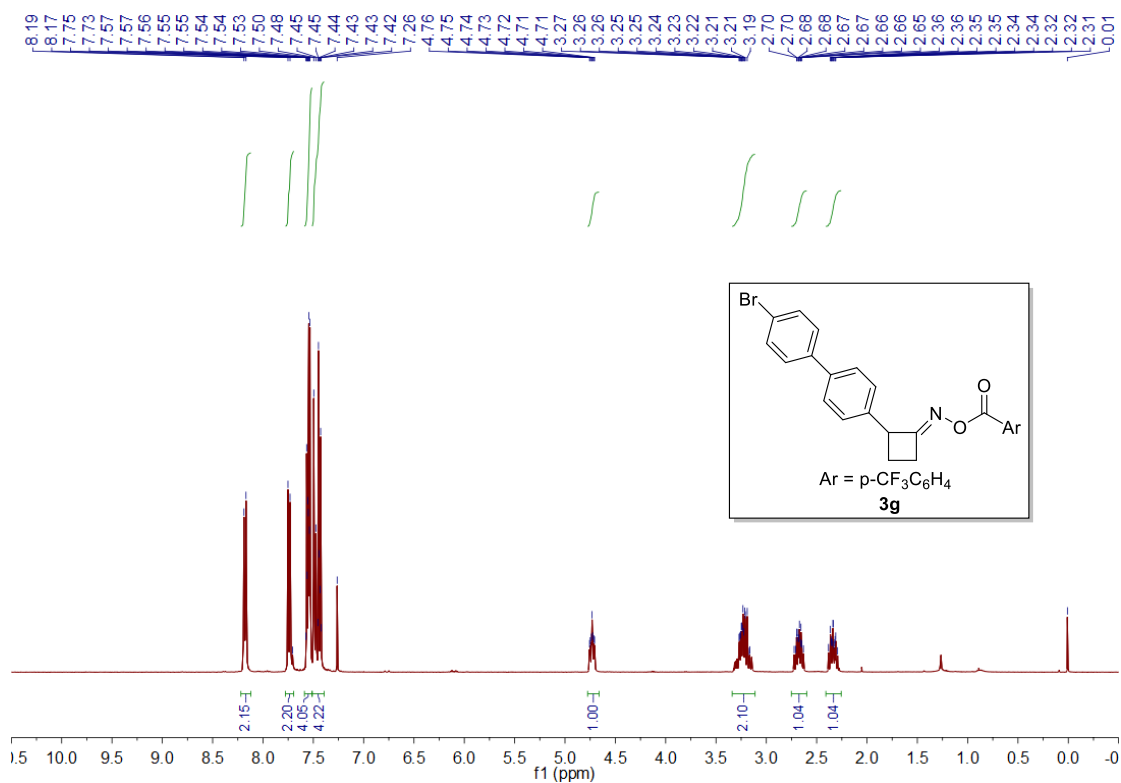
Supplementary Figure 32. ¹³C NMR of **3d**



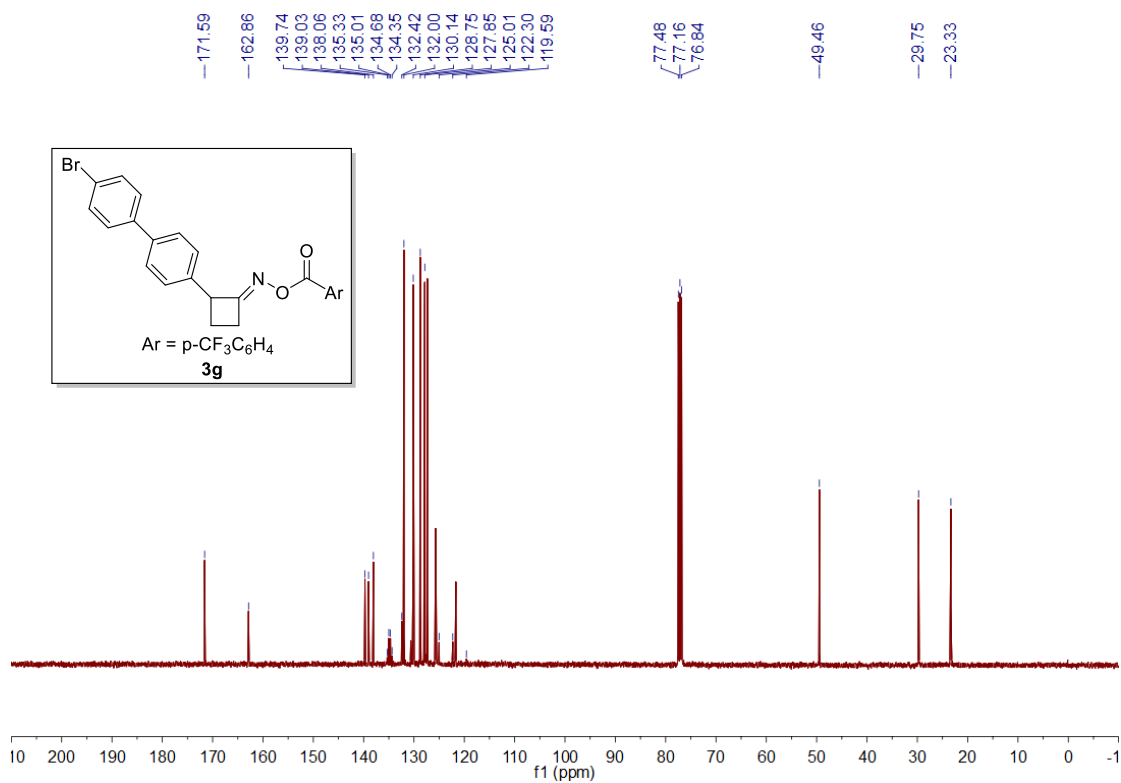
Supplementary Figure 33. ¹H NMR of **3f**



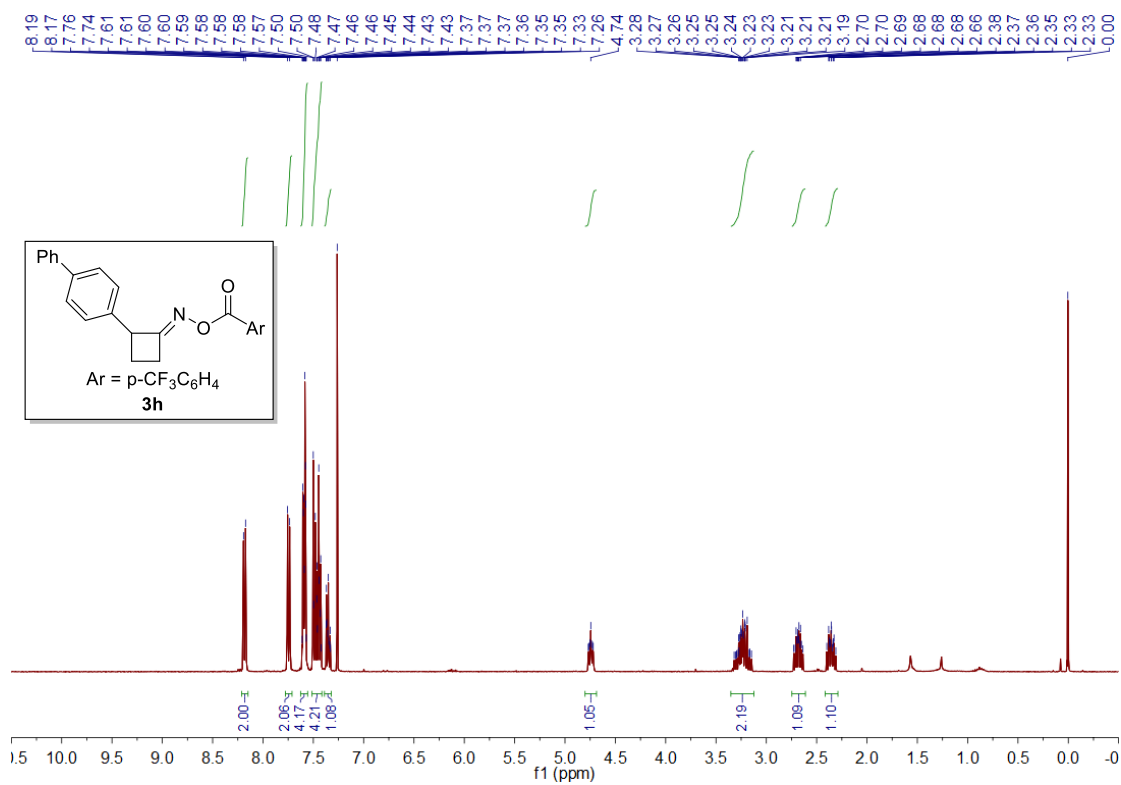
Supplementary Figure 34. ^{13}C NMR of **3f**



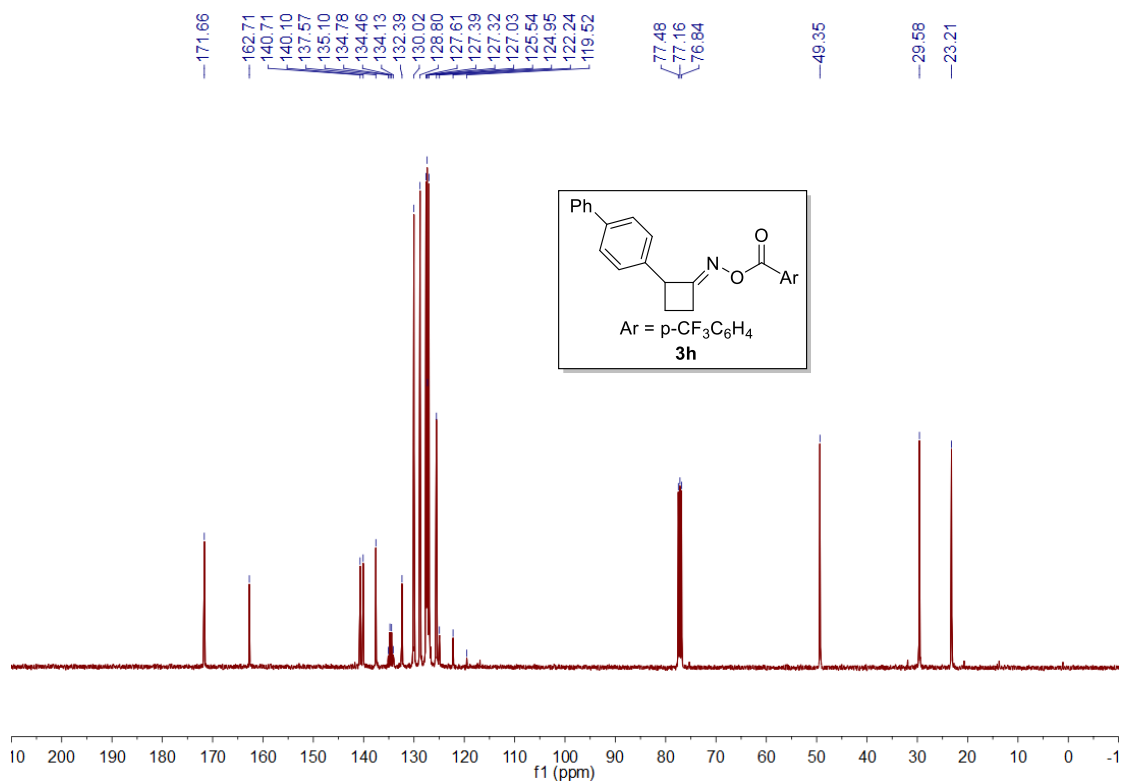
Supplementary Figure 35. ^1H NMR of **3g**



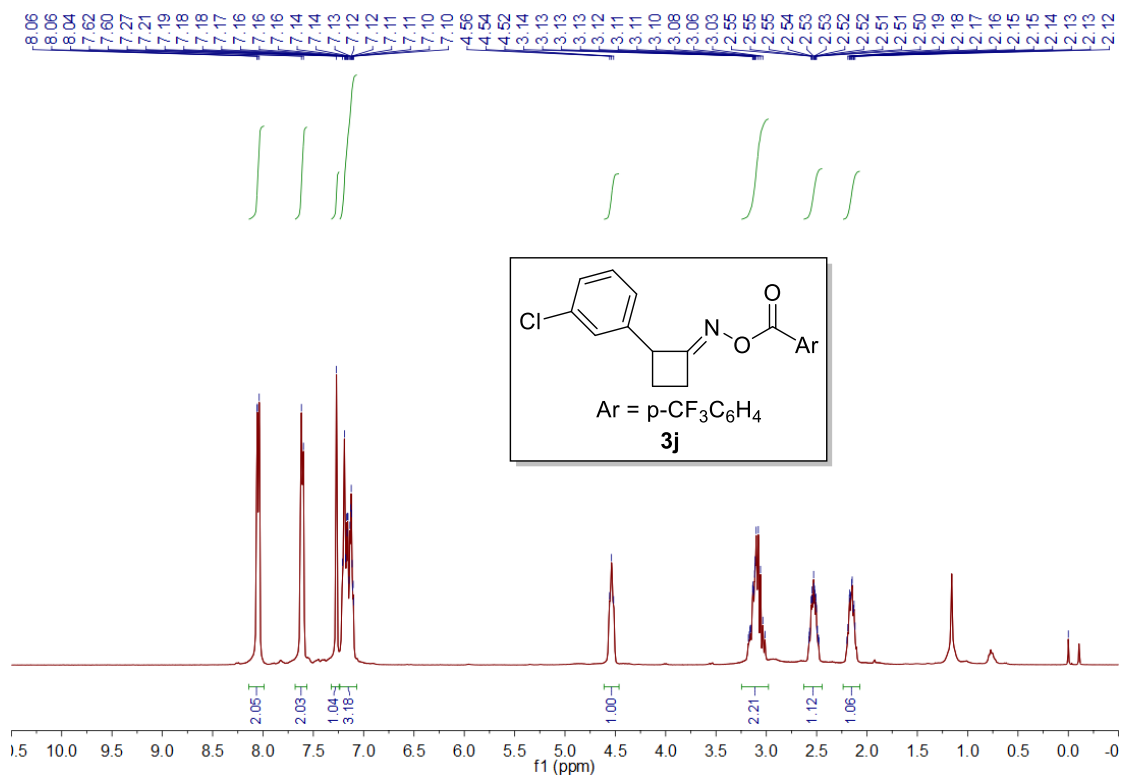
Supplementary Figure 36. ¹³C NMR of **3g**



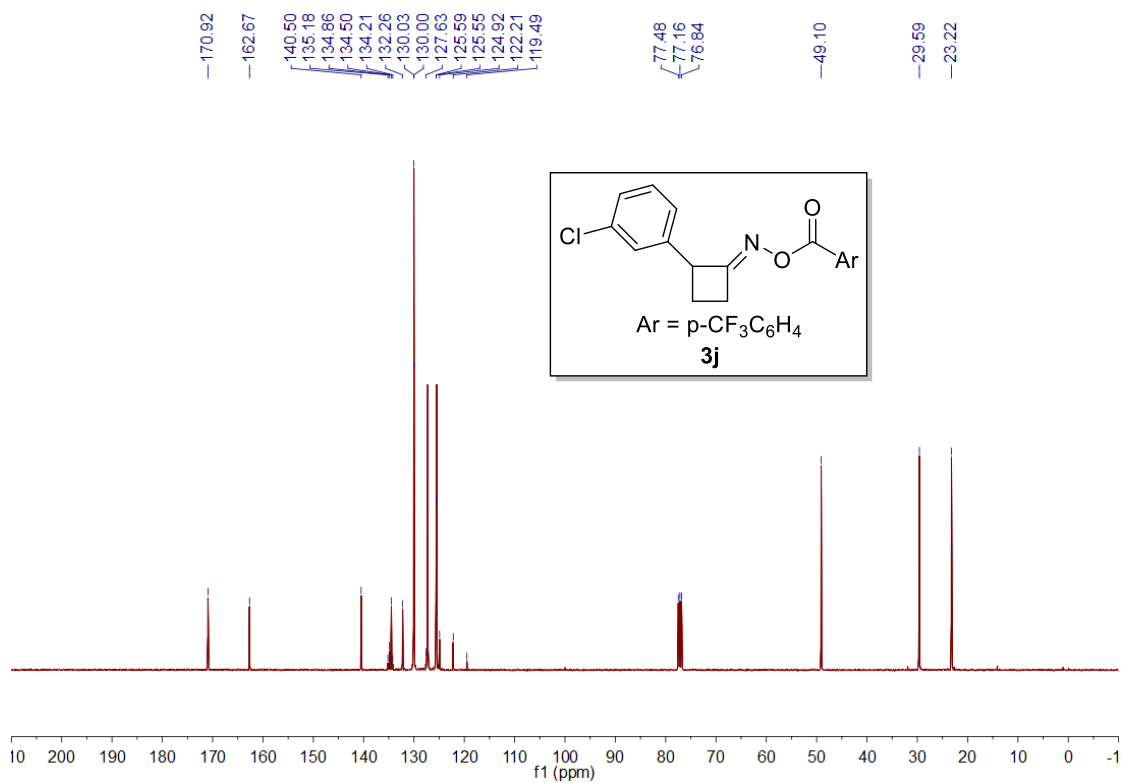
Supplementary Figure 37. ¹H NMR of **3h**



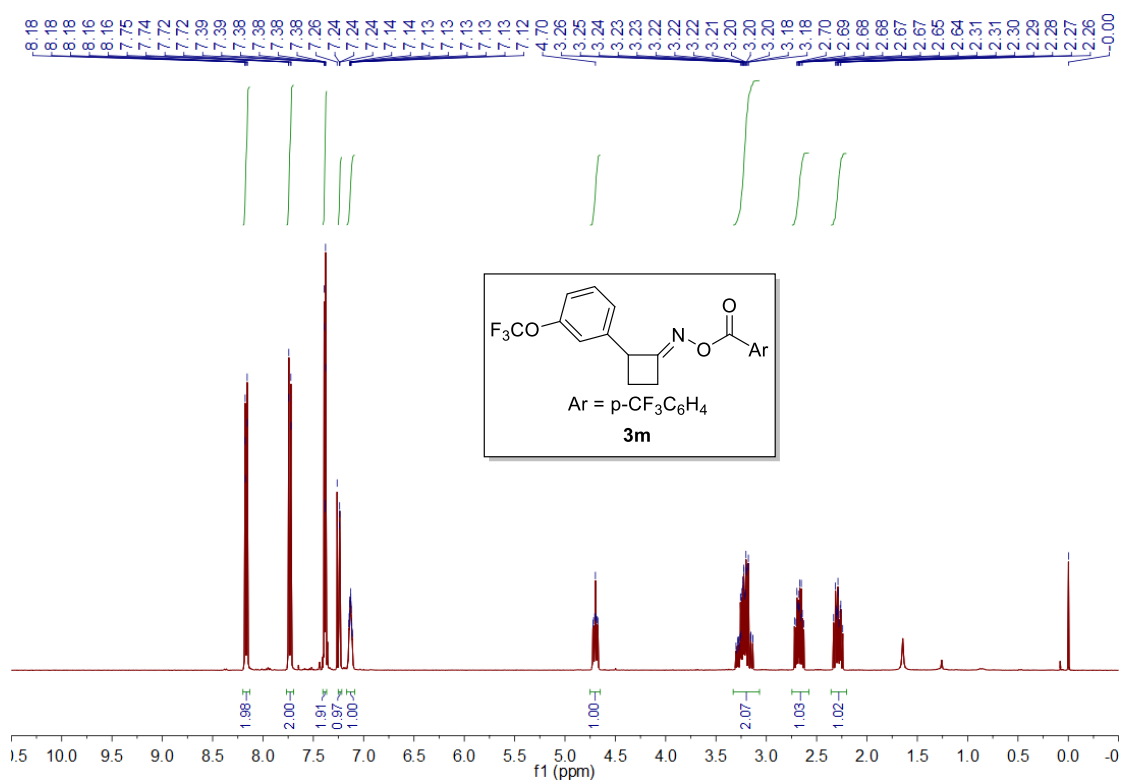
Supplementary Figure 38. ¹³C NMR of **3h**



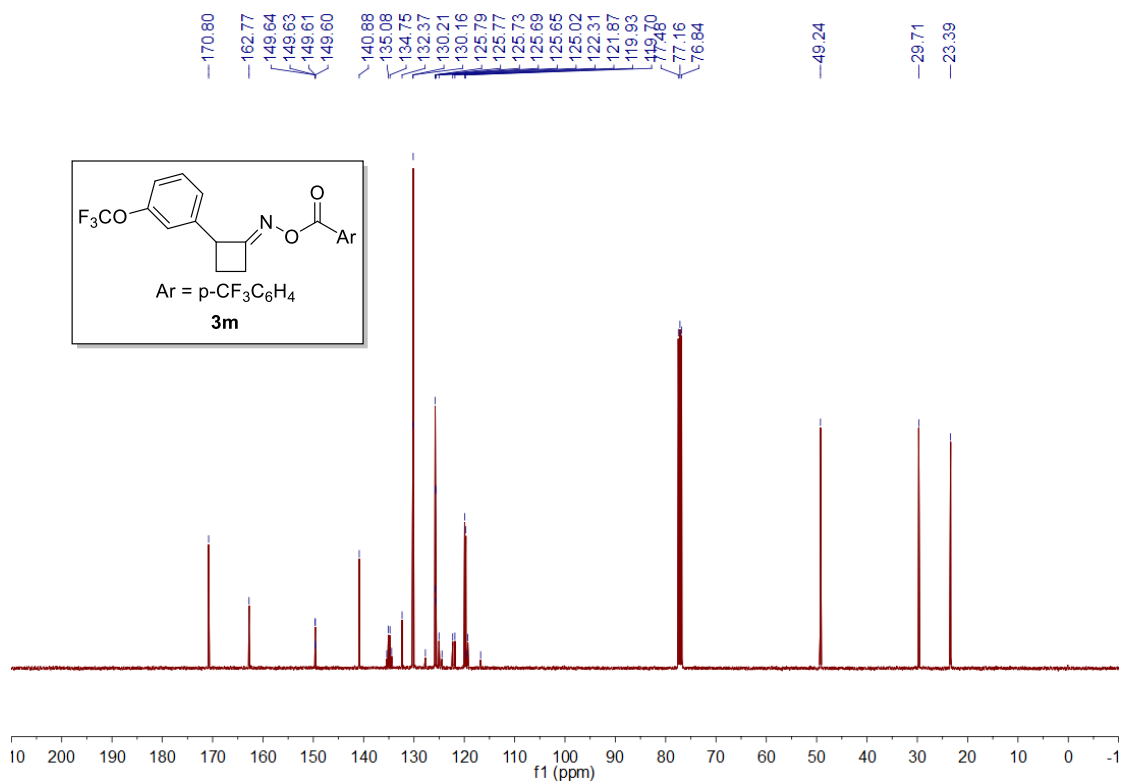
Supplementary Figure 39. ¹H NMR of **3j**



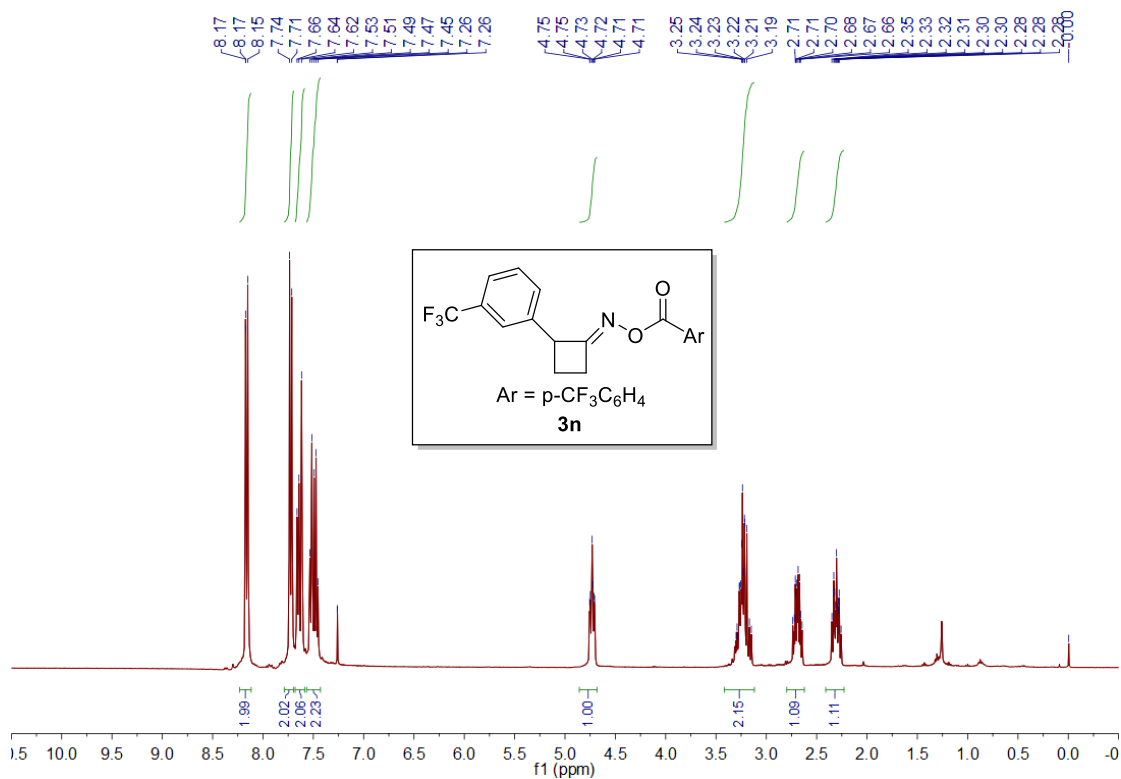
Supplementary Figure 40. ¹³C NMR of **3j**



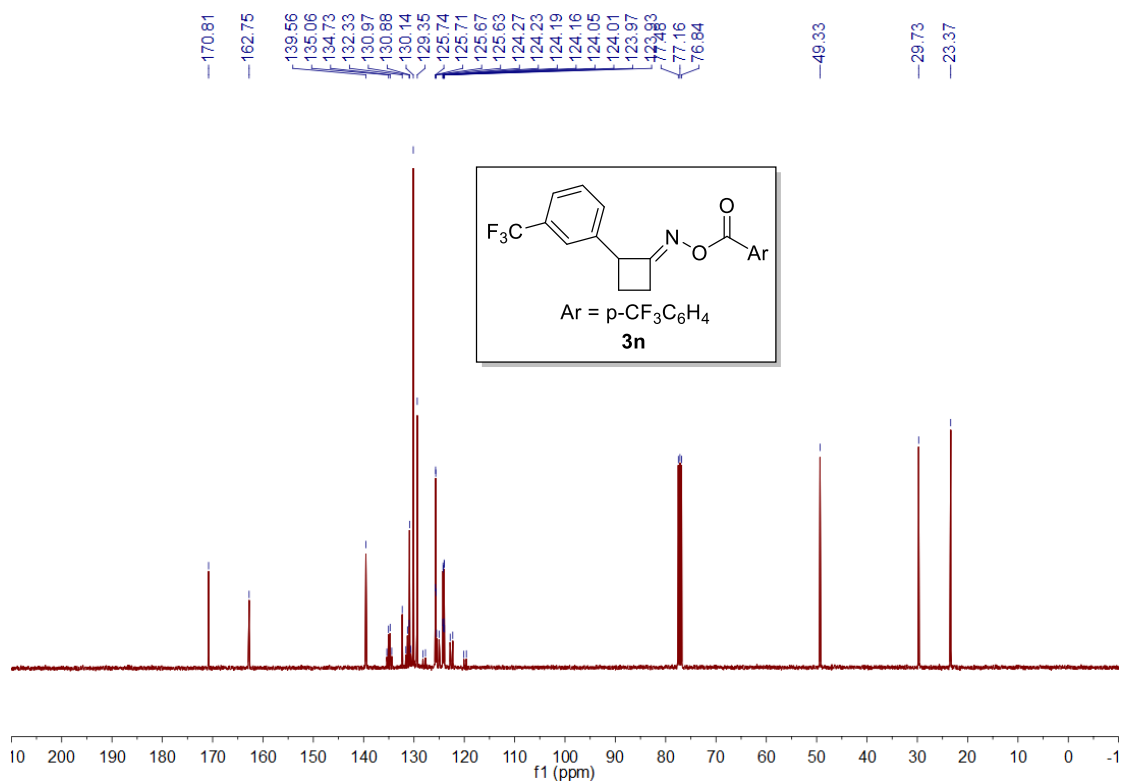
Supplementary Figure 41. ¹H NMR of **3m**



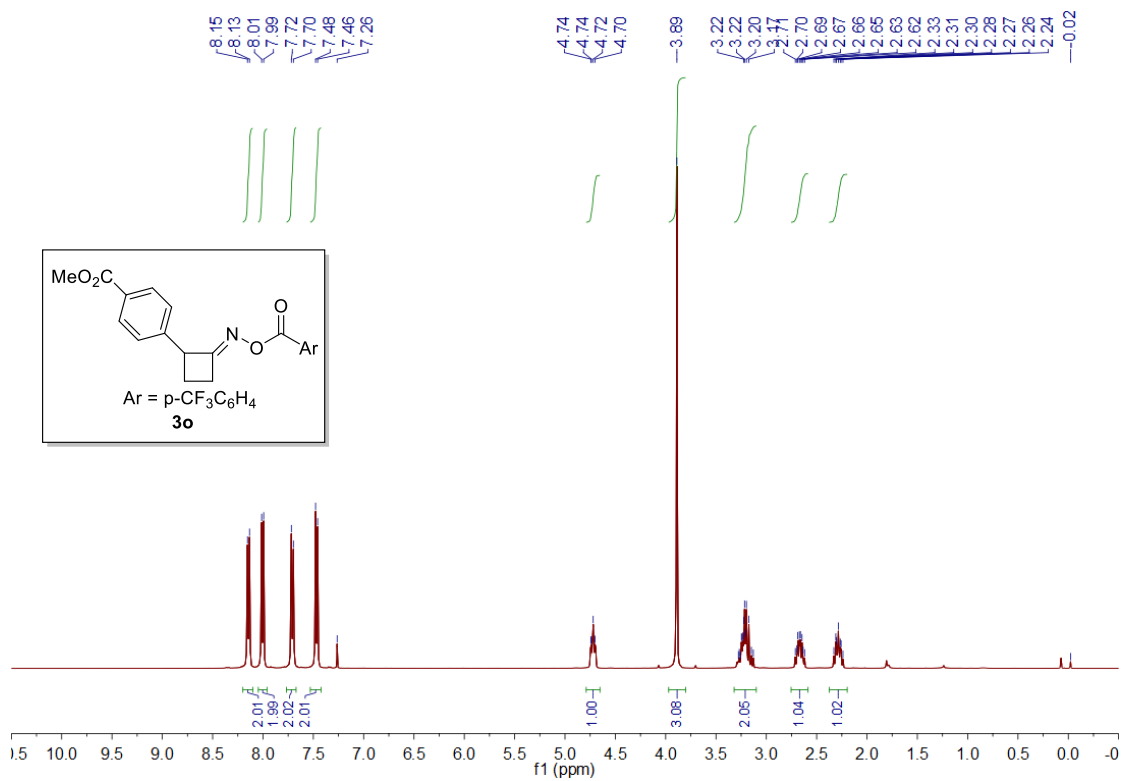
Supplementary Figure 42. ¹³C NMR of **3m**



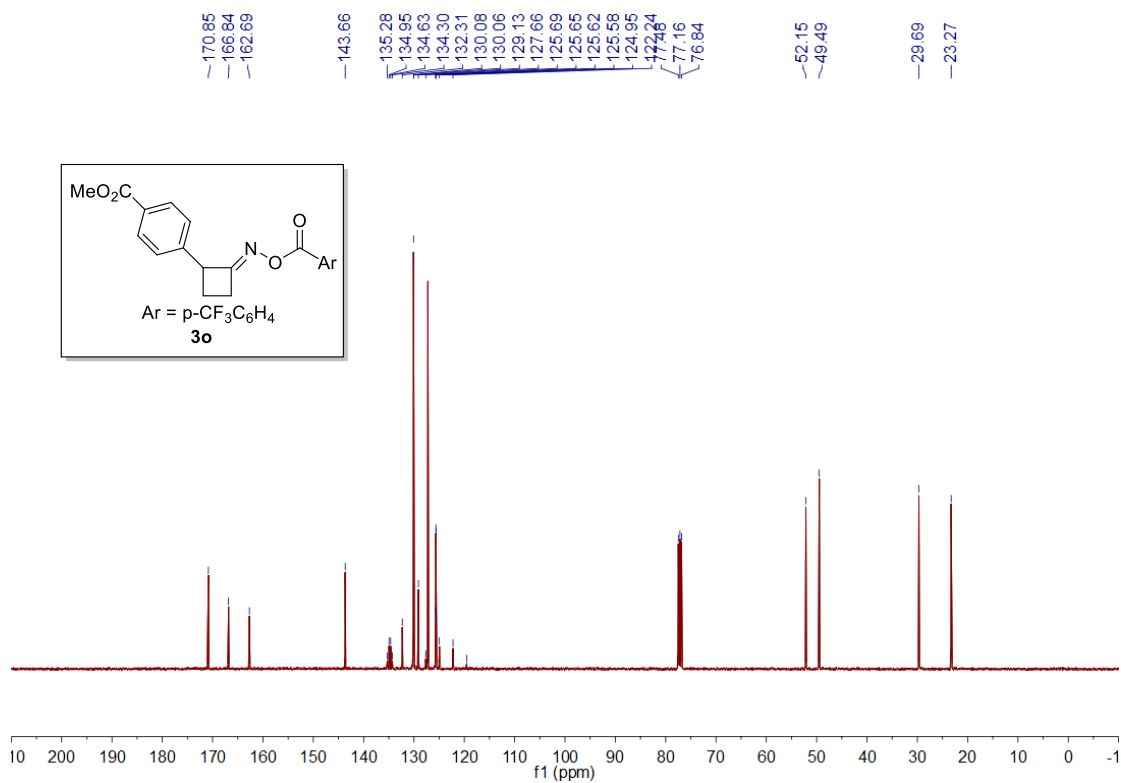
Supplementary Figure 43. ¹H NMR of **3n**



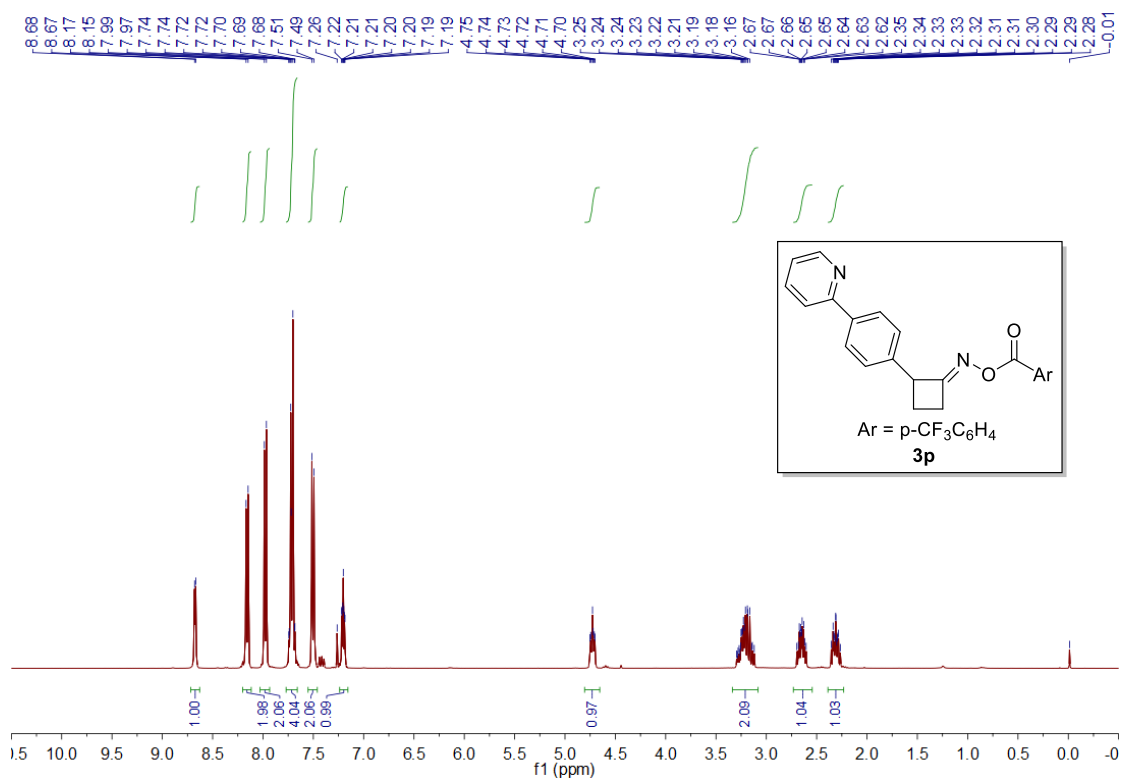
Supplementary Figure 44. ^{13}C NMR of **3n**



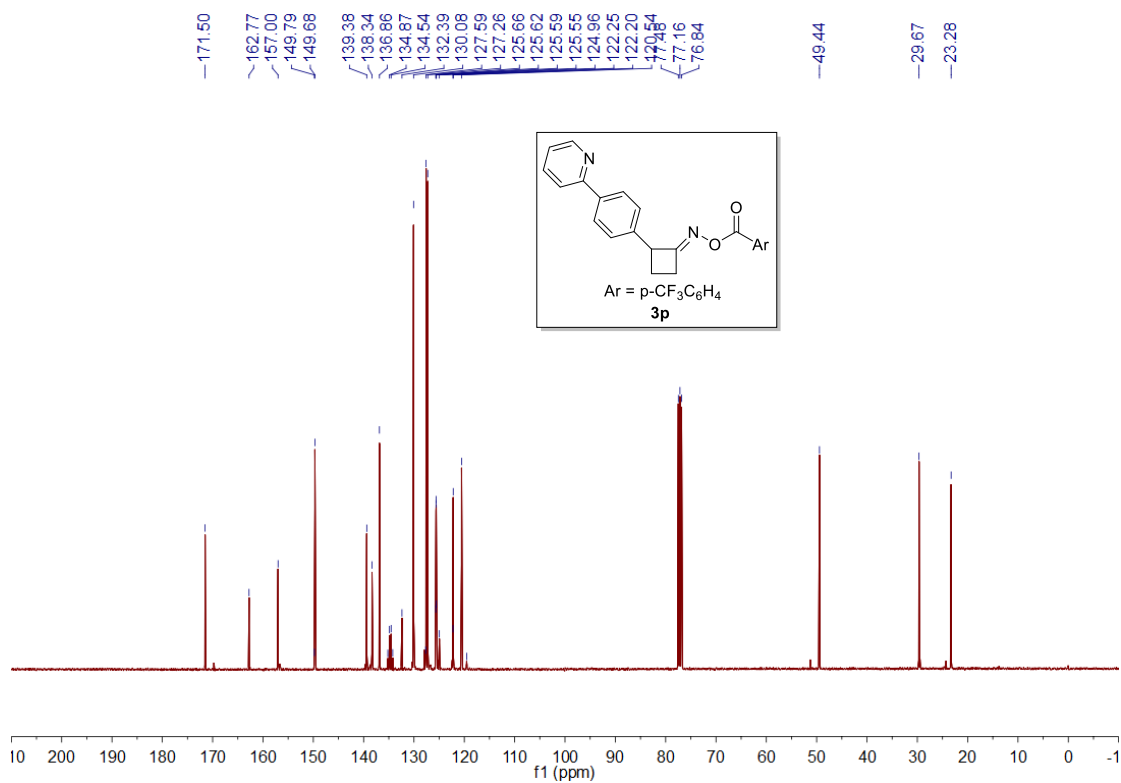
Supplementary Figure 45. ^1H NMR of **3o**



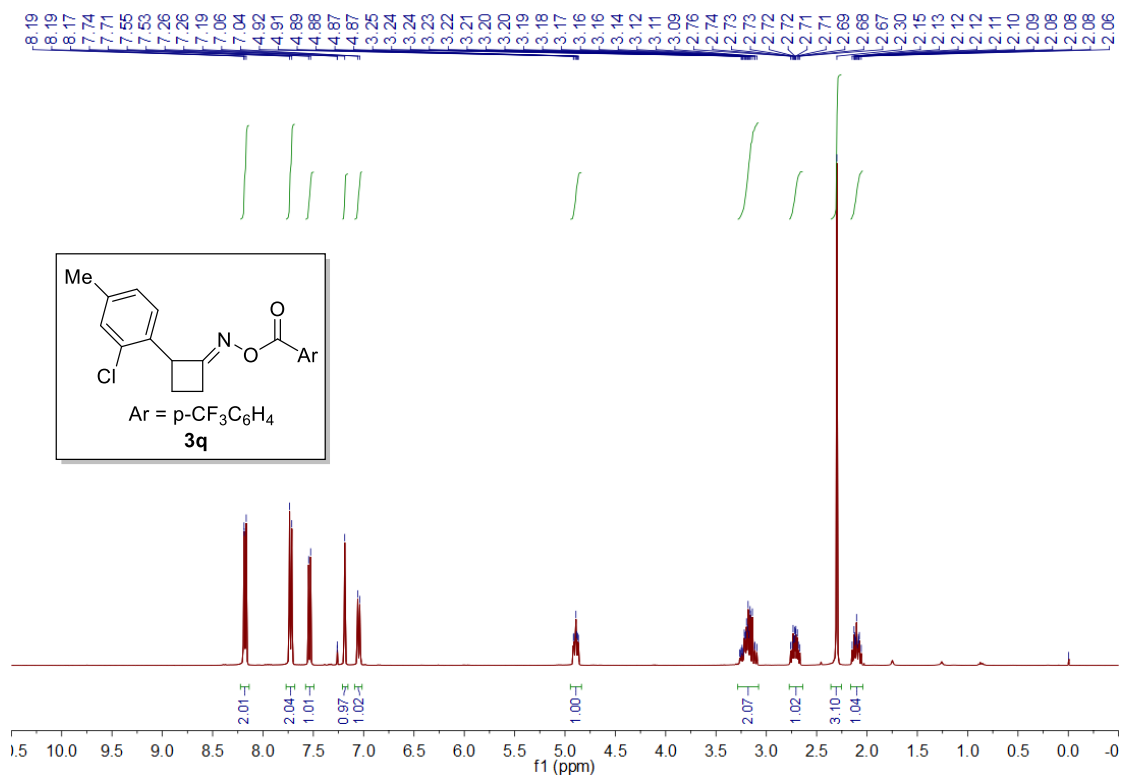
Supplementary Figure 46. ¹³C NMR of **3o**



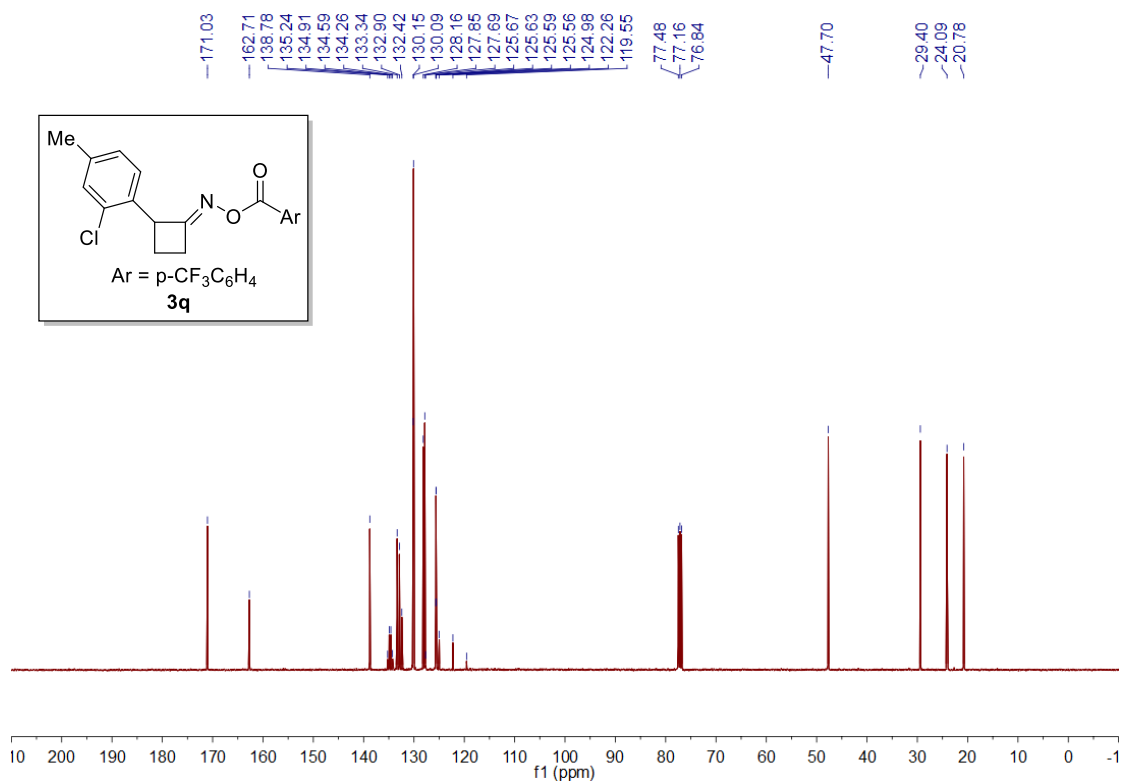
Supplementary Figure 47. ¹H NMR of **3p**



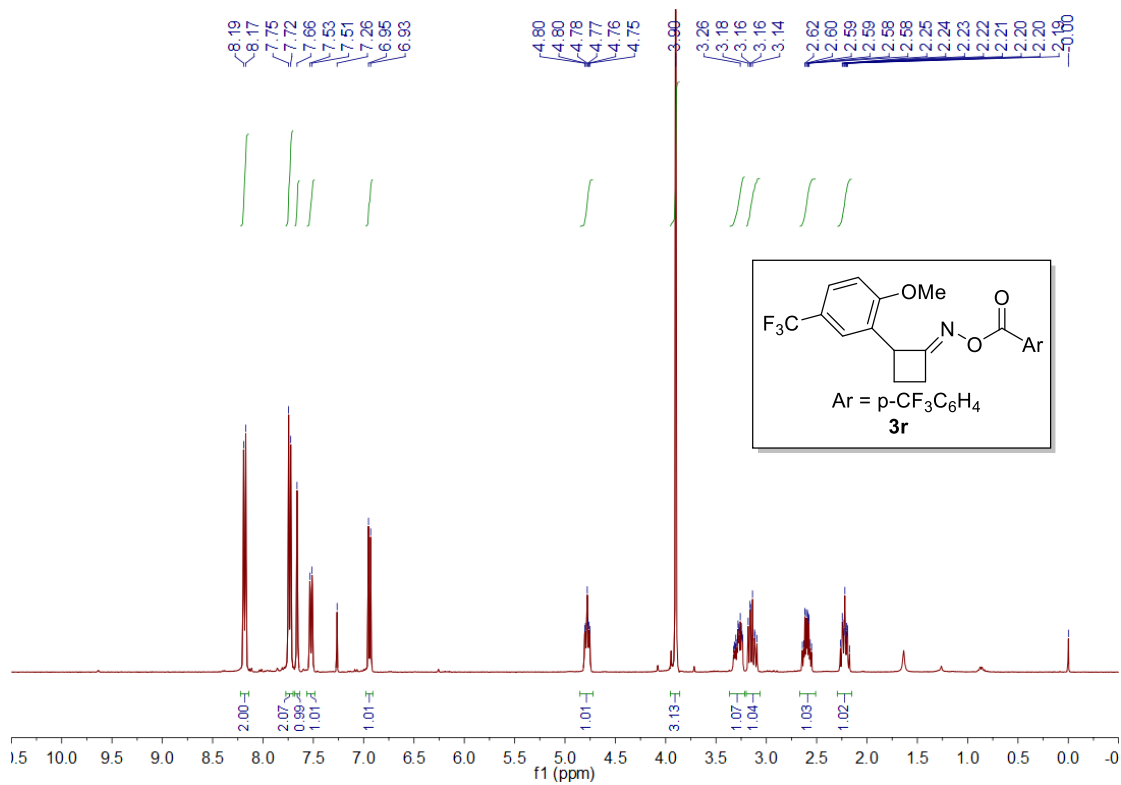
Supplementary Figure 48. ¹³C NMR of **3p**



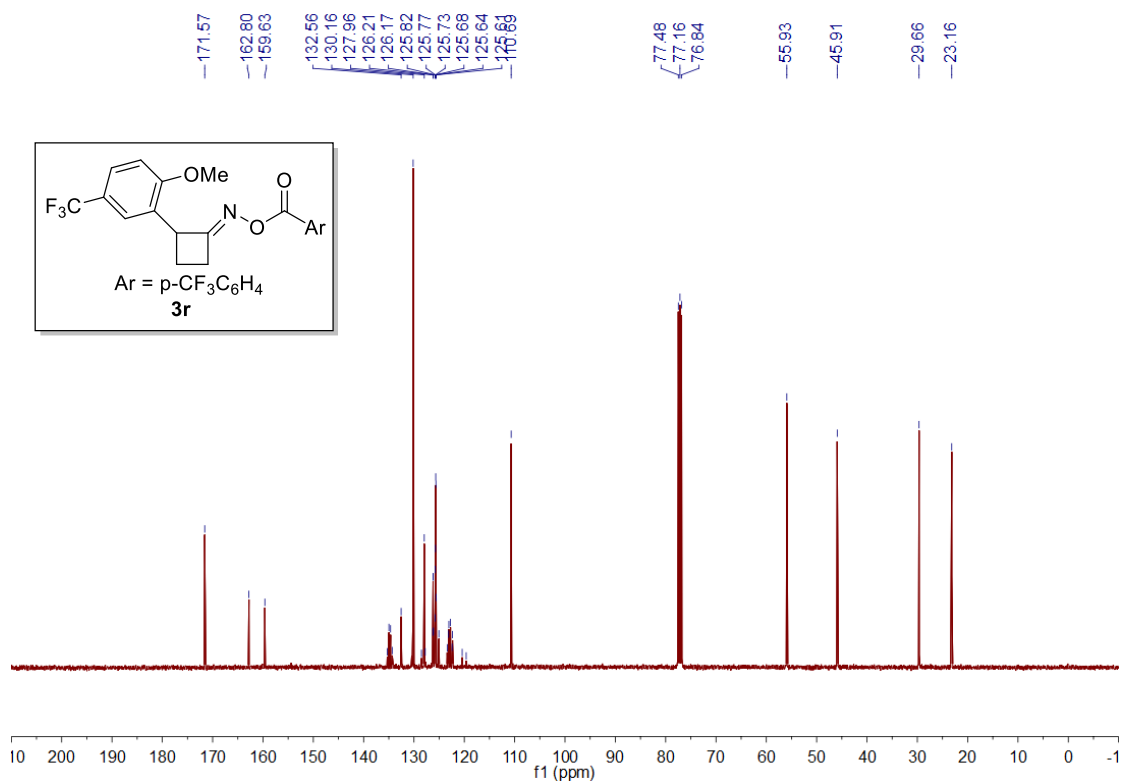
Supplementary Figure 49. ¹H NMR of **3q**



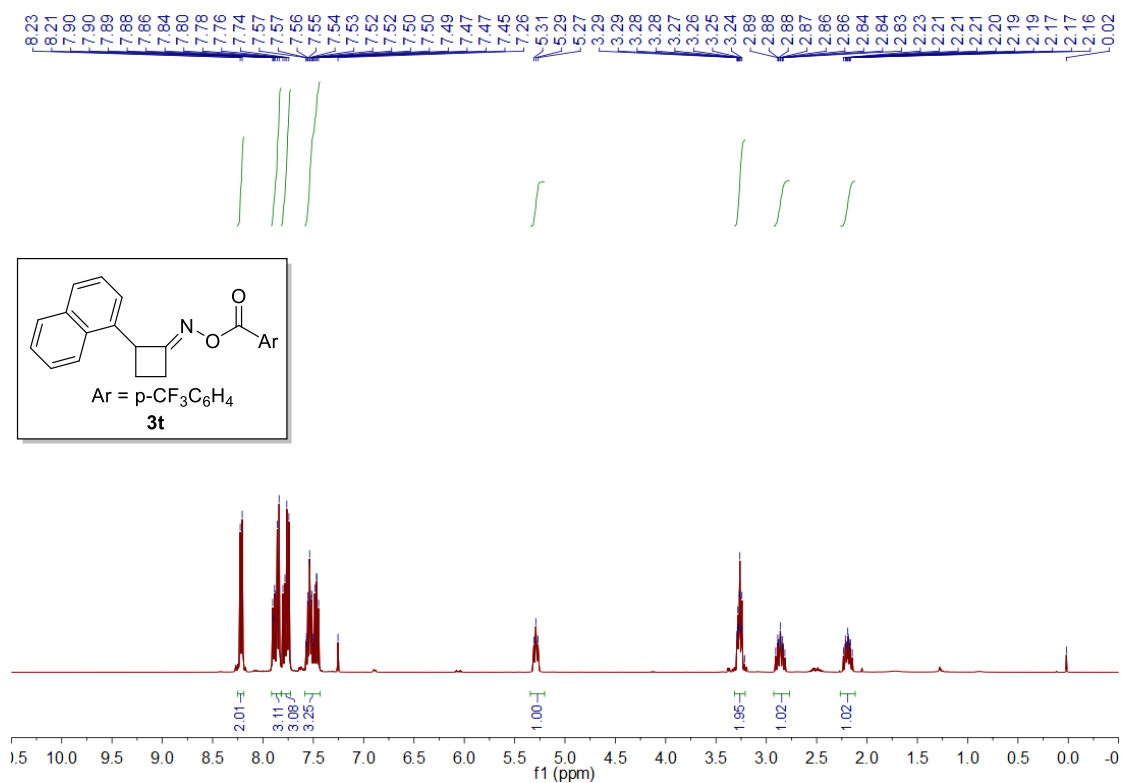
Supplementary Figure 50. ^{13}C NMR of **3q**



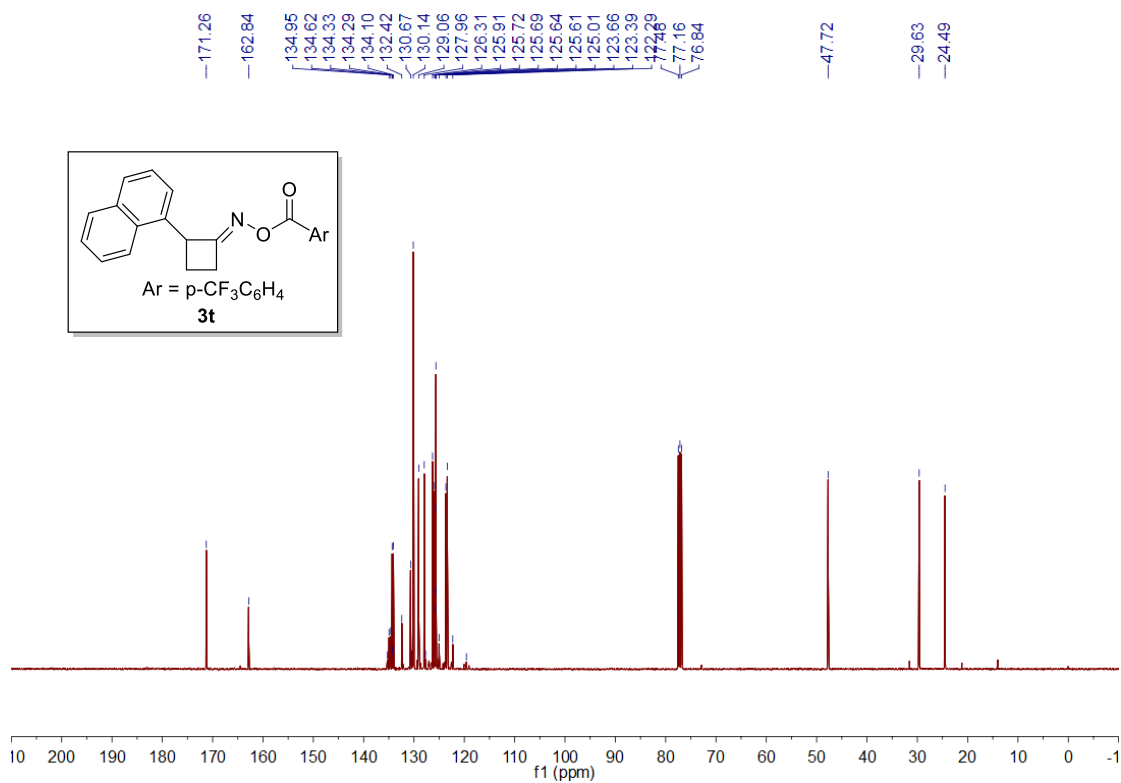
Supplementary Figure 51. ^1H NMR of **3r**



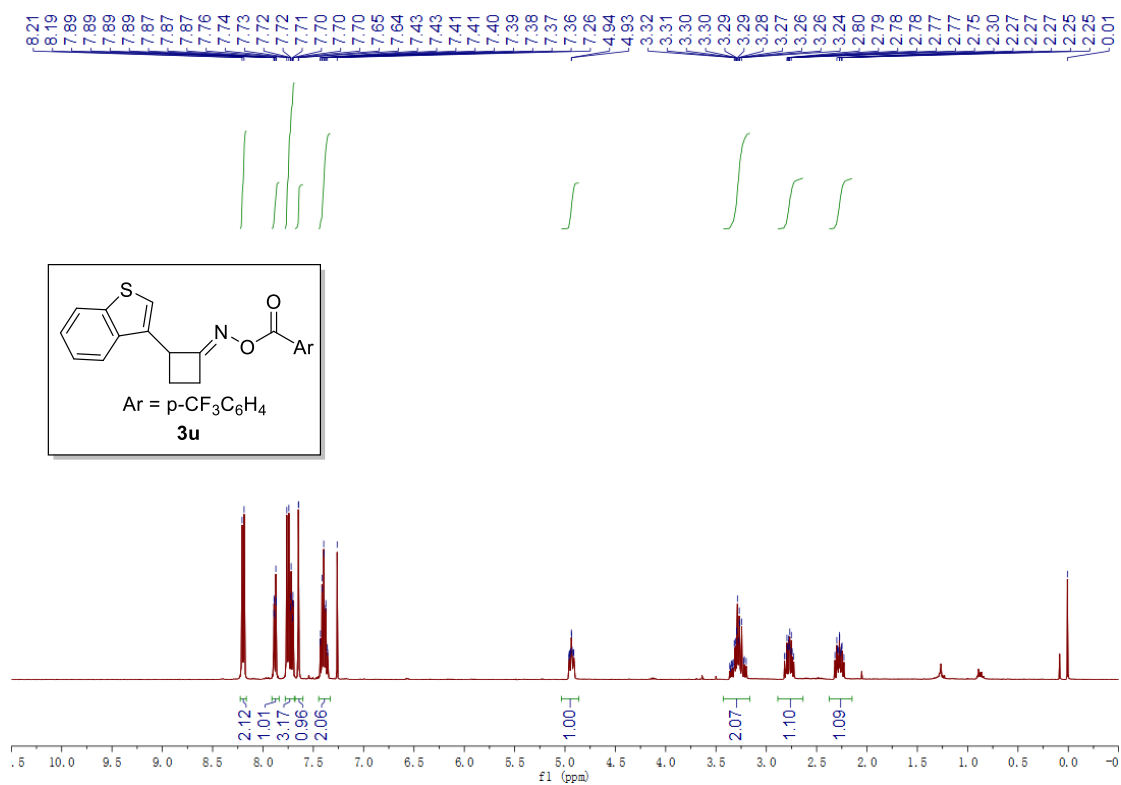
Supplementary Figure 52. ^{13}C NMR of **3r**



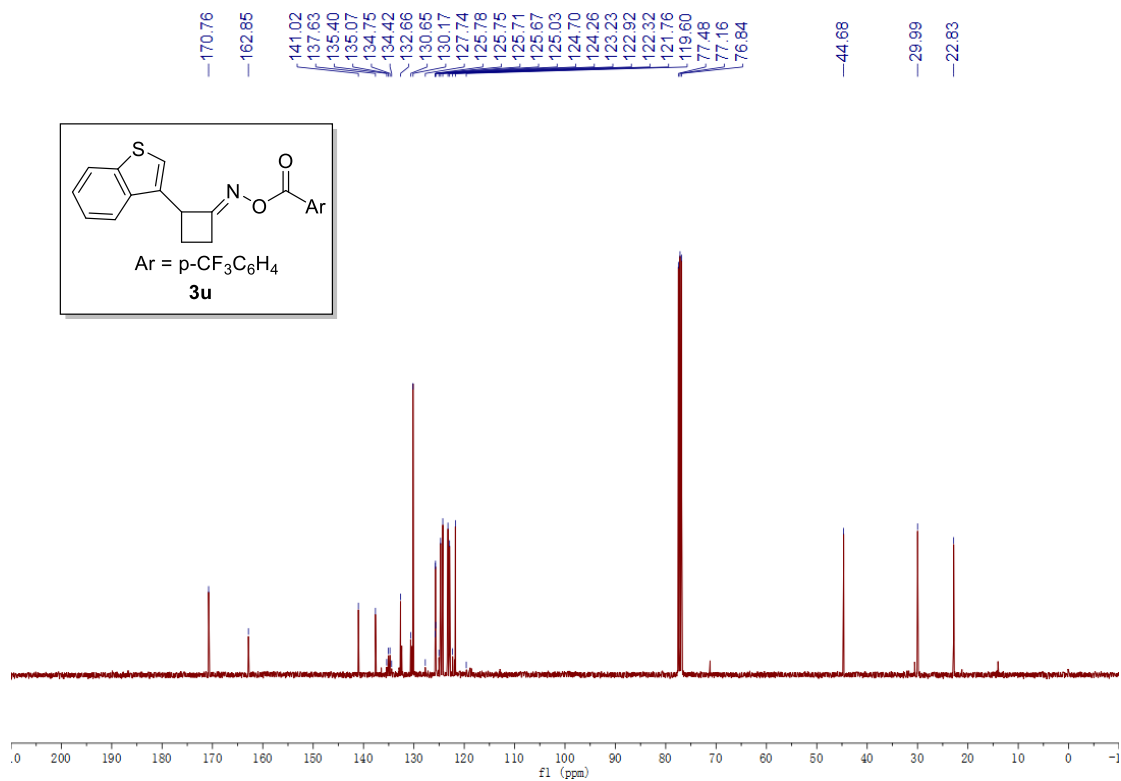
Supplementary Figure 53. ^1H NMR of **3t**



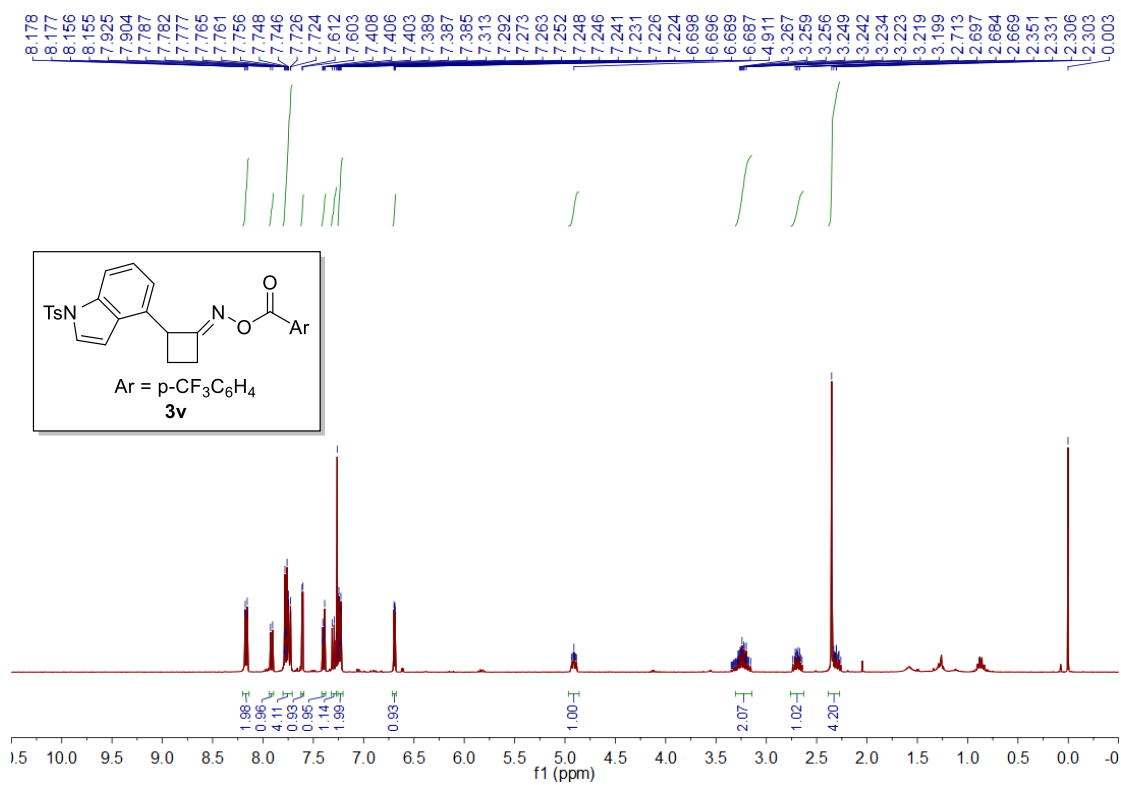
Supplementary Figure 54. ¹³C NMR of **3t**



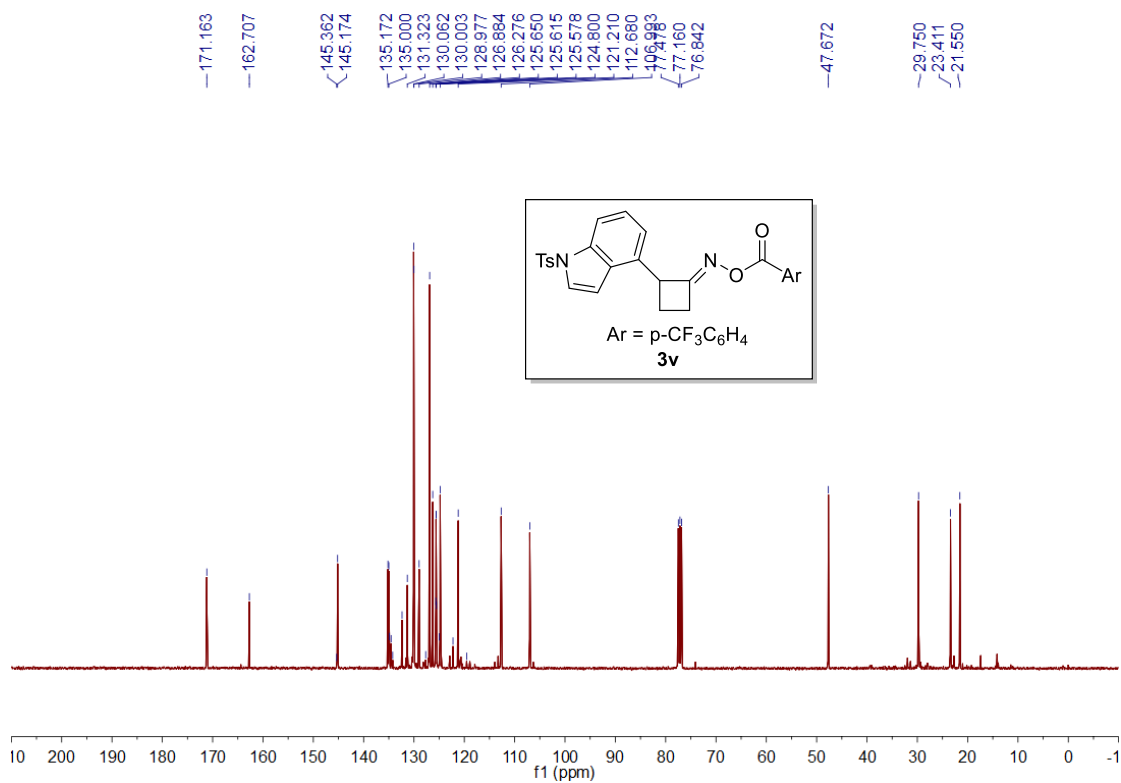
Supplementary Figure 55. ¹H NMR of **3u**



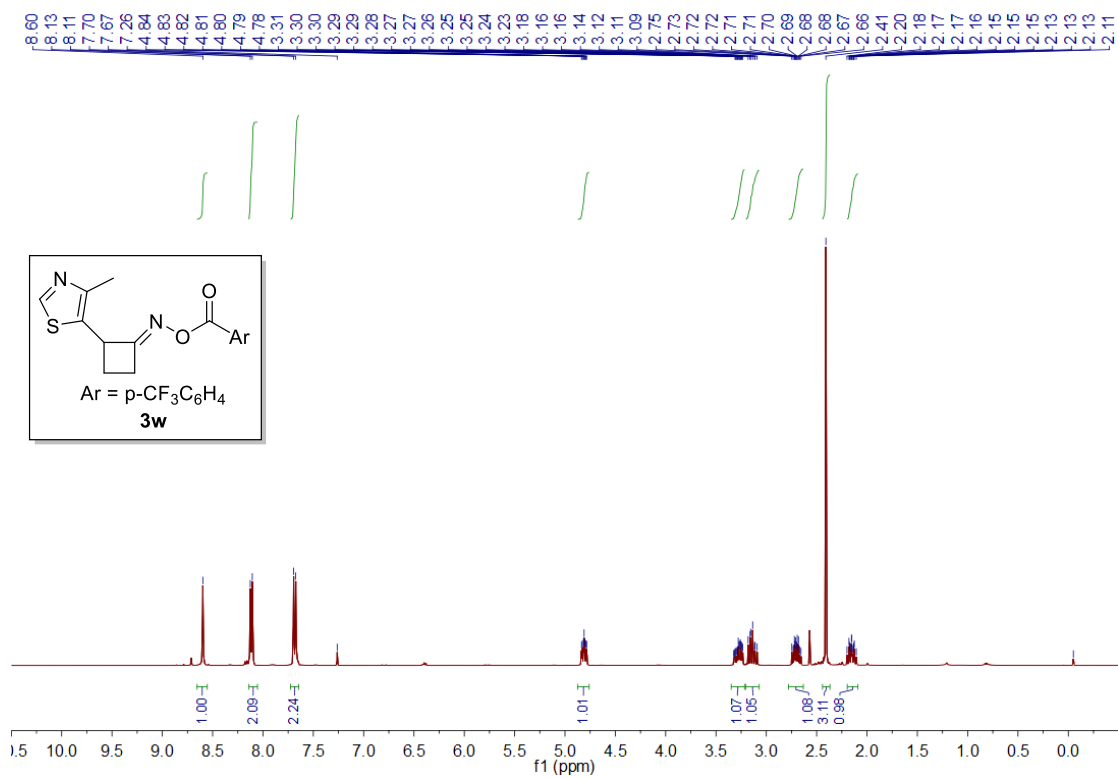
Supplementary Figure 56. ¹³C NMR of **3u**



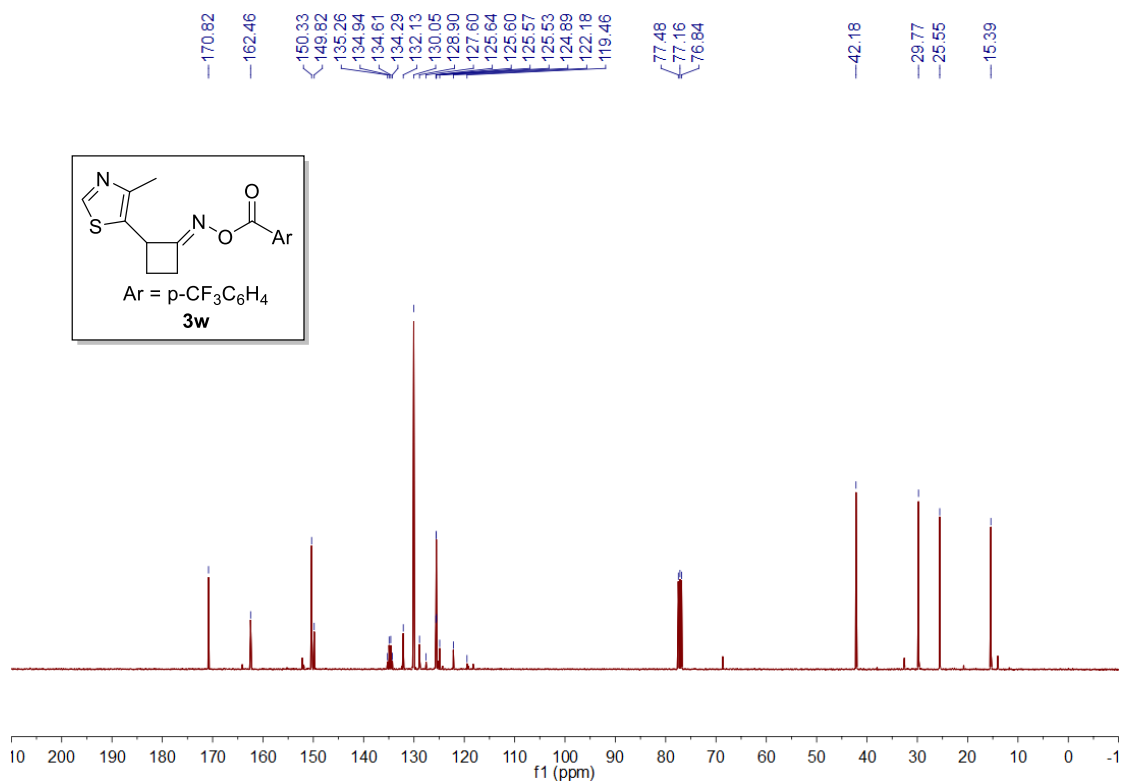
Supplementary Figure 57. ¹H NMR of **3v**



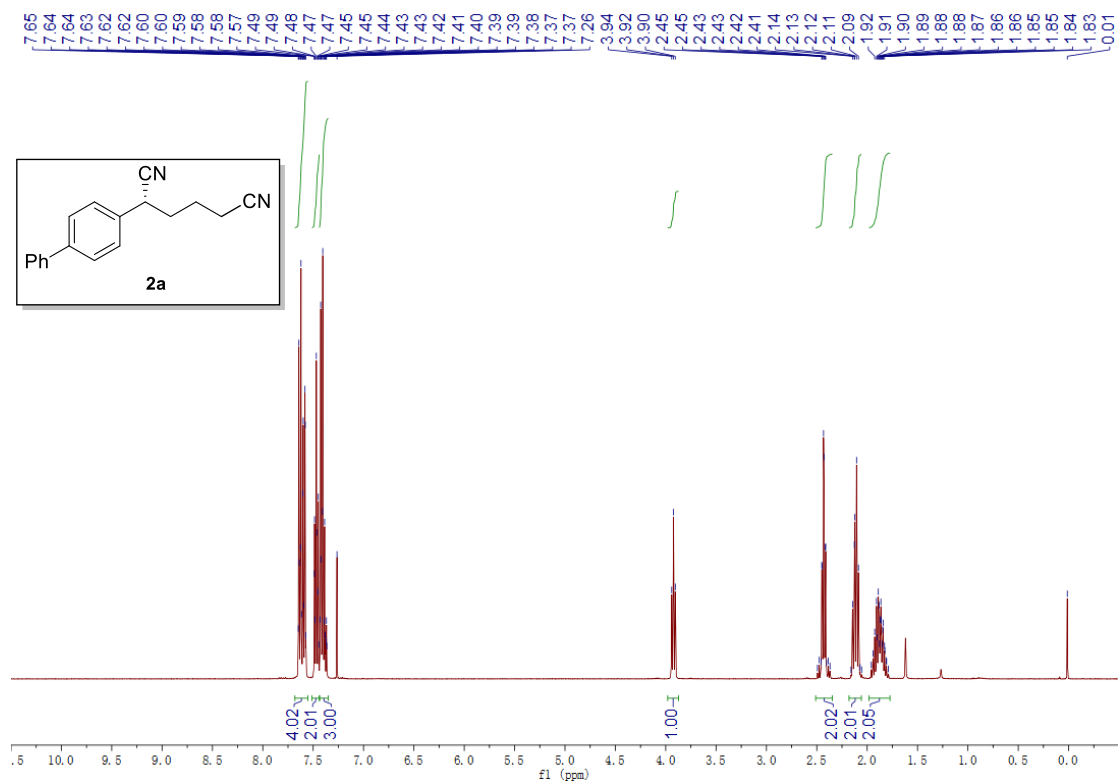
Supplementary Figure 58. ^{13}C NMR of **3v**



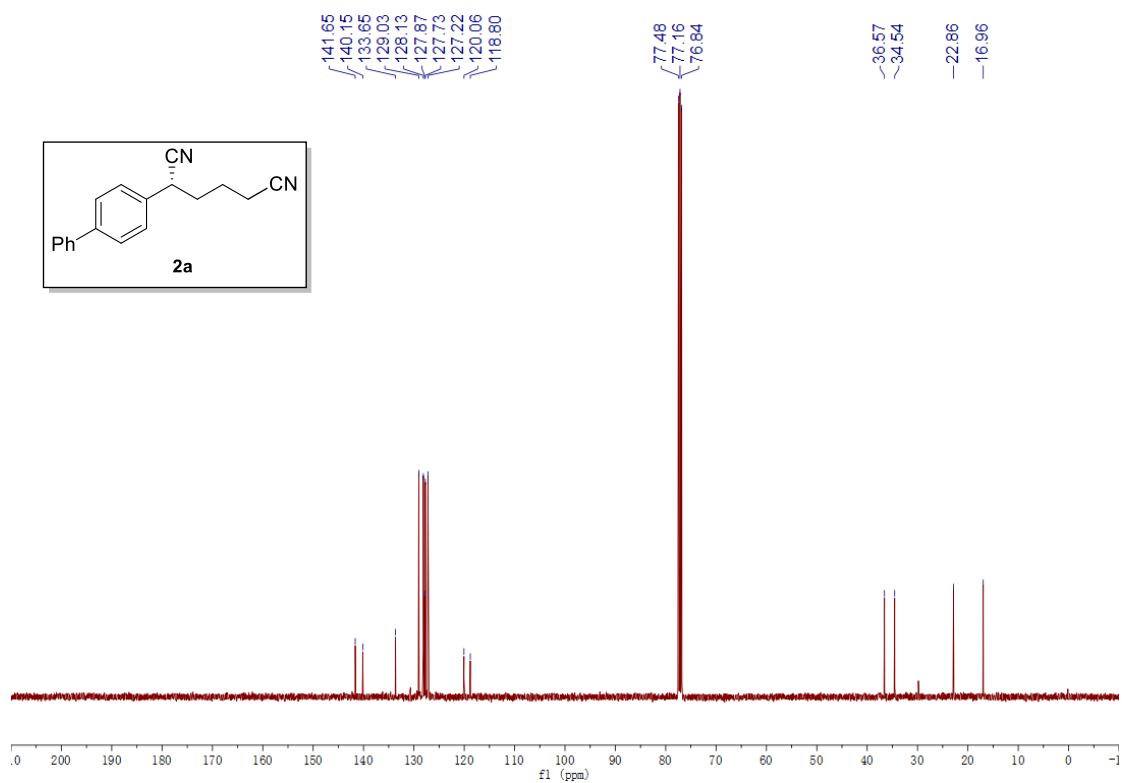
Supplementary Figure 59. ^1H NMR of **3w**



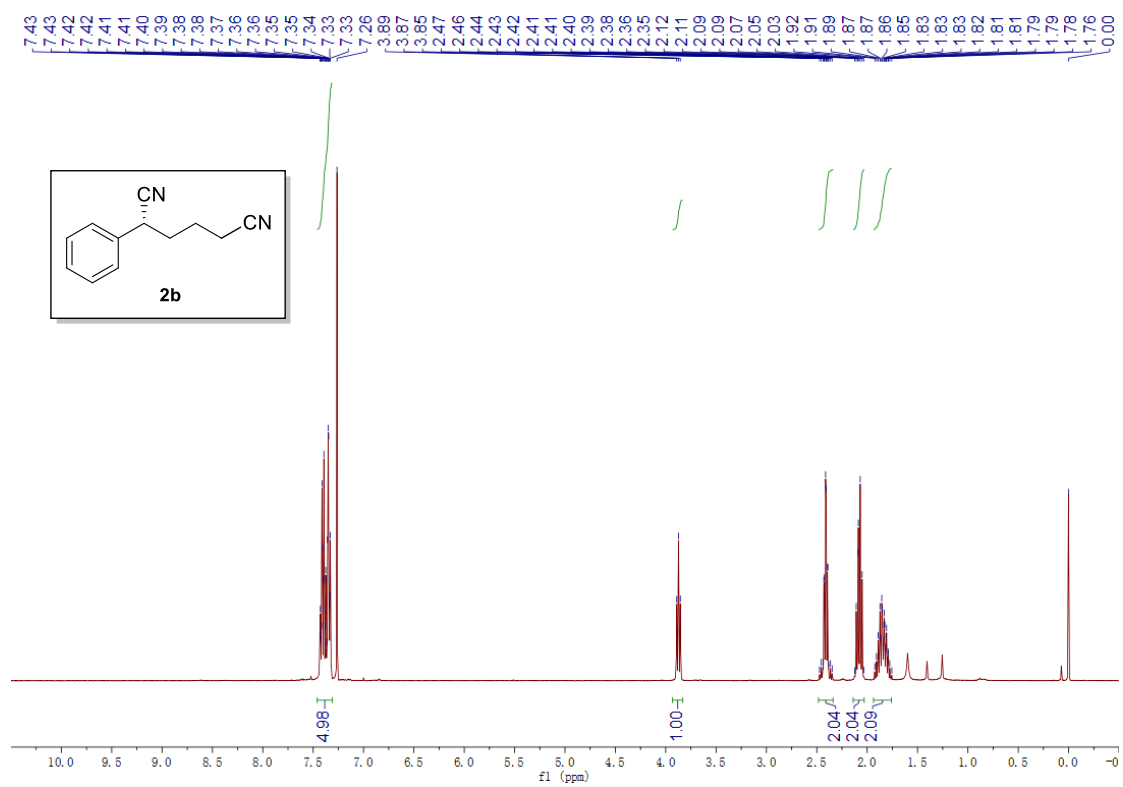
Supplementary Figure 60. ¹³C NMR of **3w**



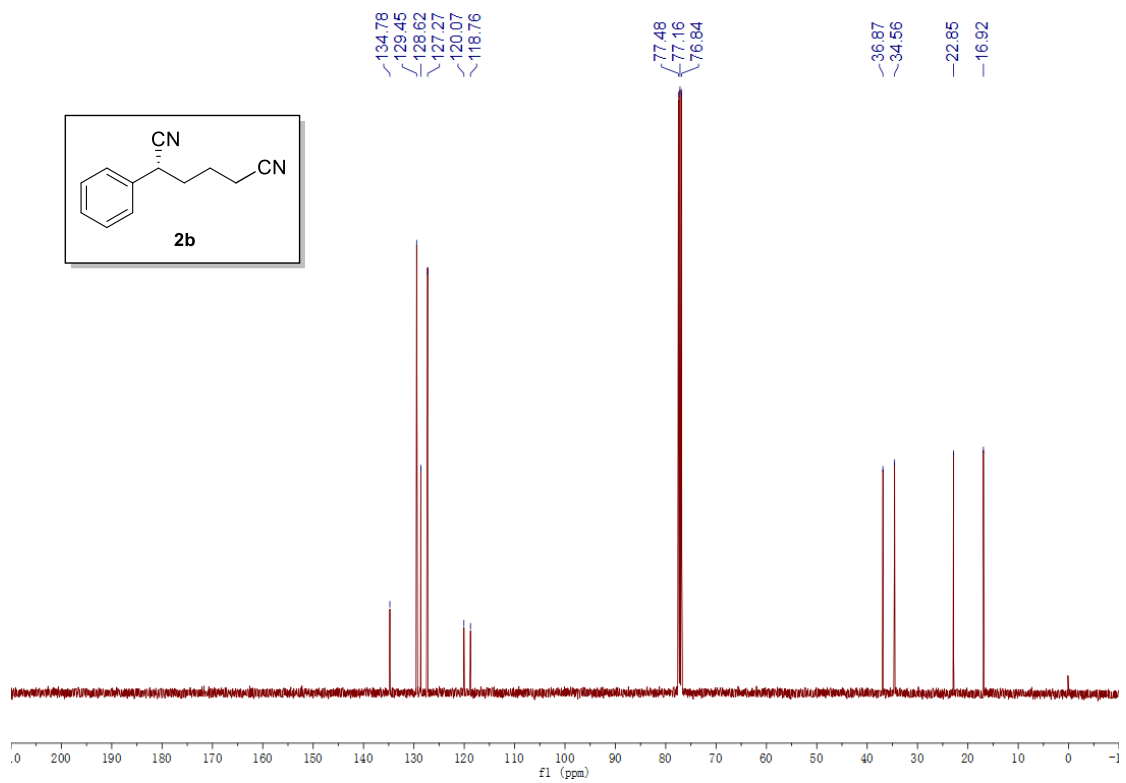
Supplementary Figure 61. ¹H NMR of **2a**



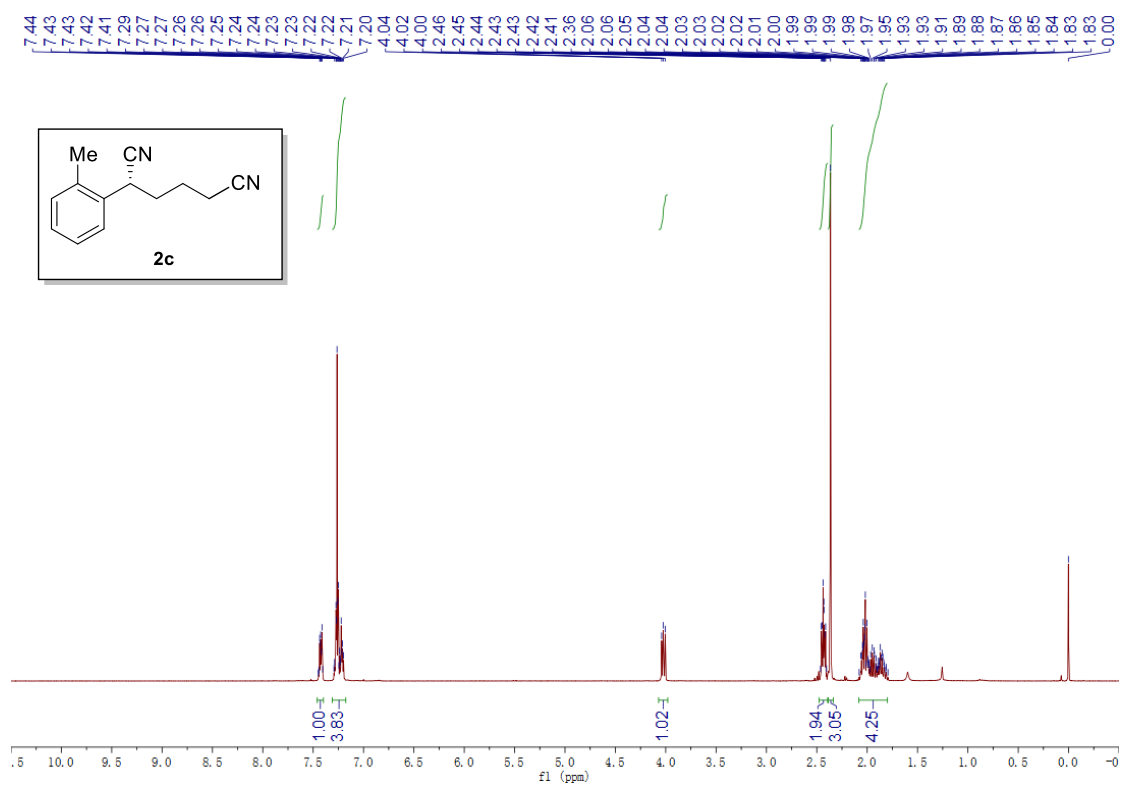
Supplementary Figure 62. ¹³C NMR of **2a**



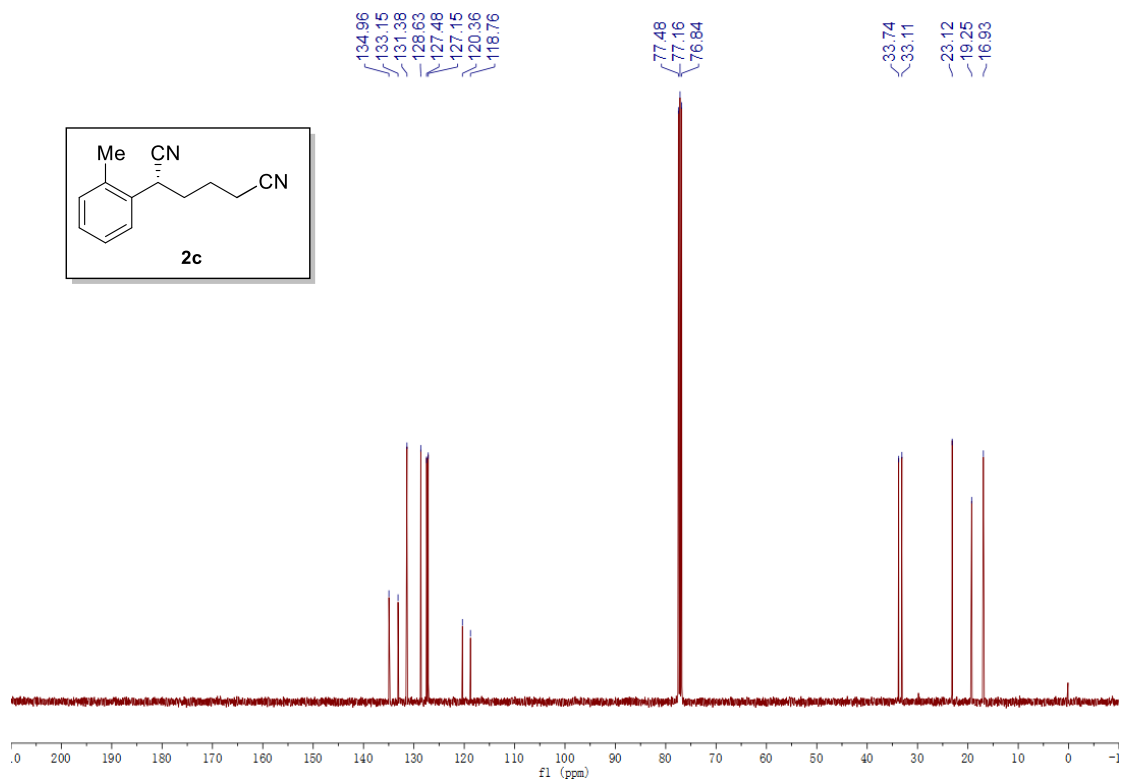
Supplementary Figure 63. ¹H NMR of **2b**



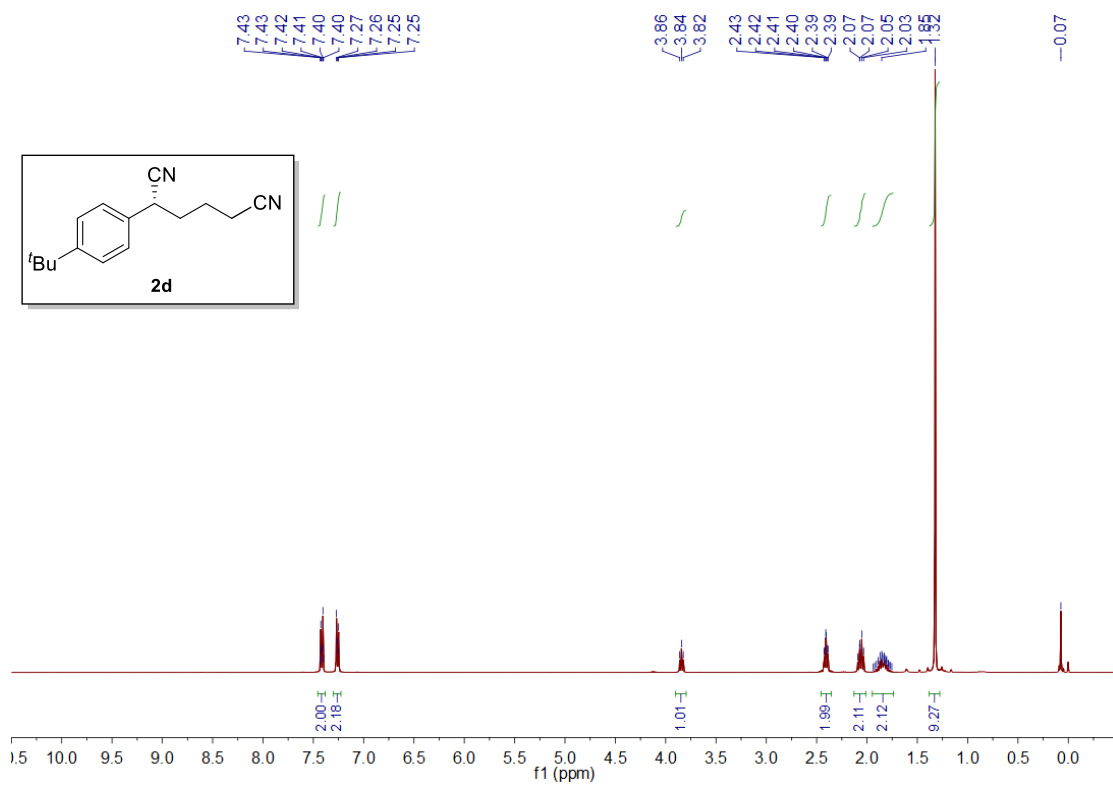
Supplementary Figure 64. ^{13}C NMR of **2b**



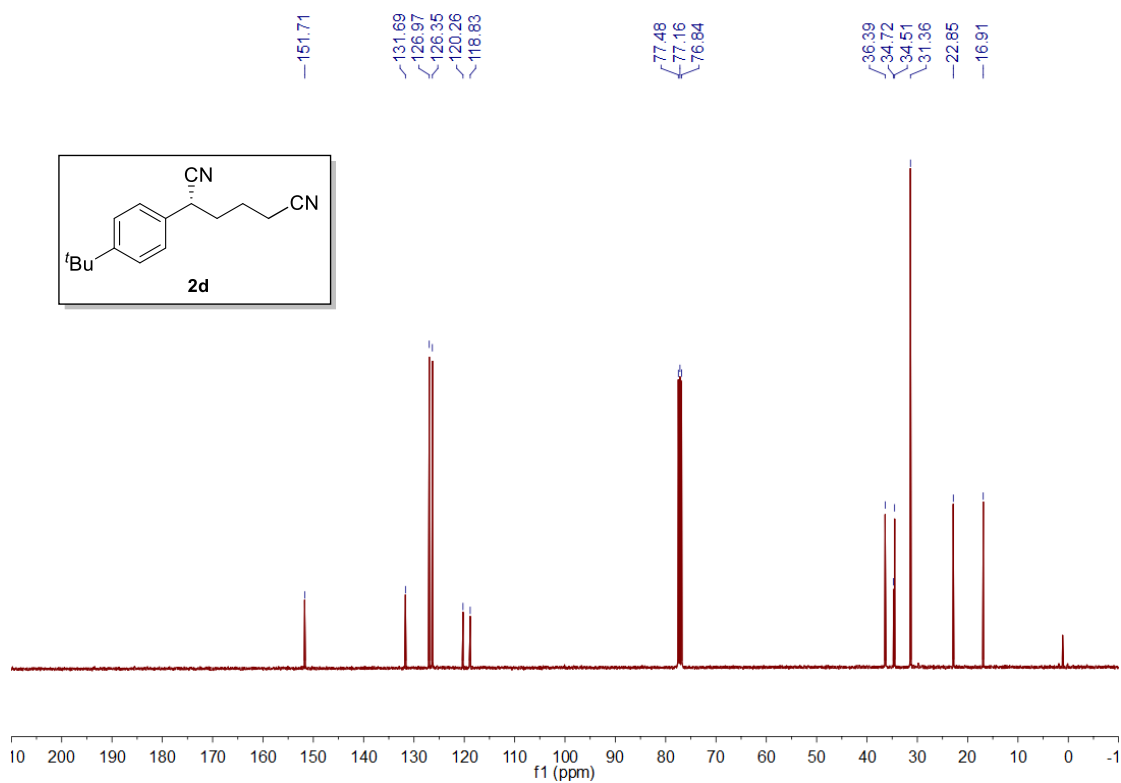
Supplementary Figure 65. ^1H NMR of **2c**



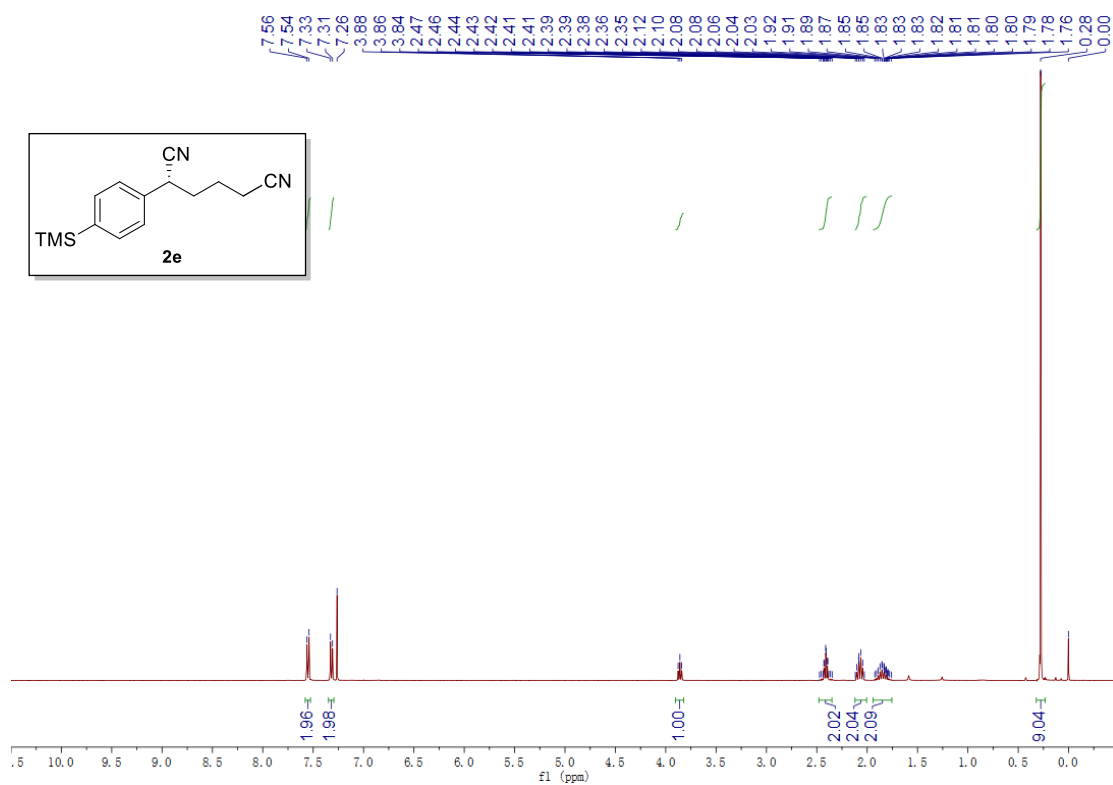
Supplementary Figure 66. ¹³C NMR of **2c**



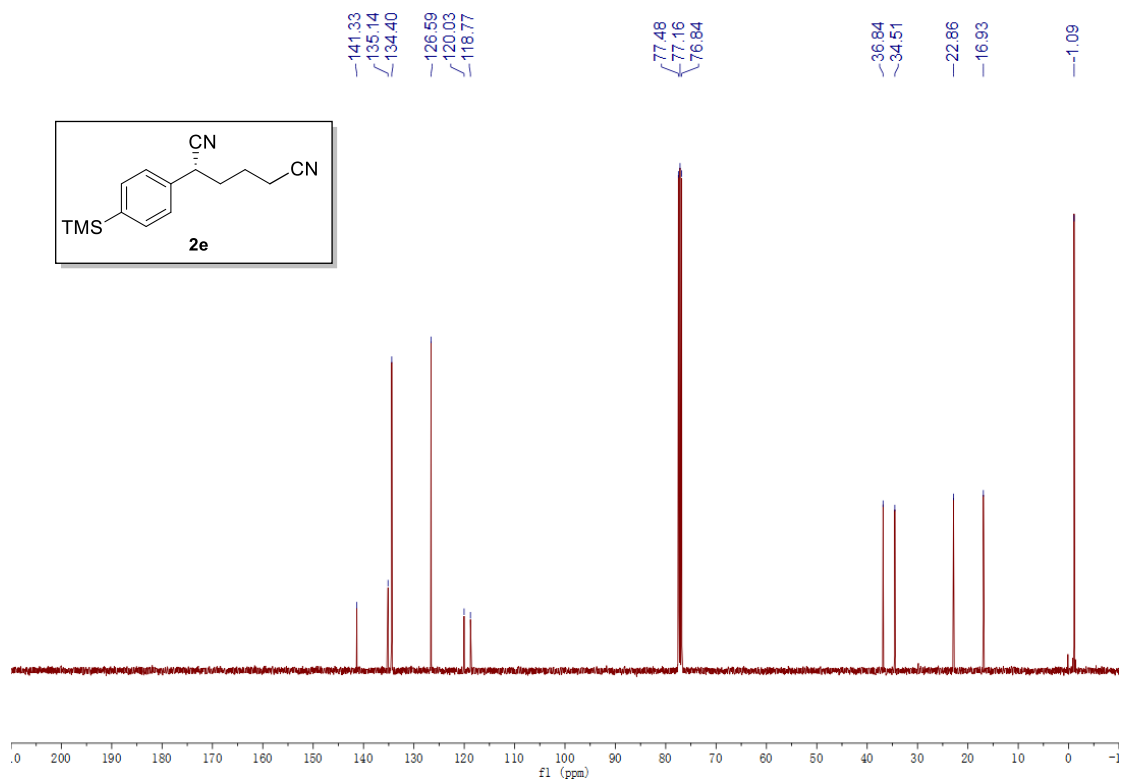
Supplementary Figure 67. ¹H NMR of **2d**



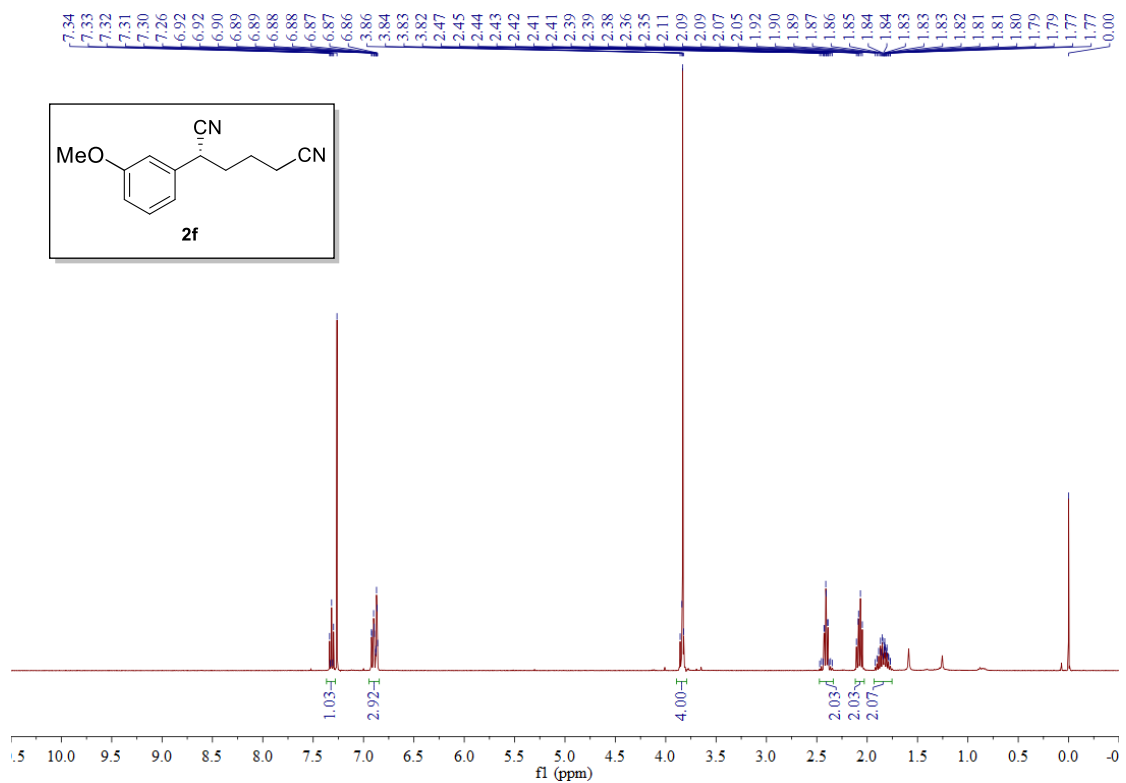
Supplementary Figure 68. ¹³C NMR of **2d**



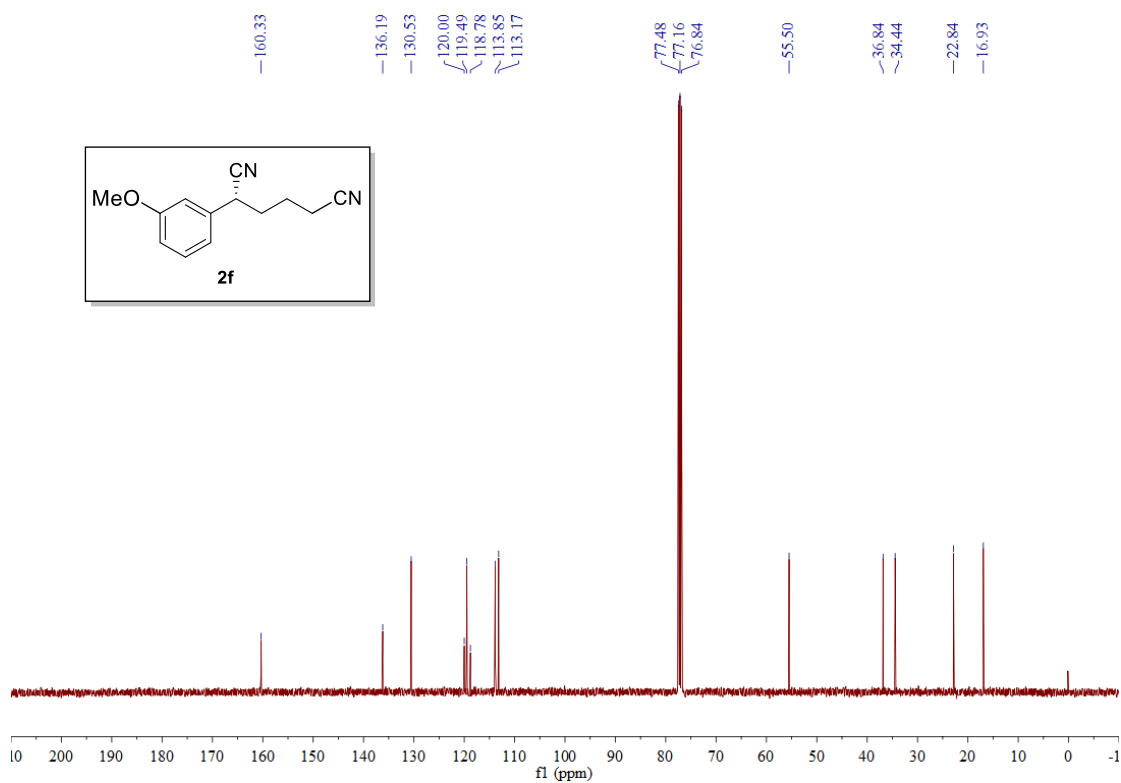
Supplementary Figure 69. ¹H NMR of **2e**



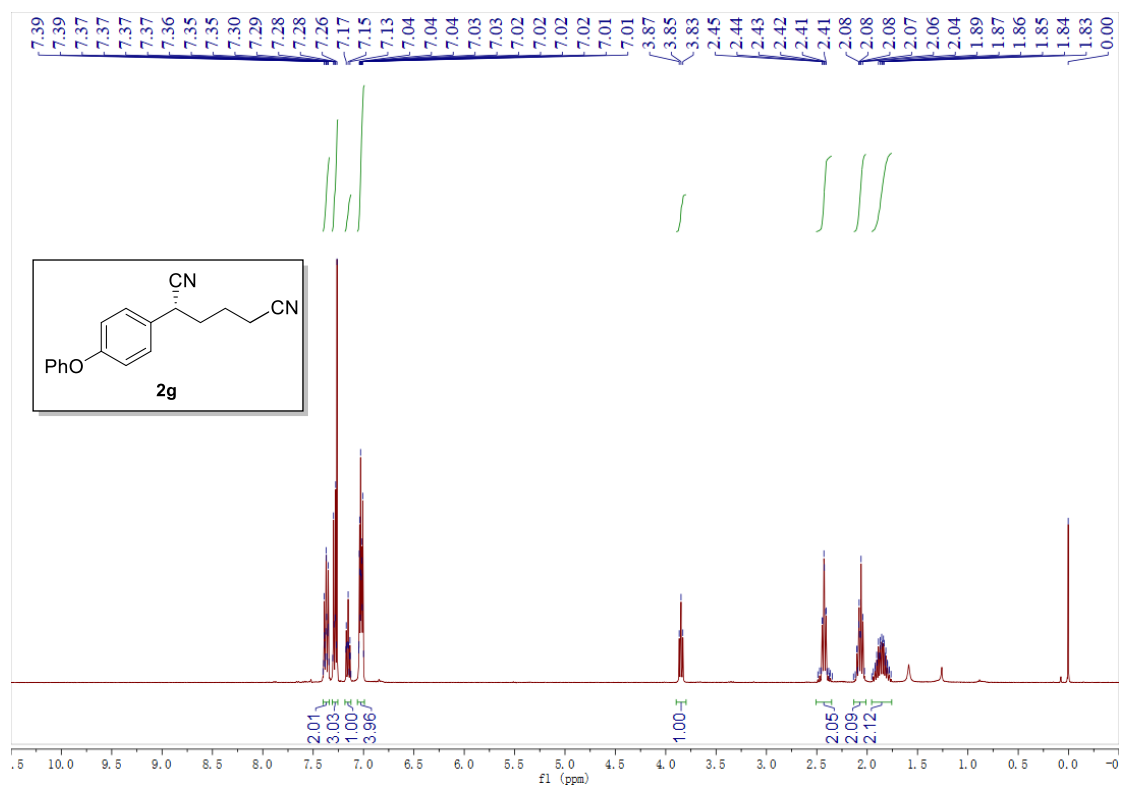
Supplementary Figure 70. ¹³C NMR of **2e**



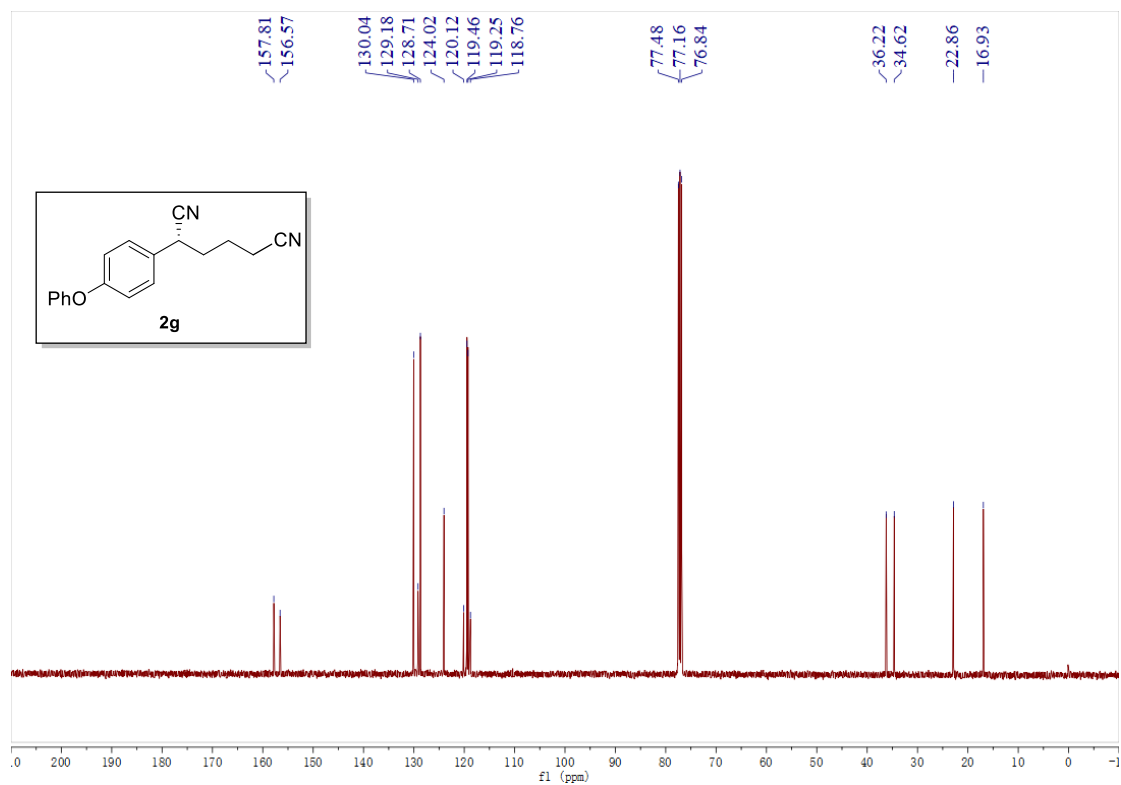
Supplementary Figure 71. ¹H NMR of **2f**



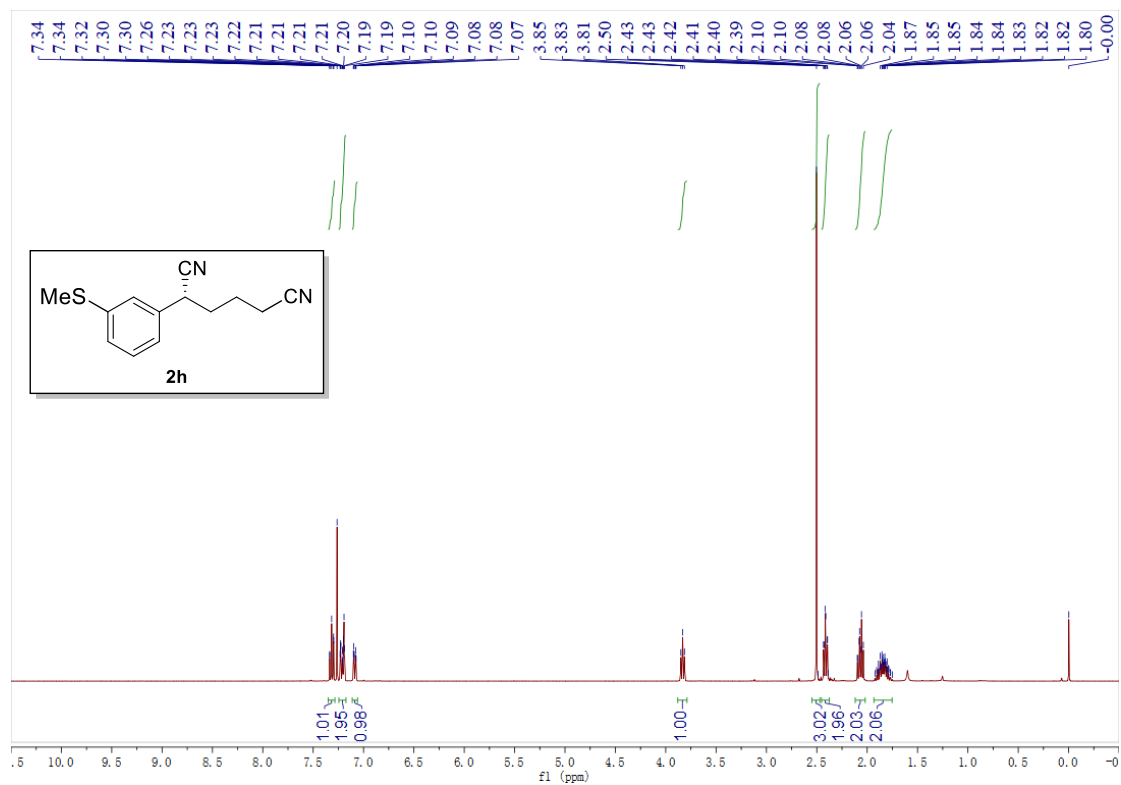
Supplementary Figure 72. ¹³C NMR of **2f**



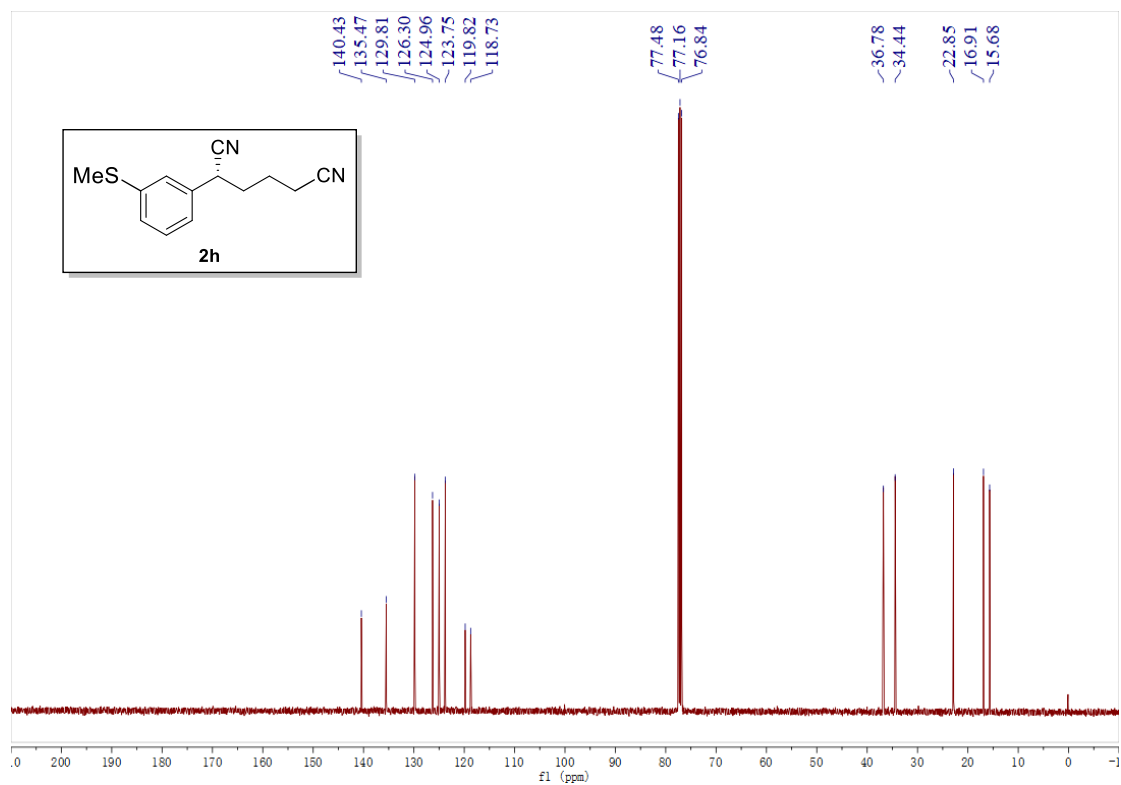
Supplementary Figure 73. ¹H NMR of **2g**



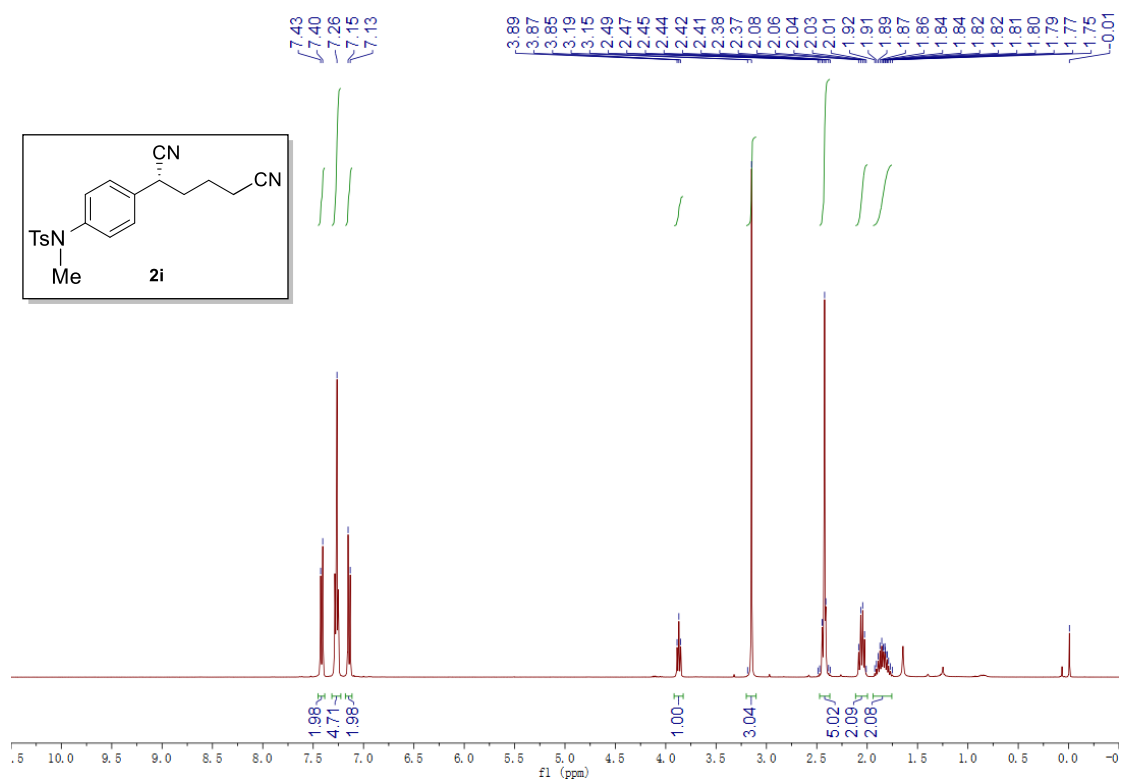
Supplementary Figure 74. ¹³C NMR of **2g**



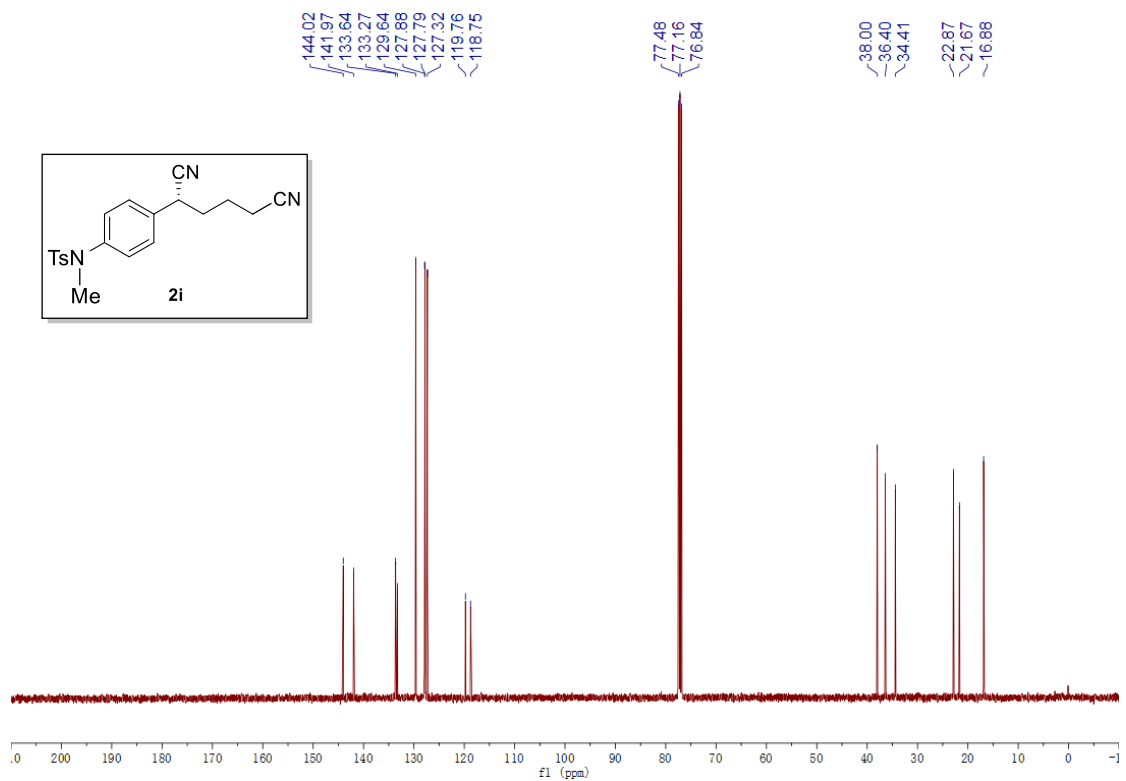
Supplementary Figure 75. ¹H NMR of **2h**



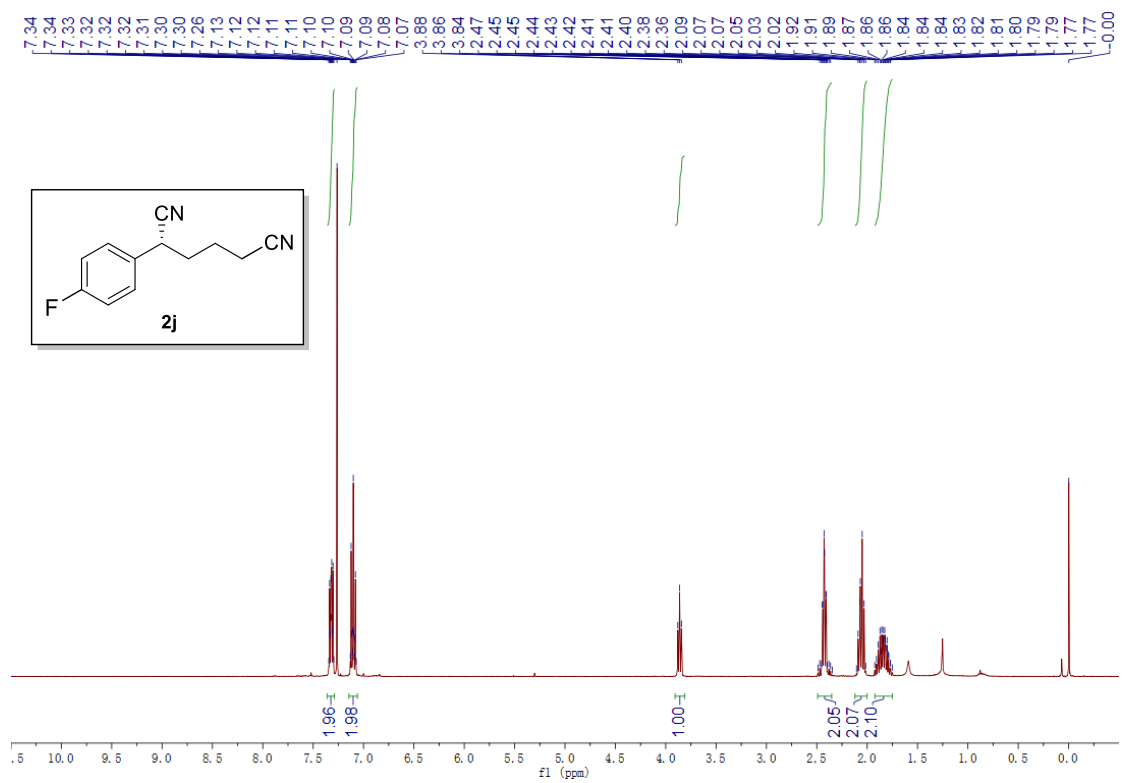
Supplementary Figure 76. ¹³C NMR of **2h**



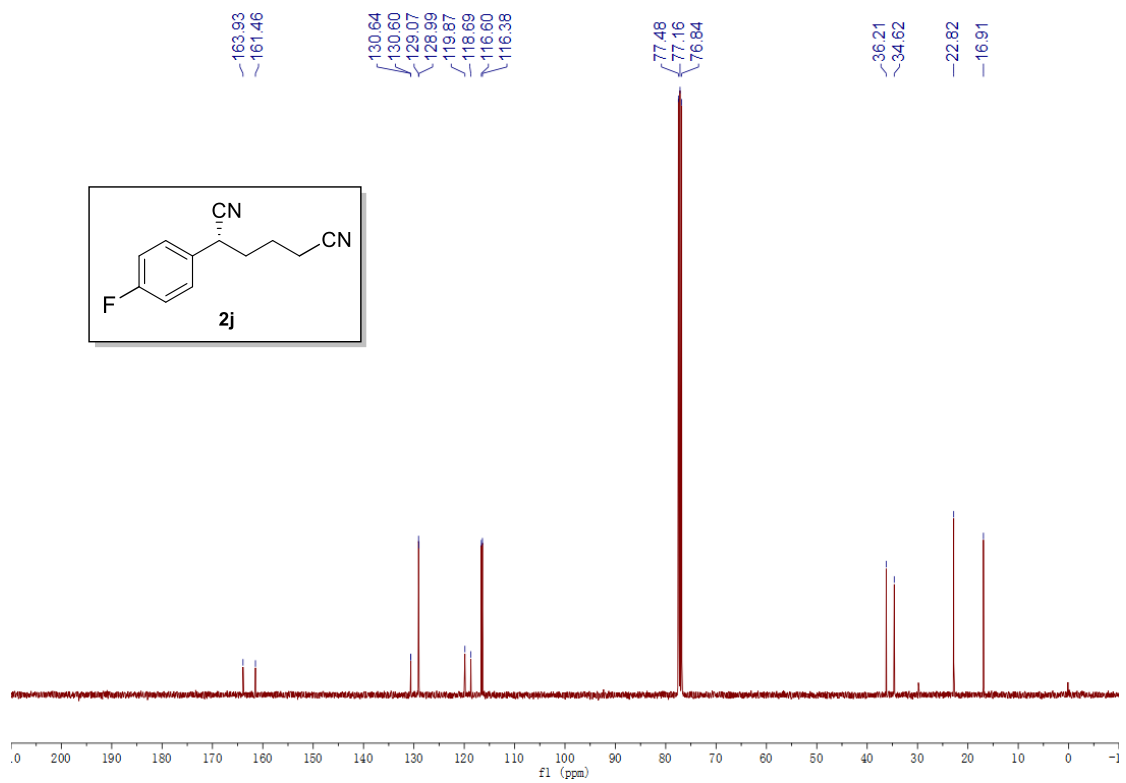
Supplementary Figure 77. ¹H NMR of **2i**



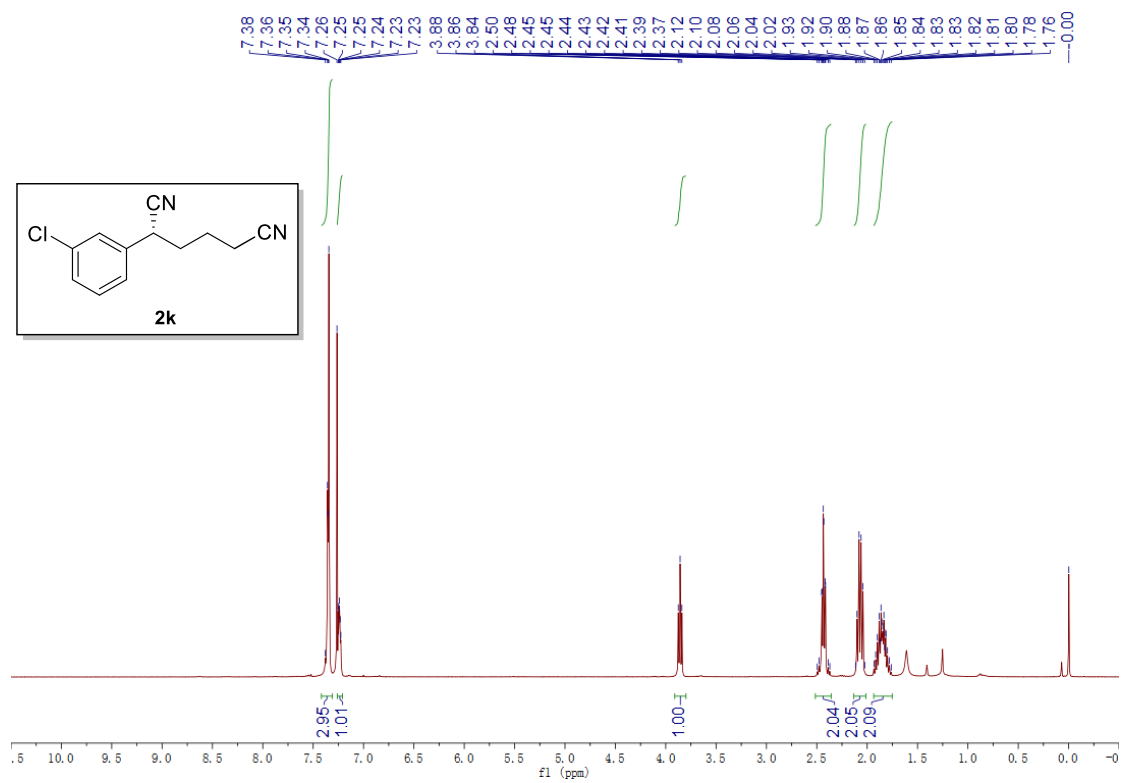
Supplementary Figure 78. ¹³C NMR of **2i**



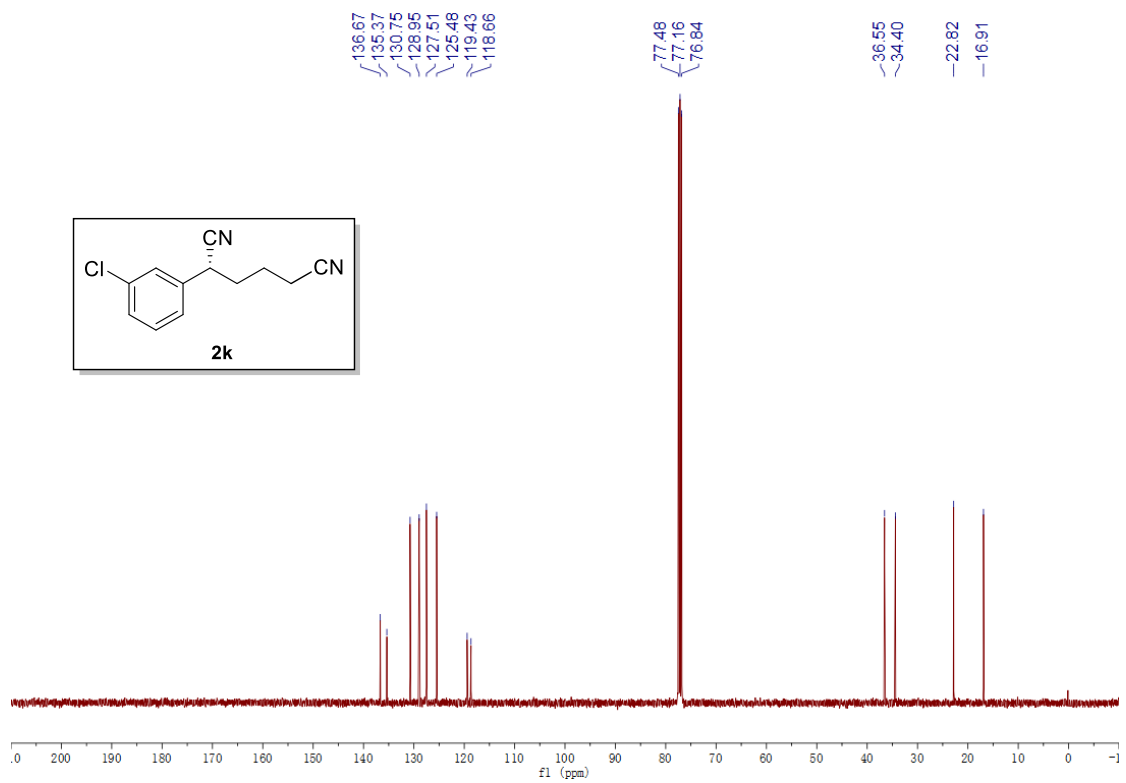
Supplementary Figure 79. ¹H NMR of **2j**



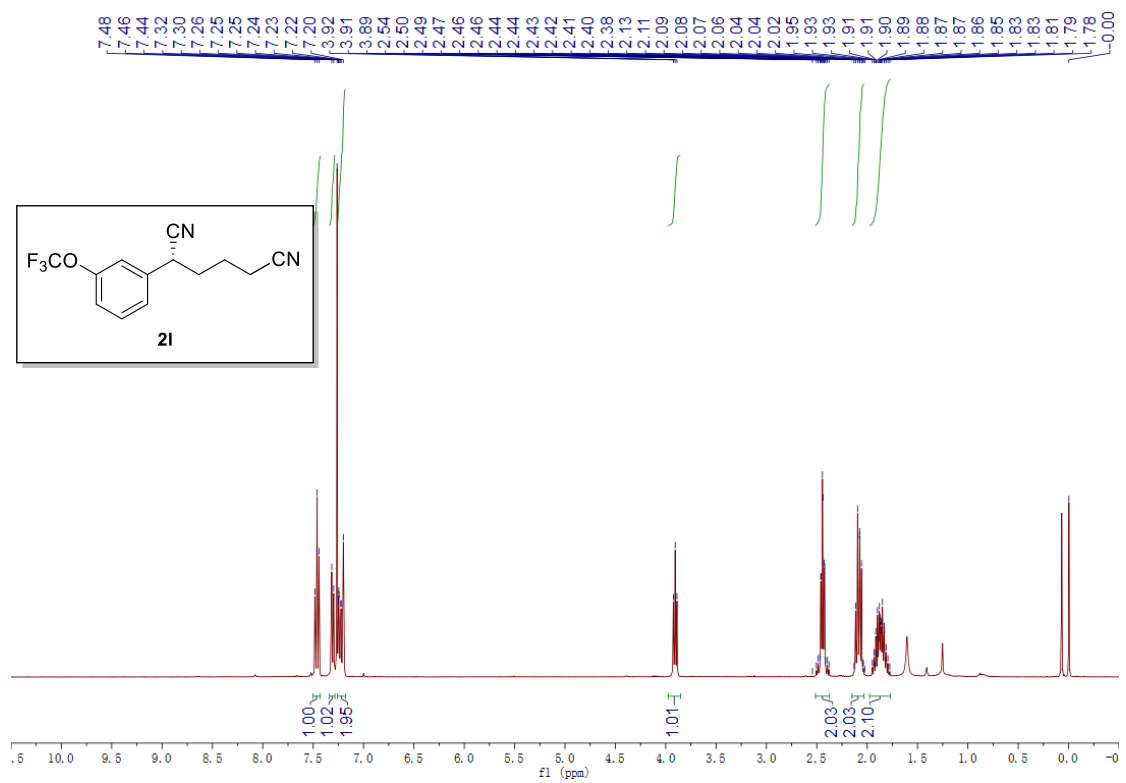
Supplementary Figure 80. ^{13}C NMR of **2j**



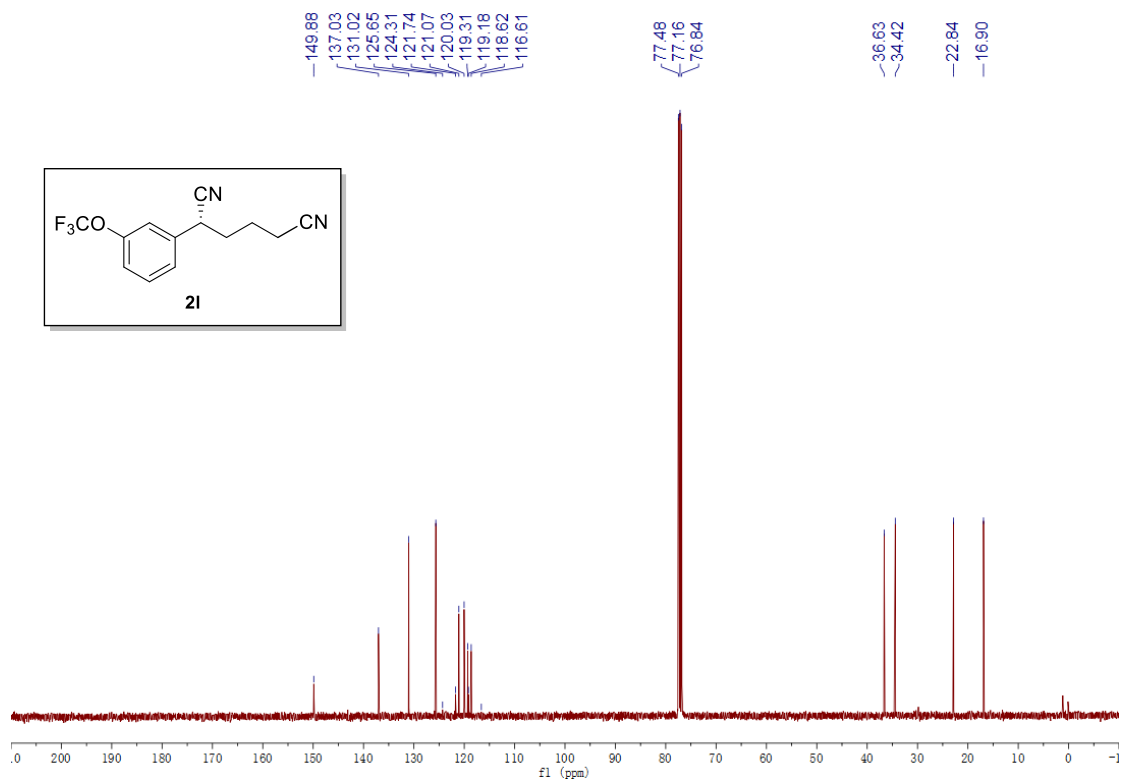
Supplementary Figure 81. ^1H NMR of **2k**



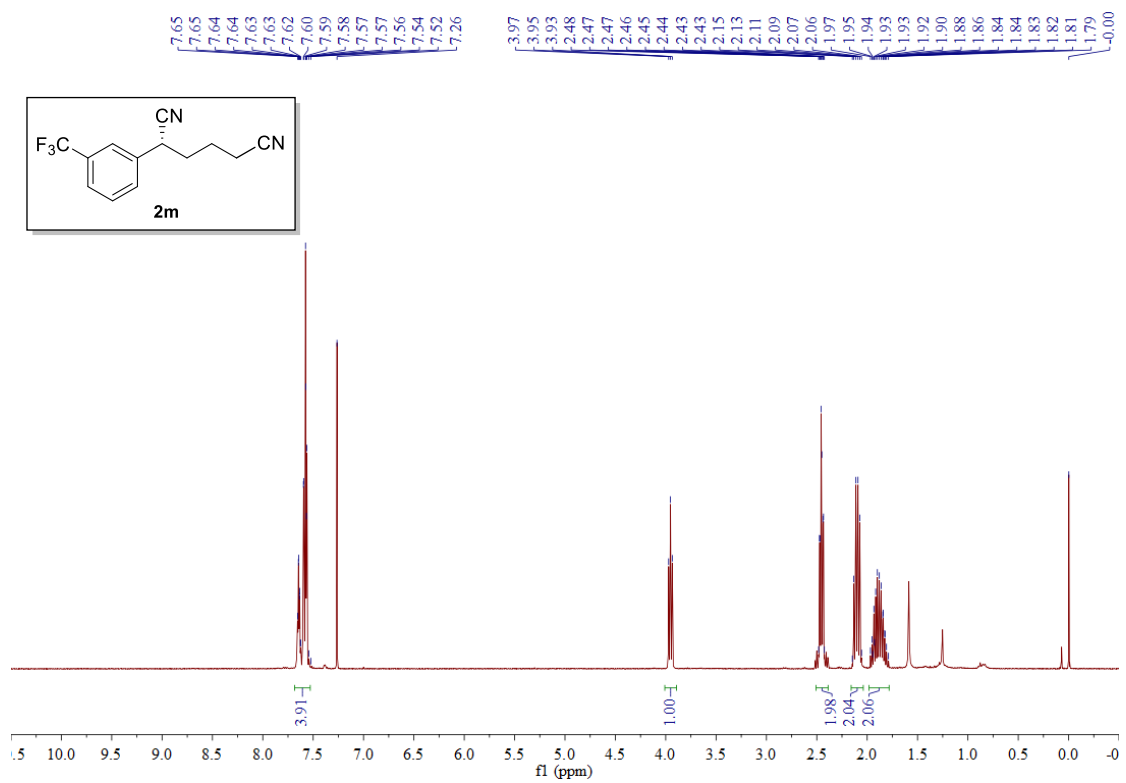
Supplementary Figure 82. ¹³C NMR of **2k**



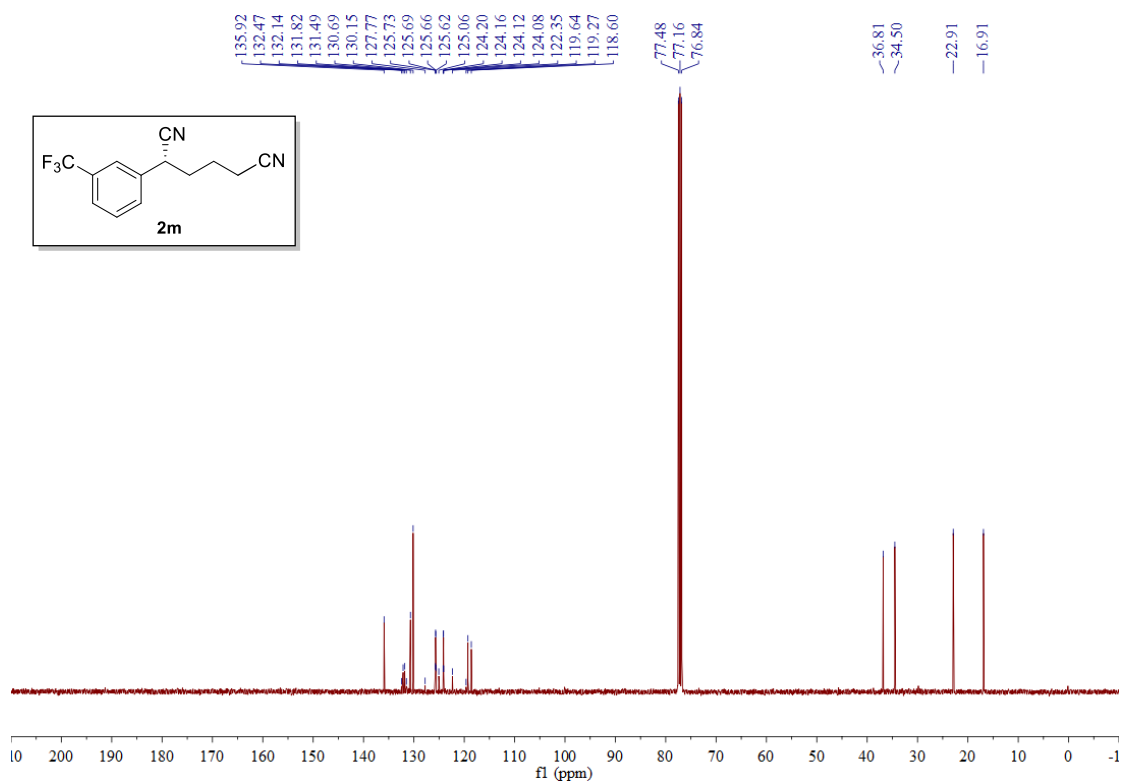
Supplementary Figure 83. ¹H NMR of **2l**



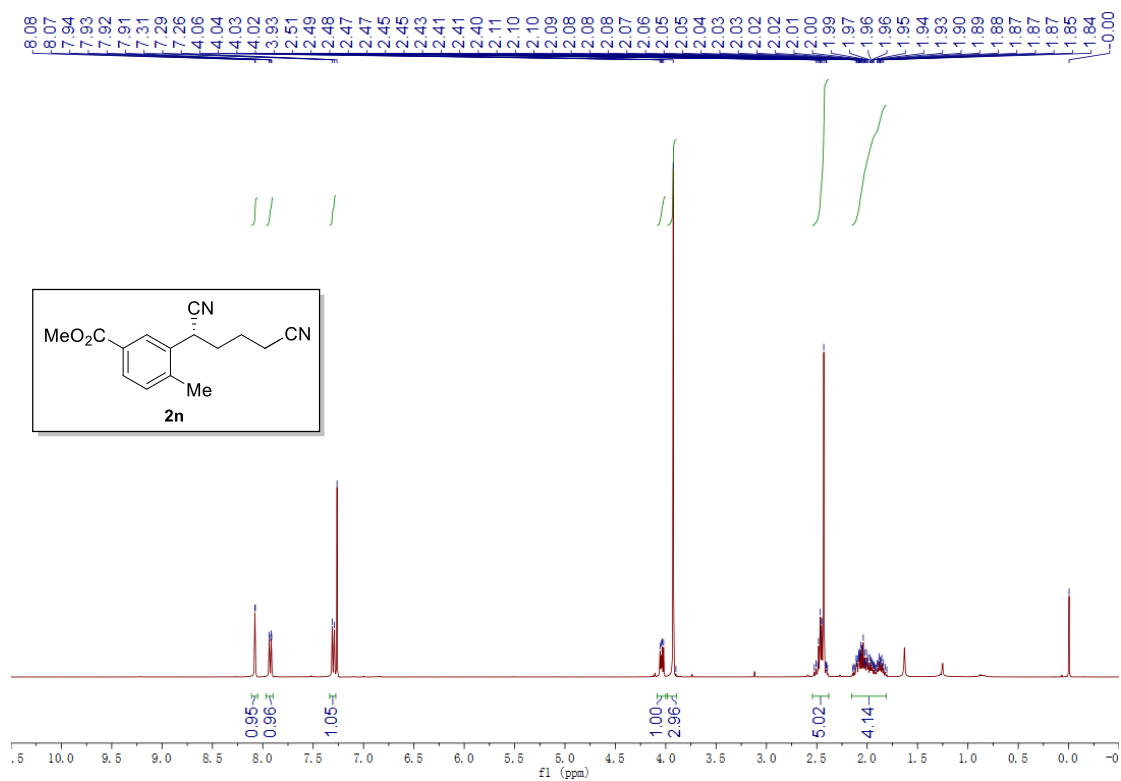
Supplementary Figure 84. ¹³C NMR of **2l**



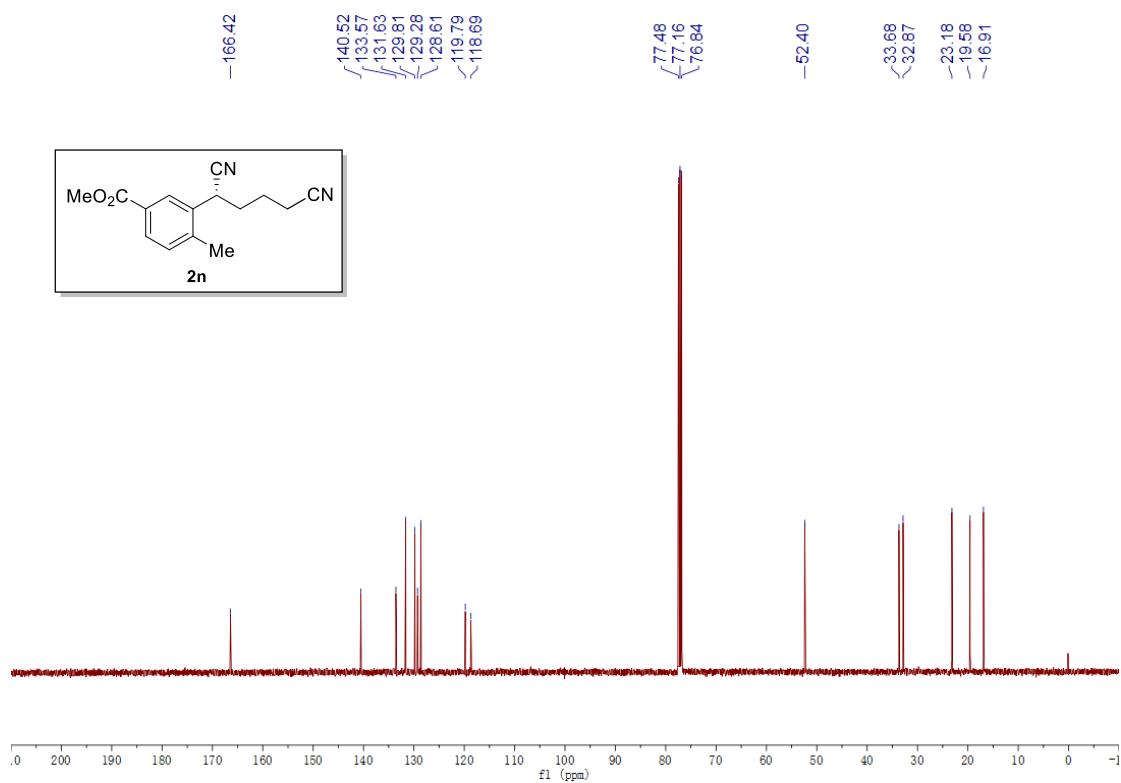
Supplementary Figure 85. ¹H NMR of **2m**



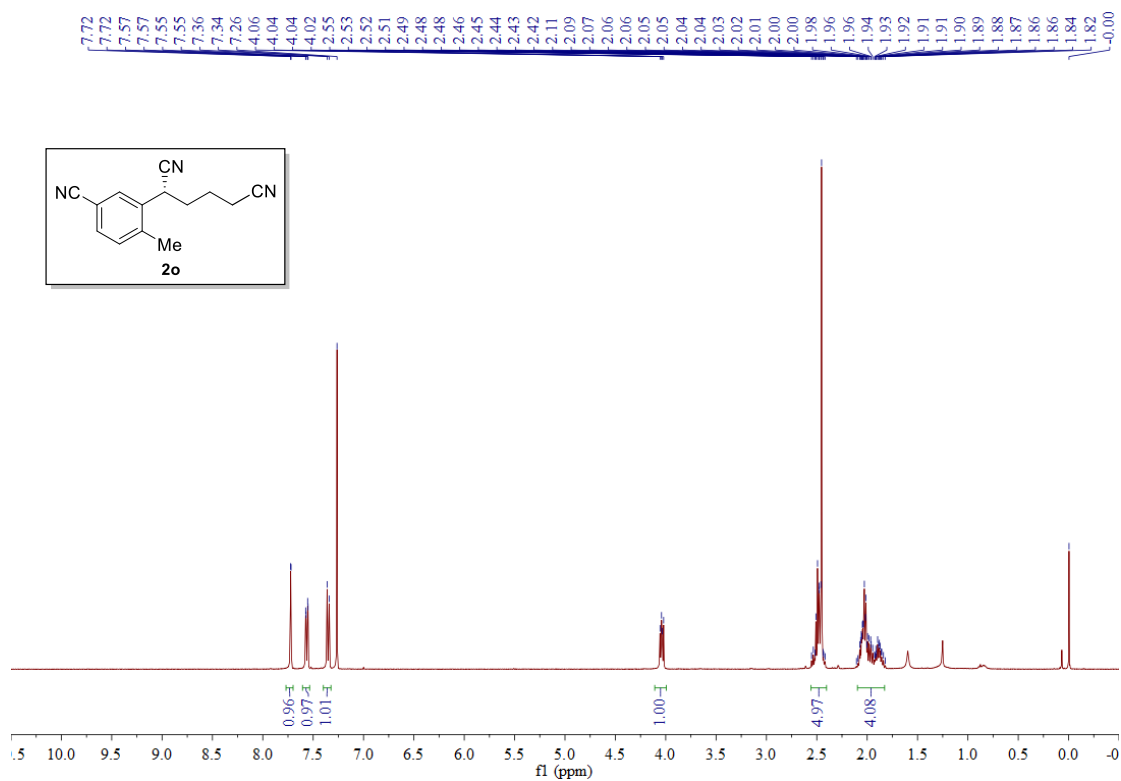
Supplementary Figure 86. ¹³C NMR of **2m**



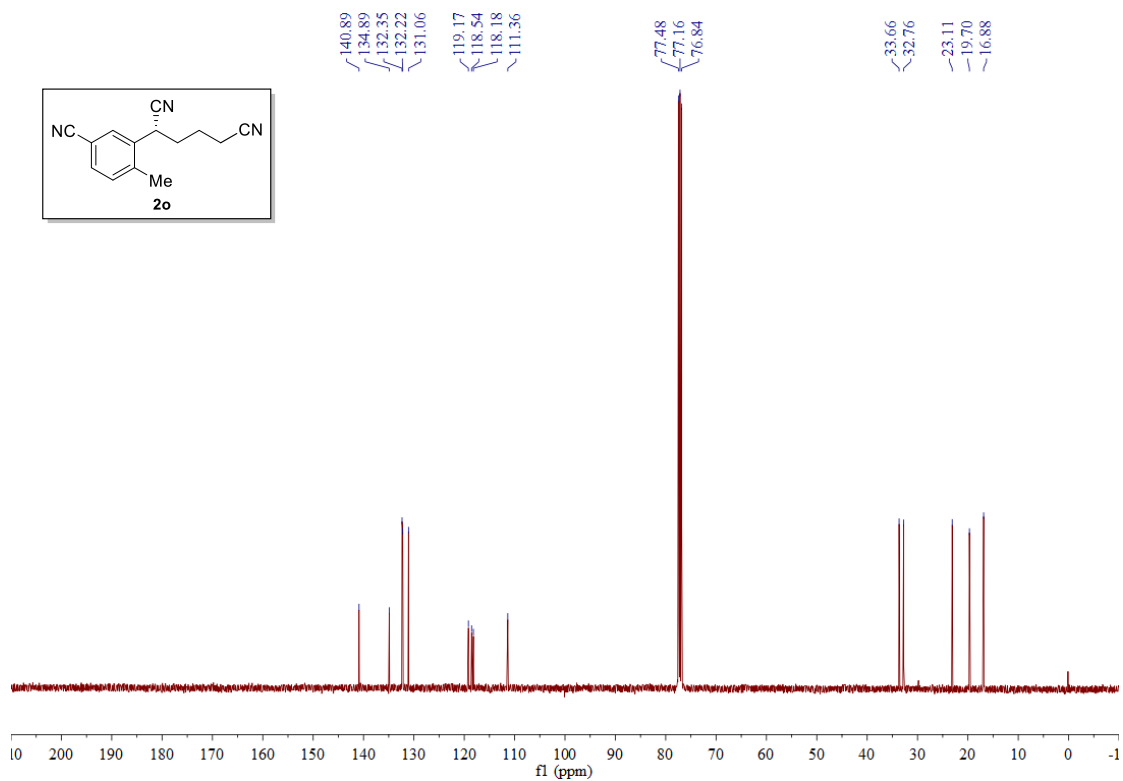
Supplementary Figure 87. ¹H NMR of **2n**



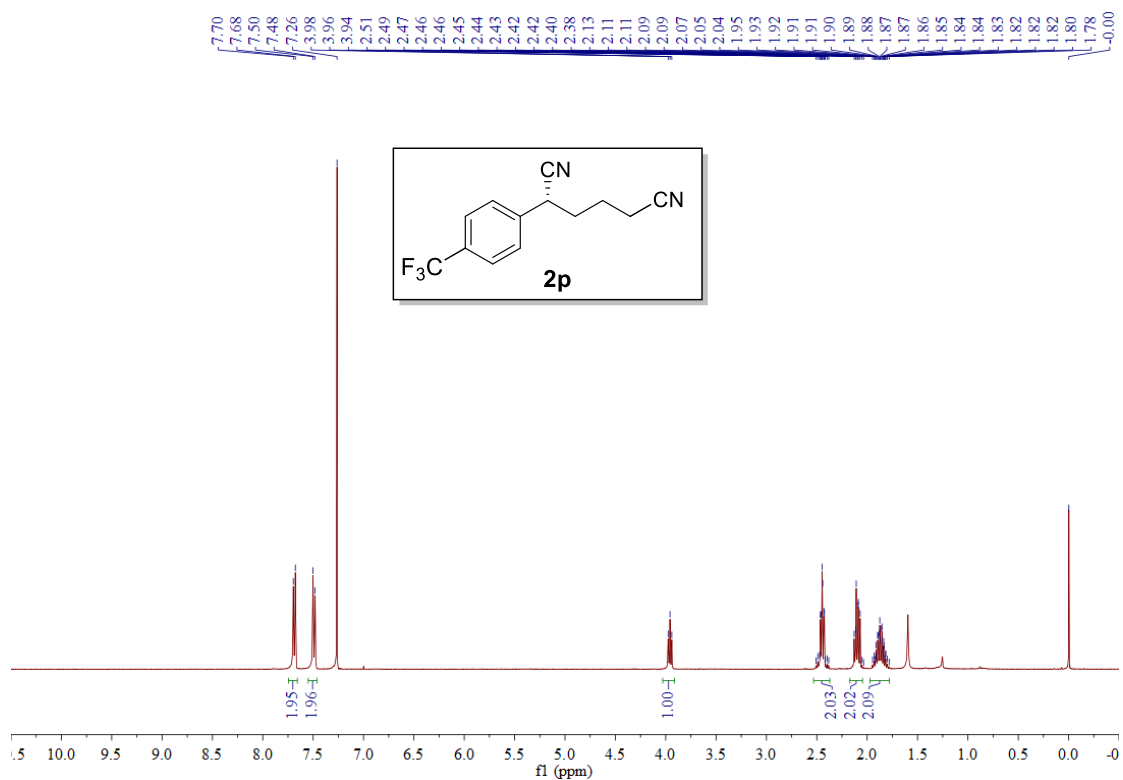
Supplementary Figure 88. ^{13}C NMR of **2n**



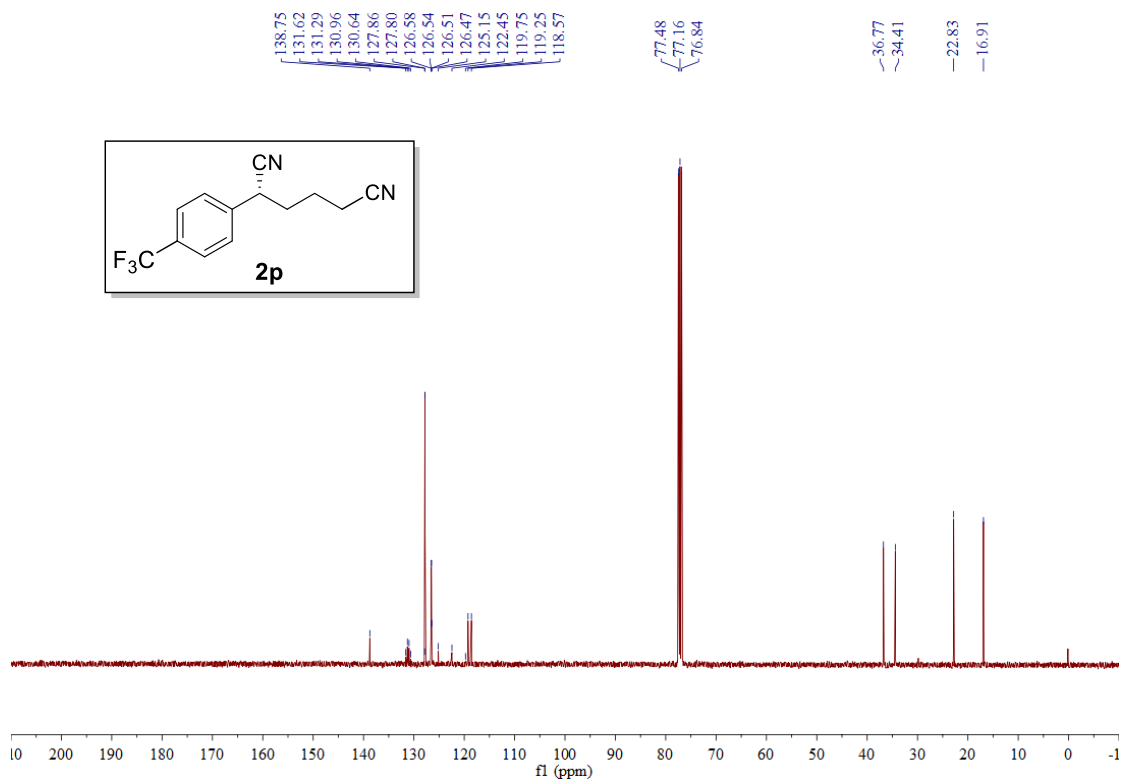
Supplementary Figure 89. ^1H NMR of **2o**



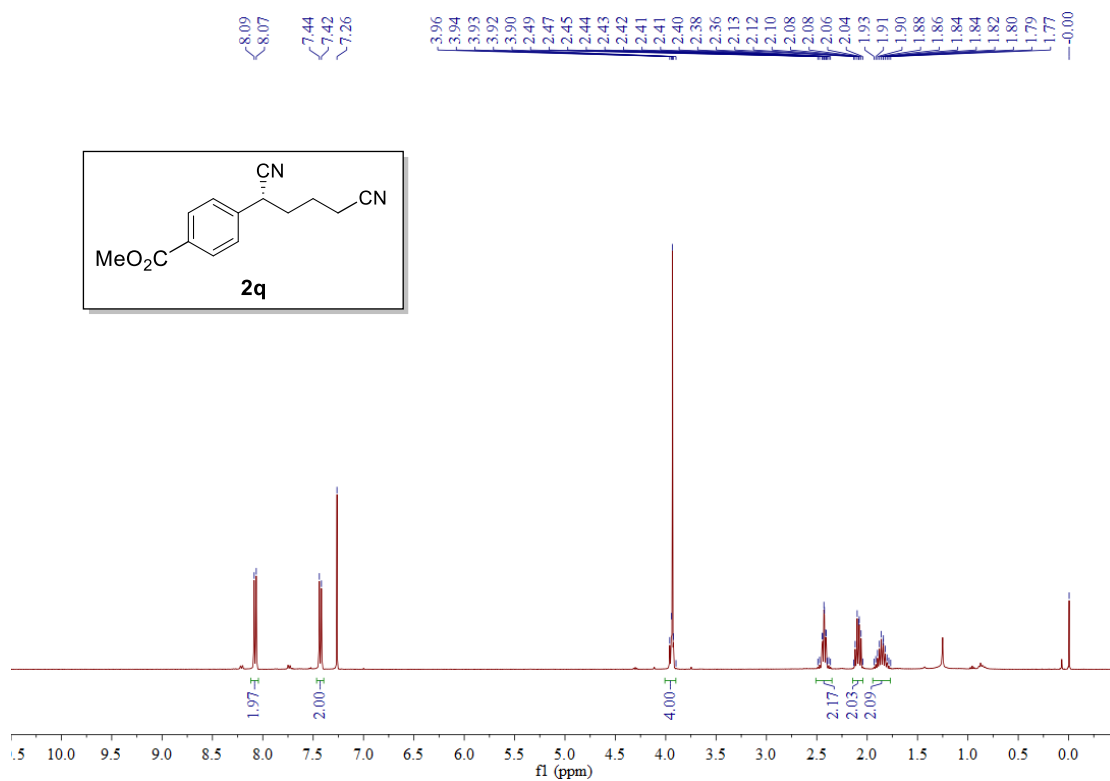
Supplementary Figure 90. ¹³C NMR of **2o**



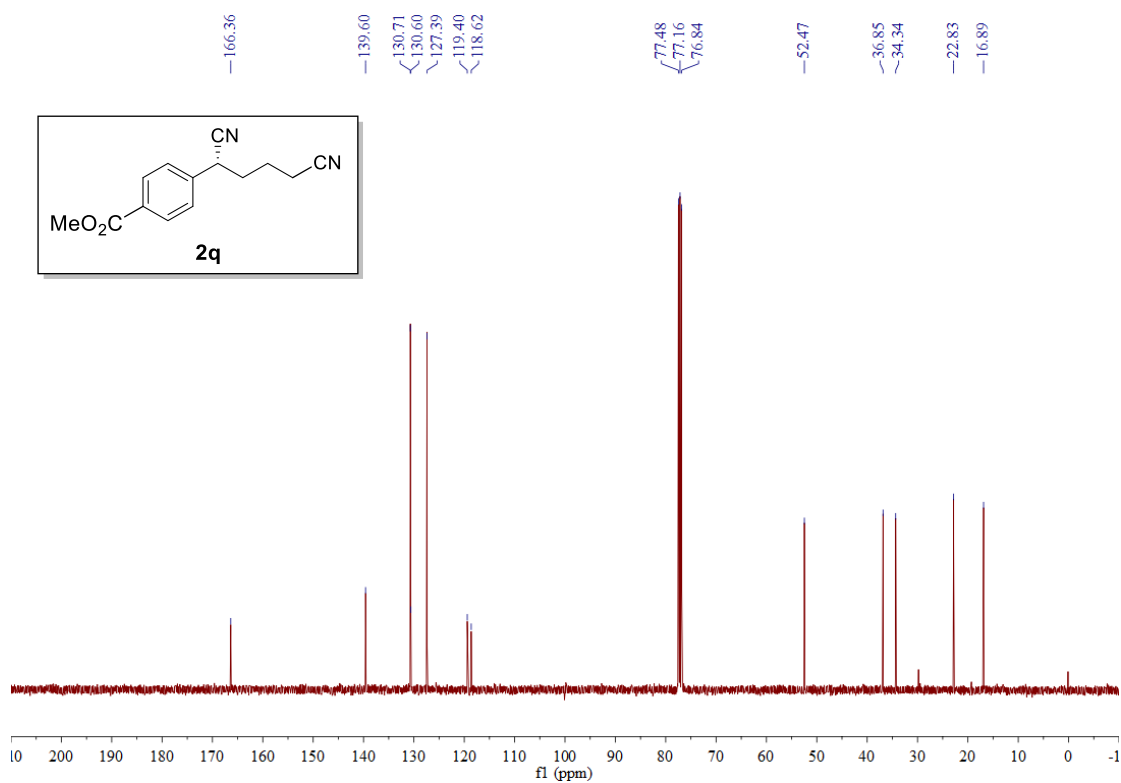
Supplementary Figure 91. ¹H NMR of **2p**



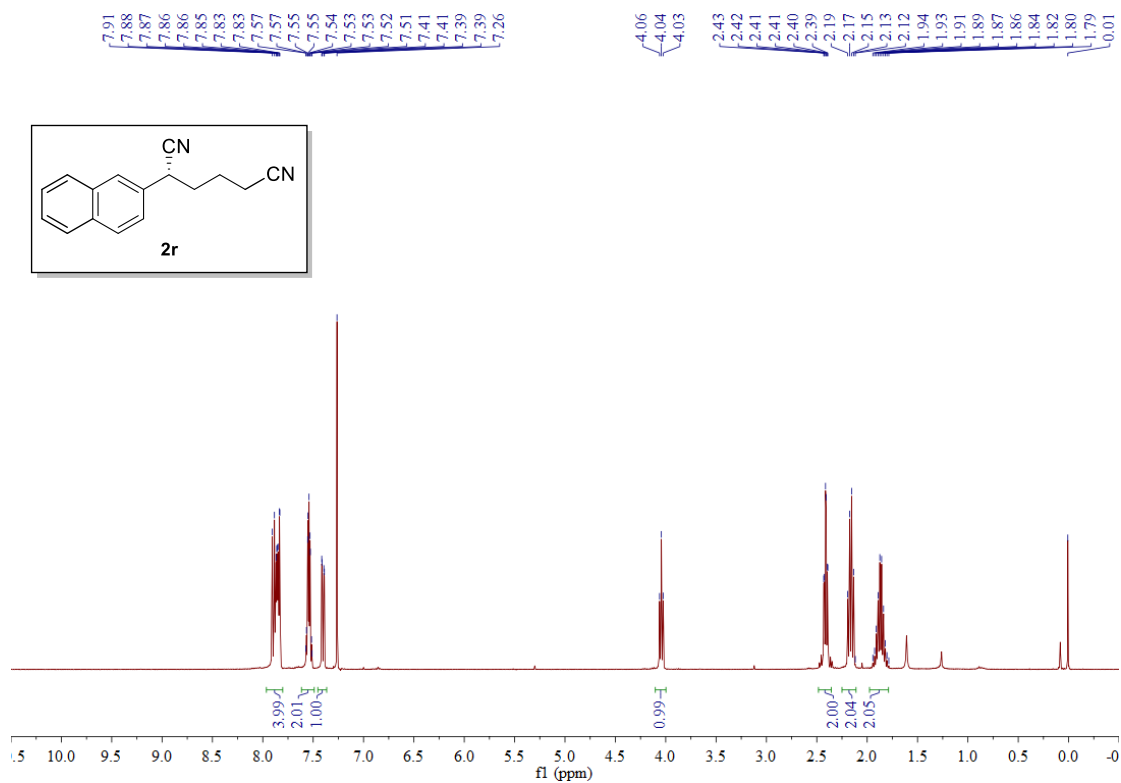
Supplementary Figure 92. ¹³C NMR of **2p**



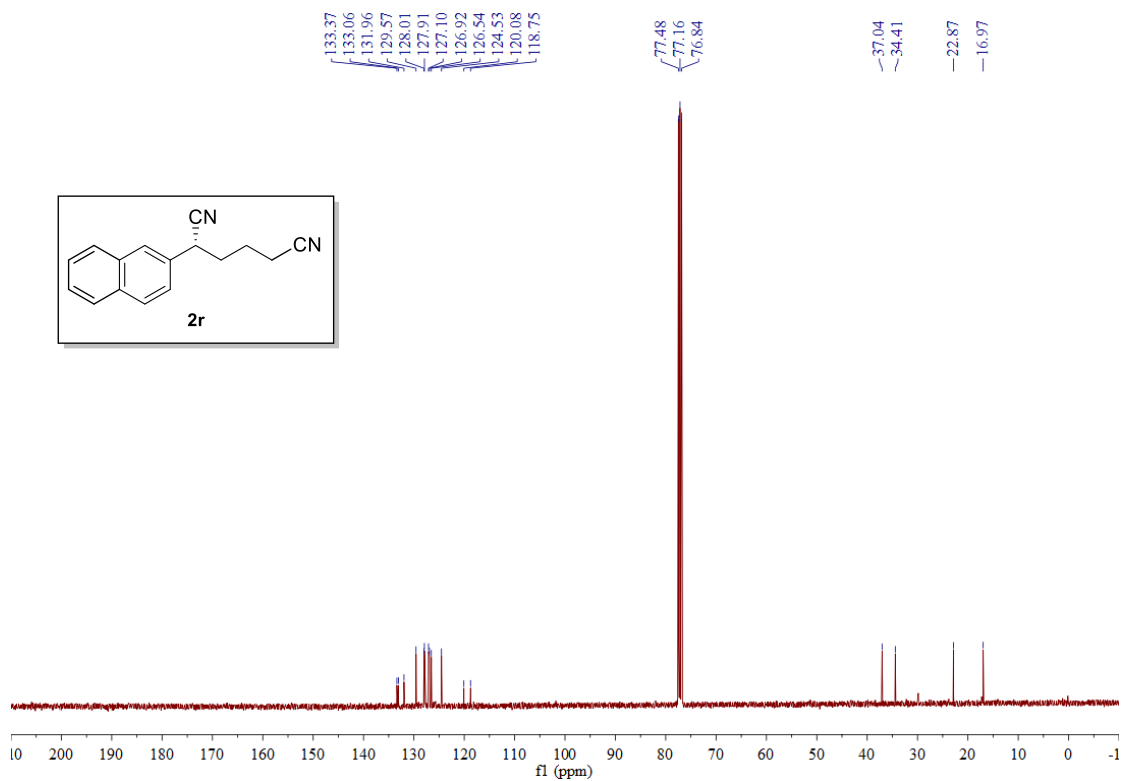
Supplementary Figure 93. ¹H NMR of **2q**



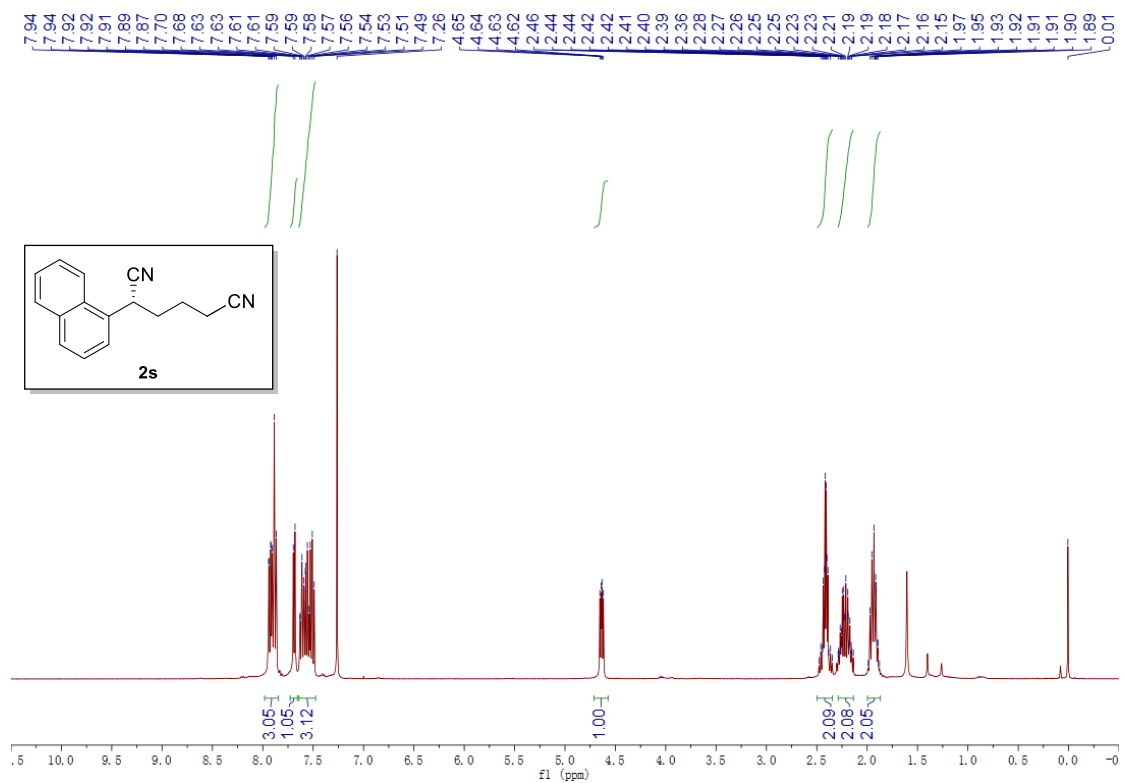
Supplementary Figure 94. ¹³C NMR of **2q**



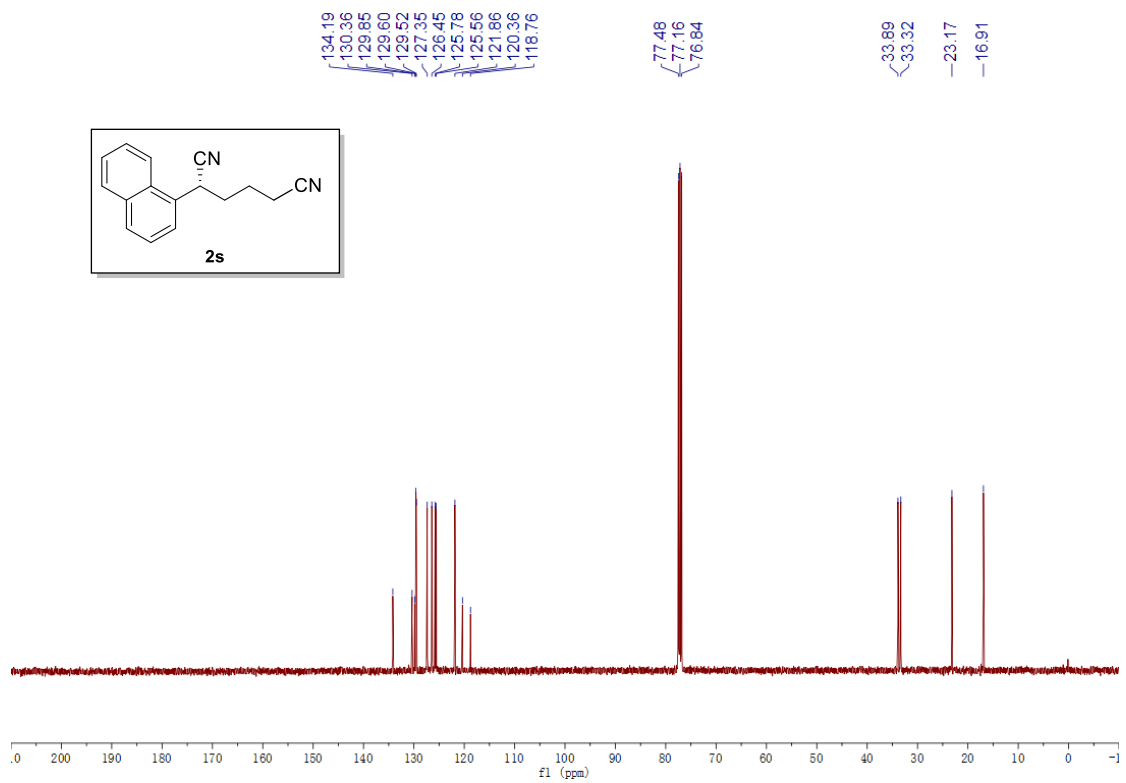
Supplementary Figure 95. ¹H NMR of **2r**



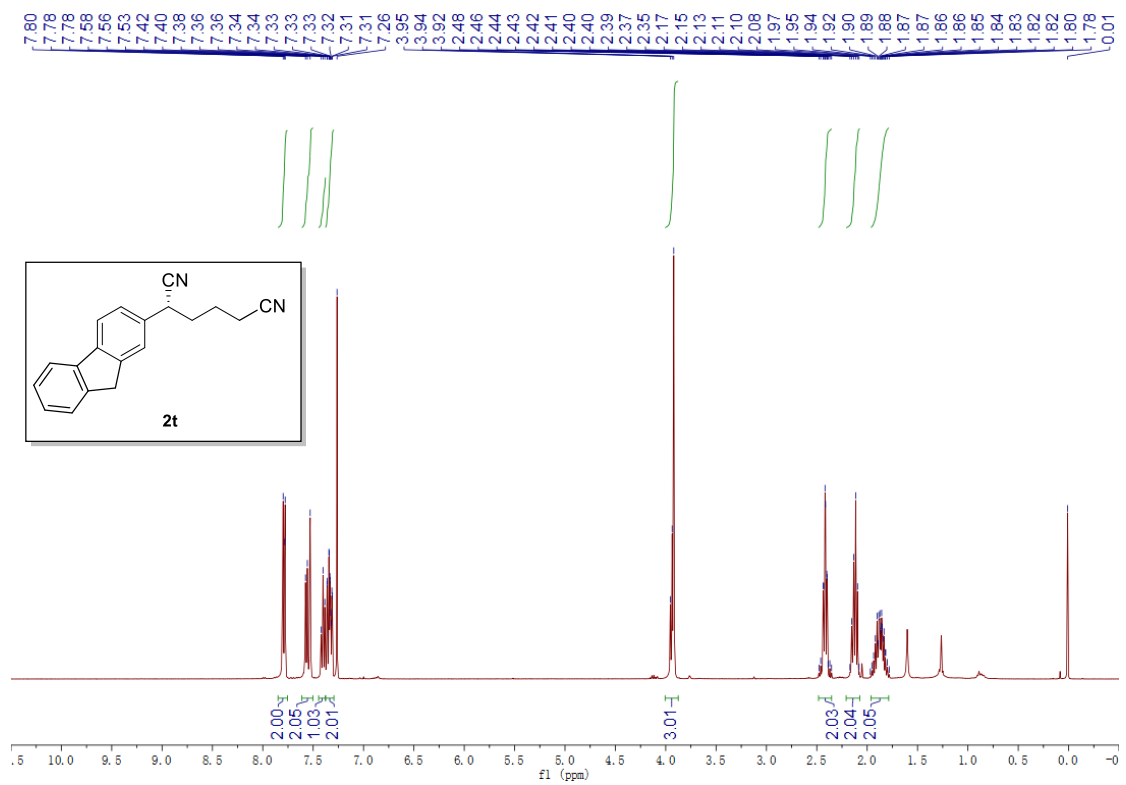
Supplementary Figure 96. ¹³C NMR of **2r**



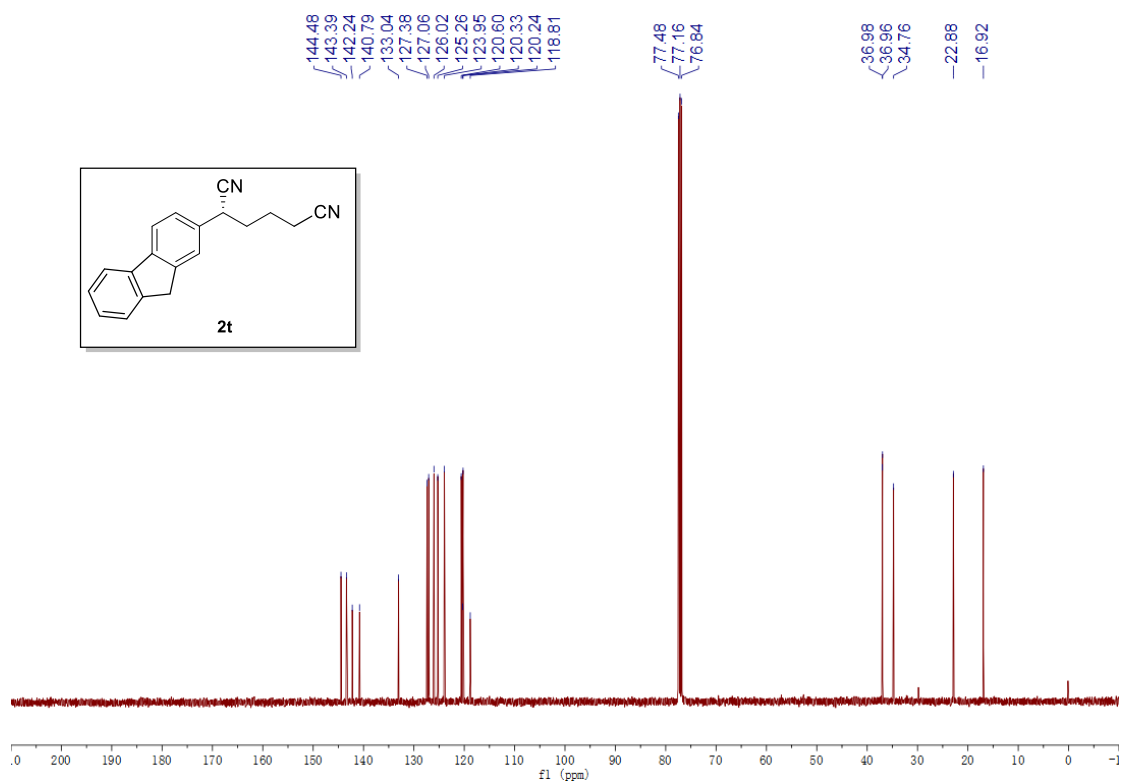
Supplementary Figure 97. ¹H NMR of **2s**



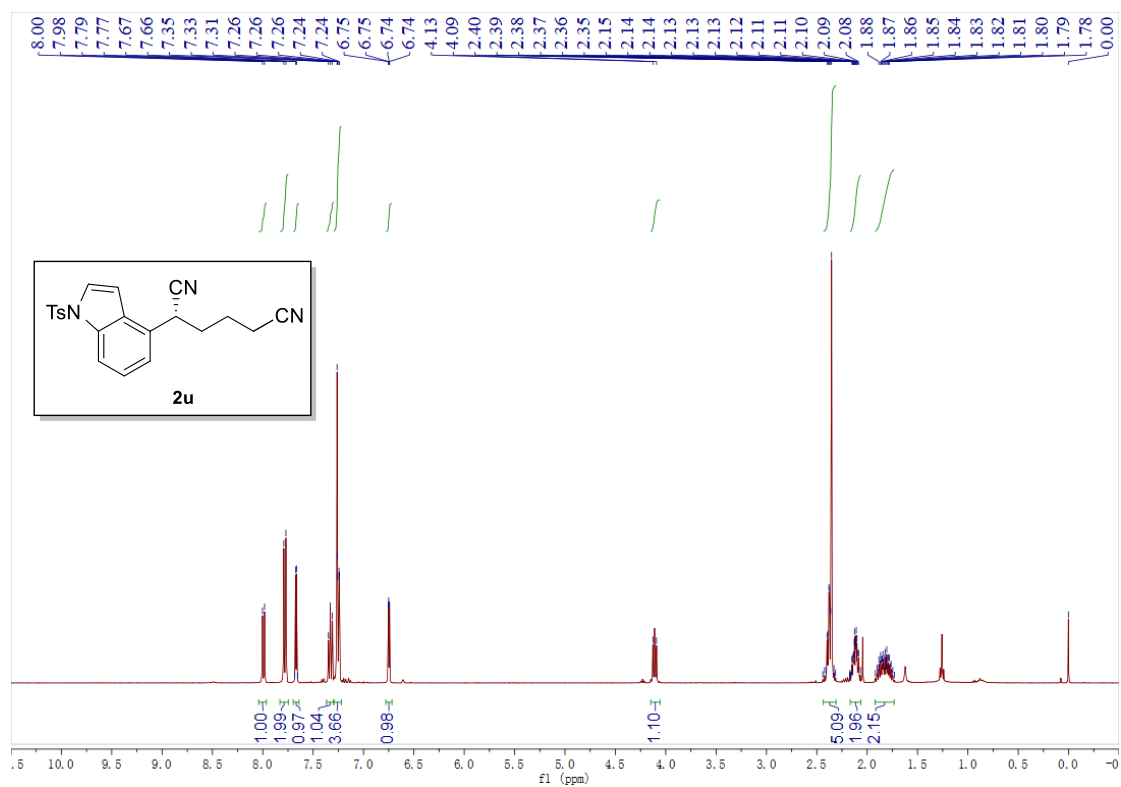
Supplementary Figure 98. ¹³C NMR of **2s**



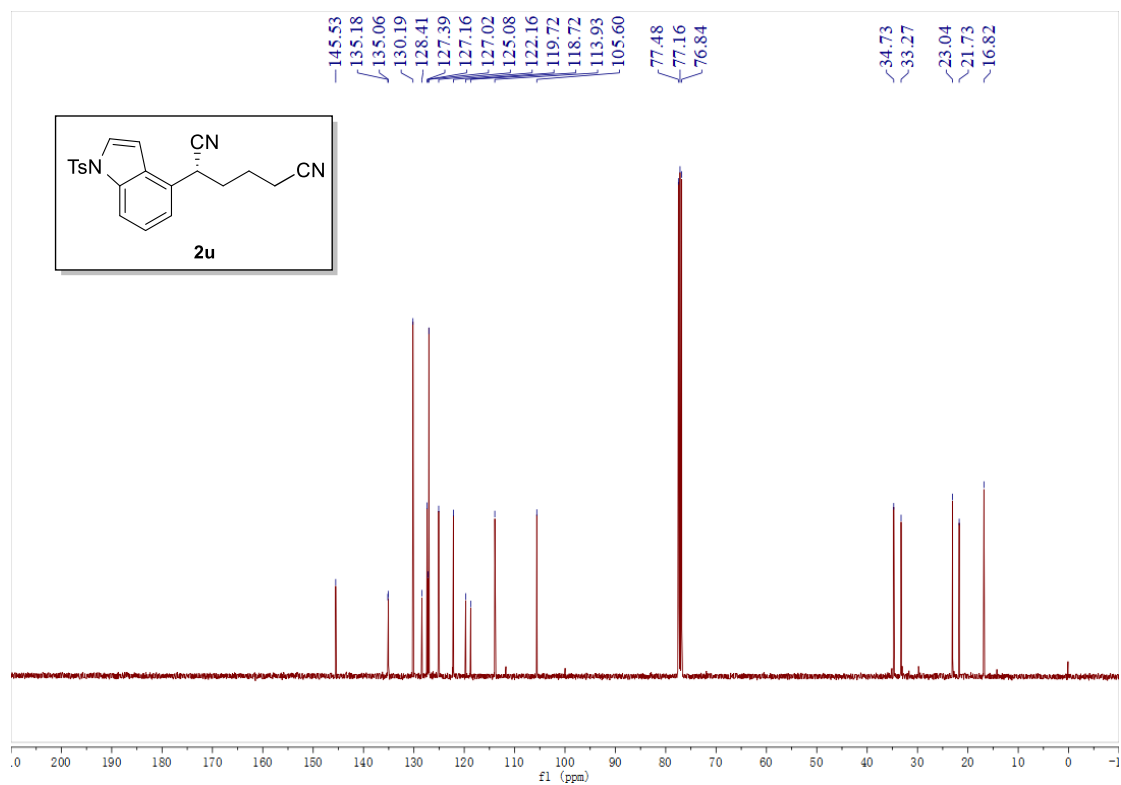
Supplementary Figure 99. ¹H NMR of **2t**



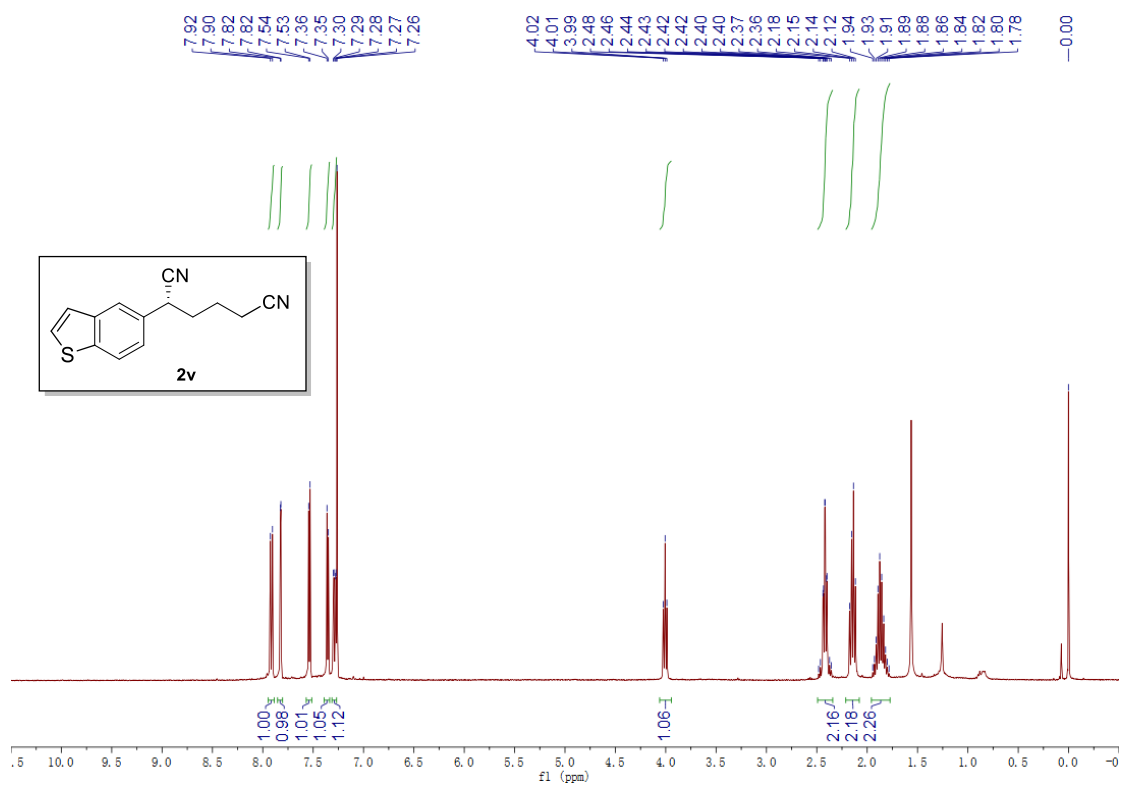
Supplementary Figure 100. ¹³C NMR of **2t**



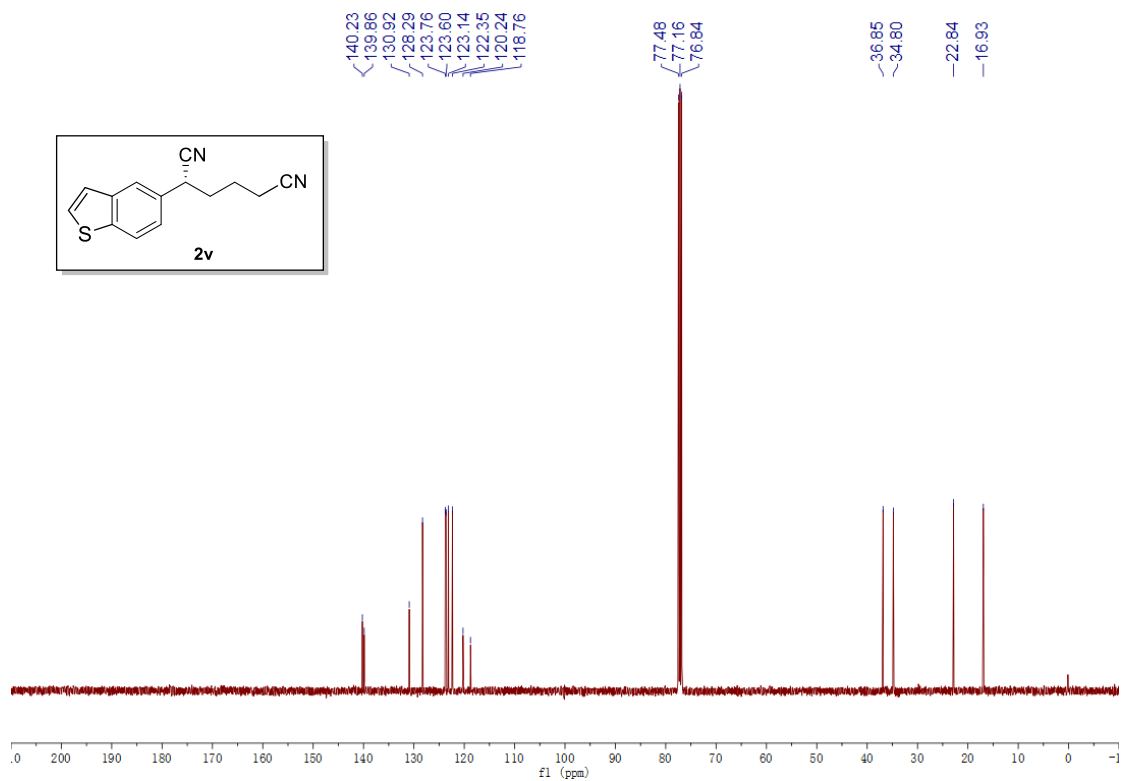
Supplementary Figure 101. ¹H NMR of **2u**



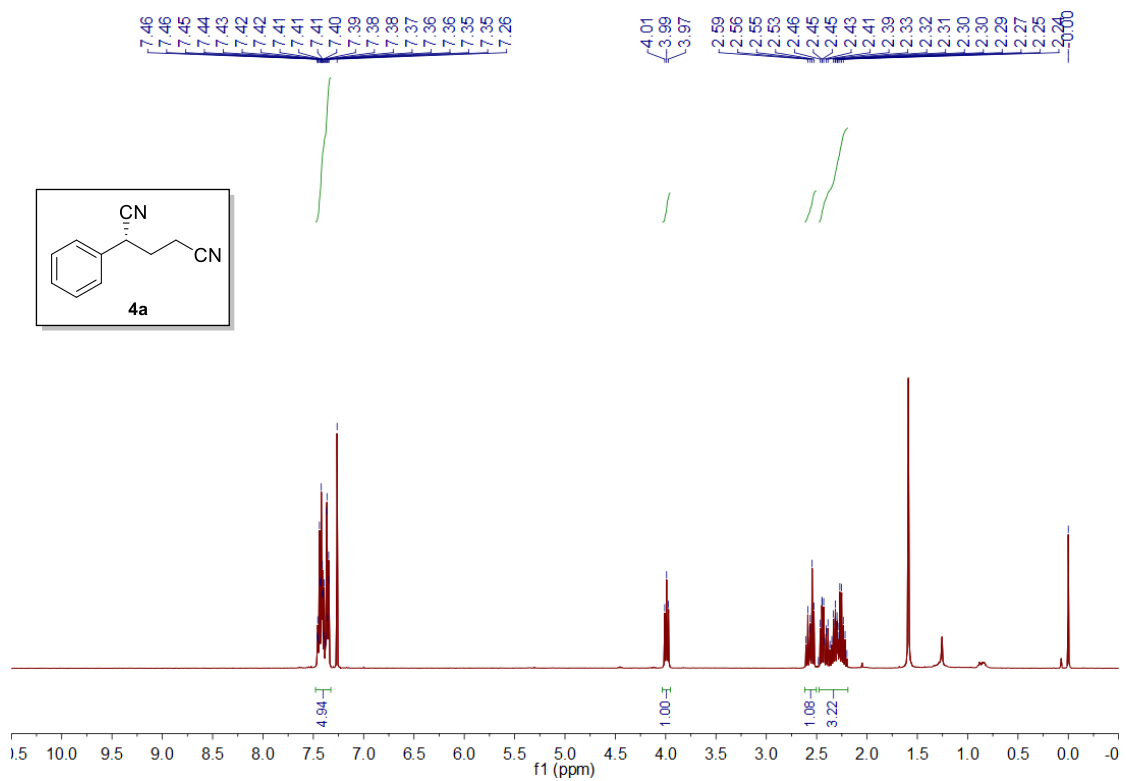
Supplementary Figure 102. ¹³C NMR of **2u**



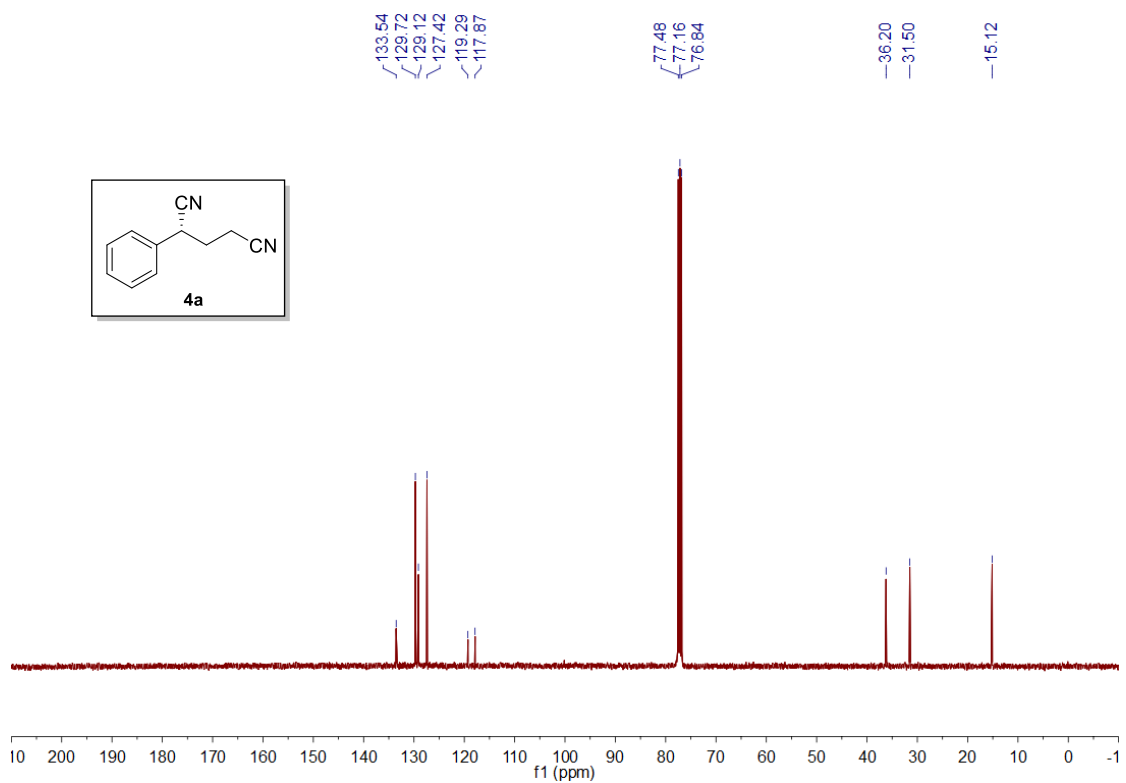
Supplementary Figure 103. ¹H NMR of **2v**



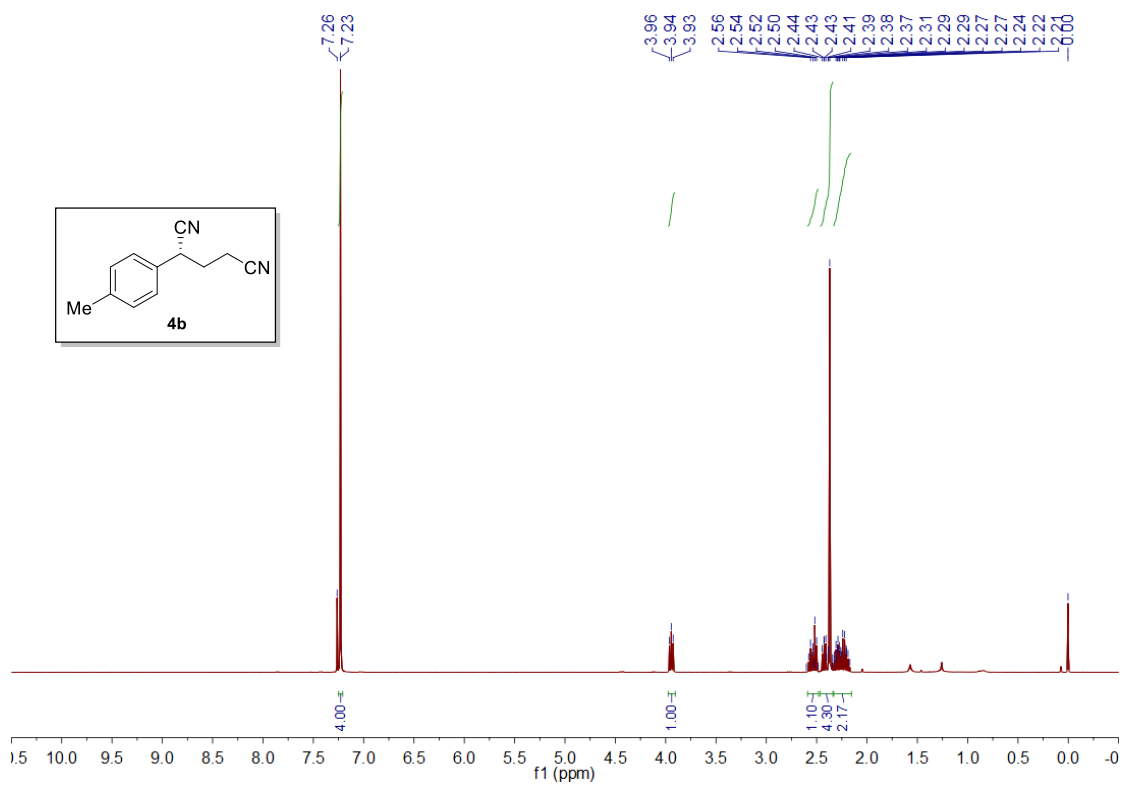
Supplementary Figure 104. ^{13}C NMR of **2v**



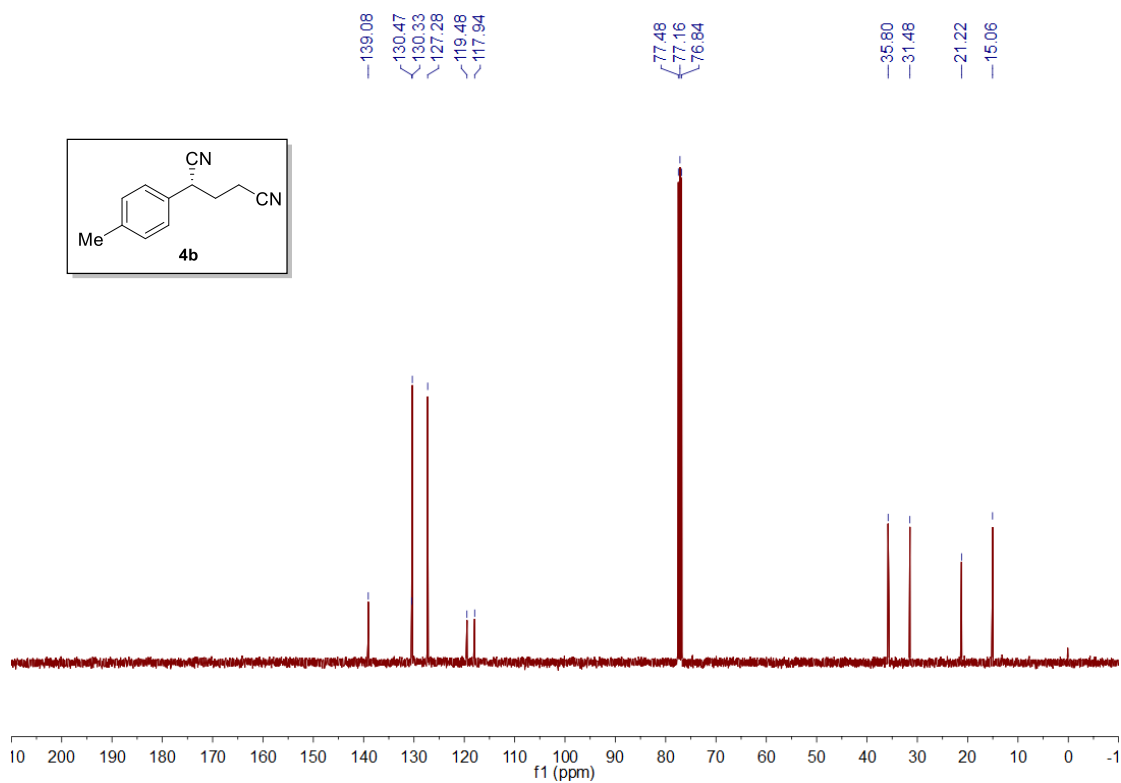
Supplementary Figure 105. ^1H NMR of **4a**



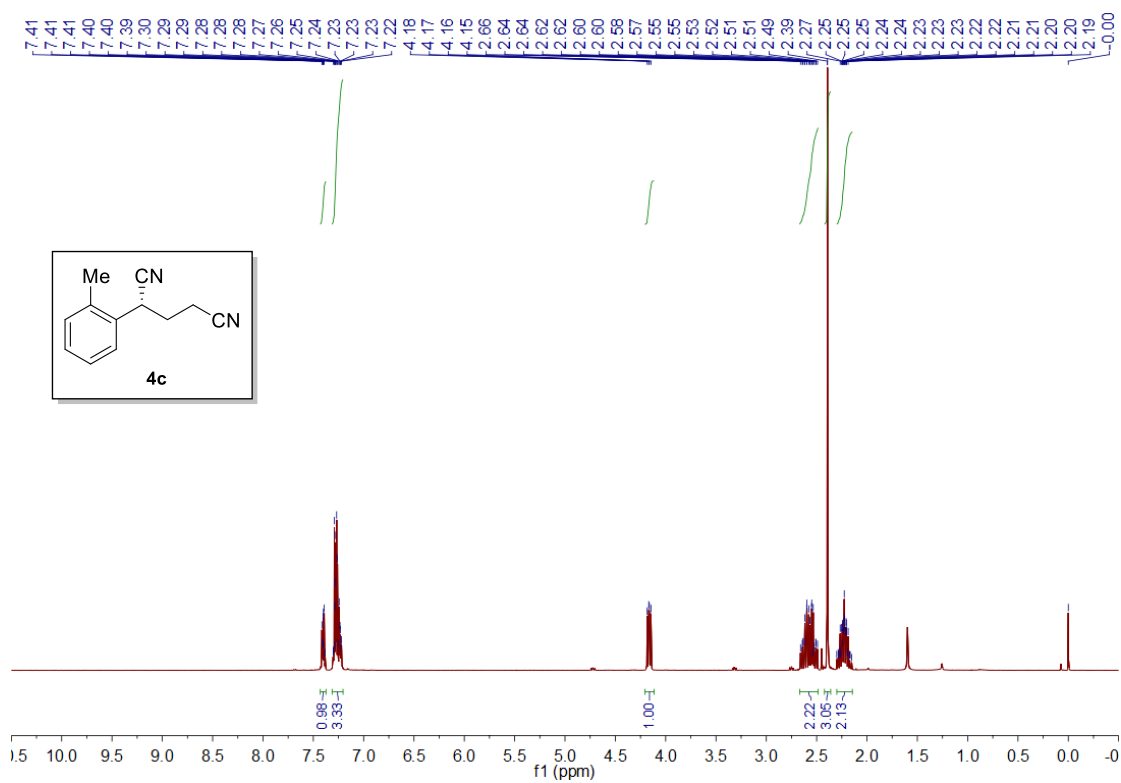
Supplementary Figure 106. ^{13}C NMR of **4a**



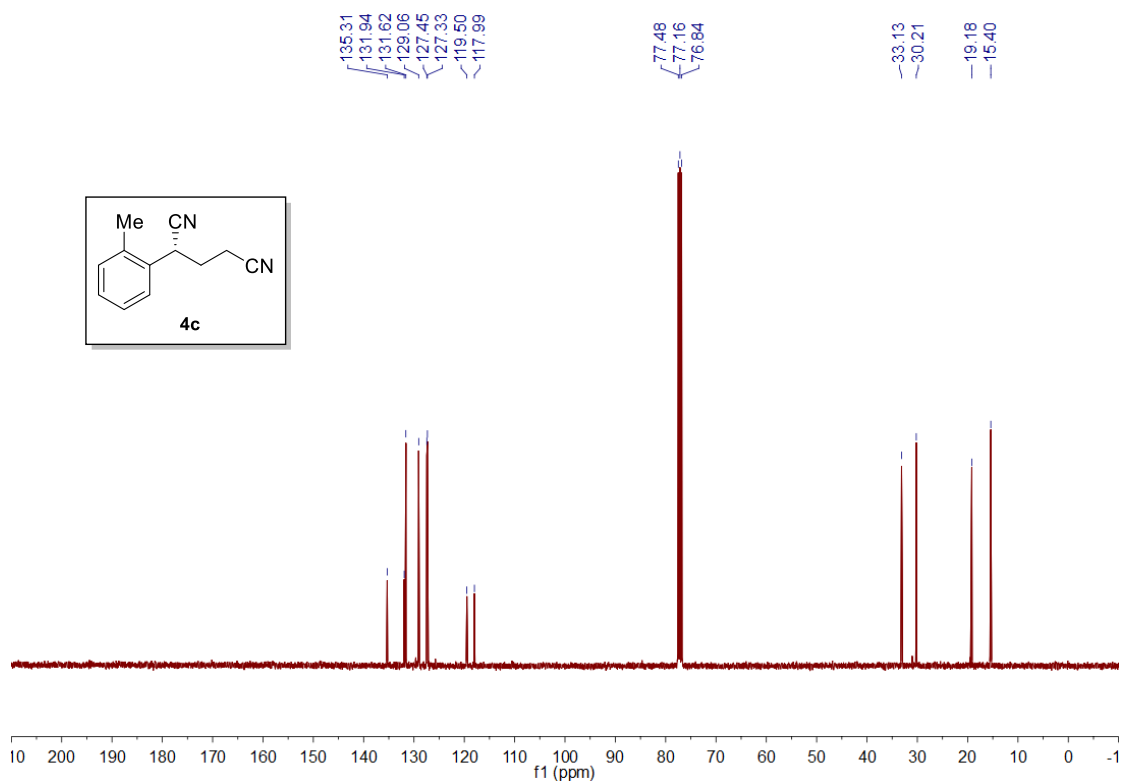
Supplementary Figure 107. ^1H NMR of **4b**



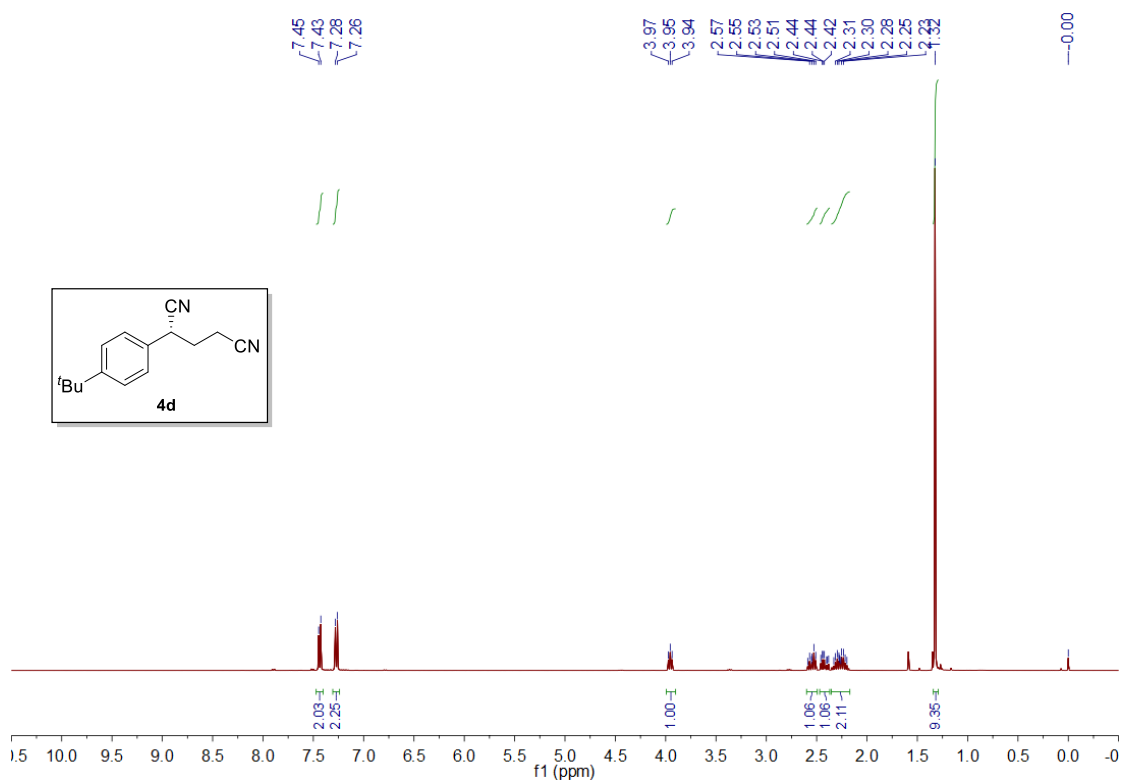
Supplementary Figure 108. ^{13}C NMR of **4b**



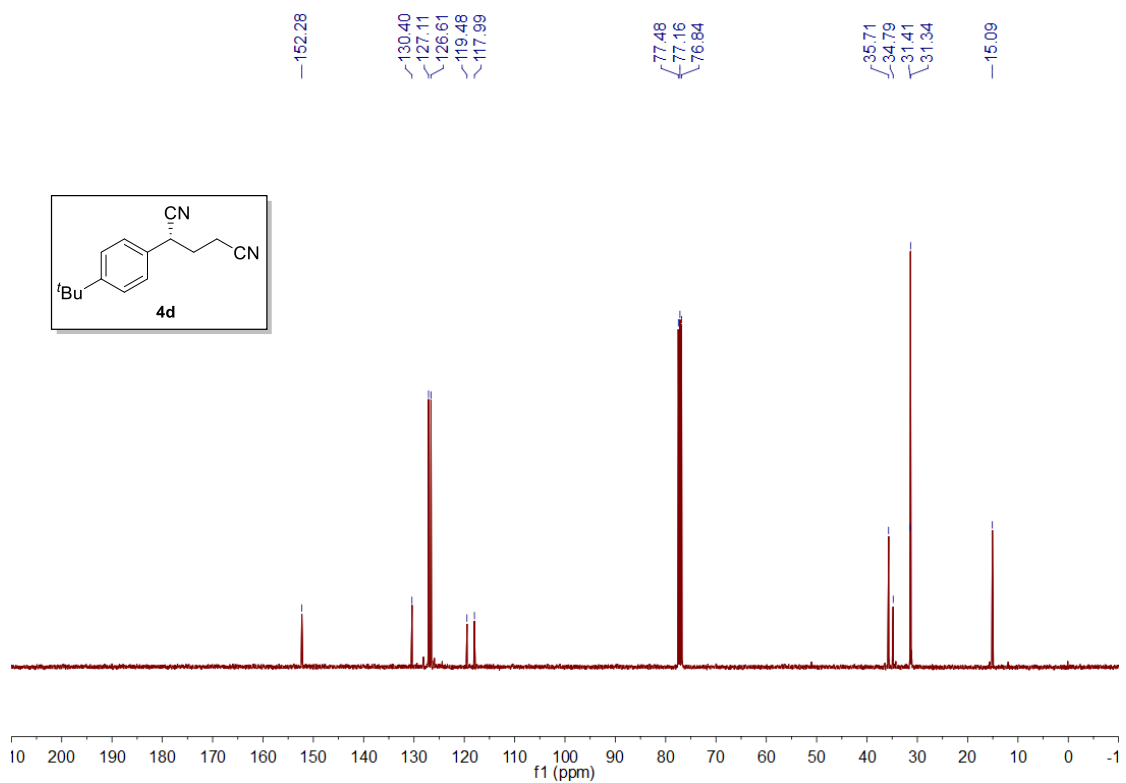
Supplementary Figure 109. ^1H NMR of **4c**



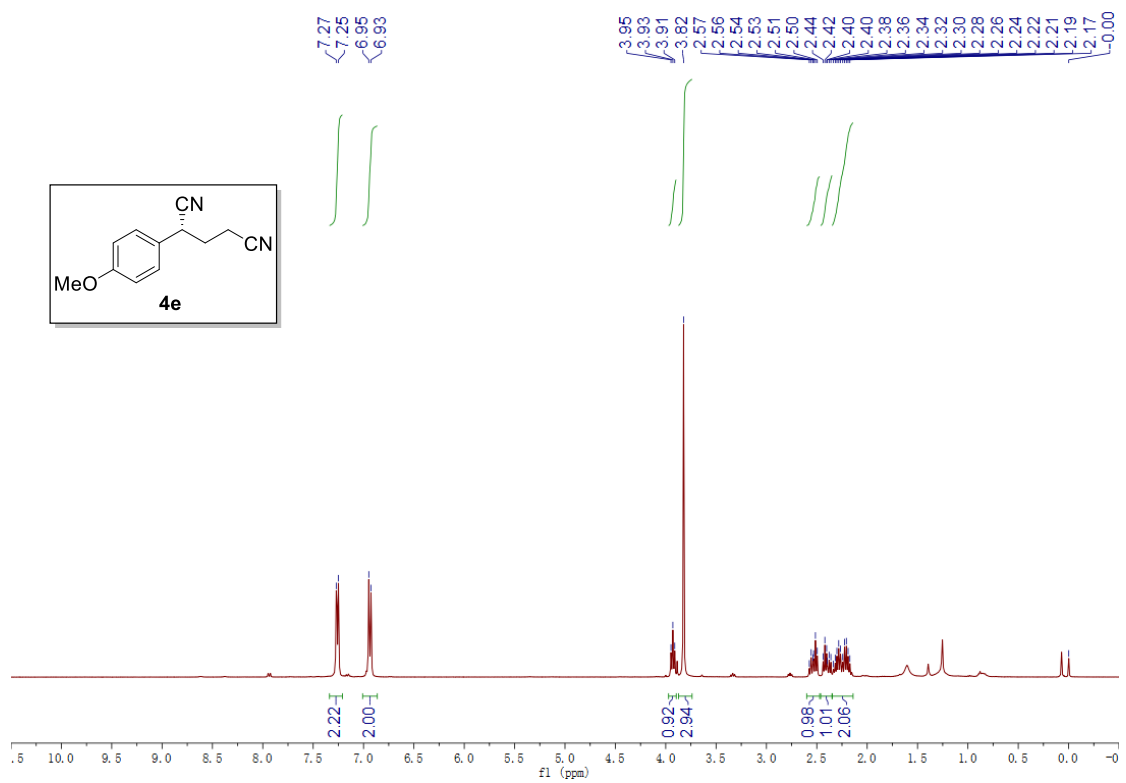
Supplementary Figure 110. ¹³C NMR of **4c**



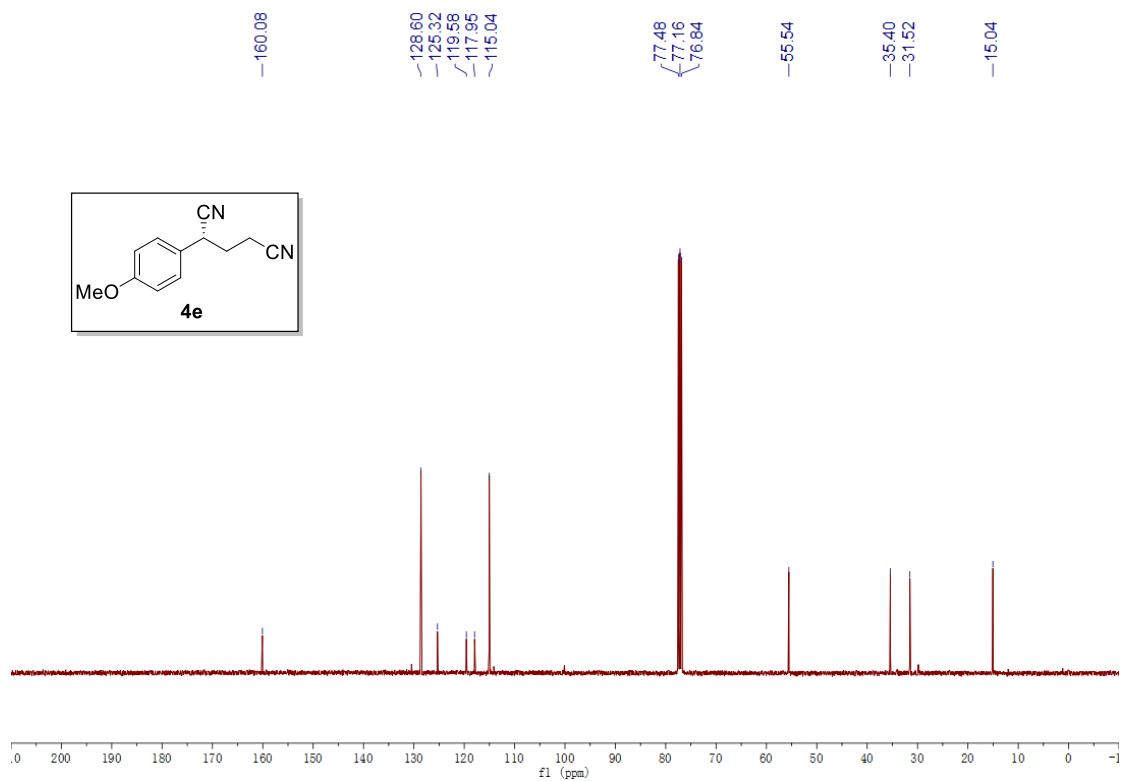
Supplementary Figure 111. ¹H NMR of **4d**



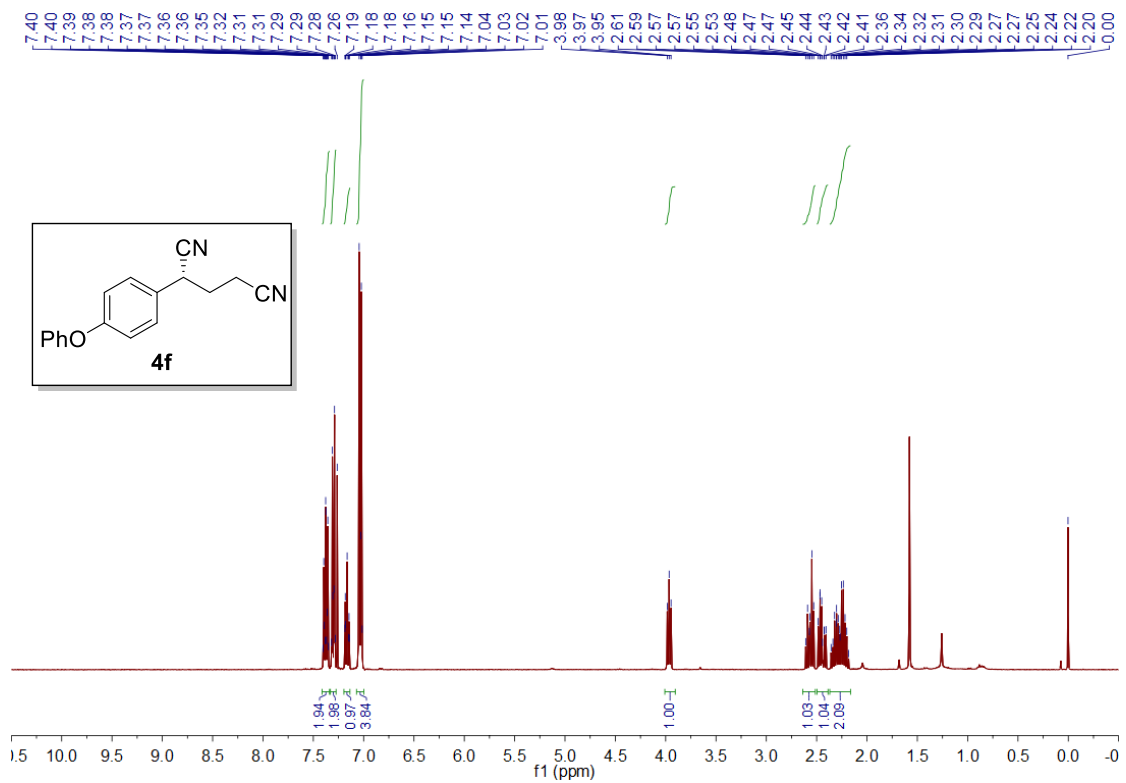
Supplementary Figure 112. ^{13}C NMR of **4d**



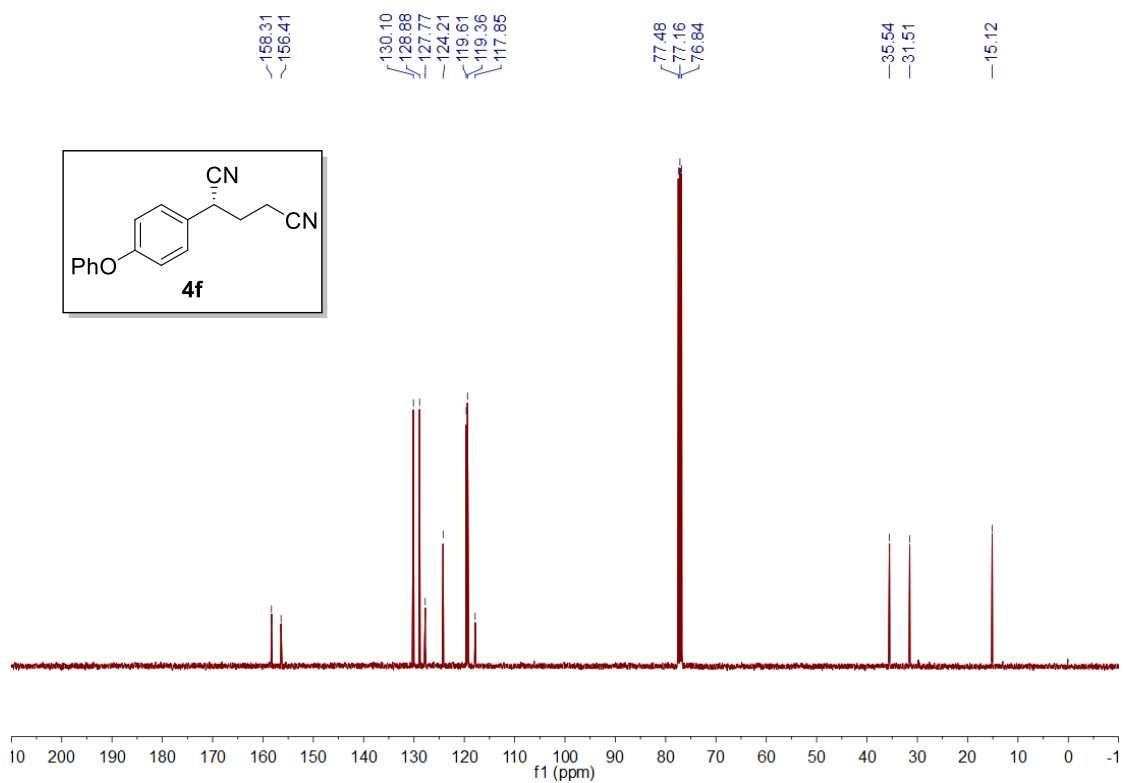
Supplementary Figure 113. ^1H NMR of **4e**



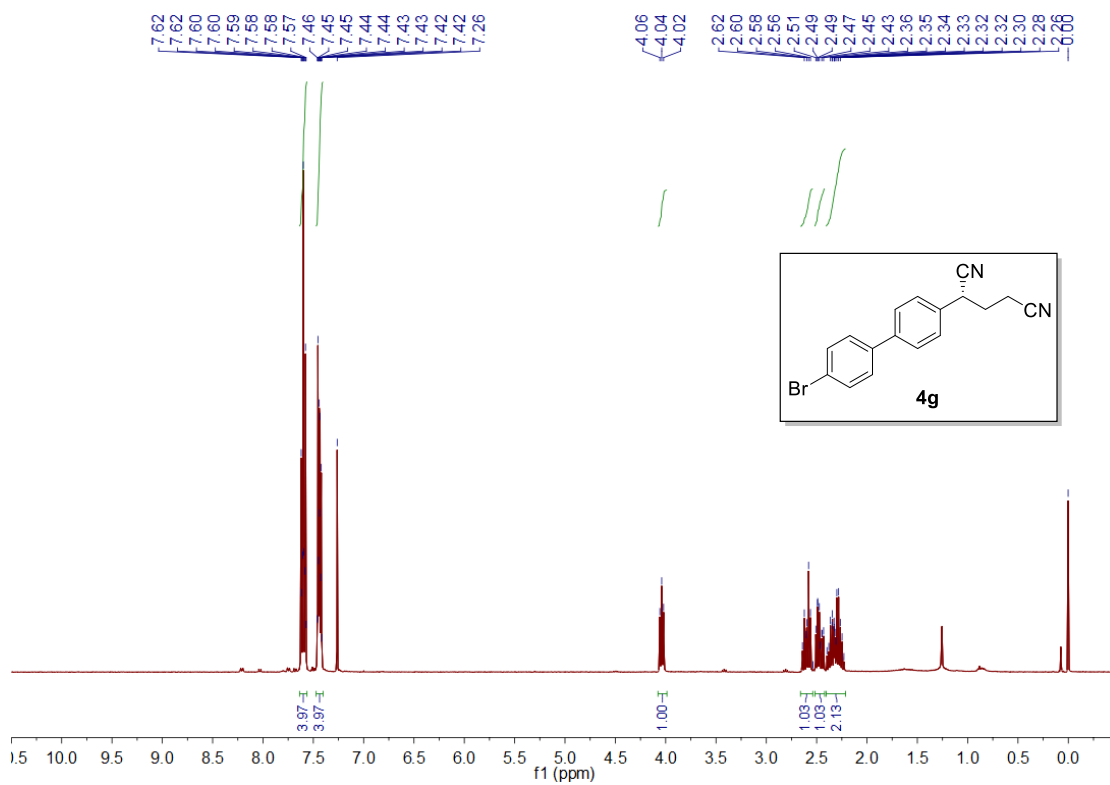
Supplementary Figure 114. ¹³C NMR of **4e**



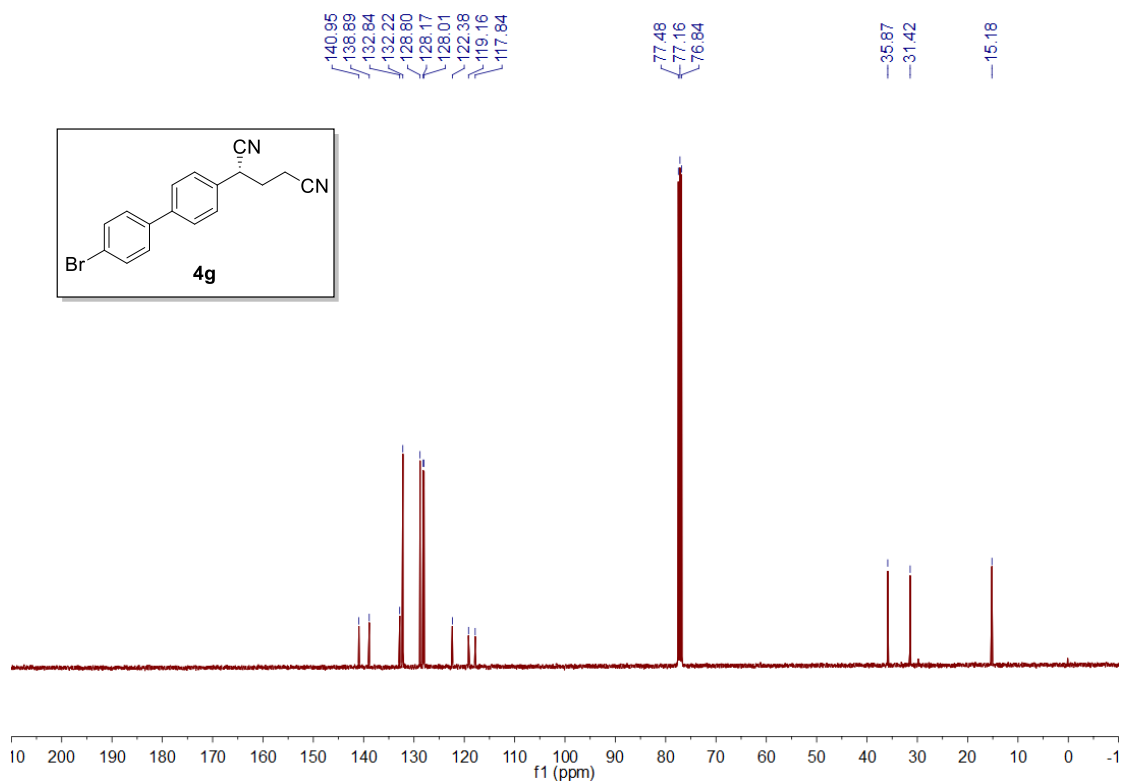
Supplementary Figure 115. ¹H NMR of **4f**



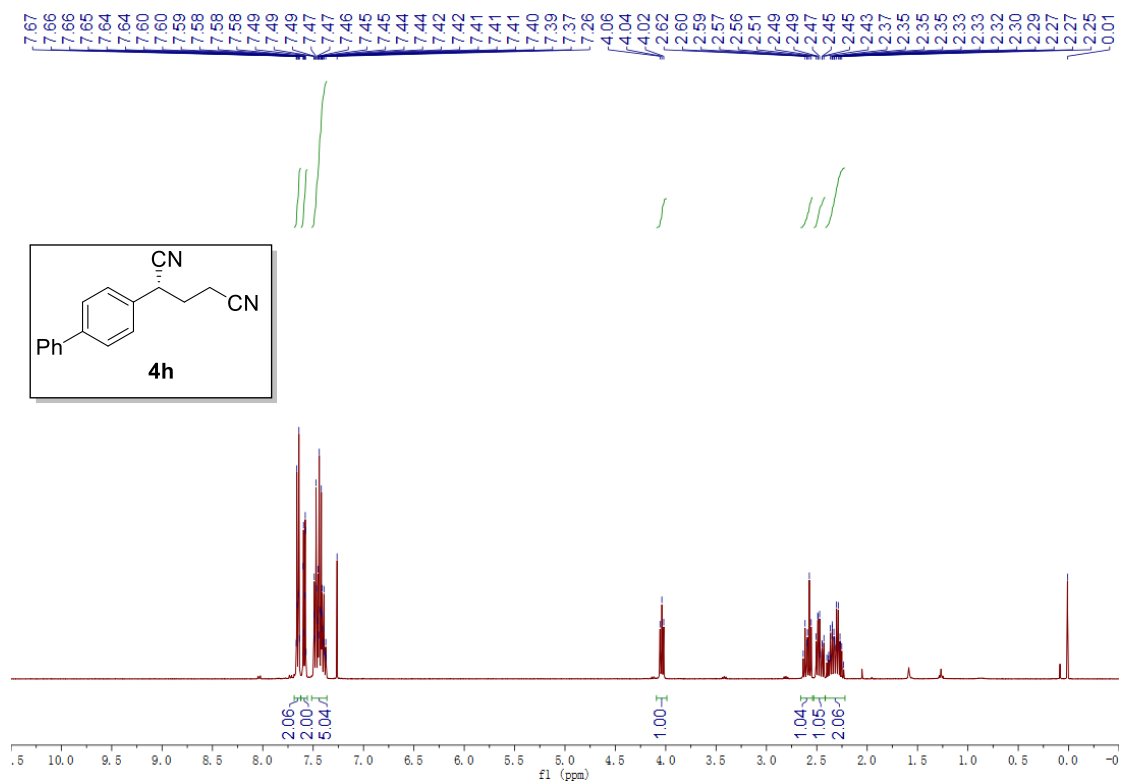
Supplementary Figure 116. ¹³C NMR of **4f**



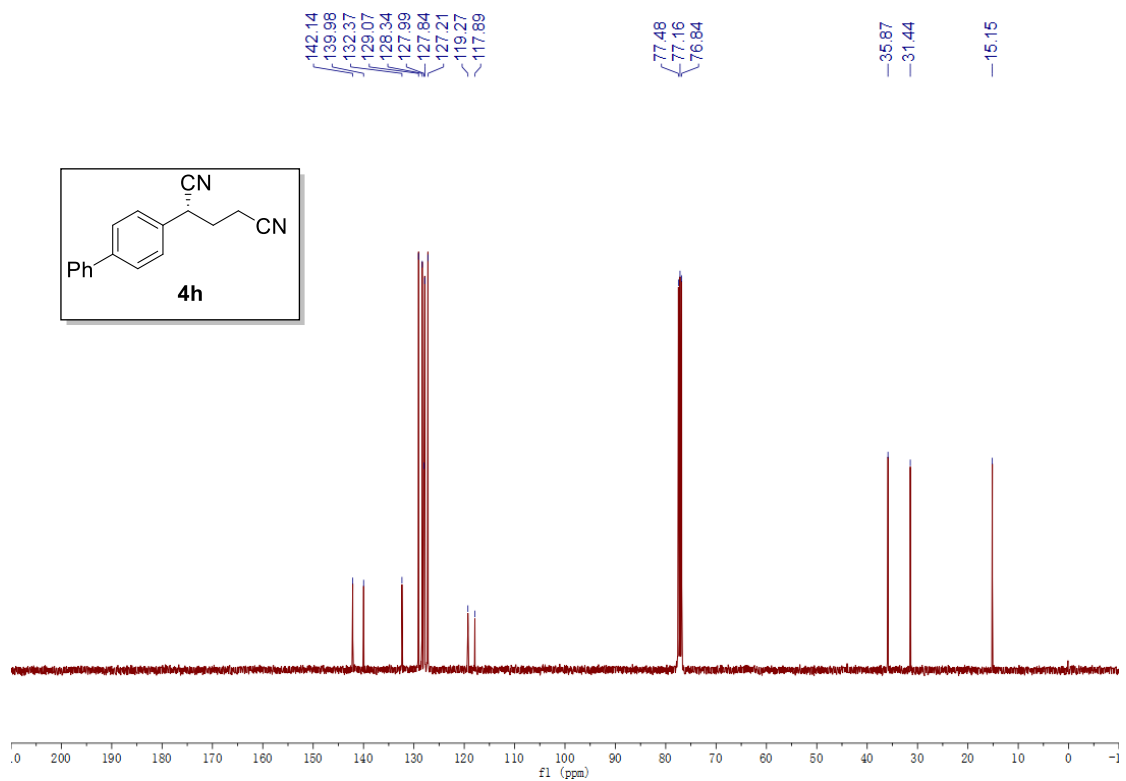
Supplementary Figure 117. ¹H NMR of **4g**



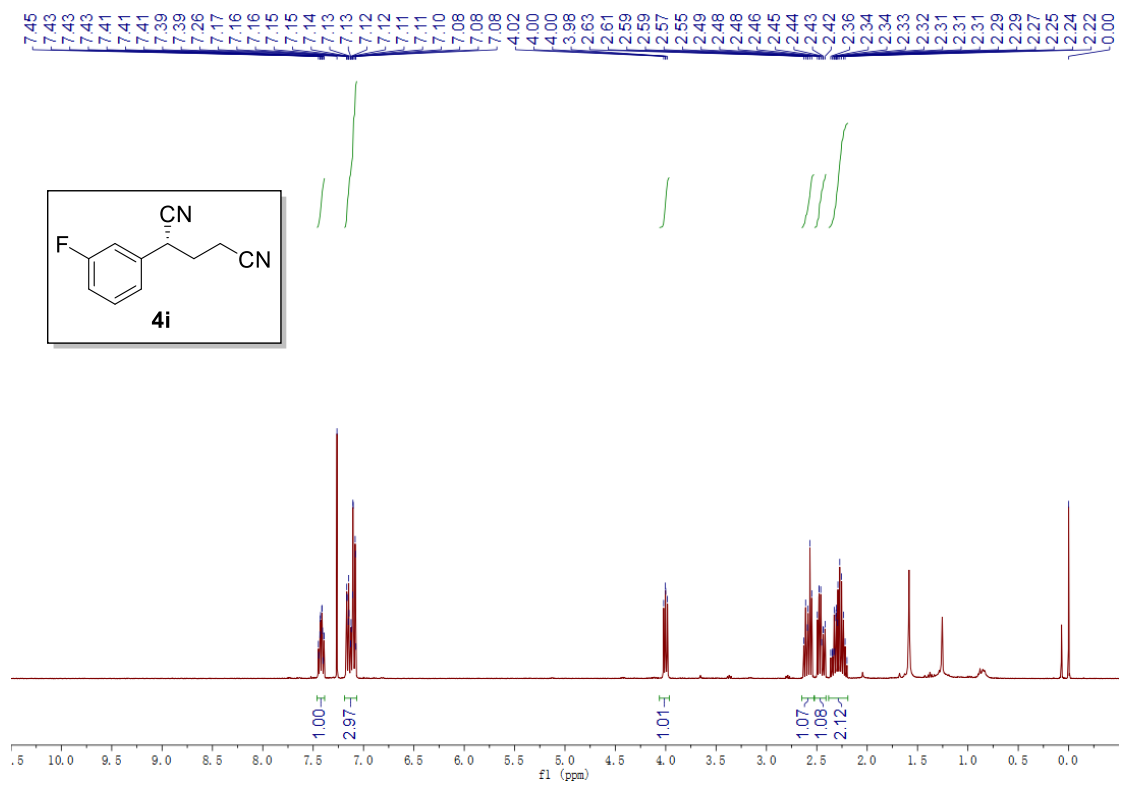
Supplementary Figure 118. ^{13}C NMR of **4g**



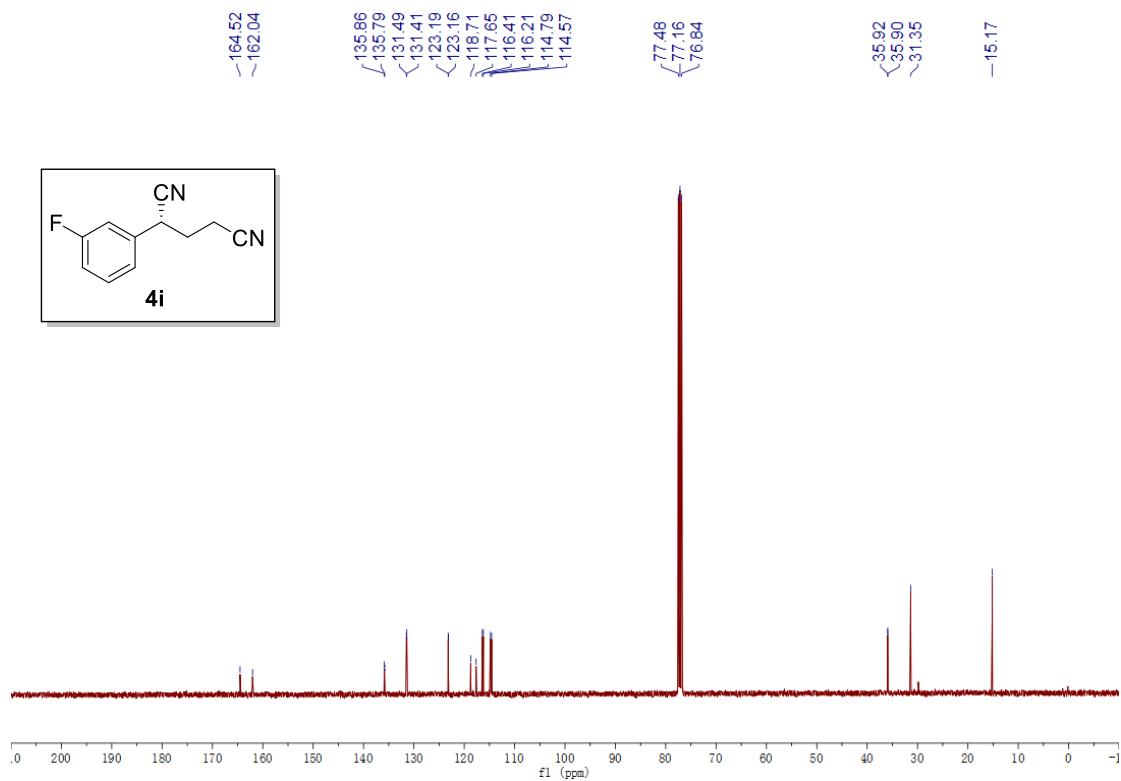
Supplementary Figure 119. ^1H NMR of **4h**



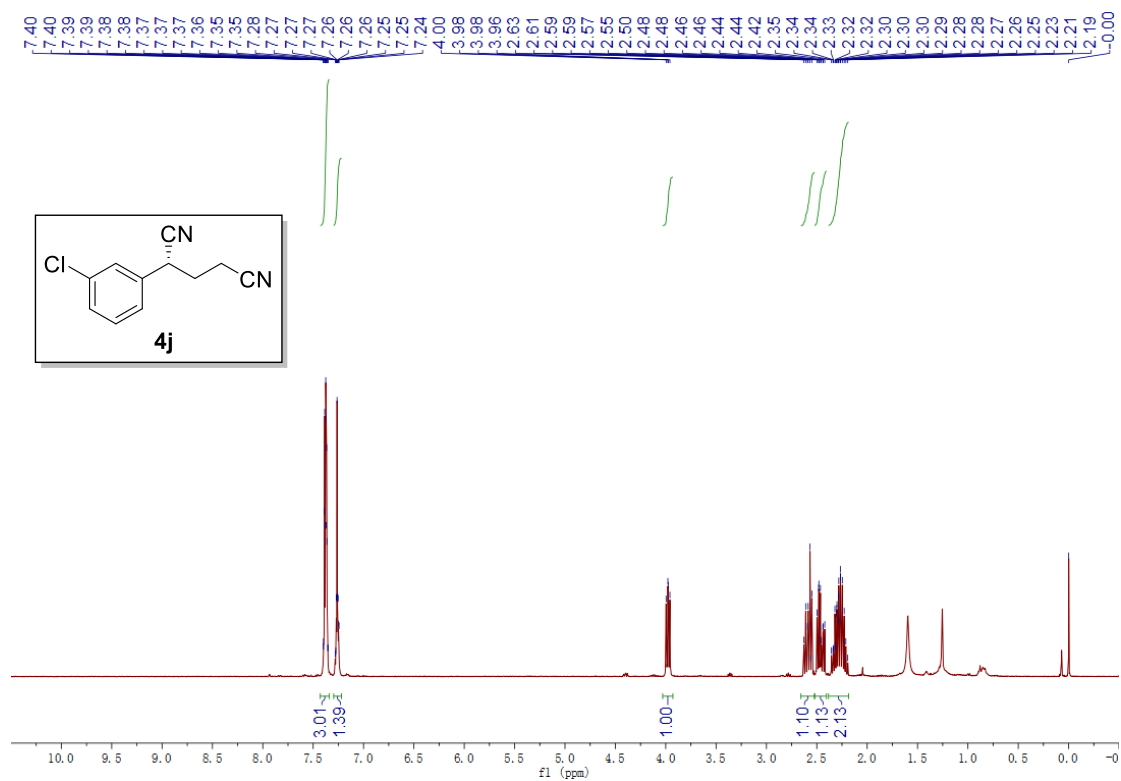
Supplementary Figure 120. ¹³C NMR of **4h**



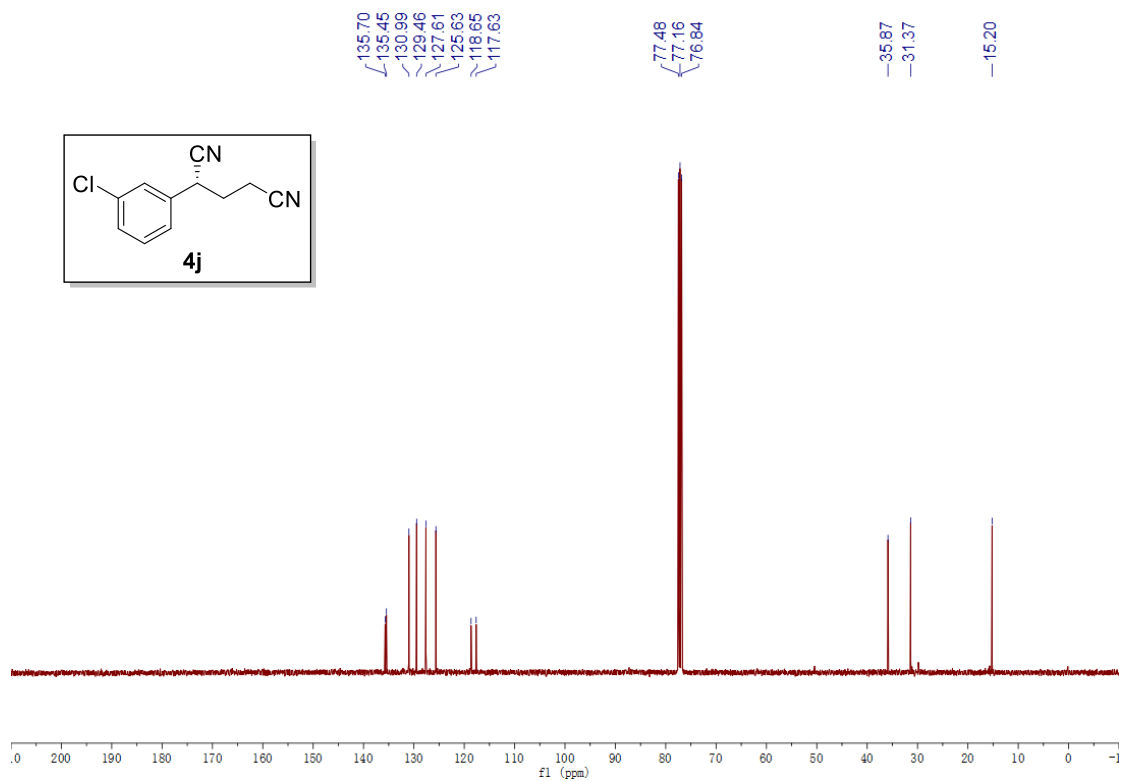
Supplementary Figure 121. ¹H NMR of **4i**



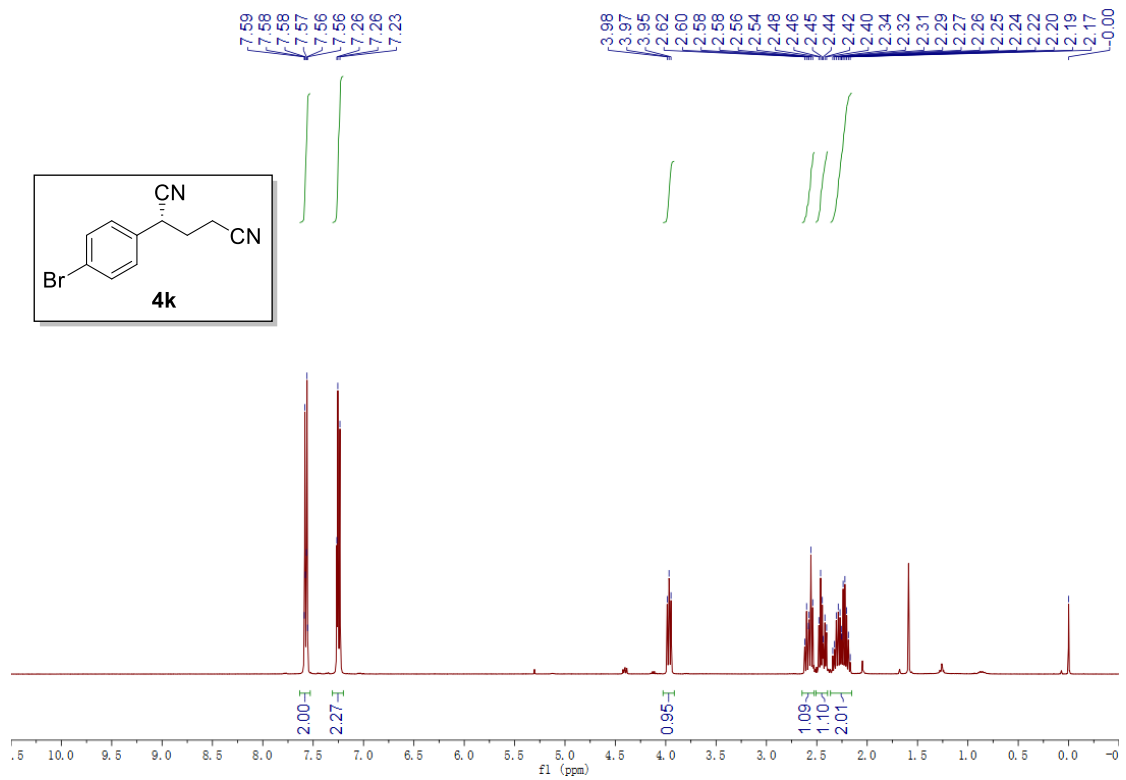
Supplementary Figure 122. ^{13}C NMR of **4i**



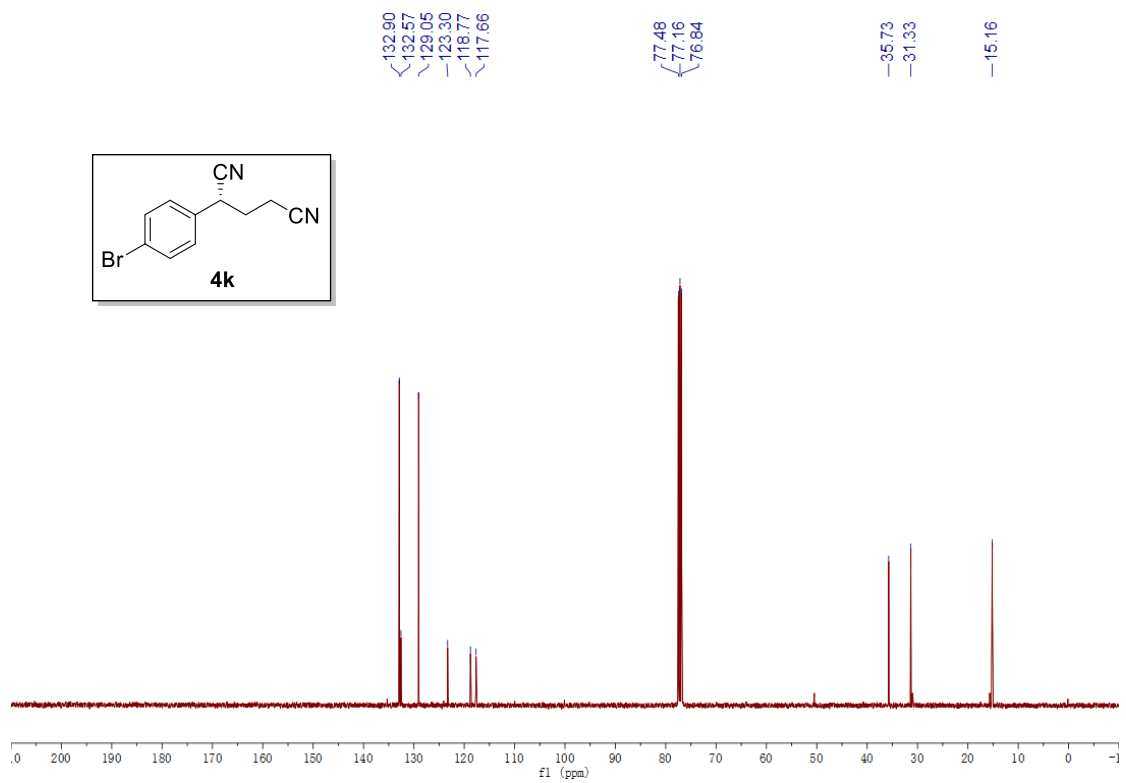
Supplementary Figure 123. ^1H NMR of **4j**



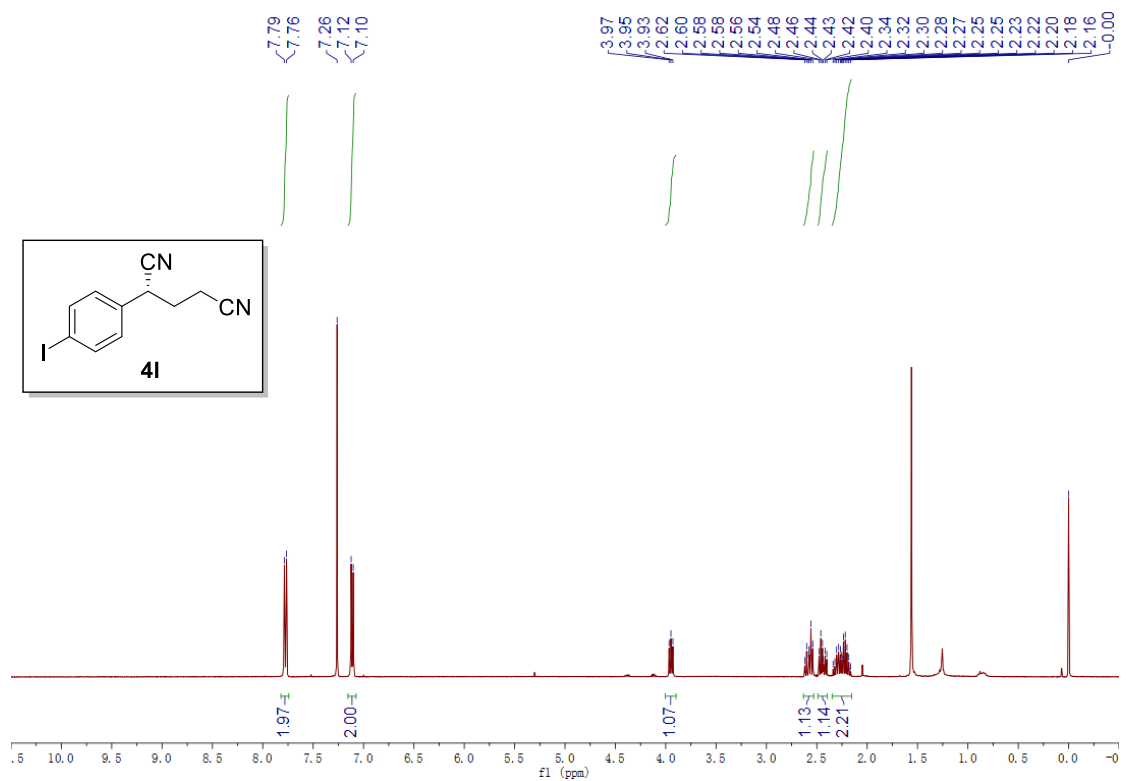
Supplementary Figure 124. ^{13}C NMR of **4j**



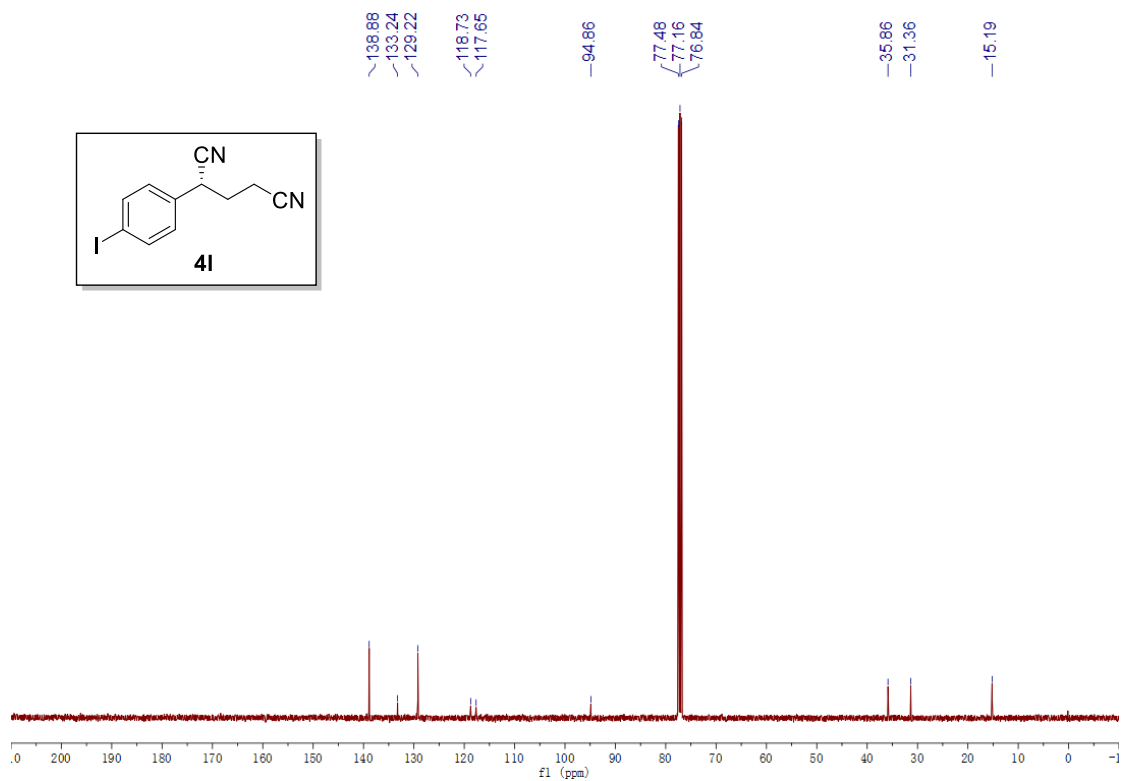
Supplementary Figure 125. ^1H NMR of **4k**



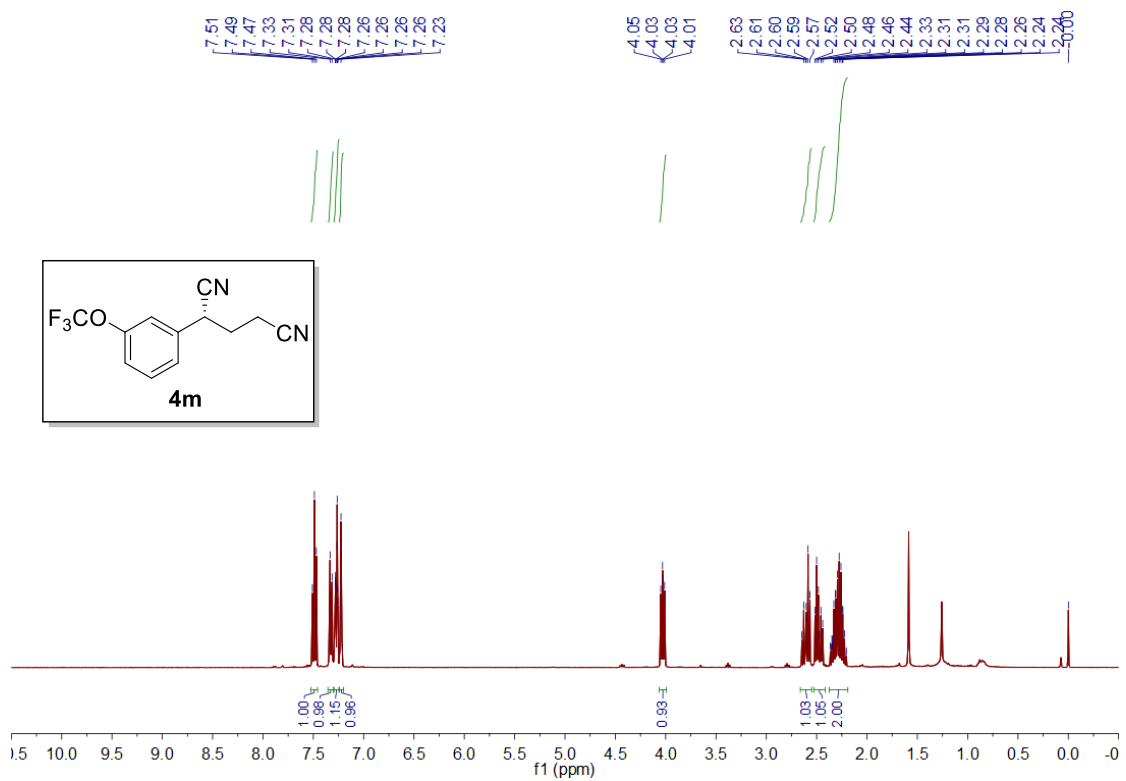
Supplementary Figure 126. ¹³C NMR of **4k**



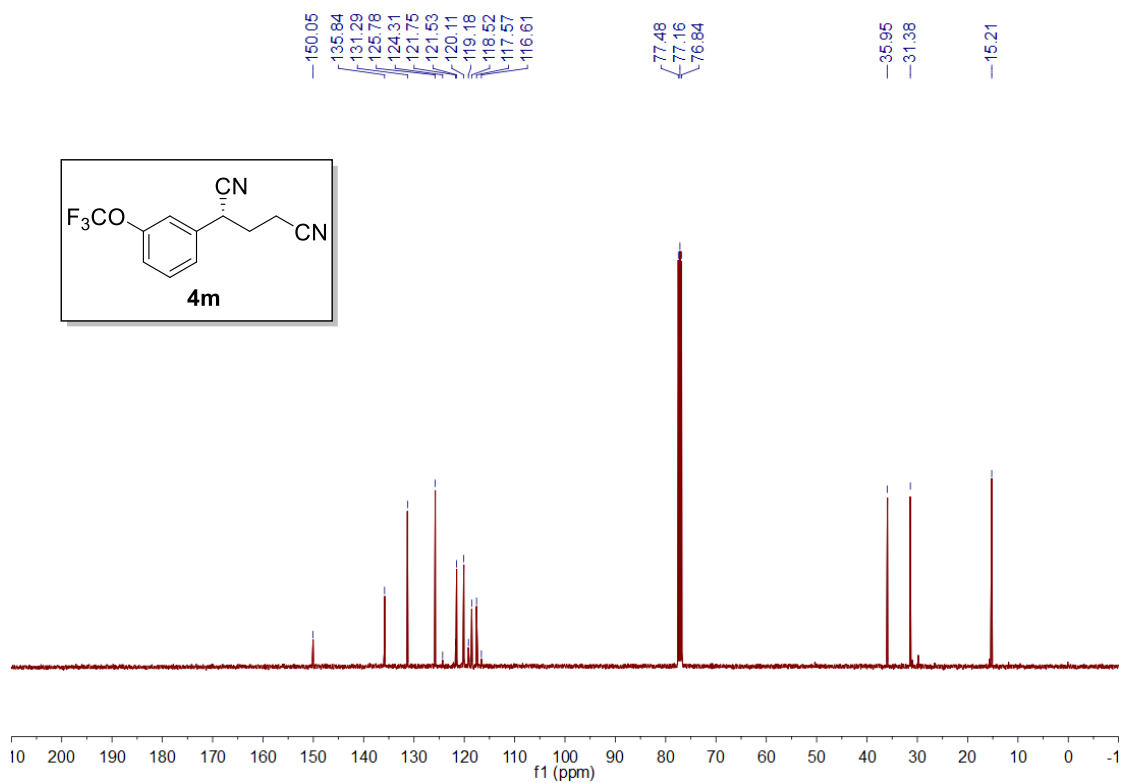
Supplementary Figure 127. ¹H NMR of **4l**



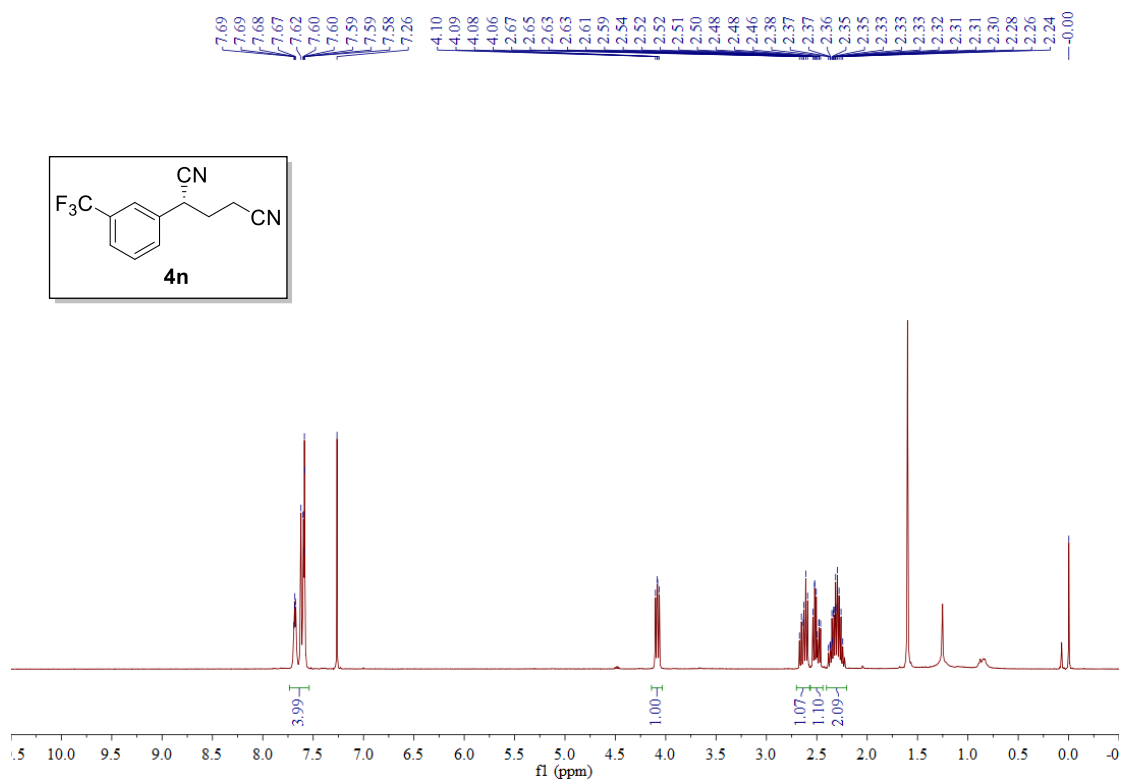
Supplementary Figure 128. ^{13}C NMR of **4l**



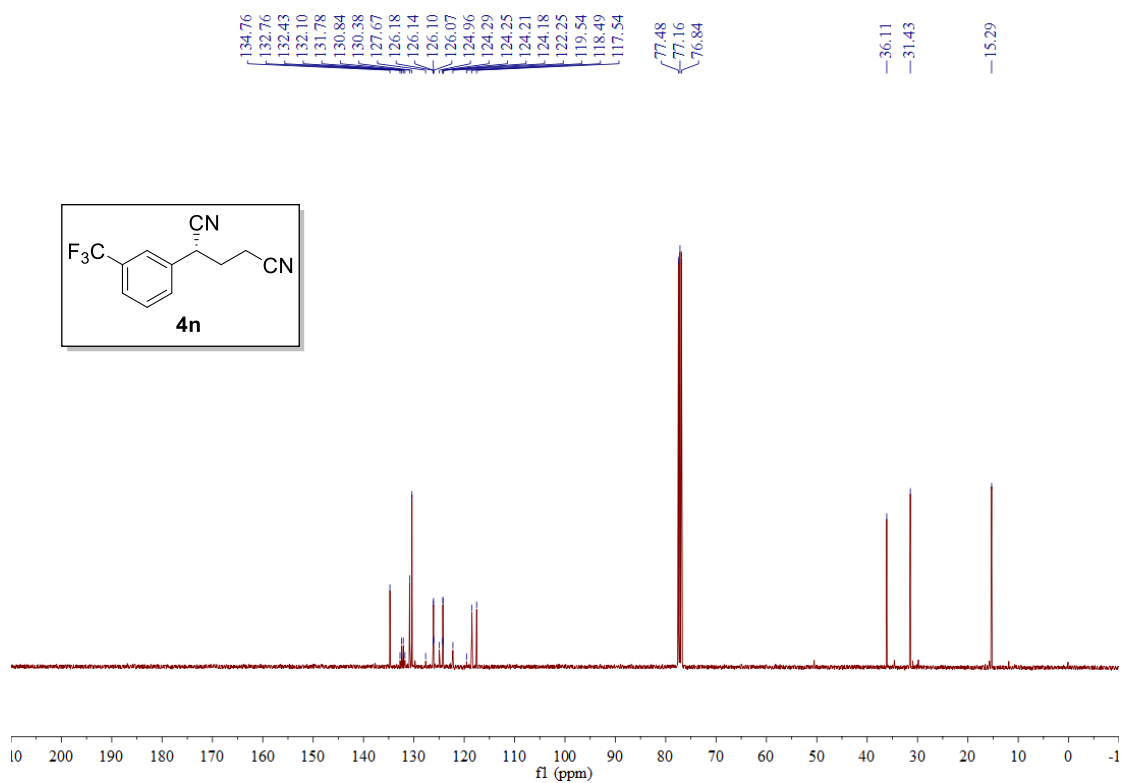
Supplementary Figure 129. ^1H NMR of **4m**



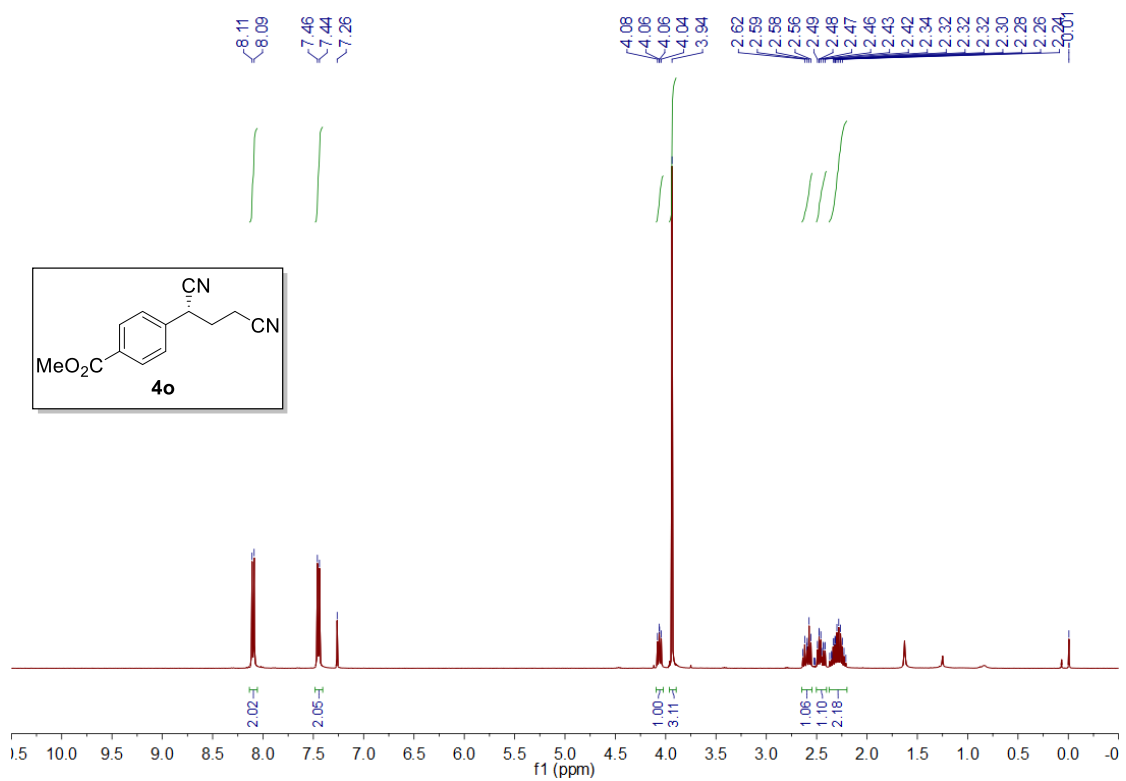
Supplementary Figure 130. ¹³C NMR of **4m**



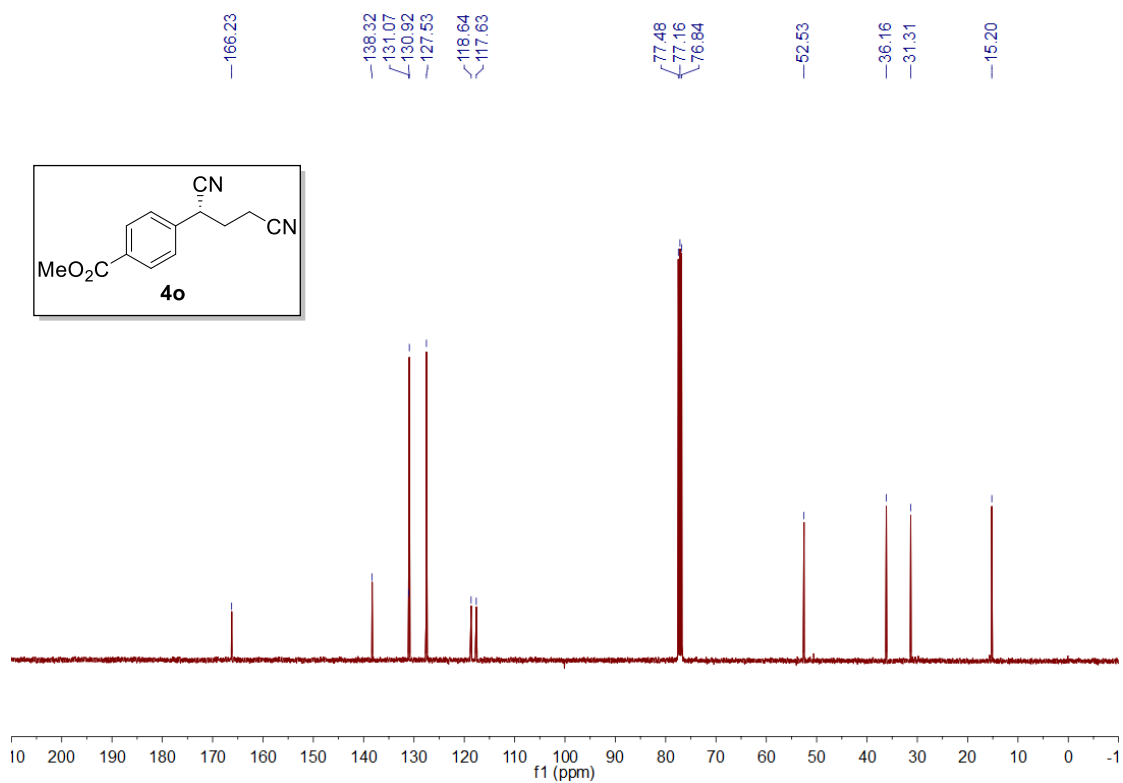
Supplementary Figure 131. ¹H NMR of **4n**



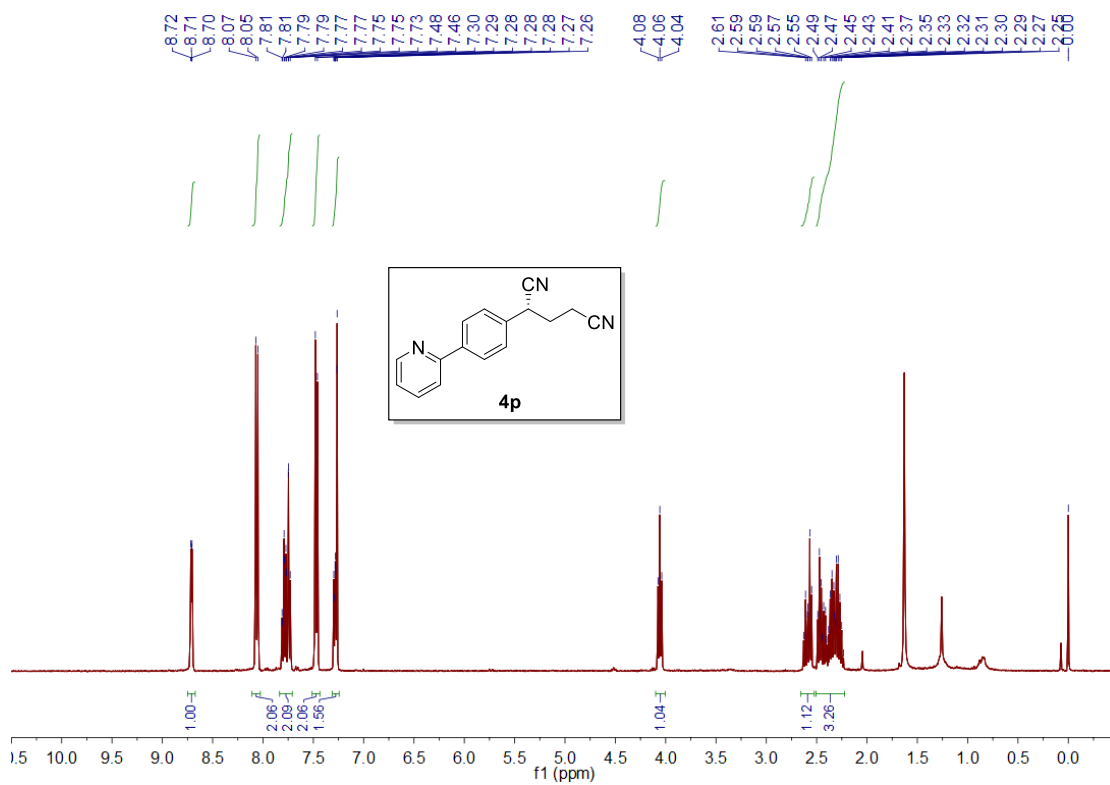
Supplementary Figure 132. ¹³C NMR of **4n**



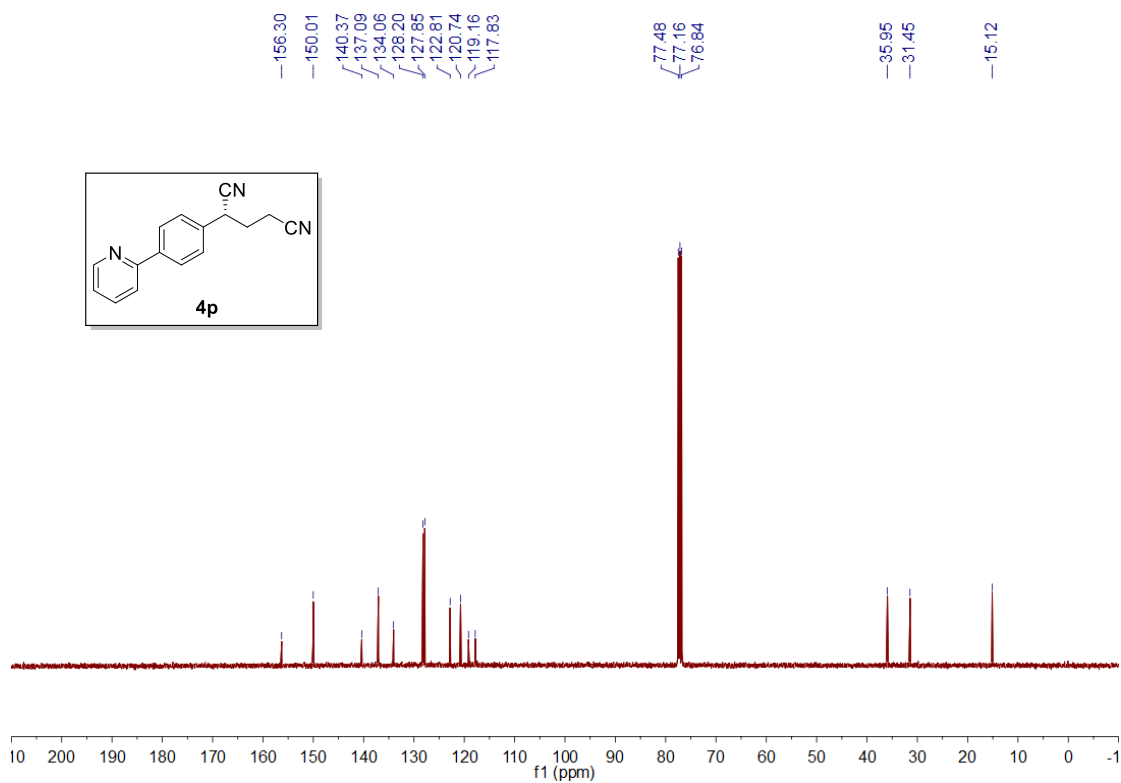
Supplementary Figure 133. ¹H NMR of **4o**



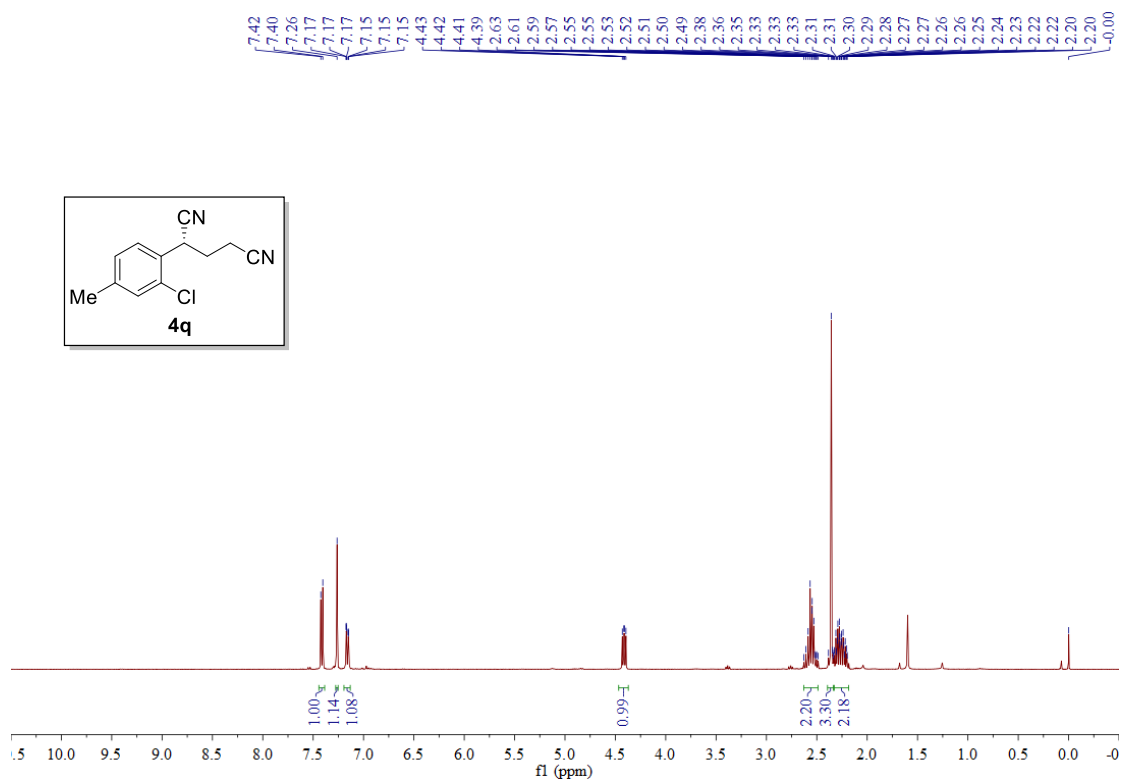
Supplementary Figure 134. ^{13}C NMR of **4o**



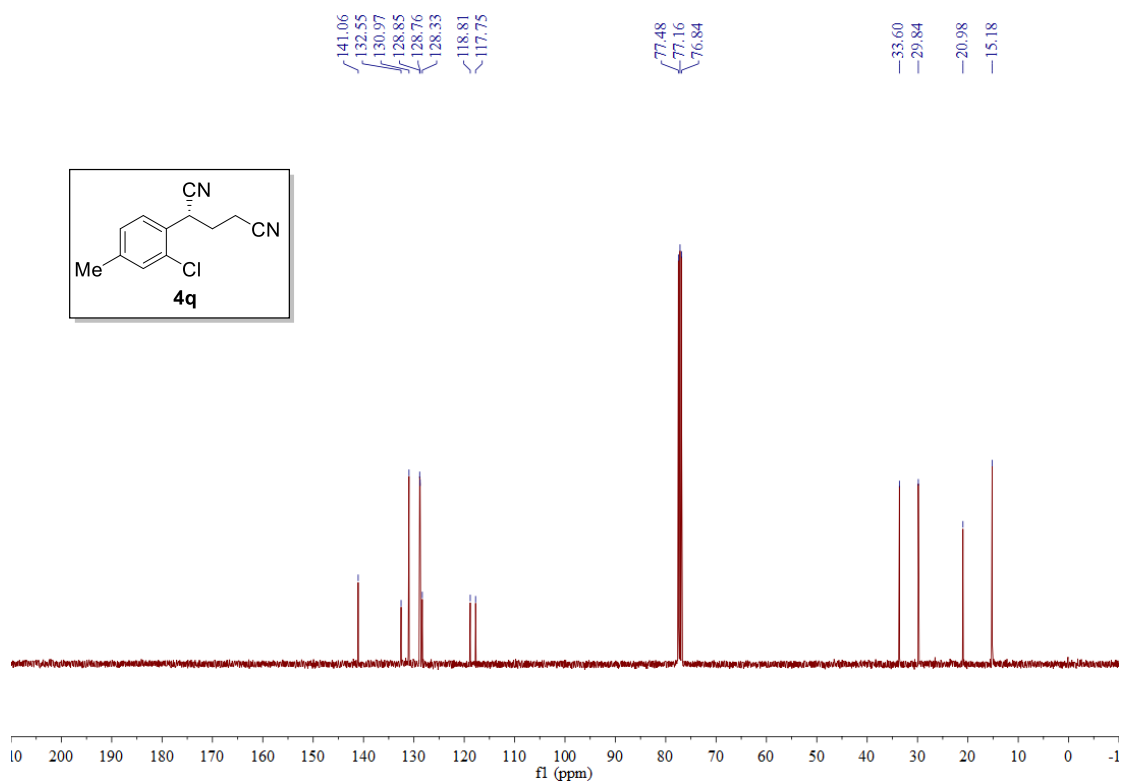
Supplementary Figure 135. ^1H NMR of **4p**



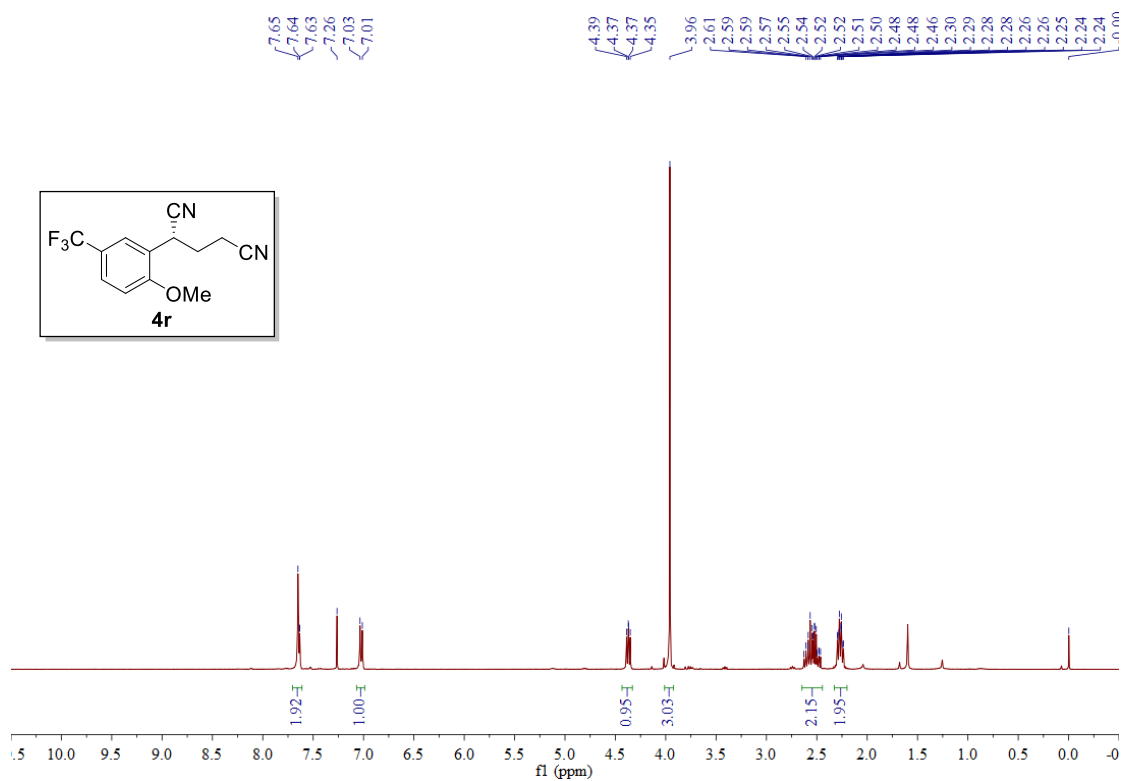
Supplementary Figure 136. ¹³C NMR of **4p**



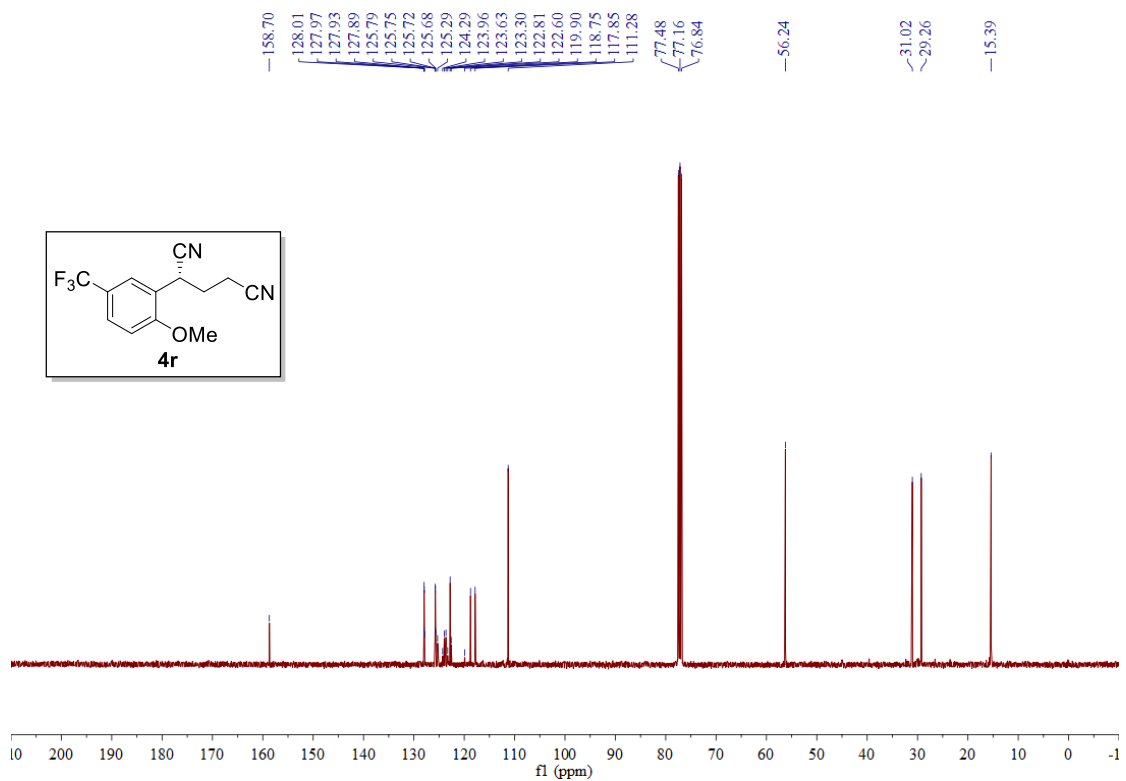
Supplementary Figure 137. ¹H NMR of **4q**



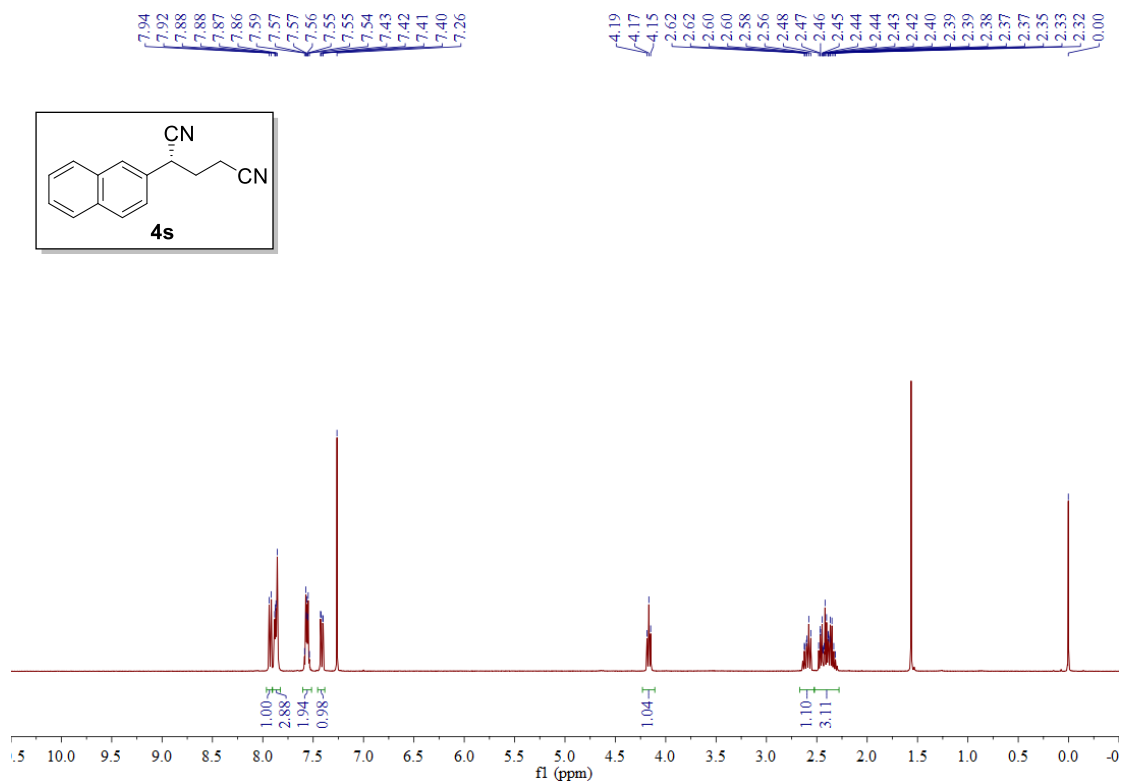
Supplementary Figure 138. ^{13}C NMR of **4q**



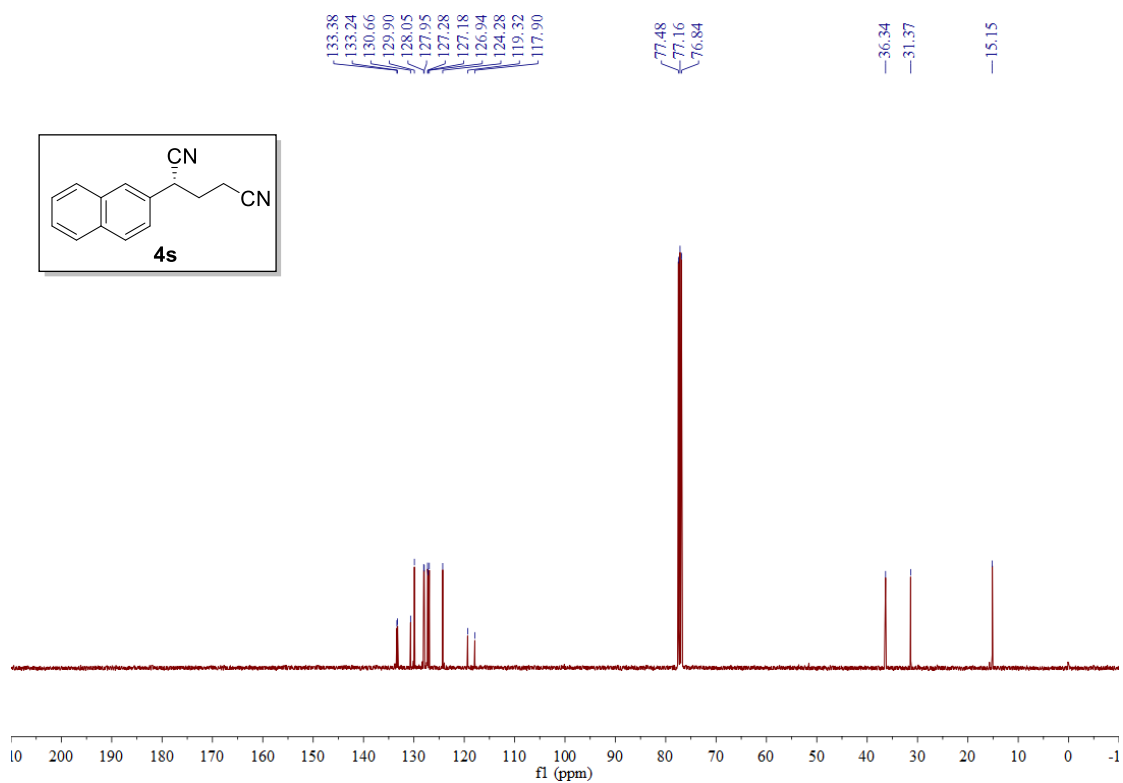
Supplementary Figure 139. ^1H NMR of **4r**



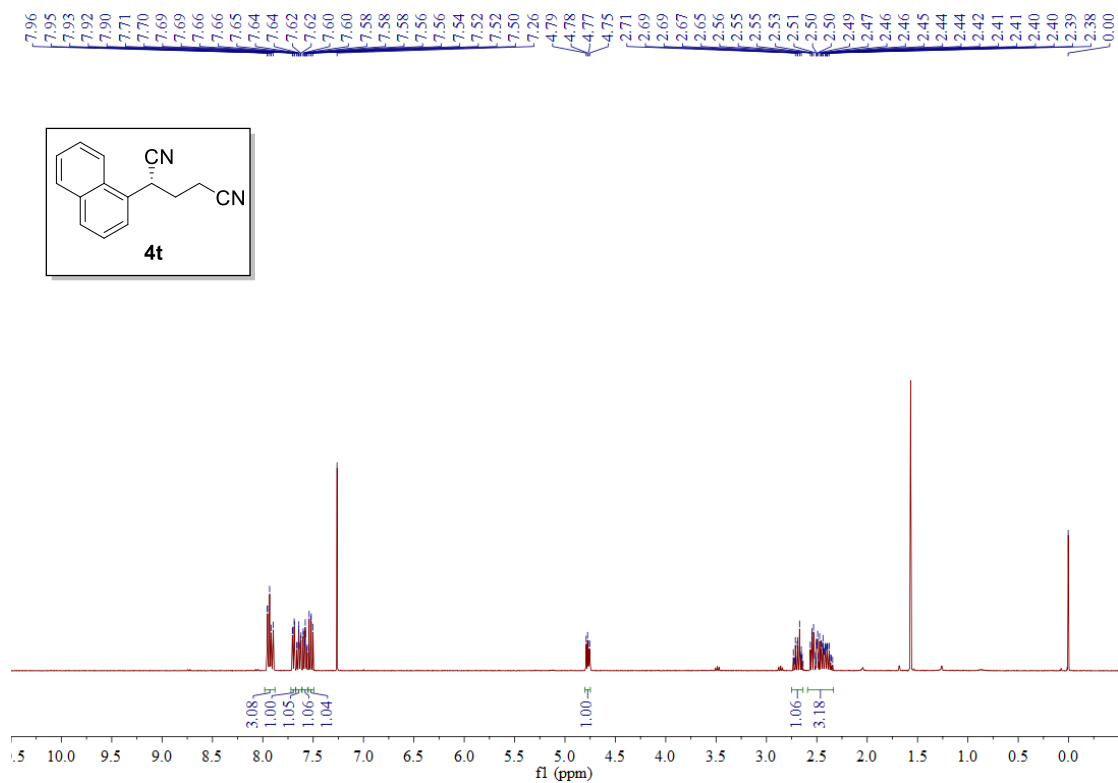
Supplementary Figure 140. ^{13}C NMR of **4r**



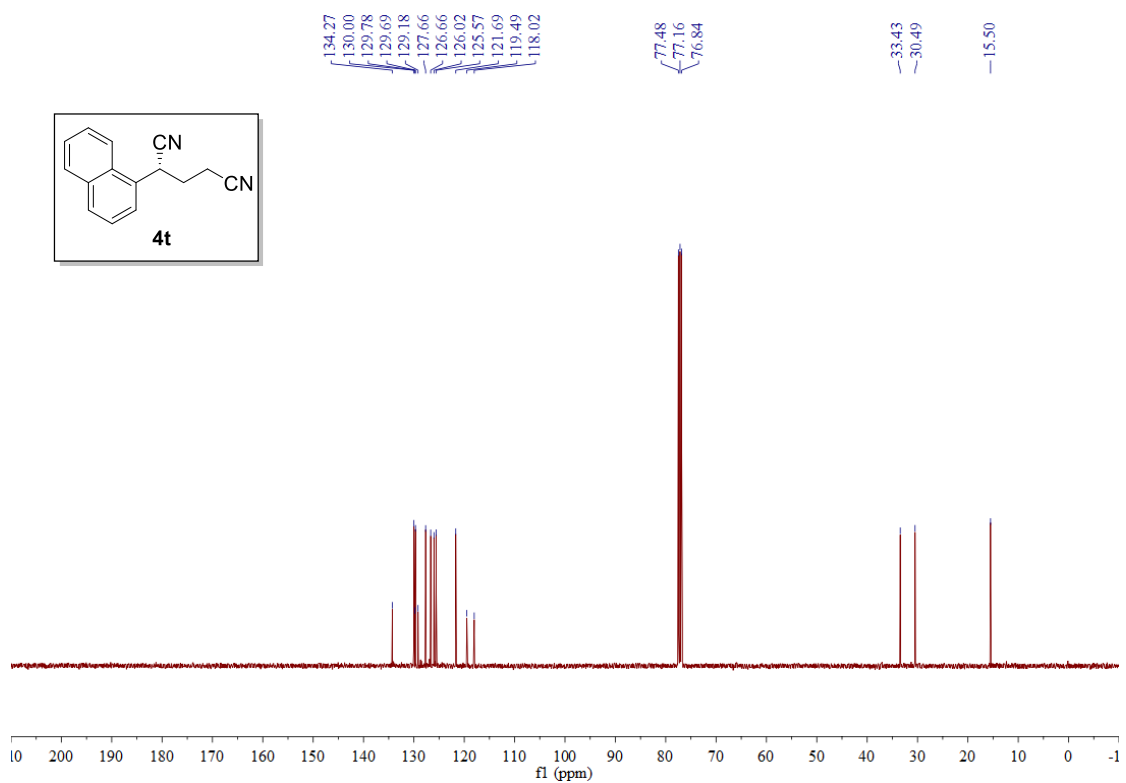
Supplementary Figure 141. ^1H NMR of **4s**



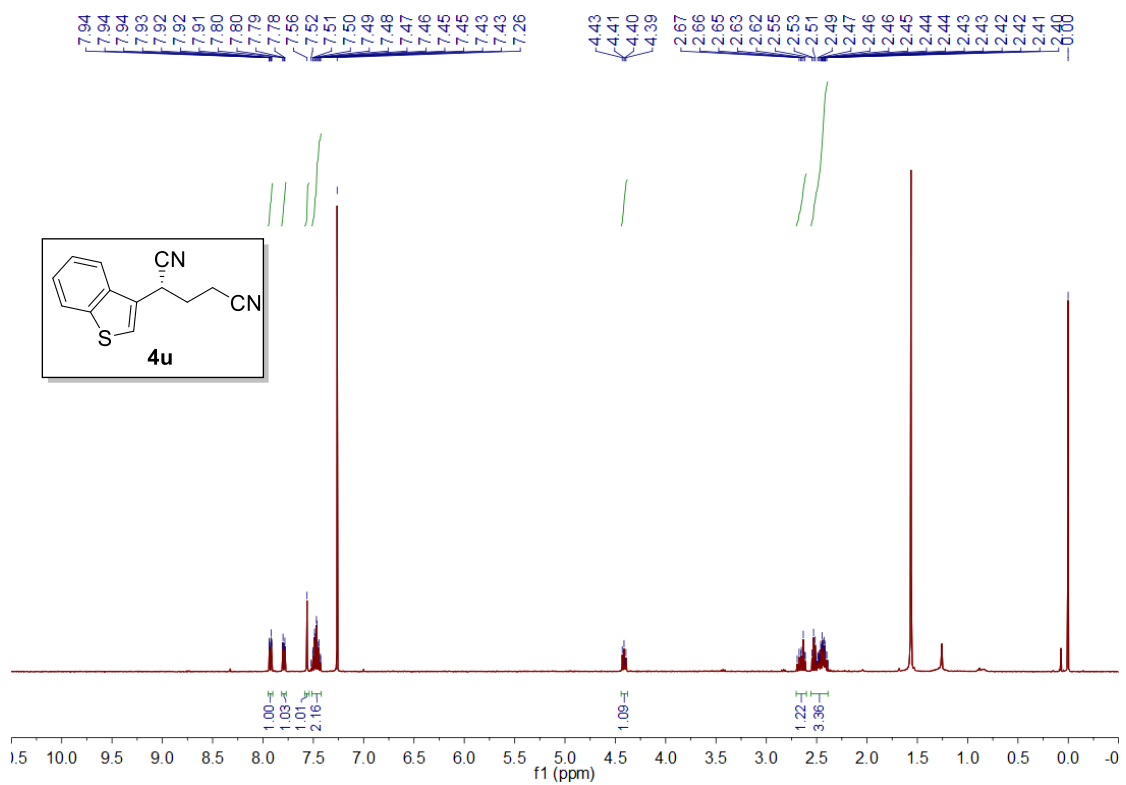
Supplementary Figure 142. ¹³C NMR of **4s**



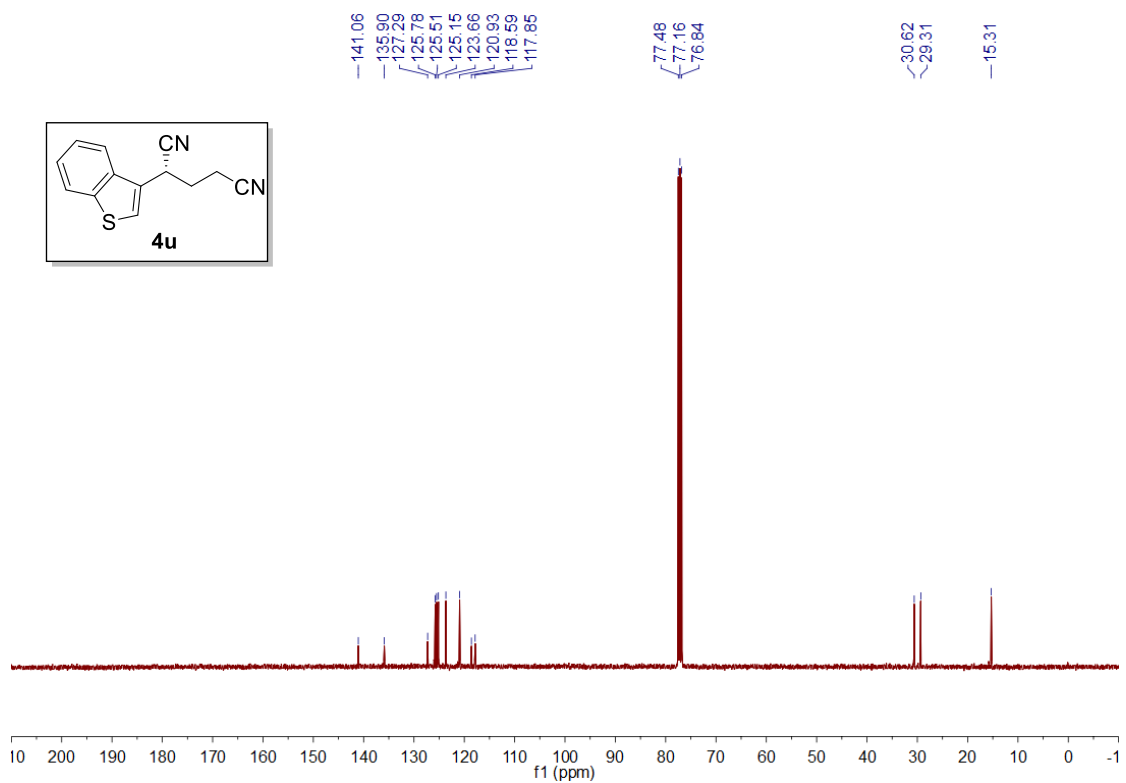
Supplementary Figure 143. ¹H NMR of **4t**



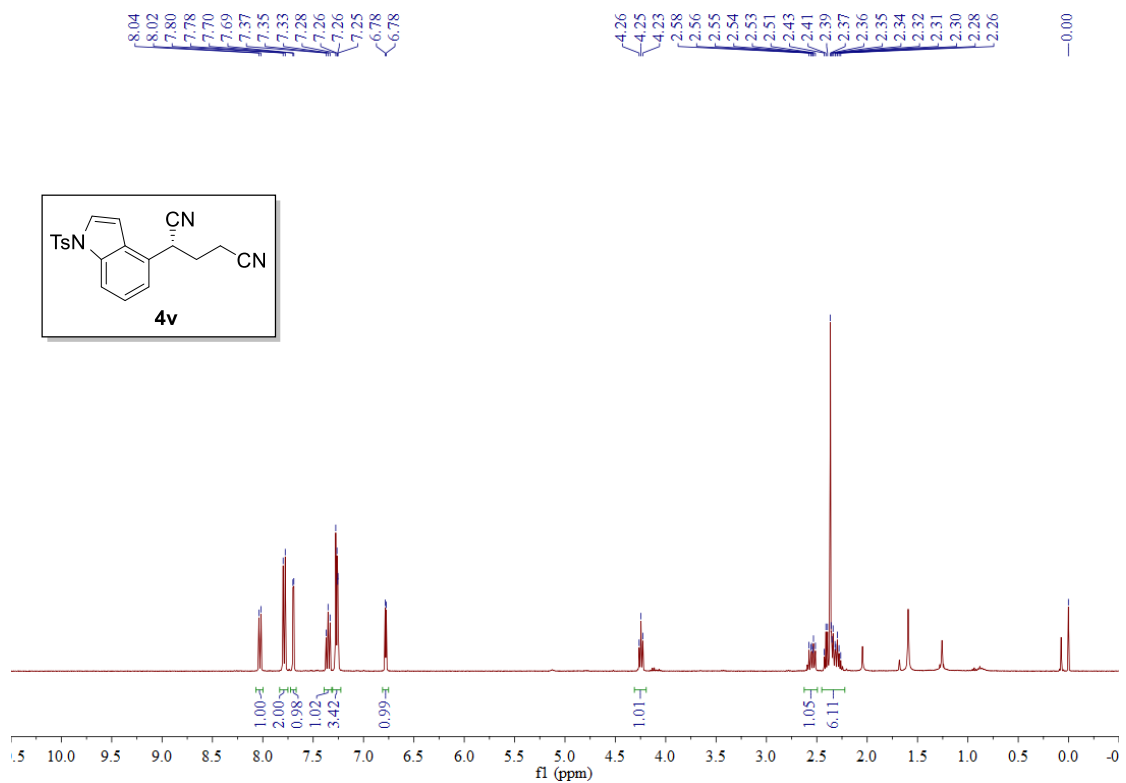
Supplementary Figure 144. ¹³C NMR of **4t**



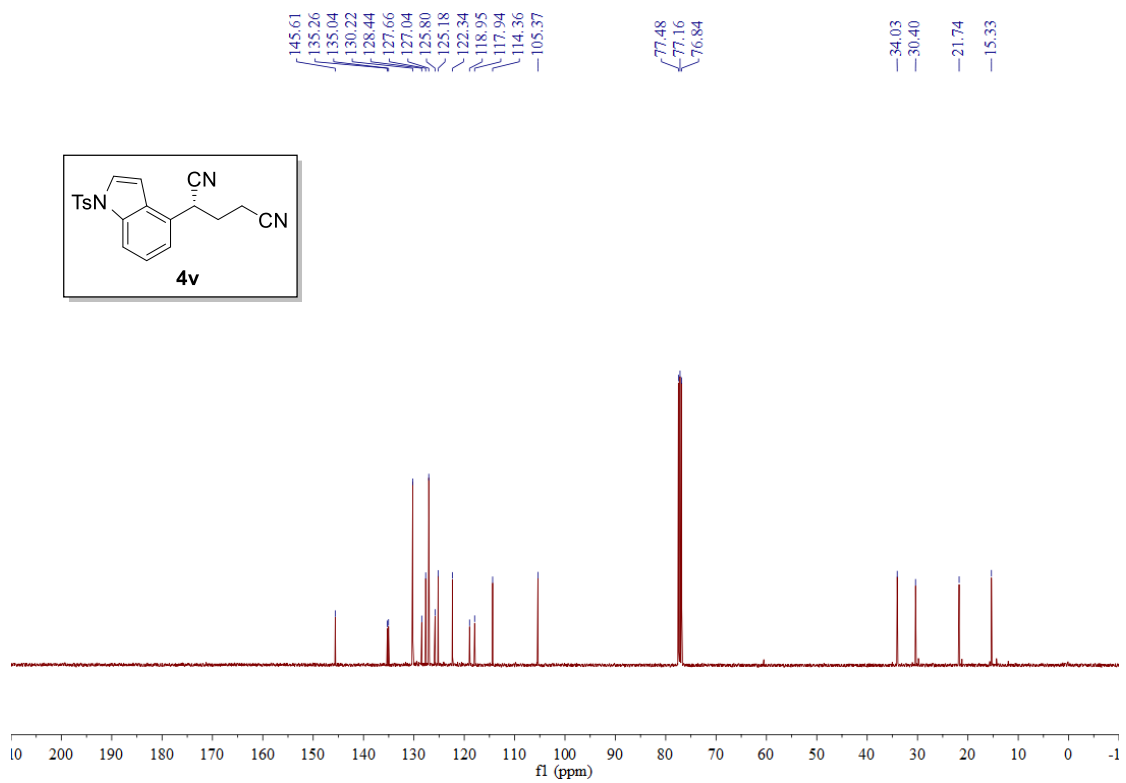
Supplementary Figure 145. ¹H NMR of **4u**



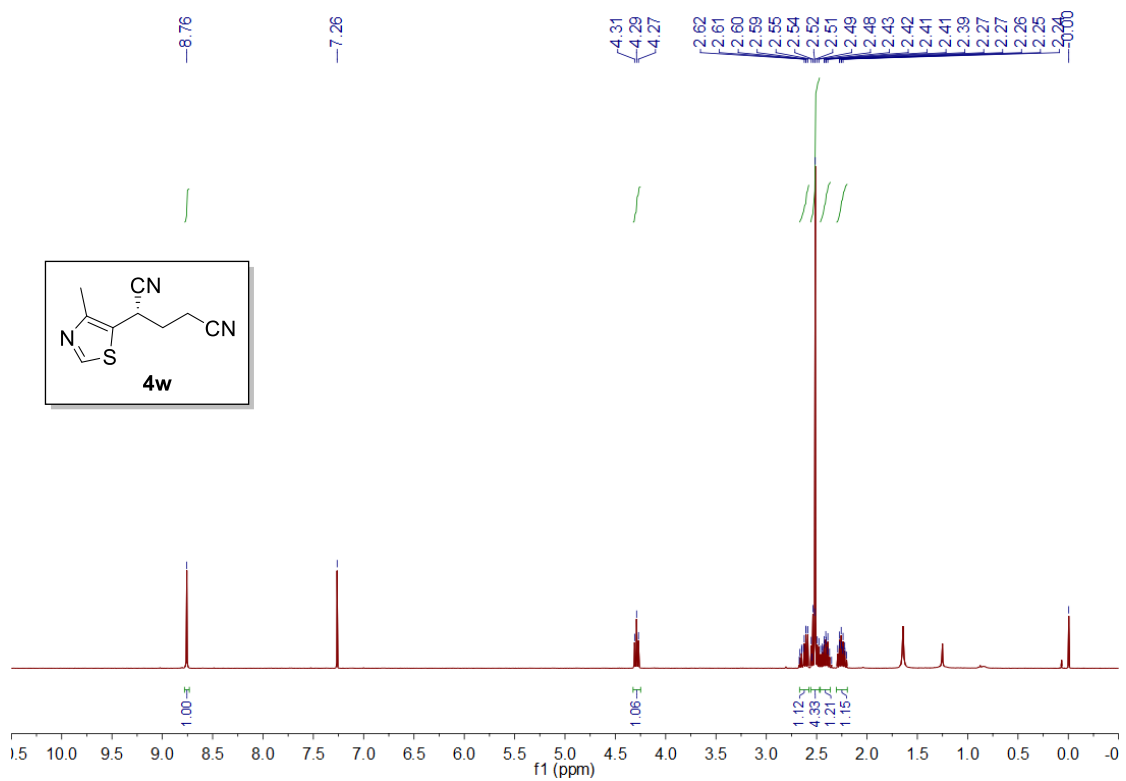
Supplementary Figure 146. ¹³C NMR of **4u**



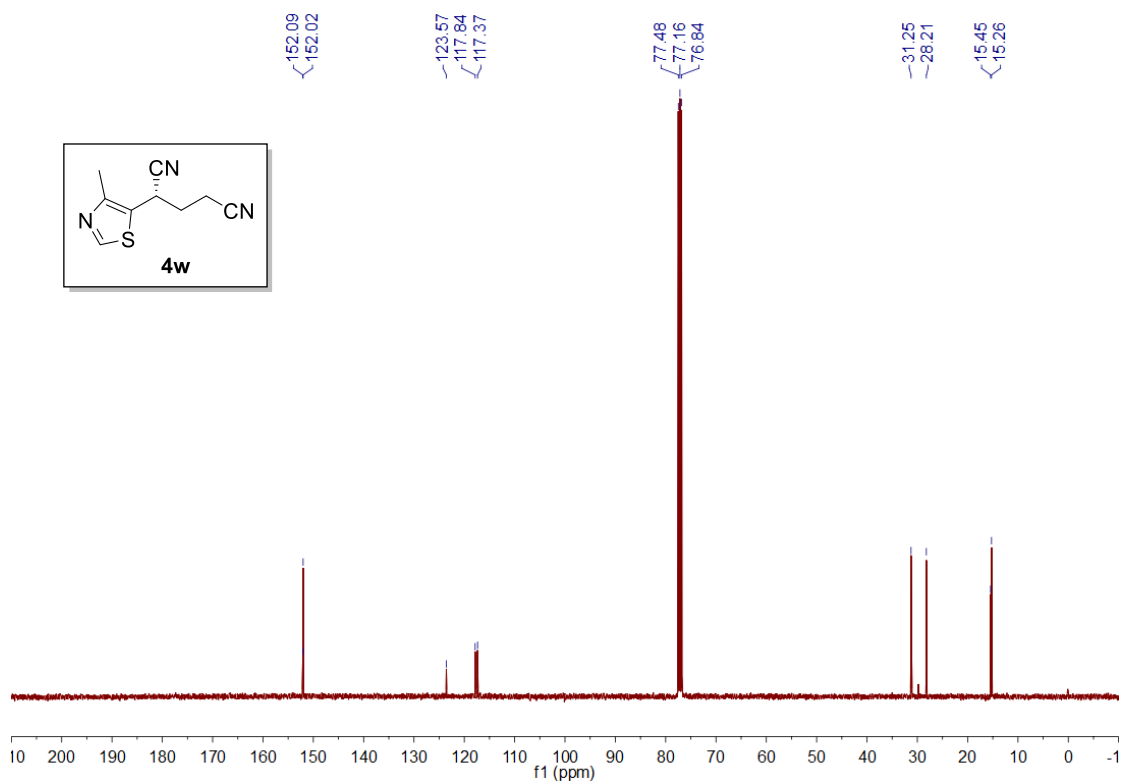
Supplementary Figure 147. ¹H NMR of **4v**



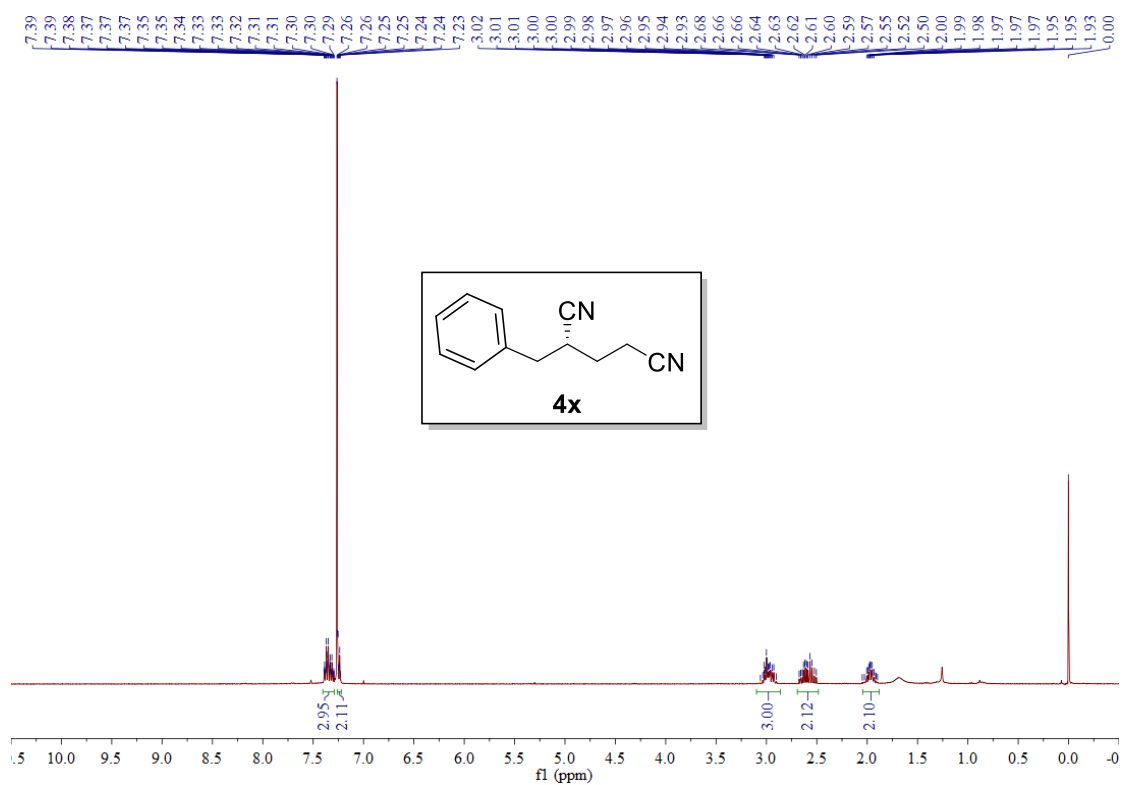
Supplementary Figure 148. ¹³C NMR of **4v**



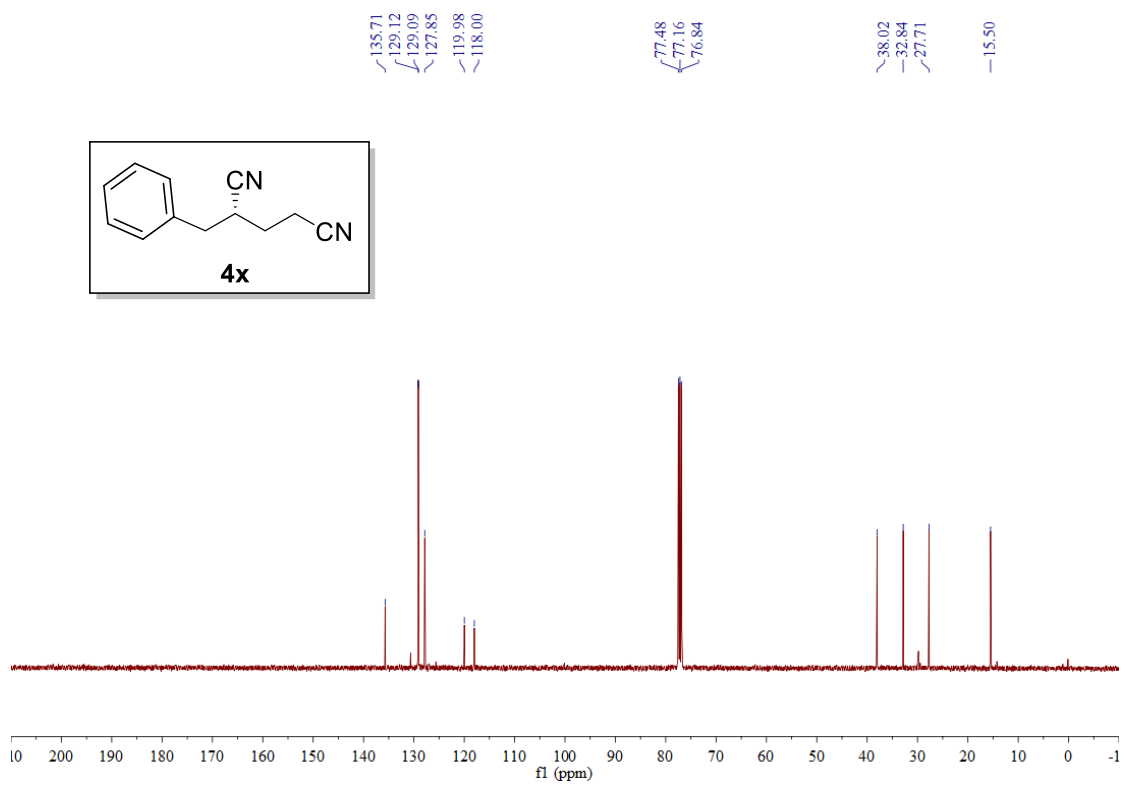
Supplementary Figure 149. ¹H NMR of **4w**



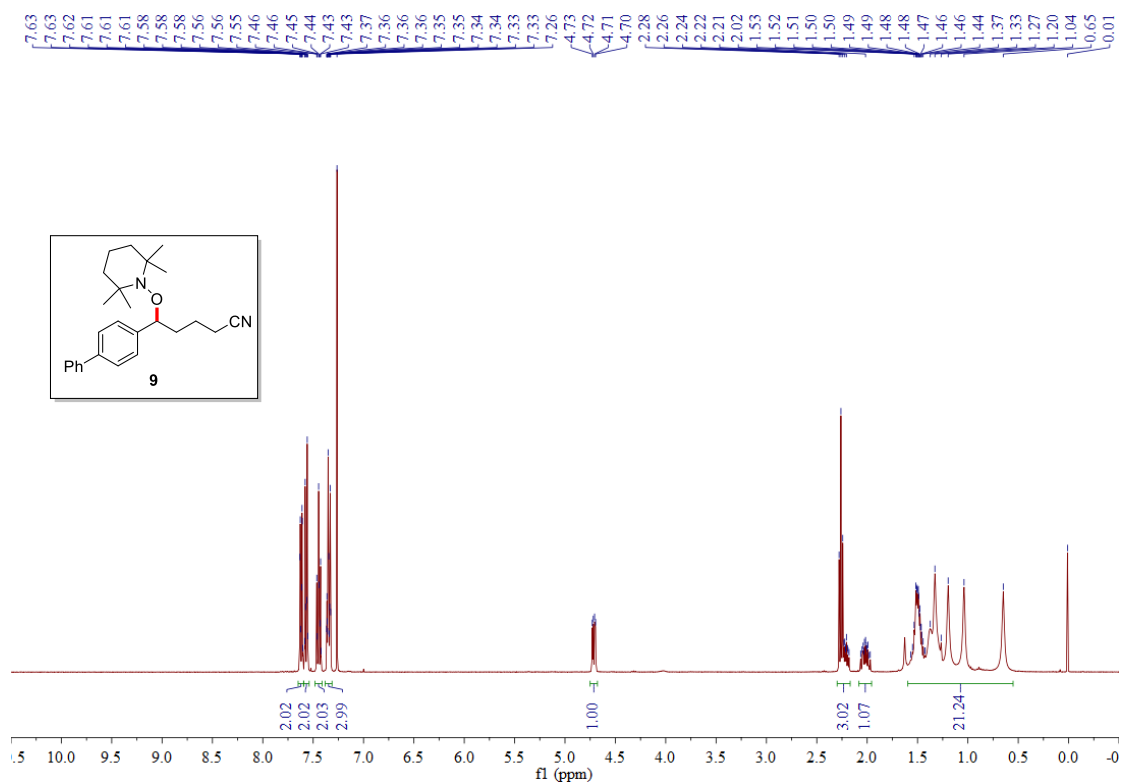
Supplementary Figure 150. ¹³C NMR of **4w**



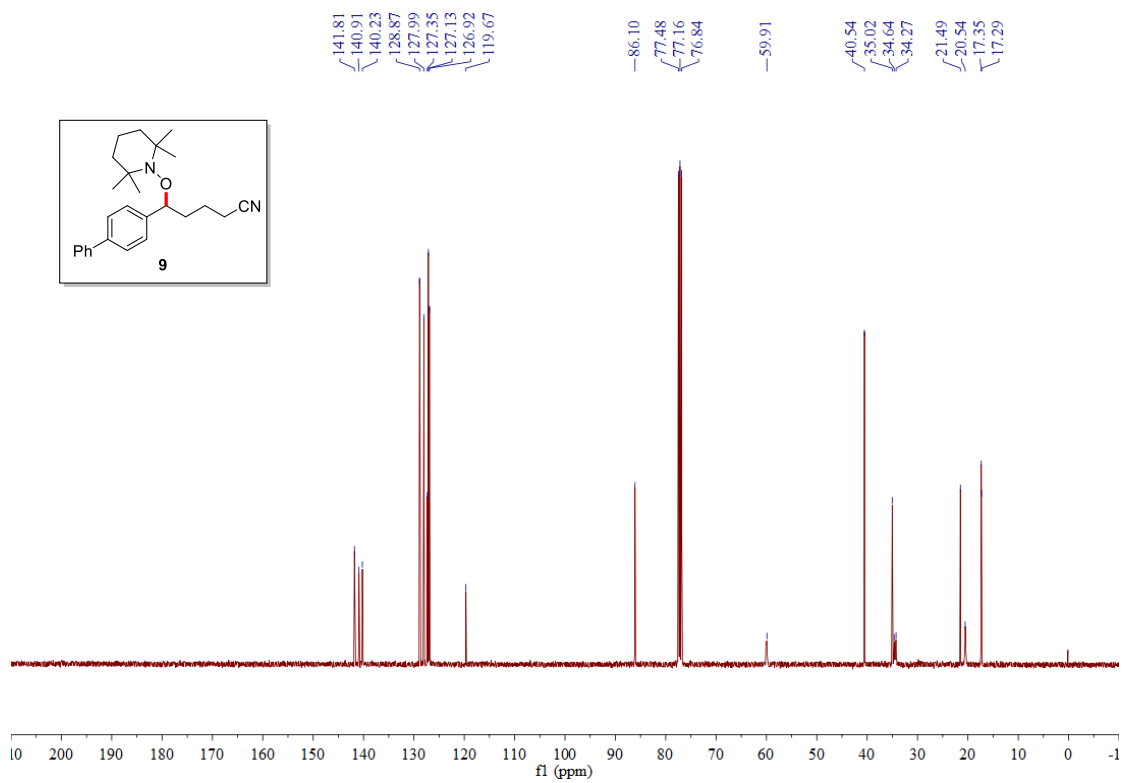
Supplementary Figure 151. ¹H NMR of **4x**



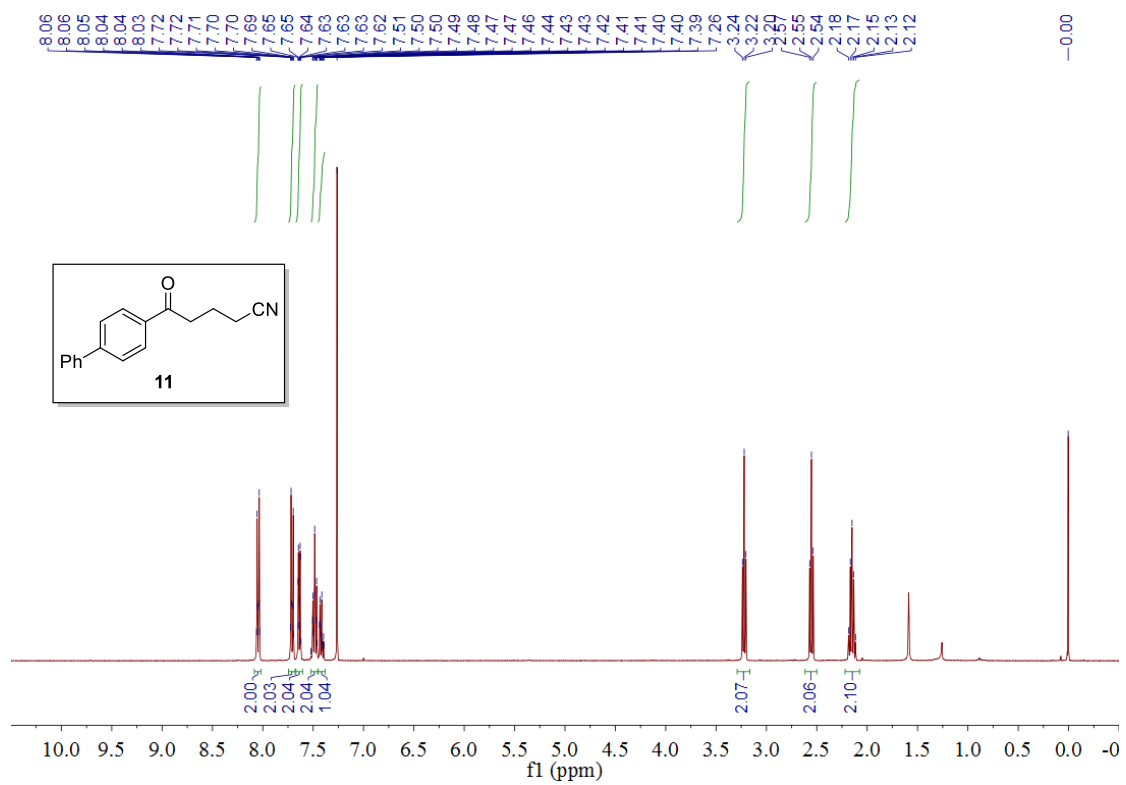
Supplementary Figure 152. ^{13}C NMR of **4x**



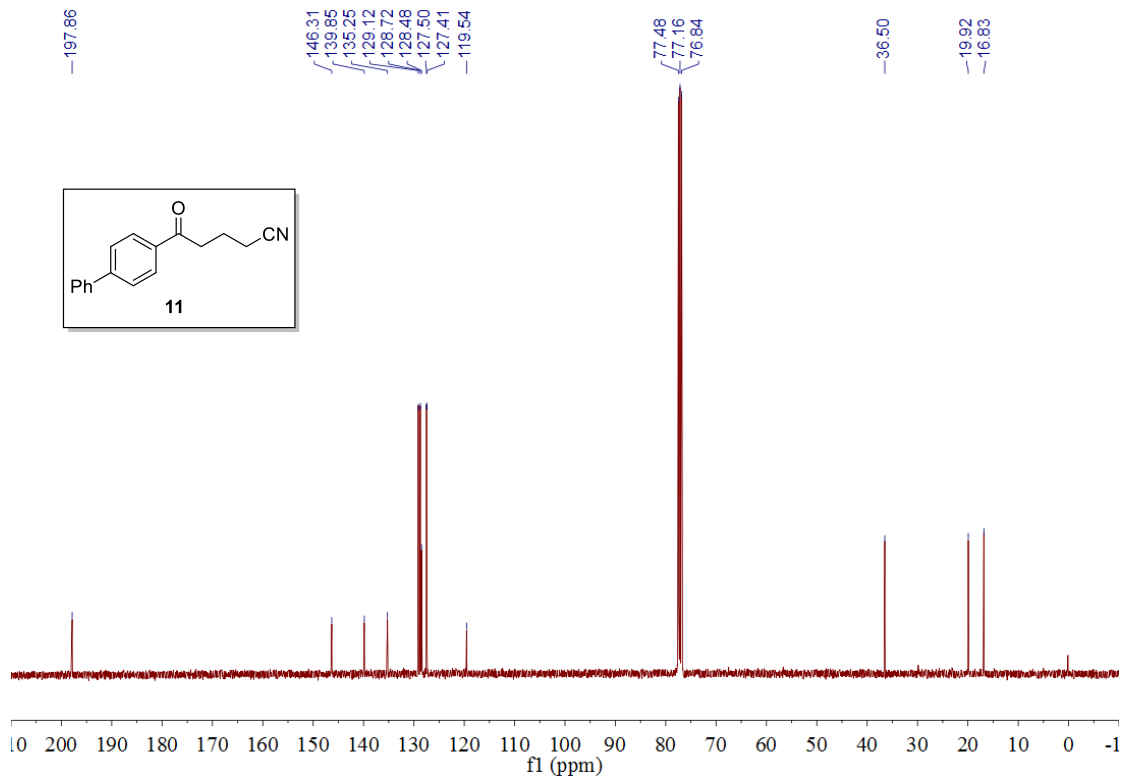
Supplementary Figure 153. ^1H NMR of **9**



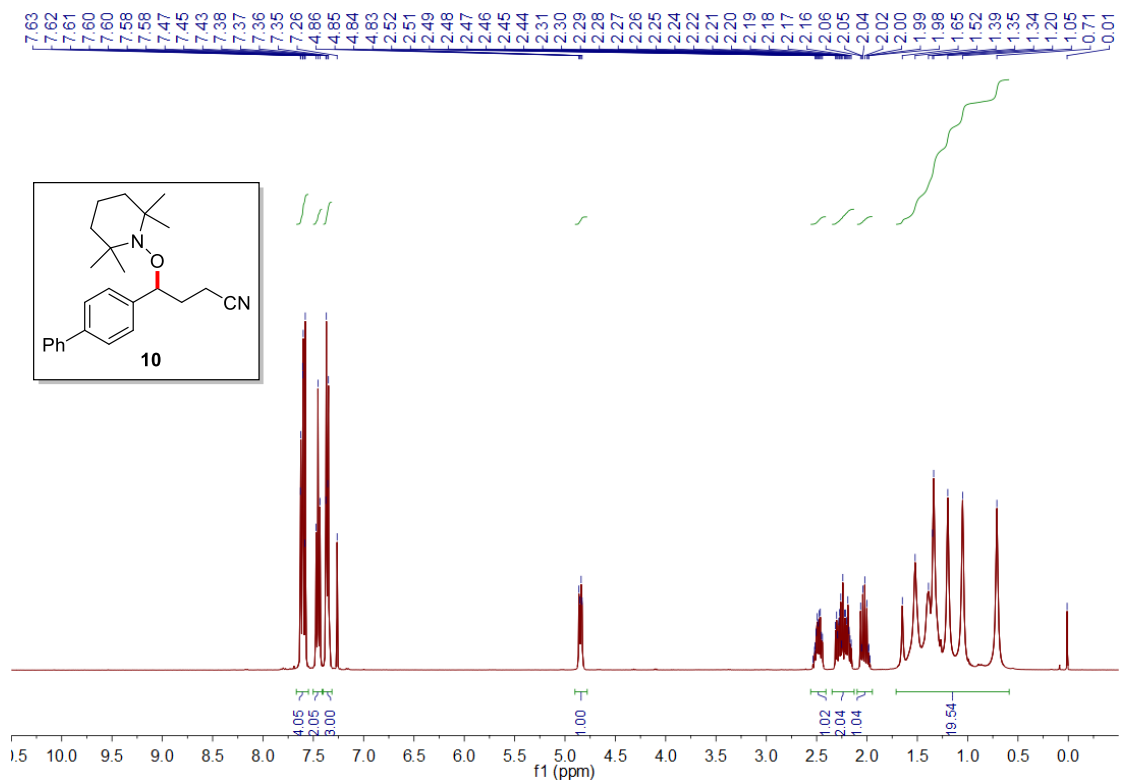
Supplementary Figure 154. ¹³C NMR of **9**



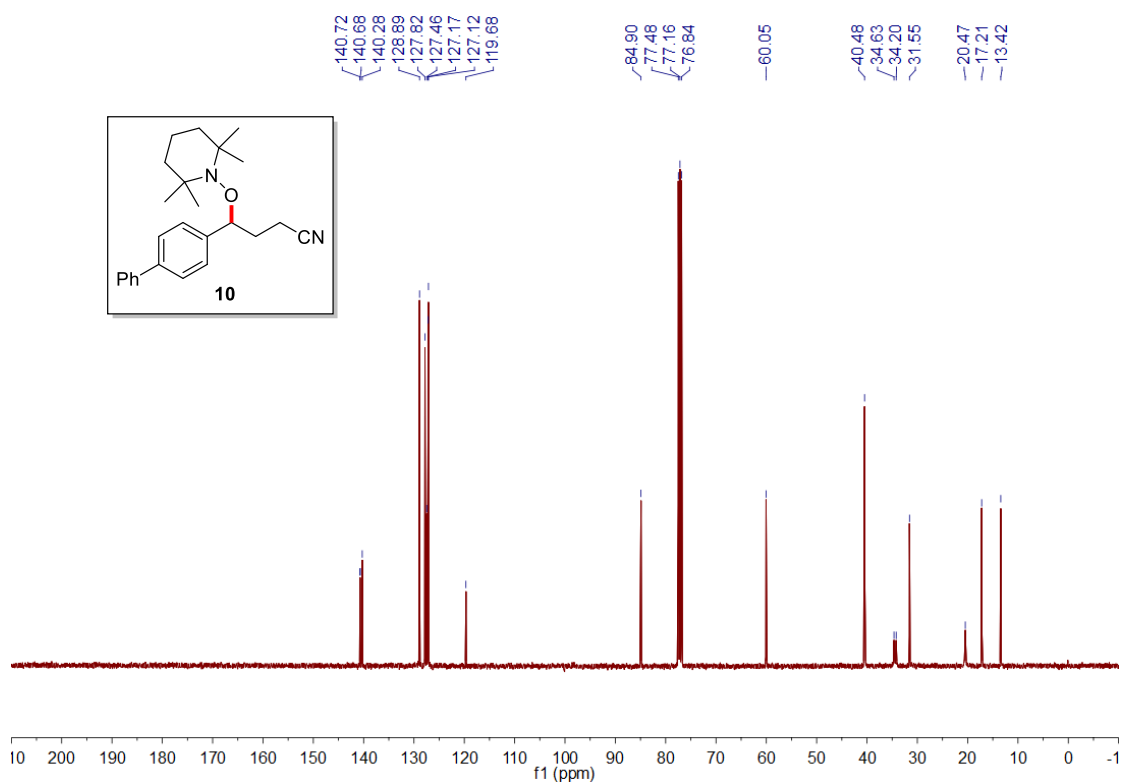
Supplementary Figure 155. ¹H NMR of **11**



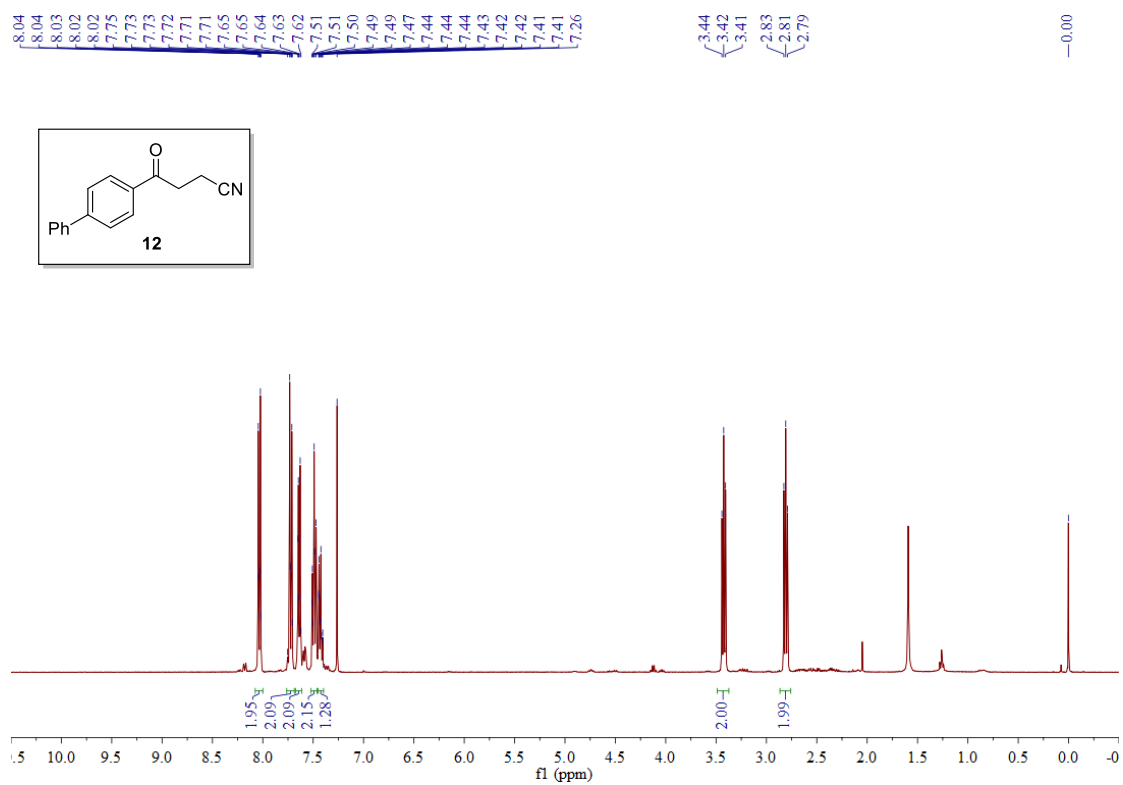
Supplementary Figure 156. ¹³C NMR of **11**



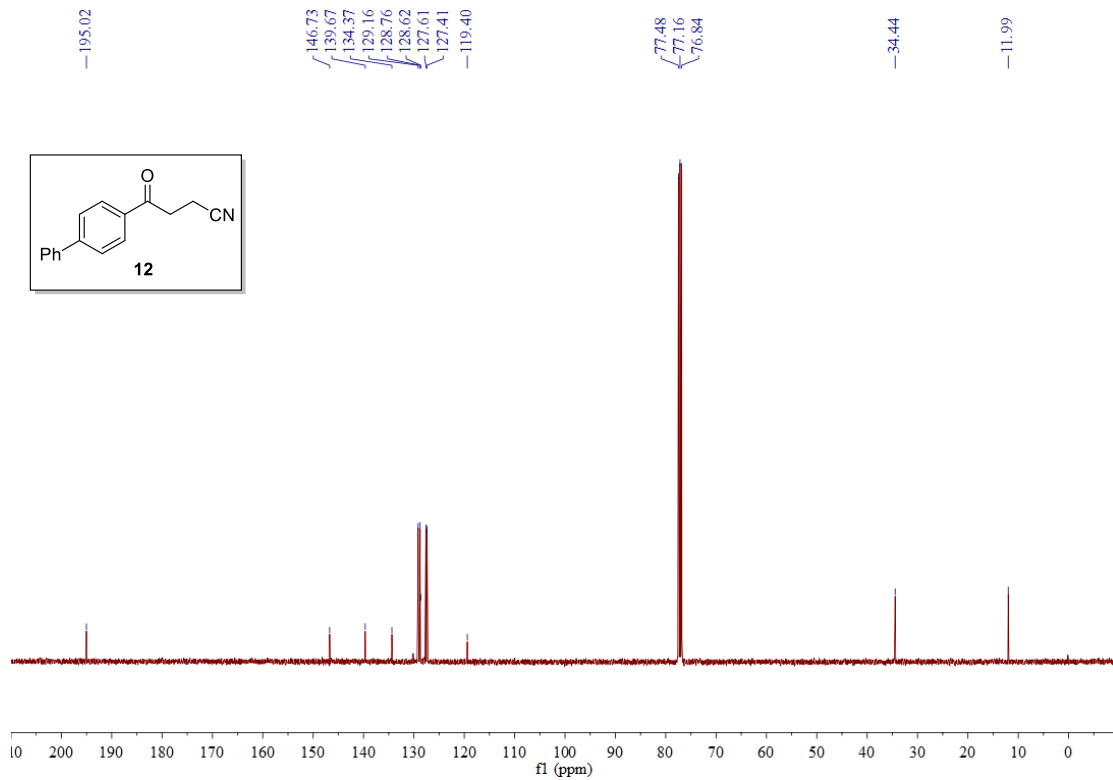
Supplementary Figure 157. ¹H NMR of **10**



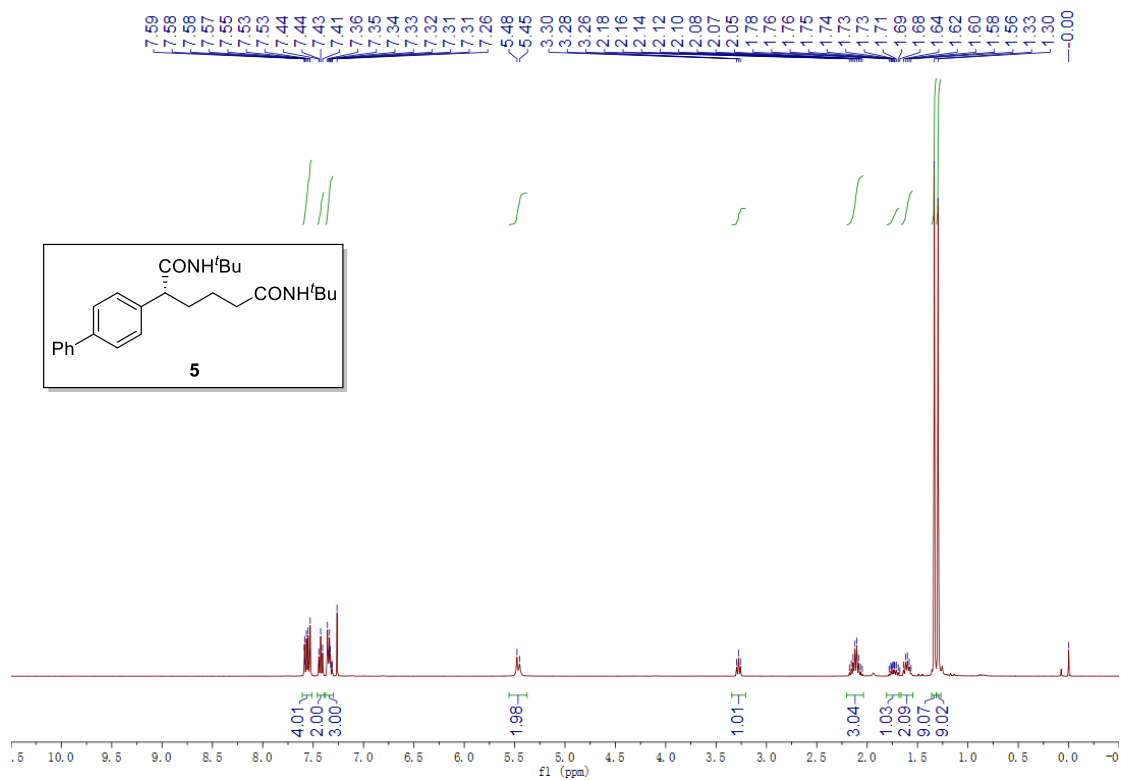
Supplementary Figure 158. ^{13}C NMR of **10**



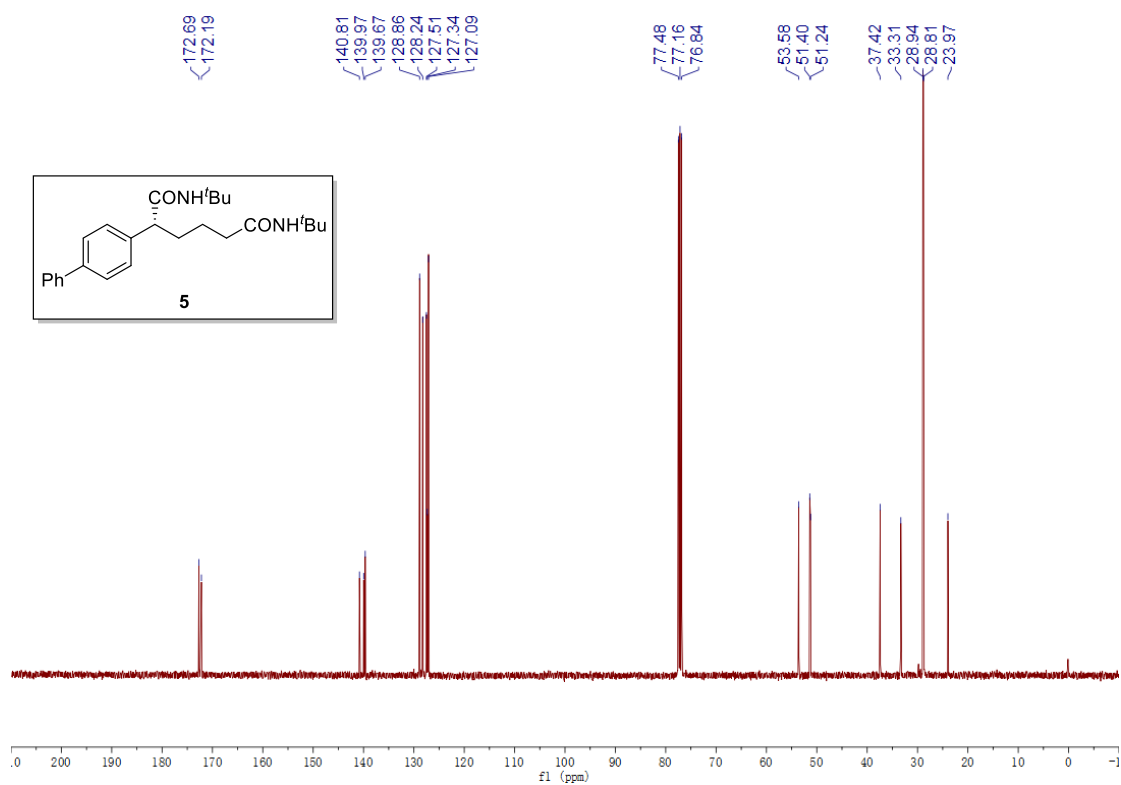
Supplementary Figure 159. ^1H NMR of **12**



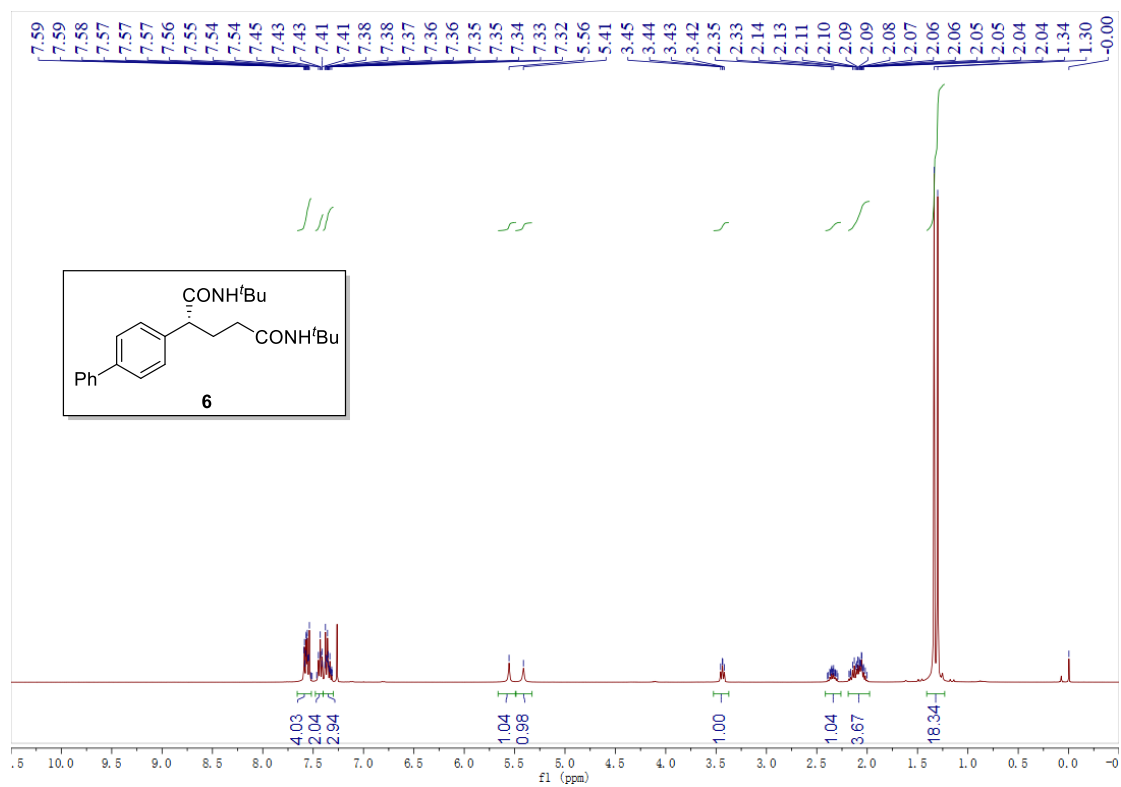
Supplementary Figure 160. ^{13}C NMR of **12**



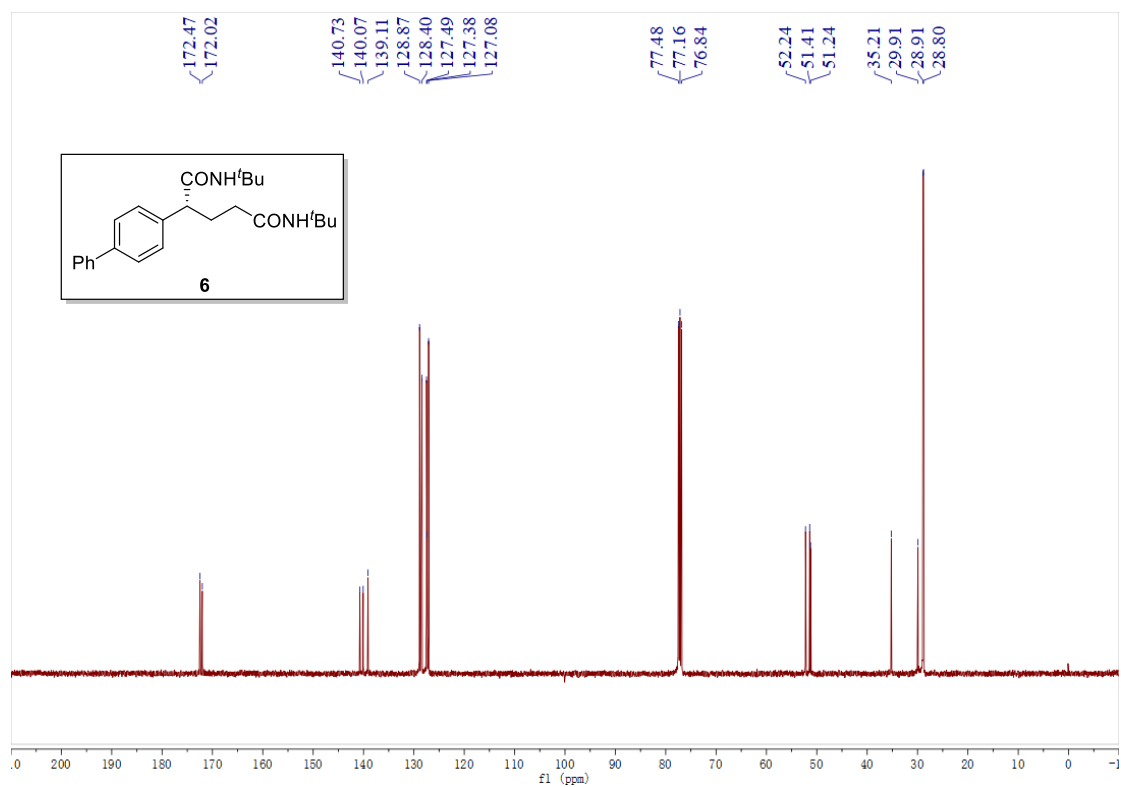
Supplementary Figure 161. ^1H NMR of **5**



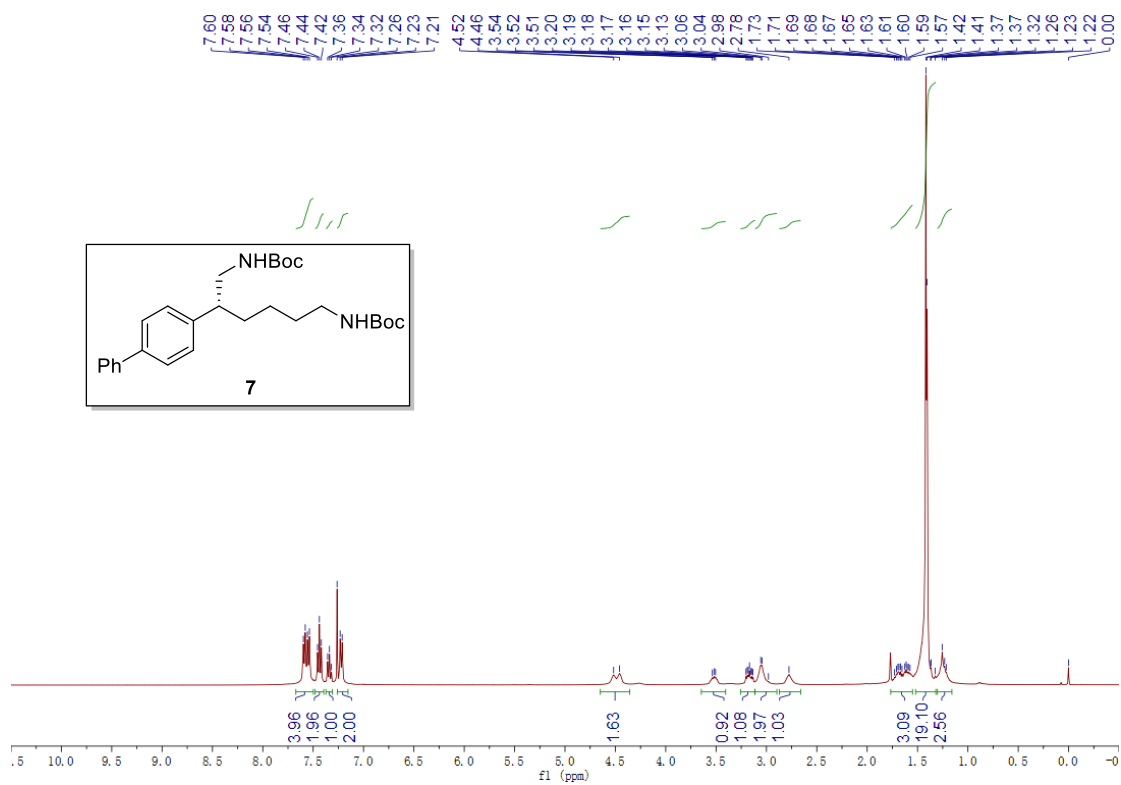
Supplementary Figure 162. ¹³C NMR of 5



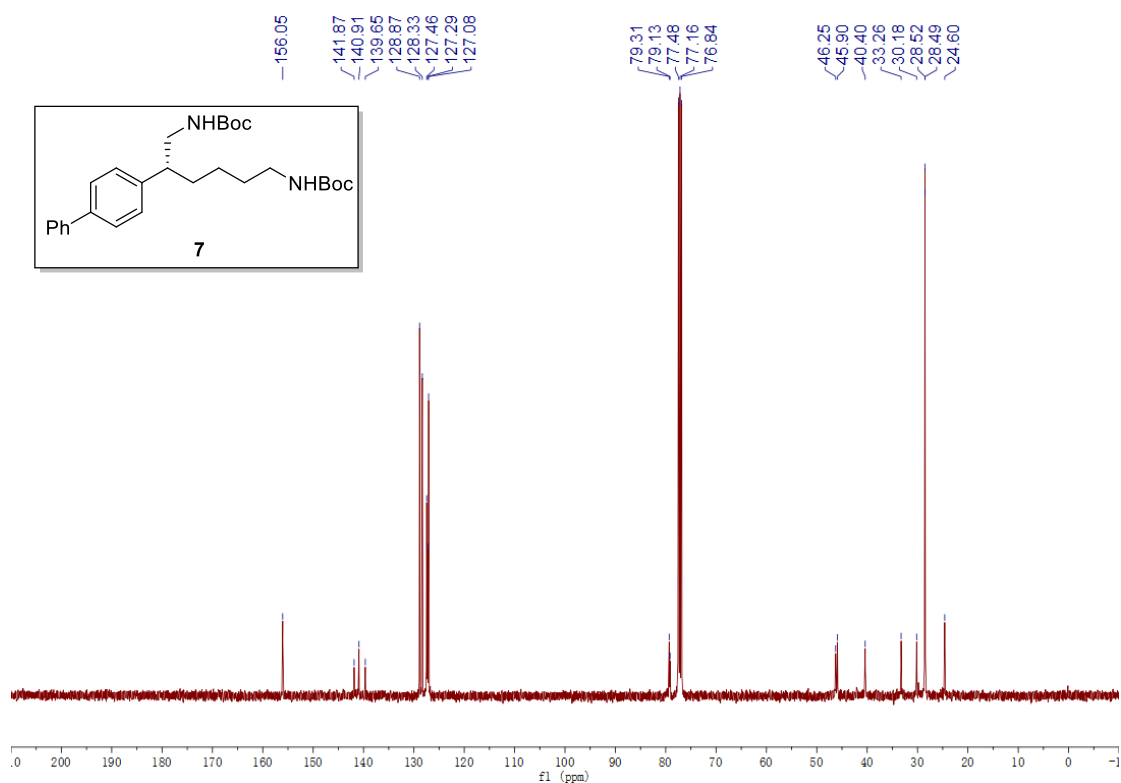
Supplementary Figure 163. ¹H NMR of 6



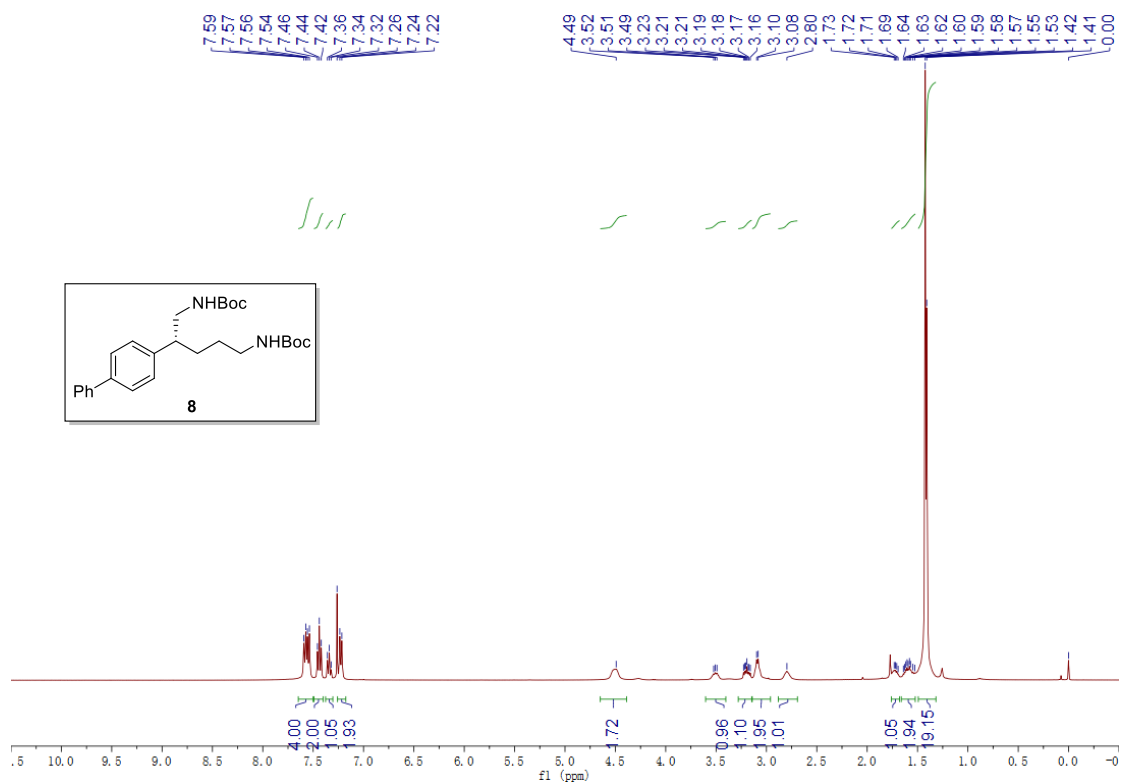
Supplementary Figure 164. ¹³C NMR of 6



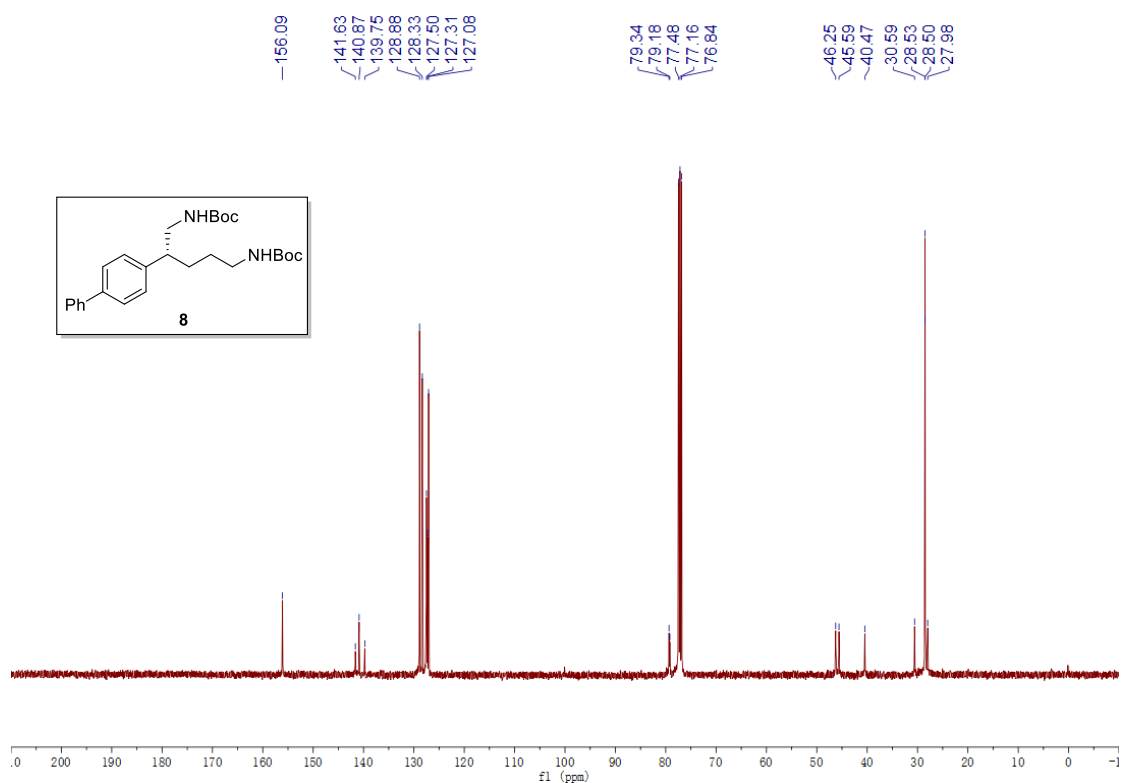
Supplementary Figure 165. ¹H NMR of 7



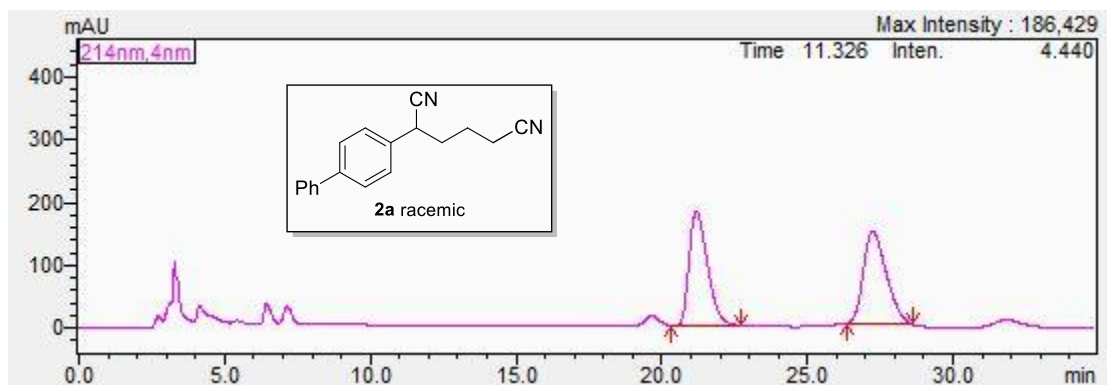
Supplementary Figure 166. ¹³C NMR of 7



Supplementary Figure 167. ¹H NMR of 8

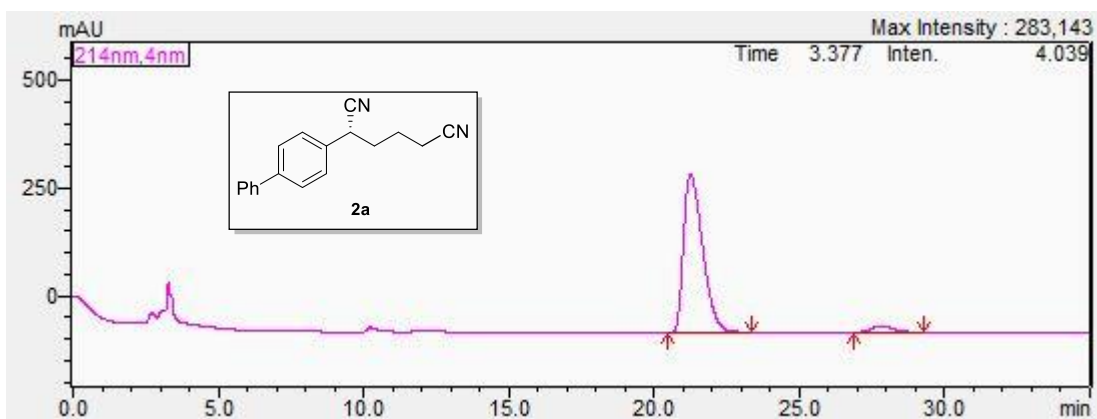


Supplementary Figure 168. ^{13}C NMR of **8**



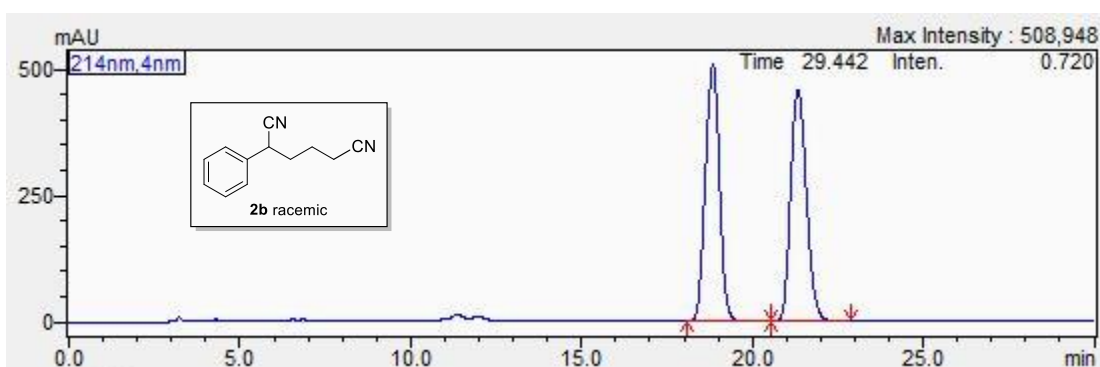
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 21.193 | 49.497 | 182329 | 7962872 |
| 2 | 27.239 | 50.503 | 147469 | 8124776 |
| Total: | | 100.000 | 329797 | 16087649 |

Supplementary Figure 169. HPLC data of rac-**2a**



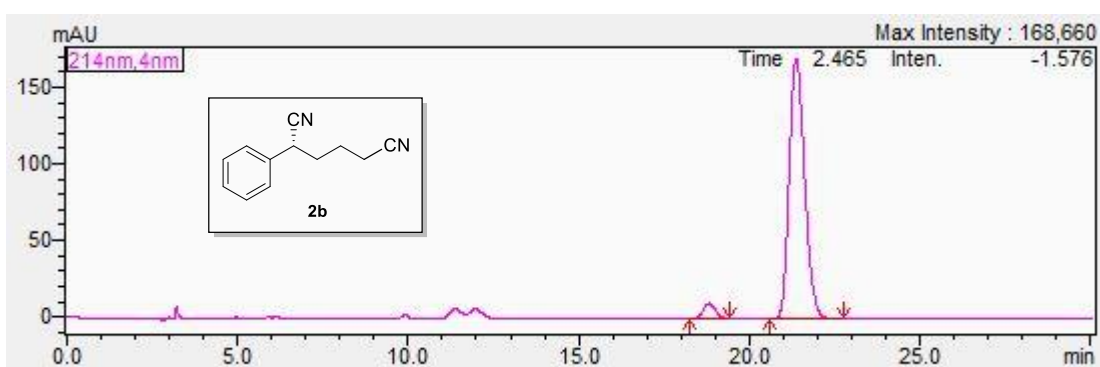
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 21.263 | 95.858 | 367380 | 16671598 |
| 2 | 27.826 | 4.142 | 13746 | 720306 |
| Total: | | 100.000 | 381126 | 17391904 |

Supplementary Figure 170. HPLC data of **2a**



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 18.849 | 49.750 | 508132 | 14665027 |
| 2 | 21.339 | 50.250 | 458583 | 14812368 |
| Total: | | 100.000 | 966715 | 29477395 |

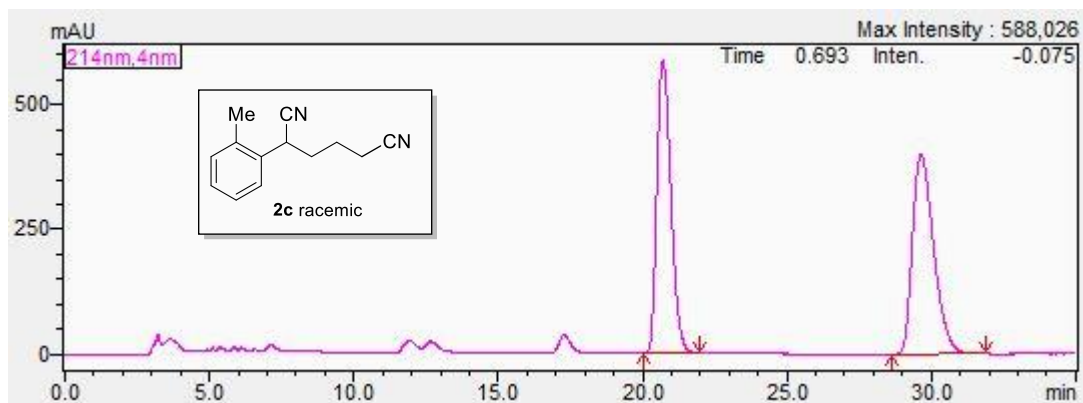
Supplementary Figure 171. HPLC data of rac-**2b**



| Peak | Ret. time | Area % | Height | Area |
|------|-----------|--------|--------|--------|
| 1 | 18.828 | 4.712 | 10158 | 264766 |

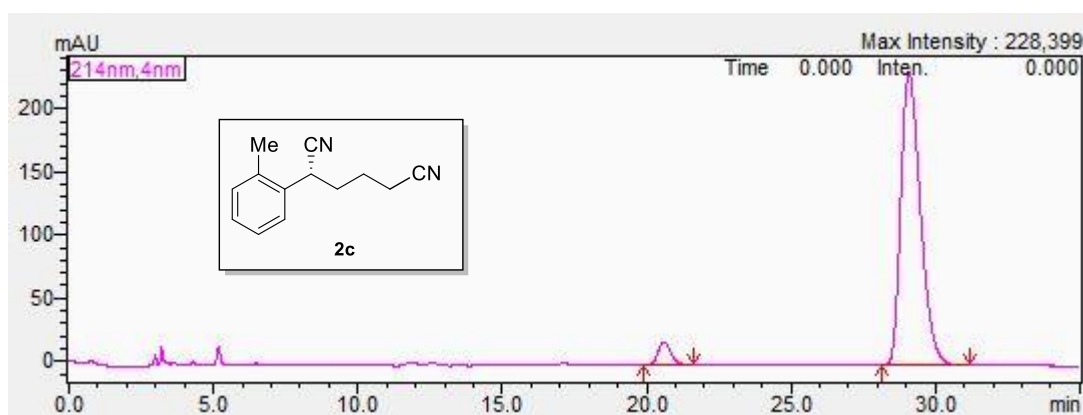
| | | | | |
|--------|--------|---------|--------|---------|
| 2 | 21.376 | 95.288 | 170088 | 5354716 |
| Totol: | | 100.000 | 180246 | 5619482 |

Supplementary Figure 172. HPLC data of **2b**



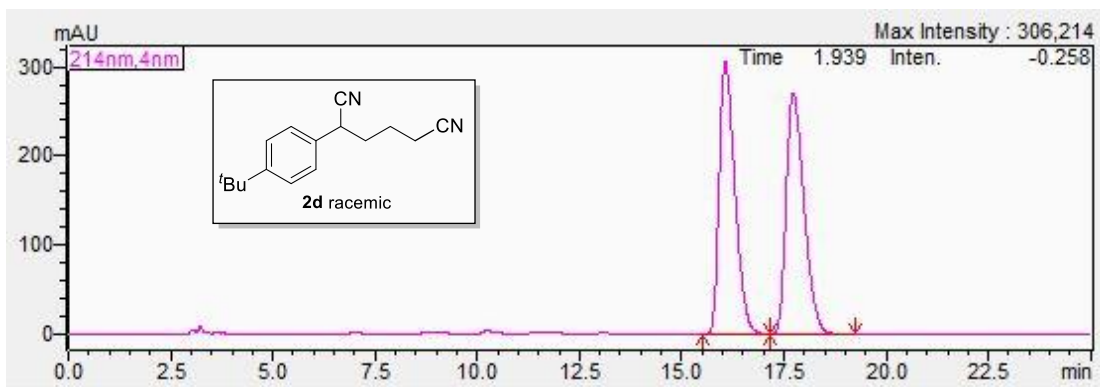
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 20.706 | 49.081 | 586181 | 19740769 |
| 2 | 29.646 | 50.919 | 399264 | 20480290 |
| Totol: | | 100.000 | 985445 | 40221059 |

Supplementary Figure 173. HPLC data of rac-**2c**



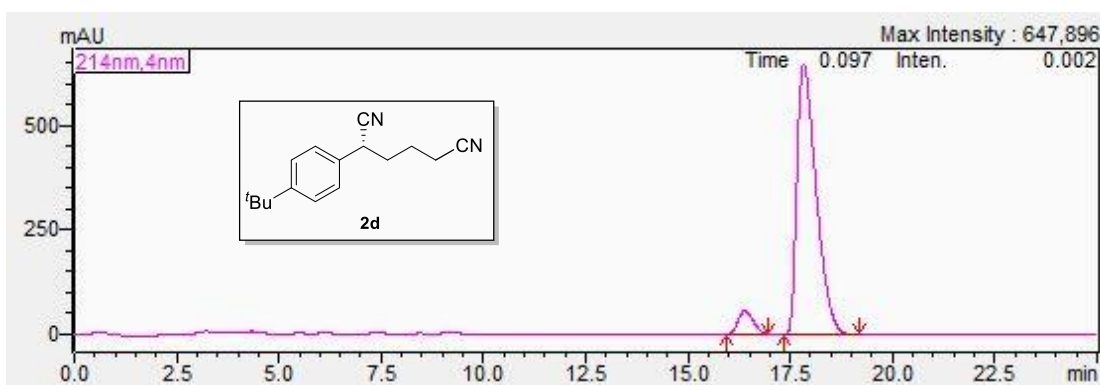
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 20.601 | 4.980 | 18454 | 573471 |
| 2 | 29.091 | 95.020 | 231685 | 10941981 |
| Totol: | | 100.000 | 250140 | 11515452 |

Supplementary Figure 174. HPLC data of **2c**



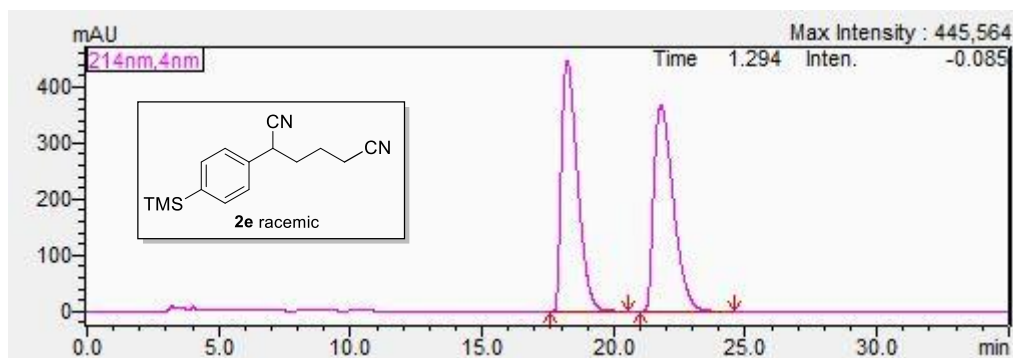
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 16.068 | 49.889 | 306448 | 8207774 |
| 2 | 17.730 | 50.111 | 271171 | 8244137 |
| Total: | | 100.000 | 577619 | 16451911 |

Supplementary Figure 175. HPLC data of rac-**2d**



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 16.393 | 6.233 | 55535 | 1407118 |
| 2 | 17.831 | 93.767 | 647704 | 21169690 |
| Total: | | 100.000 | 703240 | 22576809 |

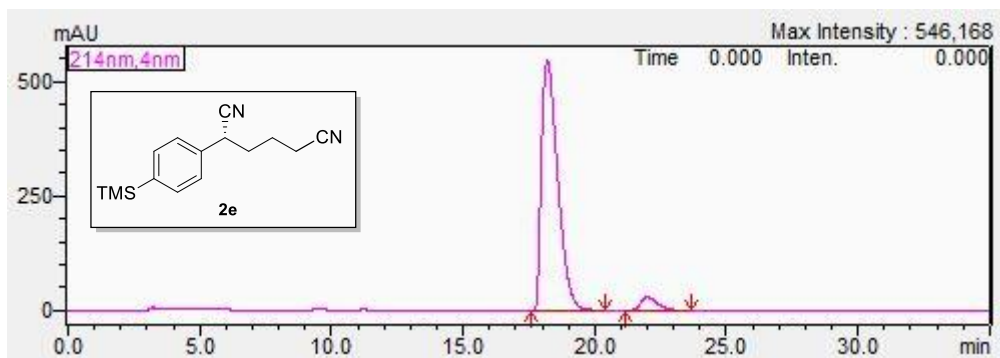
Supplementary Figure 176. HPLC data of **2d**



| Peak | Ret. time | Area % | Height | Area |
|------|-----------|--------|--------|------|
|------|-----------|--------|--------|------|

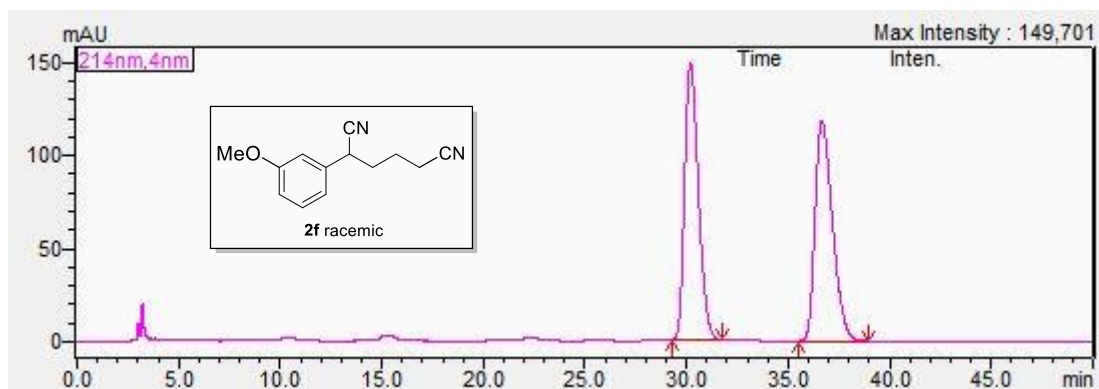
| | | | | |
|--------|--------|---------|--------|----------|
| 1 | 18.237 | 49.938 | 445578 | 18926413 |
| 2 | 21.794 | 50.062 | 369114 | 18973695 |
| Totol: | | 100.000 | 814692 | 37900108 |

Supplementary Figure 177. HPLC data of rac-2e



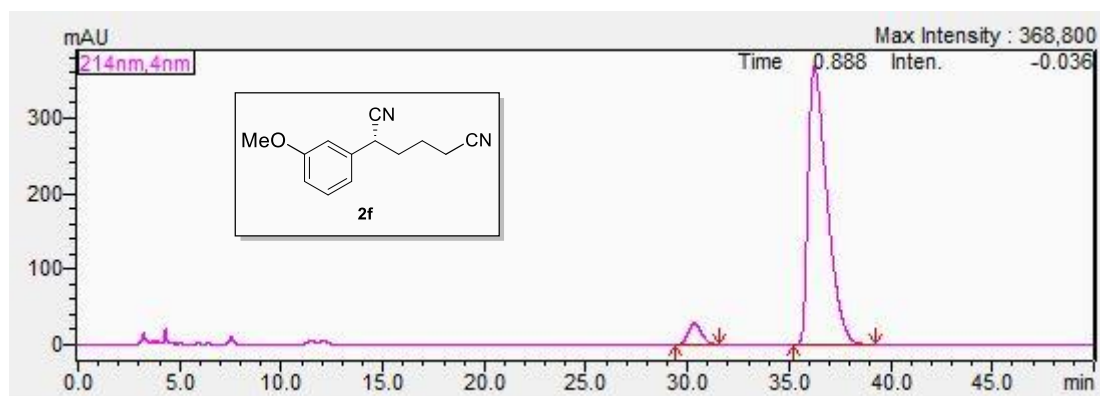
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 18.198 | 94.977 | 546278 | 23390003 |
| 2 | 22.009 | 5.023 | 28322 | 1236980 |
| Totol: | | 100.000 | 574599 | 24626983 |

Supplementary Figure 178. HPLC data of 2e



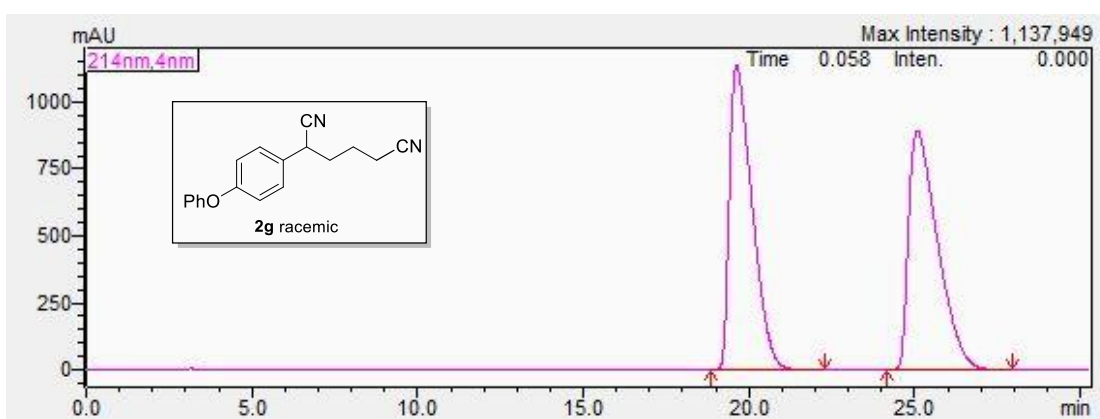
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 30.195 | 49.821 | 148727 | 6998292 |
| 2 | 36.668 | 50.179 | 118283 | 7048545 |
| Totol: | | 100.000 | 267010 | 14046838 |

Supplementary Figure 179. HPLC data of rac-2f



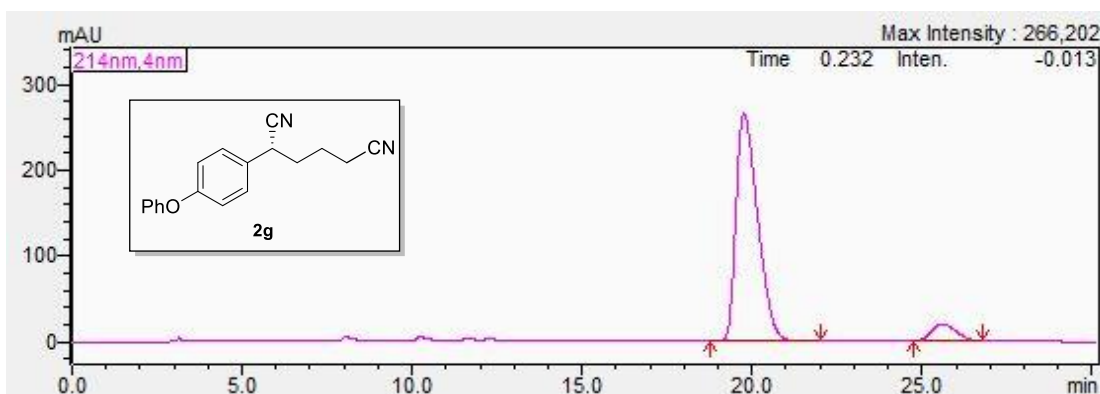
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 30.344 | 5.132 | 28150 | 1287689 |
| 2 | 36.248 | 94.868 | 369147 | 23801454 |
| Total: | | 100.000 | 397297 | 25089144 |

Supplementary Figure 180. HPLC data of **2f**



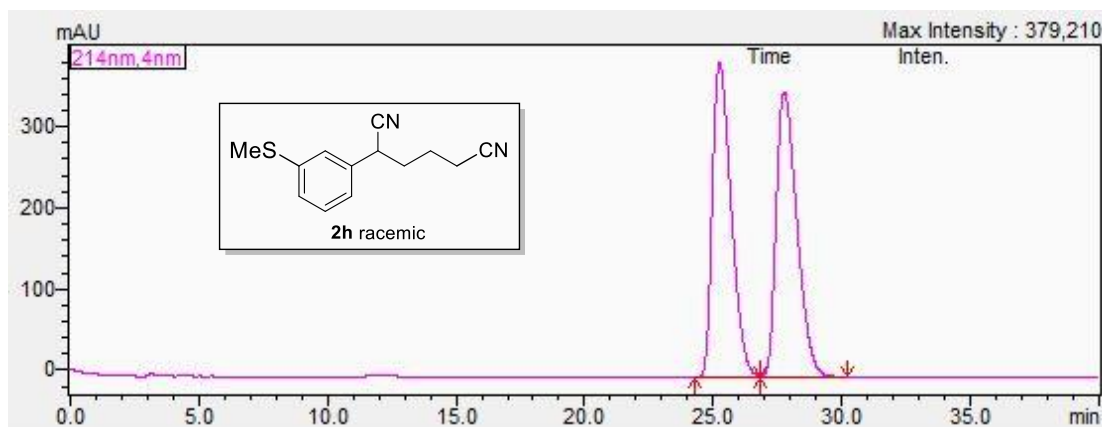
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|-----------|
| 1 | 19.626 | 50.026 | 1137173 | 54153369 |
| 2 | 25.080 | 49.974 | 892649 | 54096018 |
| Total: | | 100.000 | 2029822 | 108249387 |

Supplementary Figure 181. HPLC data of rac-**2g**



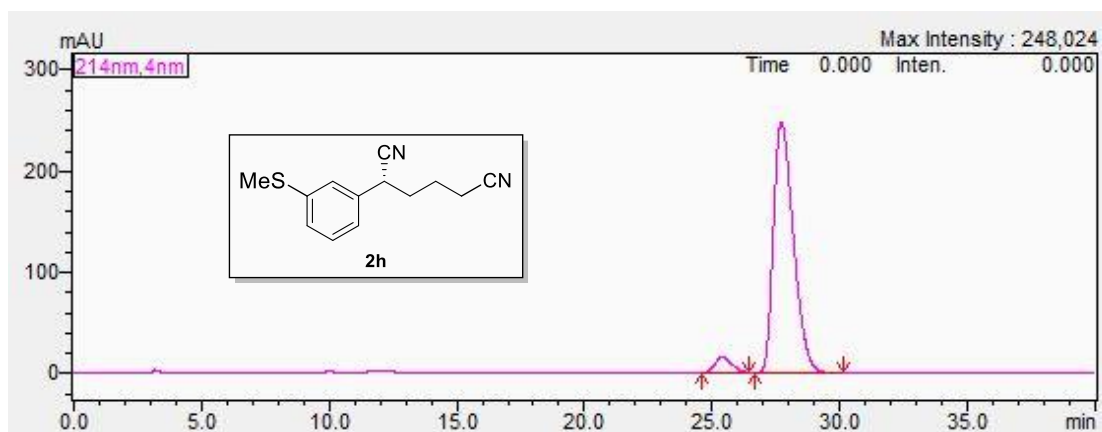
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 19.768 | 92.113 | 266013 | 11788503 |
| 2 | 25.611 | 7.887 | 19922 | 1009358 |
| Total: | | 100.000 | 285935 | 12797861 |

Supplementary Figure 182. HPLC data of **2g**



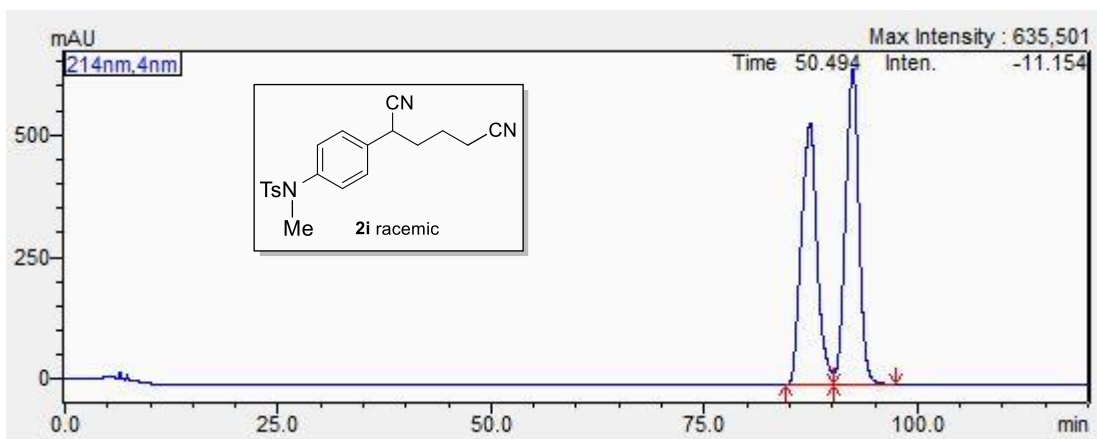
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 25.251 | 49.938 | 387603 | 19641132 |
| 2 | 27.766 | 50.062 | 351511 | 19690091 |
| Total: | | 100.000 | 739114 | 39331223 |

Supplementary Figure 183. HPLC data of rac-**2h**



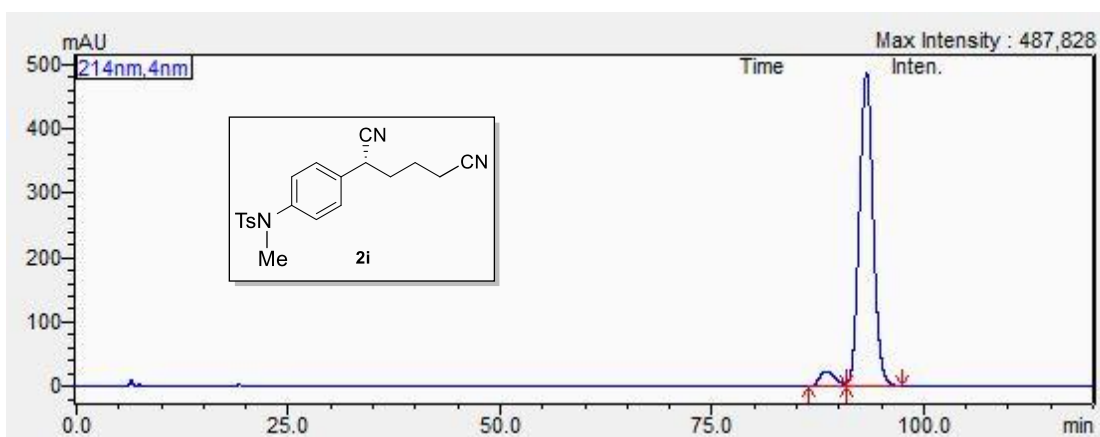
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 25.406 | 4.938 | 15150 | 699557 |
| 2 | 27.717 | 95.062 | 247380 | 13467988 |
| Total: | | 100.000 | 262530 | 14167545 |

Supplementary Figure 184. HPLC data of **2h**



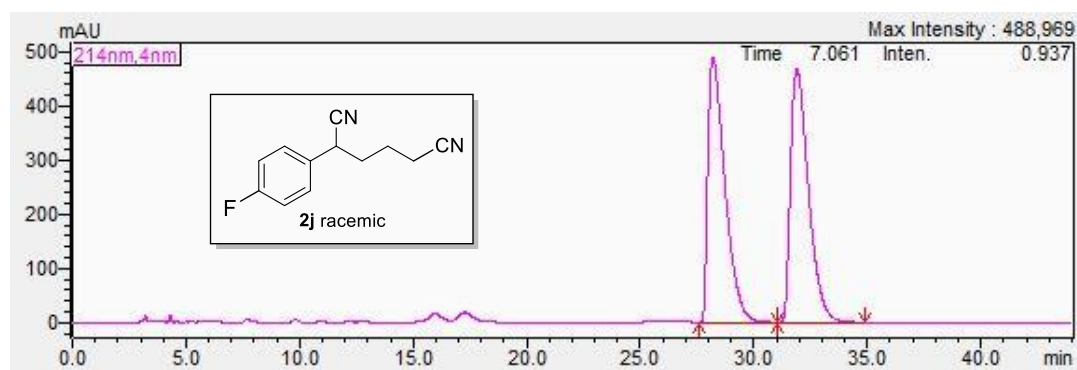
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|-----------|
| 1 | 87.405 | 49.181 | 536227 | 68926239 |
| 2 | 92.439 | 50.819 | 645825 | 71222674 |
| Total: | | 100.000 | 1182052 | 140148913 |

Supplementary Figure 185. HPLC data of rac-**2i**



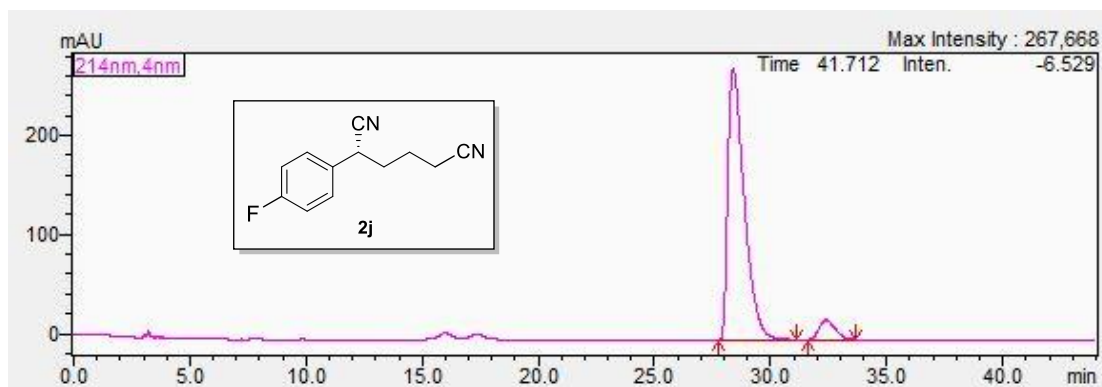
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 88.428 | 5.106 | 23141 | 2975965 |
| 2 | 93.250 | 94.894 | 487477 | 55311320 |
| Total: | | 100.000 | 510618 | 58287285 |

Supplementary Figure 186. HPLC data of **2i**



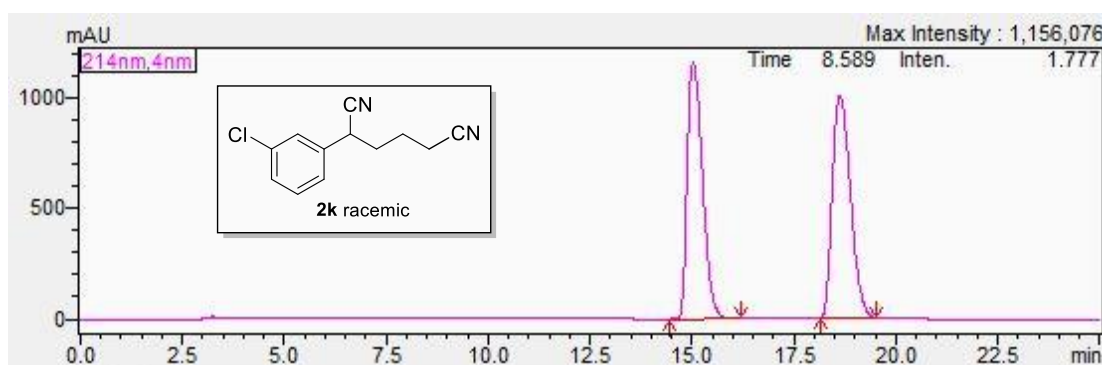
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 28.209 | 49.830 | 488000 | 25533343 |
| 2 | 31.903 | 50.170 | 468146 | 25707213 |
| Totol: | | 100.000 | 956146 | 51240556 |

Supplementary Figure 187. HPLC data of rac-2j



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 28.395 | 93.411 | 273800 | 13400075 |
| 2 | 32.390 | 6.589 | 19842 | 945145 |
| Totol: | | 100.000 | 293642 | 14345220 |

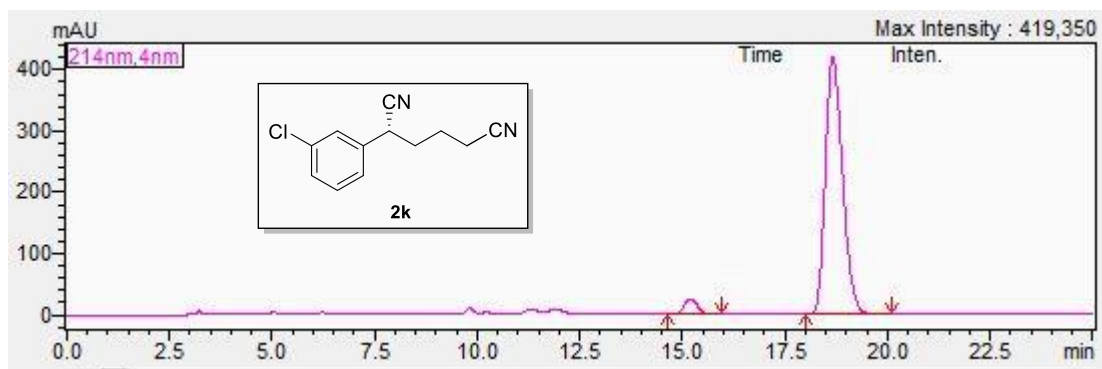
Supplementary Figure 188. HPLC data of 2j



| Peak | Ret. time | Area % | Height | Area |
|------|-----------|--------|---------|----------|
| 1 | 15.030 | 49.243 | 1154683 | 30072261 |

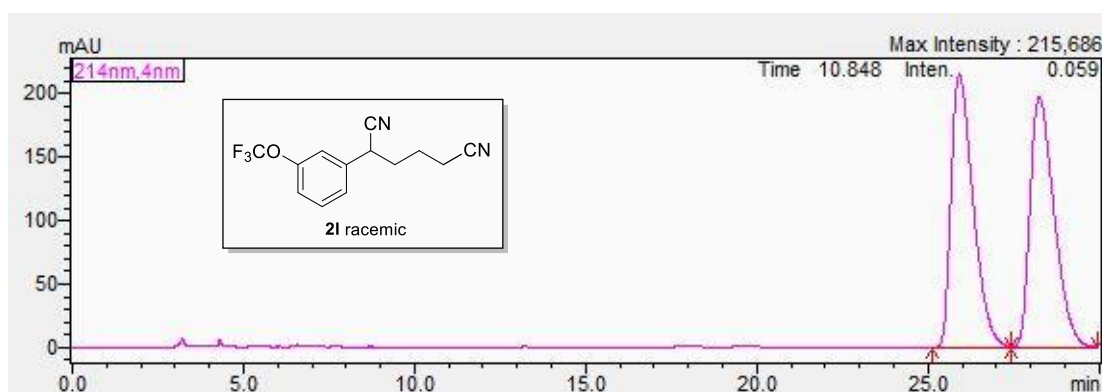
| | | | | |
|--------|--------|---------|---------|----------|
| 2 | 18.631 | 50.757 | 1001967 | 30996533 |
| Total: | | 100.000 | 2156651 | 61068795 |

Supplementary Figure 189. HPLC data of rac-2k



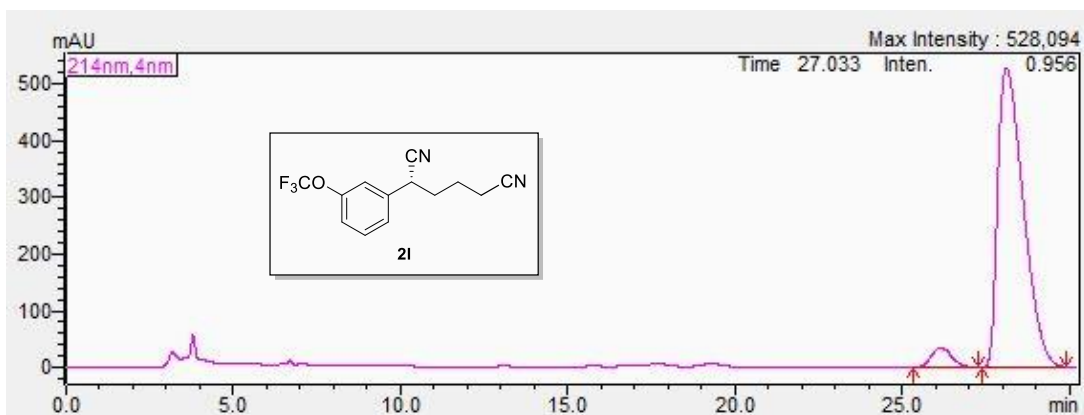
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 15.199 | 4.582 | 25682 | 588358 |
| 2 | 18.672 | 95.418 | 418529 | 12253502 |
| Total: | | 100.000 | 444211 | 12841860 |

Supplementary Figure 190. HPLC data of 2k



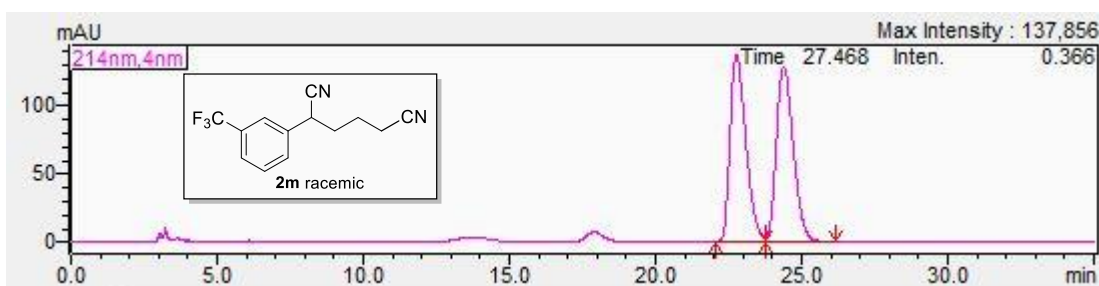
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 25.917 | 50.005 | 215726 | 9676116 |
| 2 | 28.255 | 49.995 | 197350 | 9674355 |
| Total: | | 100.000 | 413076 | 19350471 |

Supplementary Figure 191. HPLC data of rac-2l



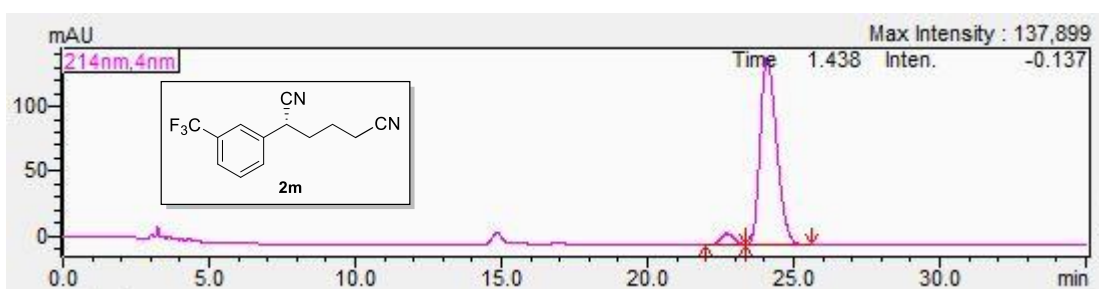
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 26.144 | 4.952 | 34865 | 1461850 |
| 2 | 28.107 | 95.048 | 527220 | 28056044 |
| Total: | | 100.000 | 562085 | 29517894 |

Supplementary Figure 192. HPLC data of **2l**



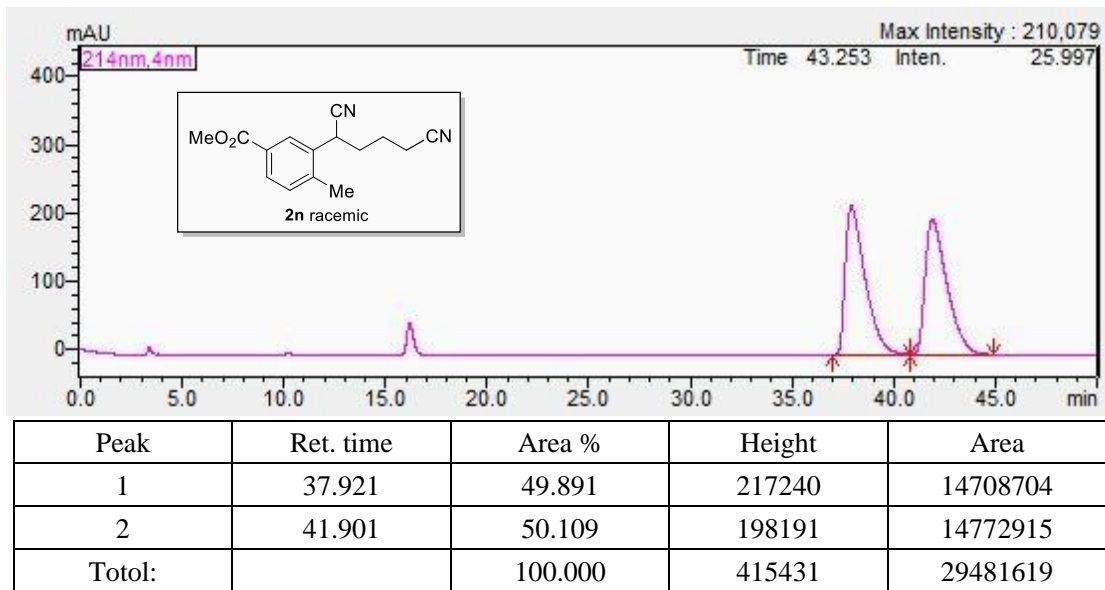
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 22.769 | 49.807 | 137238 | 5166377 |
| 2 | 24.388 | 50.193 | 128646 | 5206397 |
| Total: | | 100.000 | 265884 | 10372774 |

Supplementary Figure 193. HPLC data of rac-**2m**

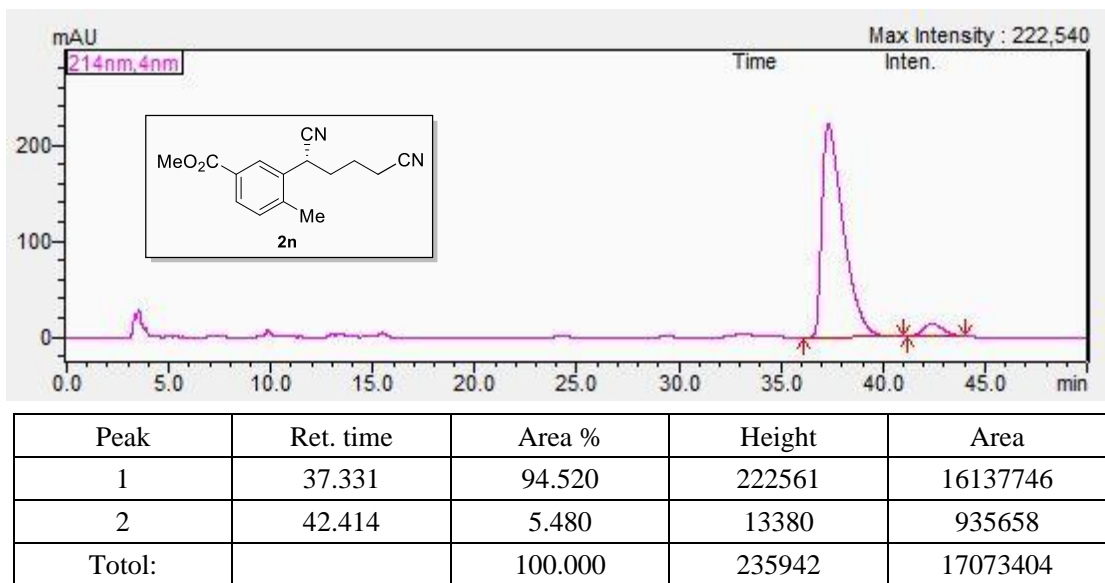


| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|---------|
| 1 | 22.711 | 5.186 | 8953 | 311115 |
| 2 | 24.099 | 94.814 | 145075 | 5687658 |
| Total: | | 100.000 | 154028 | 5998773 |

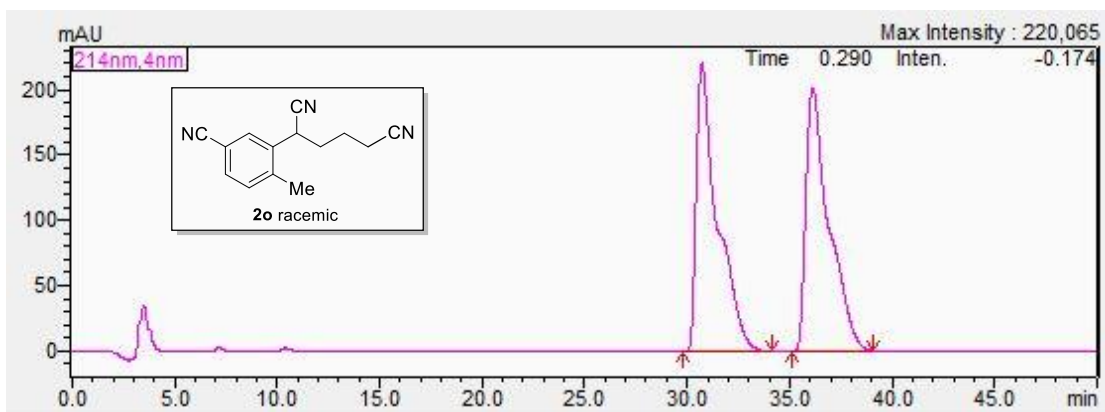
Supplementary Figure 194. HPLC data of **2m**



Supplementary Figure 195. HPLC data of rac-**2n**

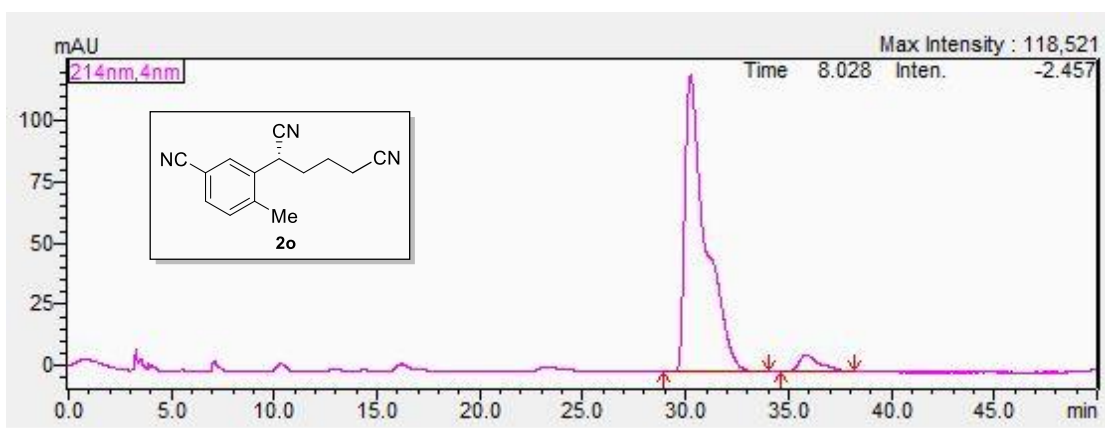


Supplementary Figure 196. HPLC data of **2n**



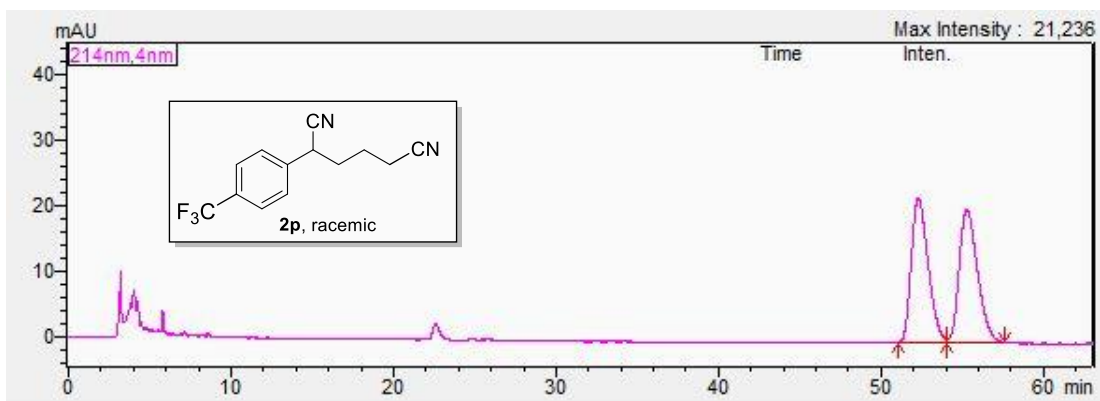
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 30.728 | 50.177 | 220568 | 15823659 |
| 2 | 36.130 | 49.823 | 202173 | 15711789 |
| Total: | | 100.000 | 422741 | 31535448 |

Supplementary Figure 197. HPLC data of rac-**2o**



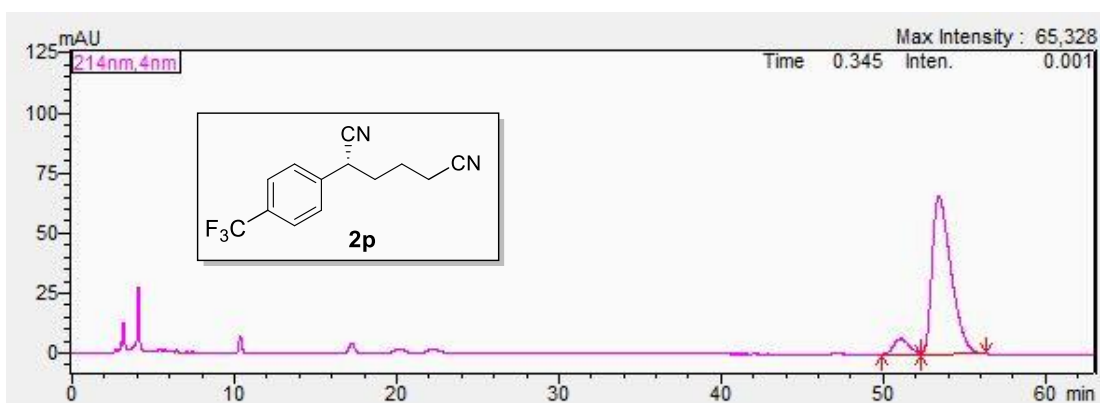
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|---------|
| 1 | 30.239 | 94.459 | 121137 | 8529419 |
| 2 | 35.870 | 5.541 | 6941 | 500312 |
| Total: | | 100.000 | 128078 | 9029731 |

Supplementary Figure 198. HPLC data of **2o**



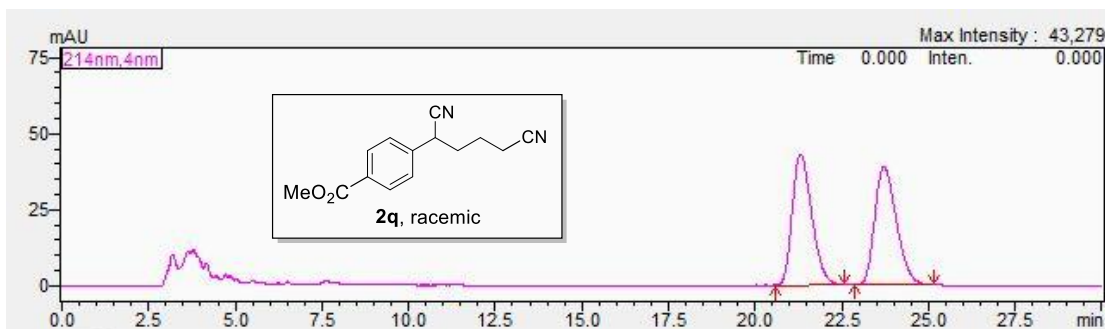
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|---------|
| 1 | 52.292 | 49.880 | 22051 | 1593710 |
| 2 | 55.272 | 50.120 | 20413 | 1601383 |
| Total: | | 100.000 | 42464 | 3195093 |

Supplementary Figure 199. HPLC data of rac-**2p**



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|---------|
| 1 | 51.073 | 7.321 | 6332 | 420558 |
| 2 | 53.432 | 92.679 | 65675 | 5324199 |
| Total: | | 100.000 | 72007 | 5744757 |

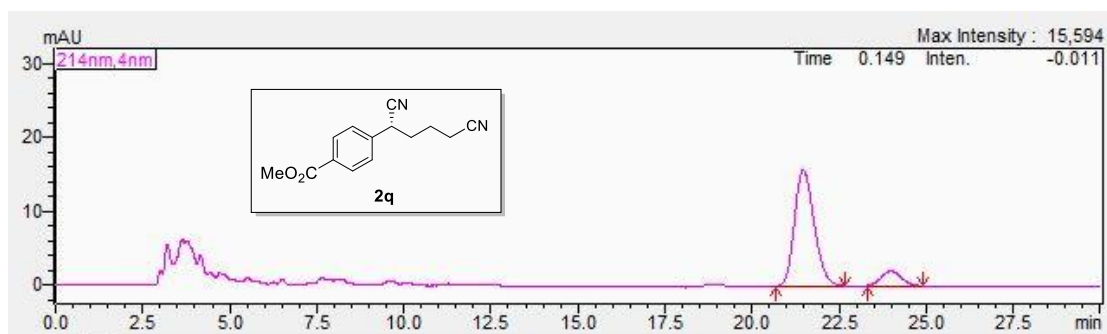
Supplementary Figure 200. HPLC data of **2p**



| Peak | Ret. time | Area % | Height | Area |
|------|-----------|--------|--------|---------|
| 1 | 21.314 | 49.879 | 43090 | 1687552 |

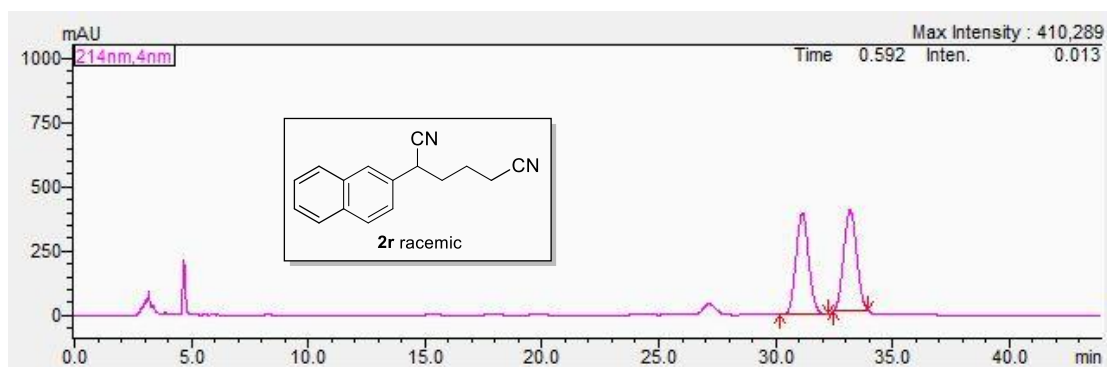
| | | | | |
|--------|--------|---------|-------|---------|
| 2 | 23.719 | 50.121 | 38950 | 1695773 |
| Total: | | 100.000 | 82040 | 3383326 |

Supplementary Figure 201. HPLC data of rac-**2q**



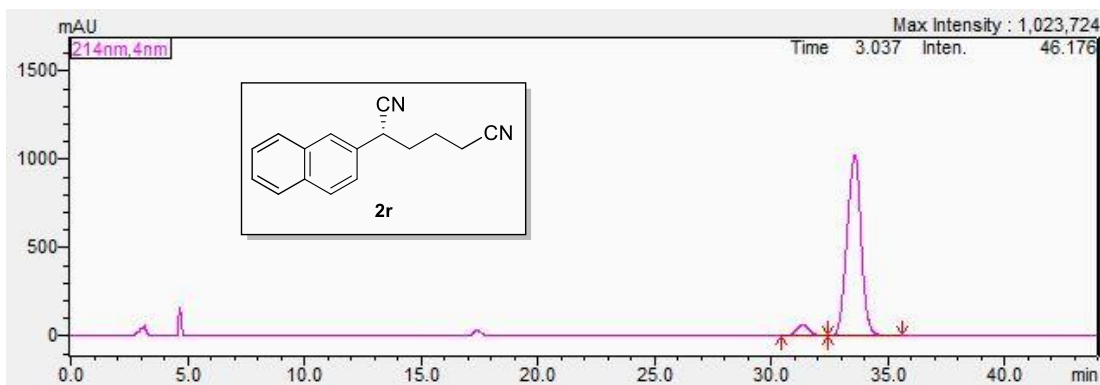
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|--------|
| 1 | 21.474 | 88.187 | 15690 | 615179 |
| 2 | 23.997 | 11.813 | 2002 | 82407 |
| Total: | | 100.000 | 17692 | 697585 |

Supplementary Figure 202. HPLC data of **2q**



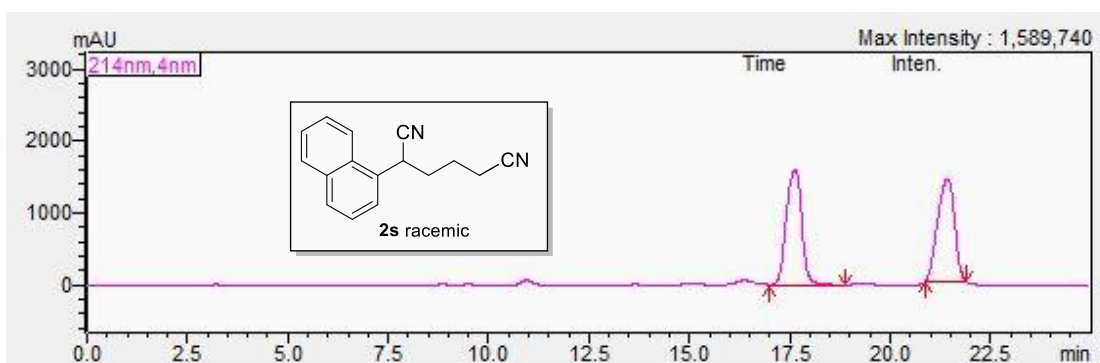
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 31.144 | 49.478 | 394152 | 14766900 |
| 2 | 33.190 | 50.522 | 392866 | 15078214 |
| Total: | | 100.000 | 787018 | 29845114 |

Supplementary Figure 203. HPLC data of rac-**2r**



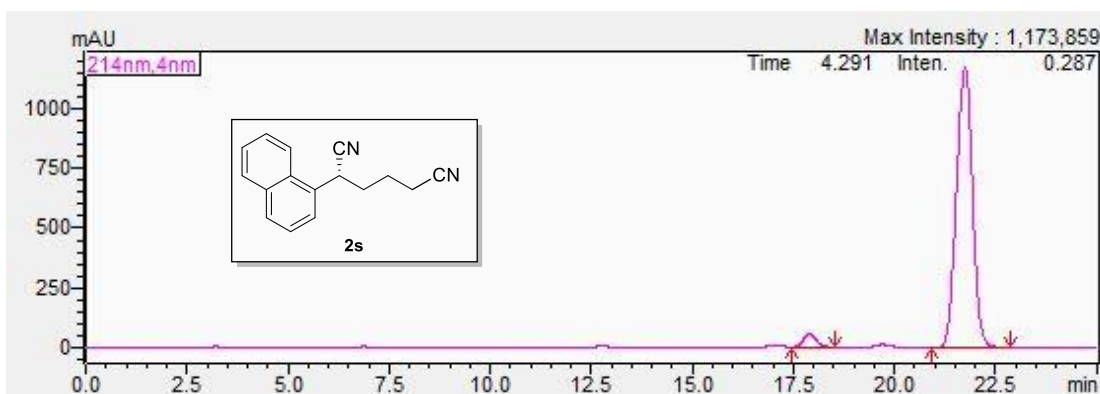
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 31.357 | 5.119 | 60979 | 2325608 |
| 2 | 33.606 | 94.881 | 1022333 | 43101992 |
| Total: | | 100.000 | 1083312 | 45427600 |

Supplementary Figure 204. HPLC data of **2r**



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 17.630 | 49.920 | 1587922 | 42303287 |
| 2 | 21.429 | 50.080 | 1433941 | 42439452 |
| Total: | | 100.000 | 3021863 | 84742739 |

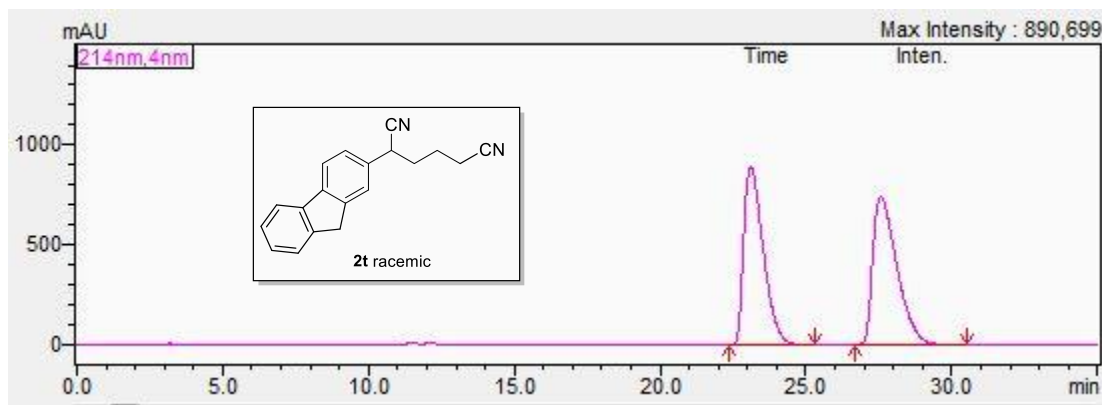
Supplementary Figure 205. HPLC data of rac-**2s**



| Peak | Ret. time | Area % | Height | Area |
|------|-----------|--------|---------|---------|
| 1 | 4.291 | 0.287 | 1173859 | 1173859 |

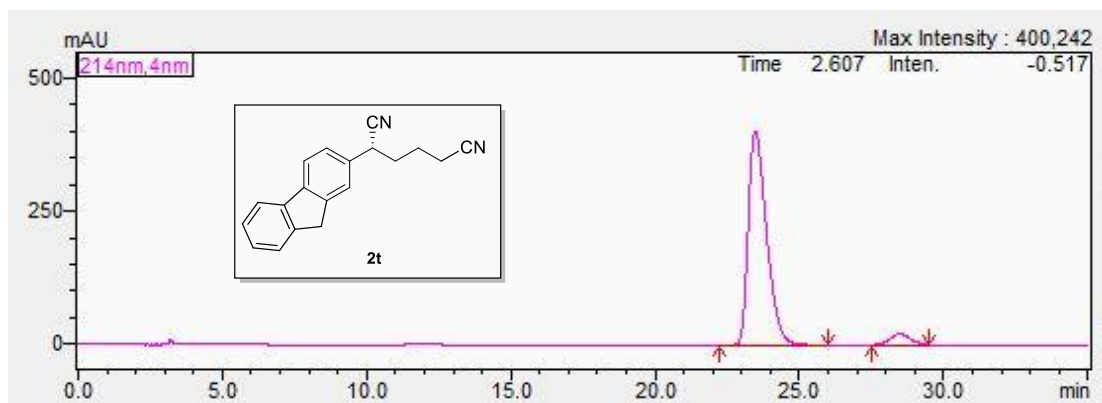
| | | | | |
|--------|--------|---------|---------|----------|
| 1 | 17.893 | 3.429 | 55504 | 1150657 |
| 2 | 21.748 | 96.571 | 1173178 | 32410824 |
| Totol: | | 100.000 | 1228682 | 33561482 |

Supplementary Figure 206. HPLC data of **2s**



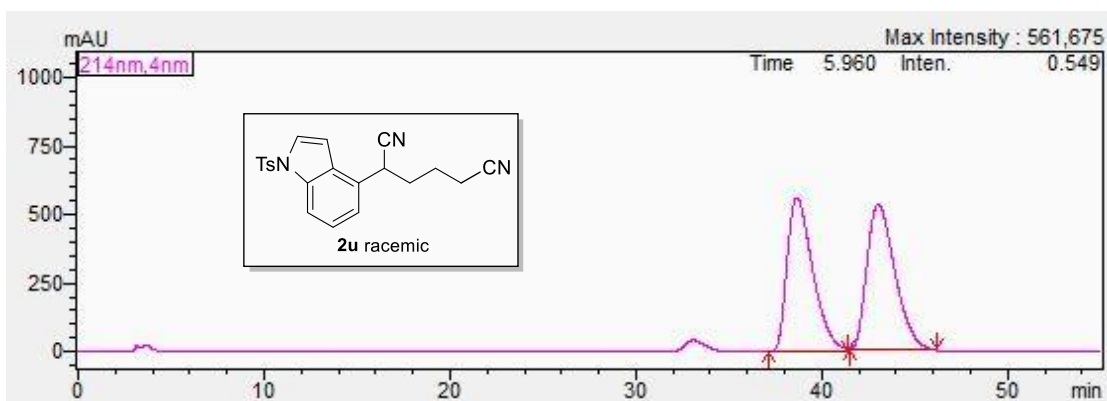
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 23.109 | 49.471 | 890346 | 41525070 |
| 2 | 27.571 | 50.529 | 740700 | 42412671 |
| Totol: | | 100.000 | 1631047 | 83937741 |

Supplementary Figure 207. HPLC data of rac-**2t**



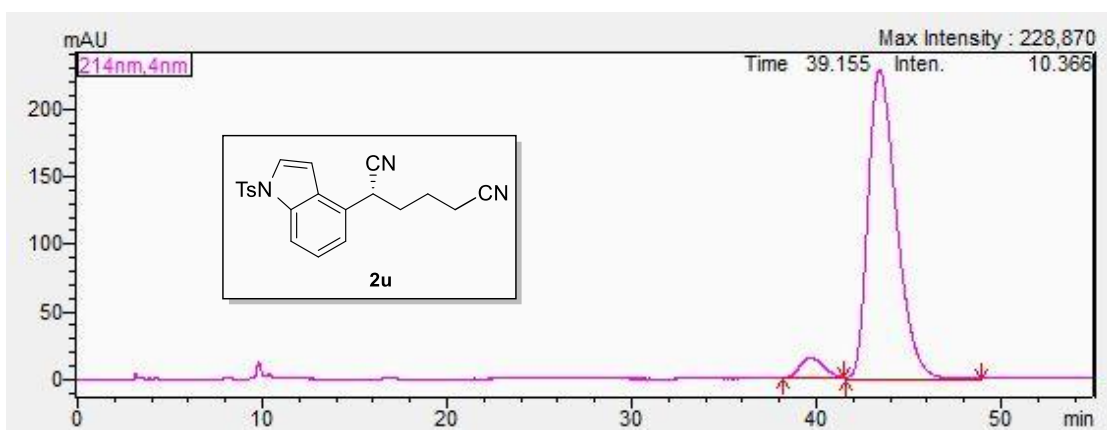
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 23.479 | 94.782 | 401146 | 17629646 |
| 2 | 28.475 | 5.218 | 19786 | 970623 |
| Totol: | | 100.000 | 420933 | 18600268 |

Supplementary Figure 208. HPLC data of **2t**



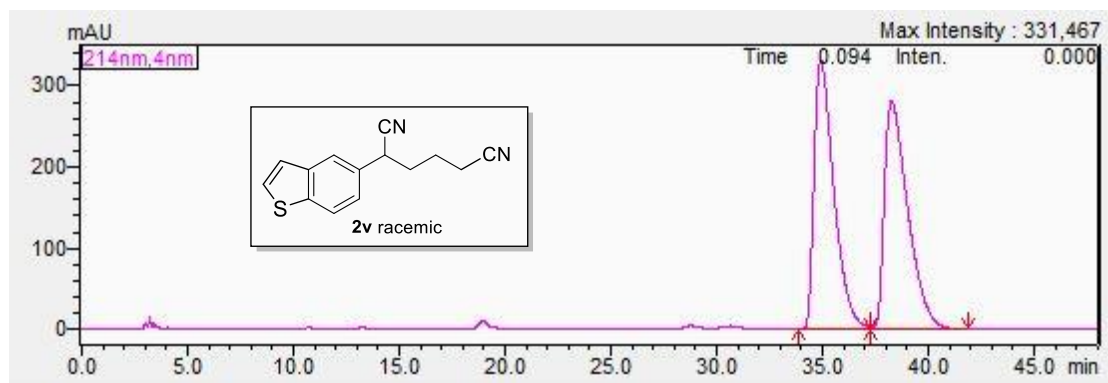
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|-----------|
| 1 | 38.653 | 49.412 | 561909 | 54258269 |
| 2 | 43.032 | 50.588 | 529866 | 55550667 |
| Total: | | 100.000 | 1091775 | 109808937 |

Supplementary Figure 209. HPLC data of rac-**2u**



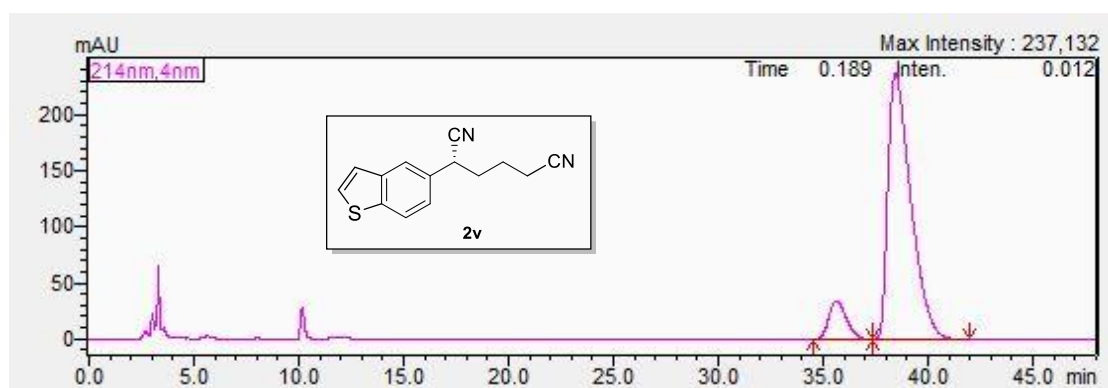
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 39.691 | 5.490 | 15362 | 1418277 |
| 2 | 43.442 | 94.510 | 228499 | 24413604 |
| Total: | | 100.000 | 243861 | 25831880 |

Supplementary Figure 210. HPLC data of **2u**



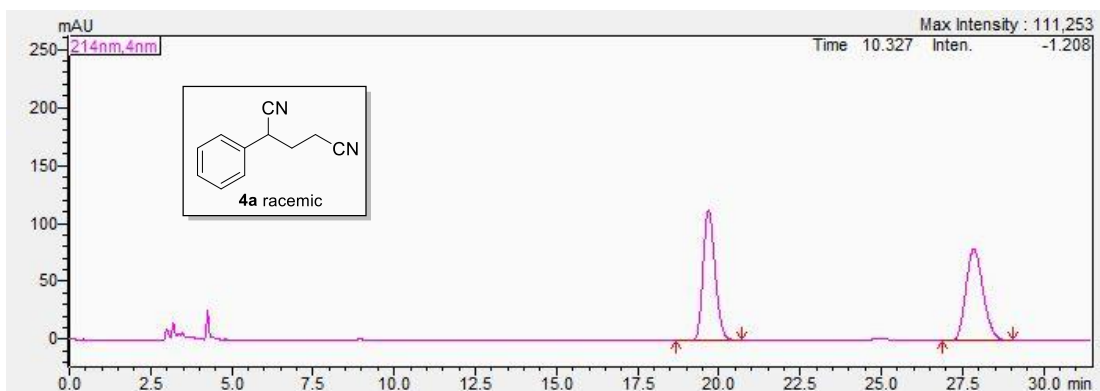
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 34.918 | 49.918 | 330654 | 21469335 |
| 2 | 38.271 | 50.082 | 280114 | 21540206 |
| Total: | | 100.000 | 610768 | 43009541 |

Supplementary Figure 211. HPLC data of rac-**2v**



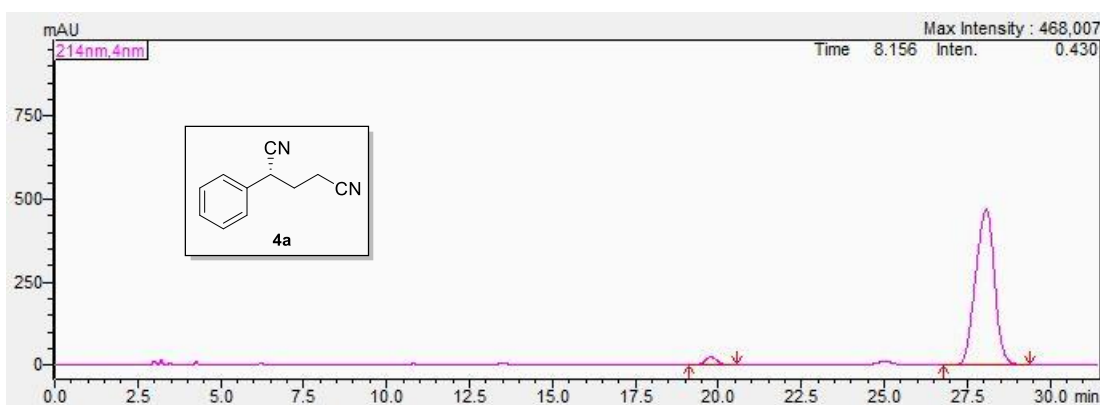
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 35.620 | 10.200 | 34455 | 2050545 |
| 2 | 38.441 | 89.800 | 237835 | 18051911 |
| Total: | | 100.000 | 272290 | 20102456 |

Supplementary Figure 212. HPLC data of **2v**



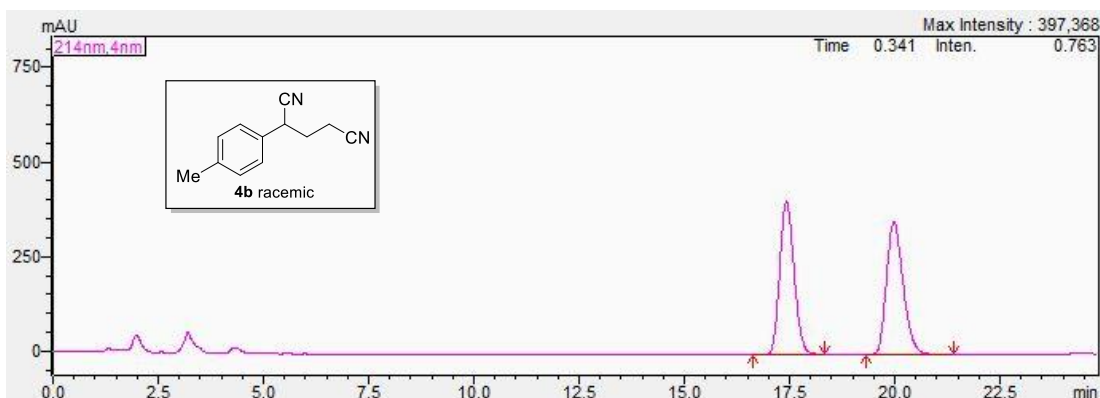
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|---------|
| 1 | 19.683 | 49.936 | 112292 | 2893054 |
| 2 | 27.849 | 50.064 | 79035 | 2900517 |
| Total: | | 100.000 | 191327 | 5793570 |

Supplementary Figure 213. HPLC data of rac-4a



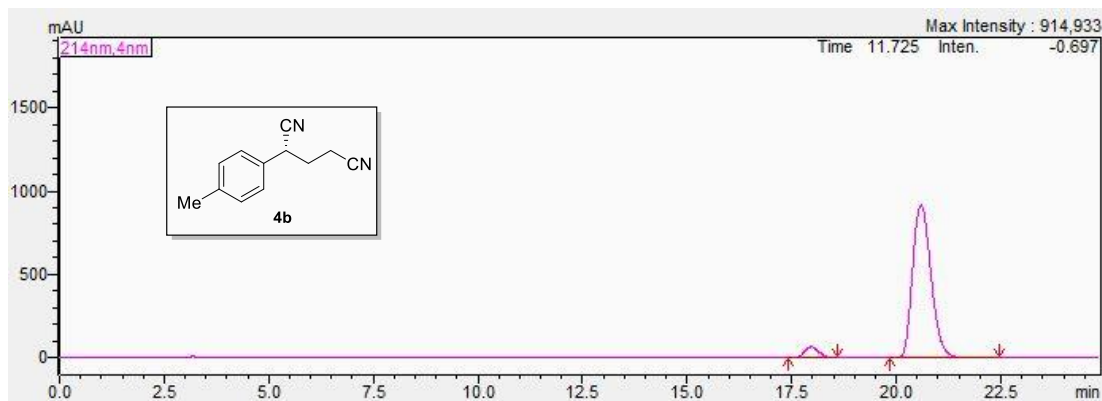
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 19.769 | 3.218 | 23998 | 615140 |
| 2 | 28.074 | 96.782 | 467582 | 18502967 |
| Total: | | 100.000 | 491579 | 19118107 |

Supplementary Figure 214. HPLC data of 4a



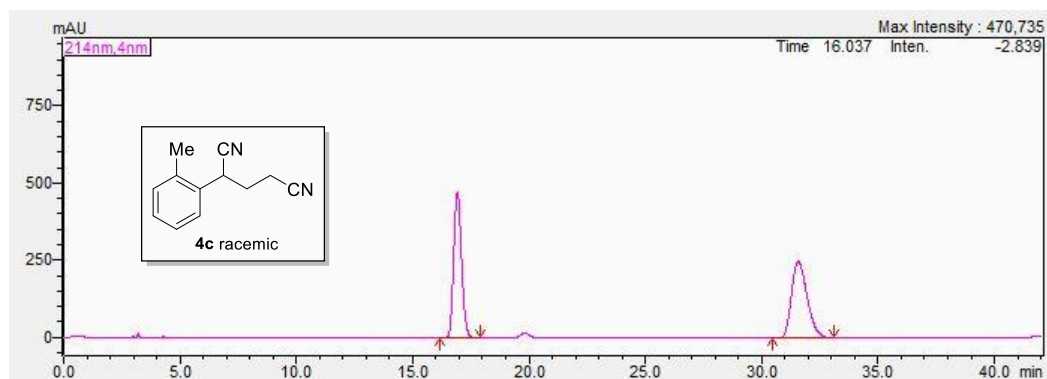
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 17.428 | 49.776 | 405705 | 9525417 |
| 2 | 19.982 | 50.224 | 348130 | 9611207 |
| Total: | | 100.000 | 753835 | 19136624 |

Supplementary Figure 215. HPLC data of rac-4b



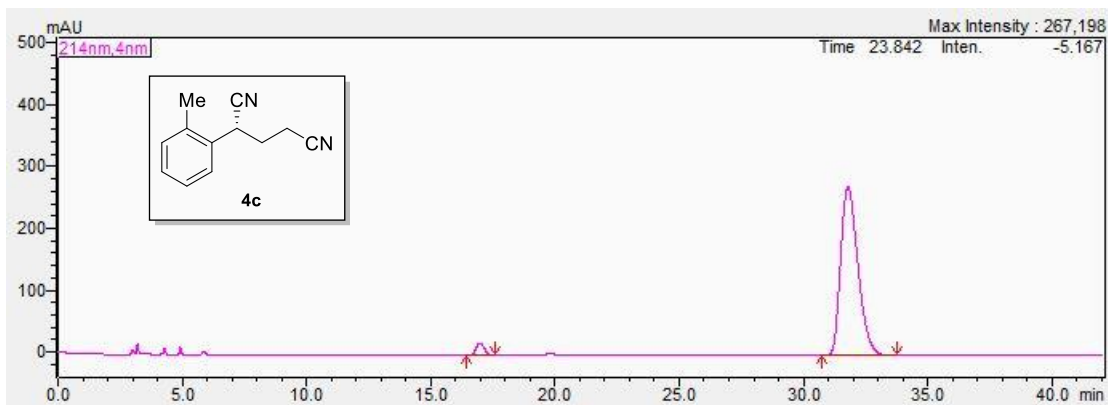
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 17.970 | 5.286 | 65706 | 1575165 |
| 2 | 20.592 | 94.714 | 915454 | 28221854 |
| Total: | | 100.000 | 981160 | 29797019 |

Supplementary Figure 216. HPLC data of 4b



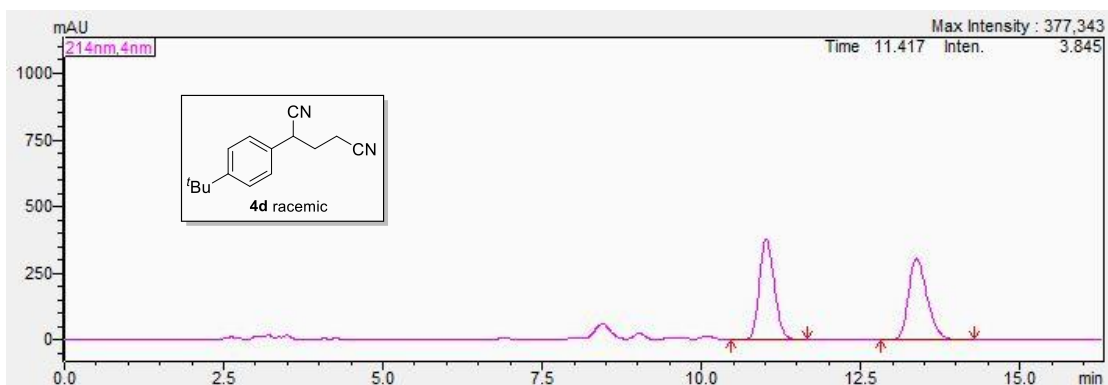
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 16.919 | 49.429 | 473320 | 11367944 |
| 2 | 31.567 | 50.571 | 248812 | 11630440 |
| Total: | | 100.000 | 722132 | 22998384 |

Supplementary Figure 217. HPLC data of rac-4c



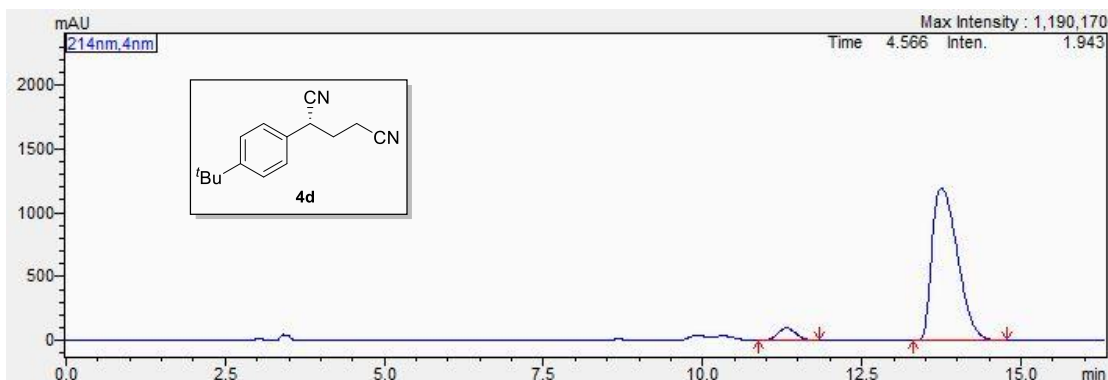
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 16.984 | 3.445 | 19793 | 468556 |
| 2 | 31.797 | 96.555 | 272589 | 13133932 |
| Total: | | 100.000 | 292382 | 13602487 |

Supplementary Figure 218. HPLC data of **4c**



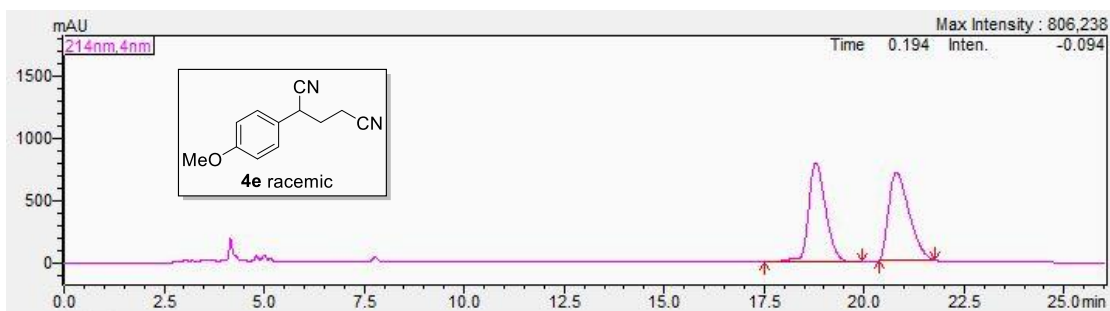
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 11.017 | 49.818 | 376890 | 6143830 |
| 2 | 13.377 | 50.182 | 303617 | 6188718 |
| Total: | | 100.000 | 680507 | 12332548 |

Supplementary Figure 219. HPLC data of rac-**4d**



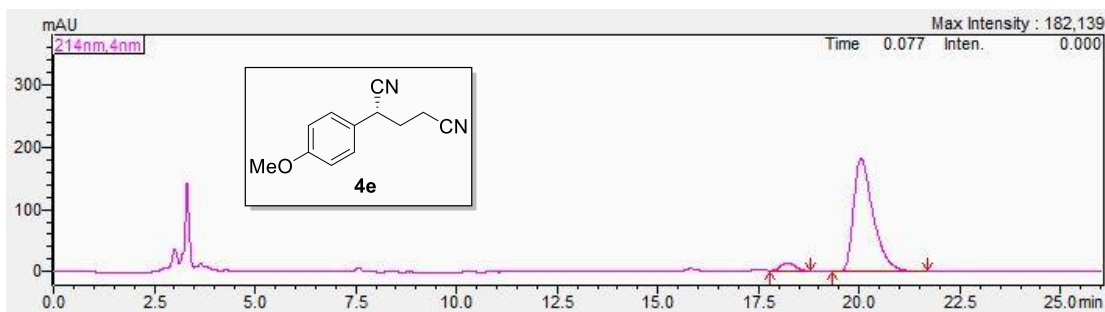
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 11.314 | 5.472 | 96692 | 1928361 |
| 2 | 13.752 | 94.528 | 1189249 | 33312362 |
| Total: | | 100.000 | 1285941 | 35240723 |

Supplementary Figure 220. HPLC data of **4d**



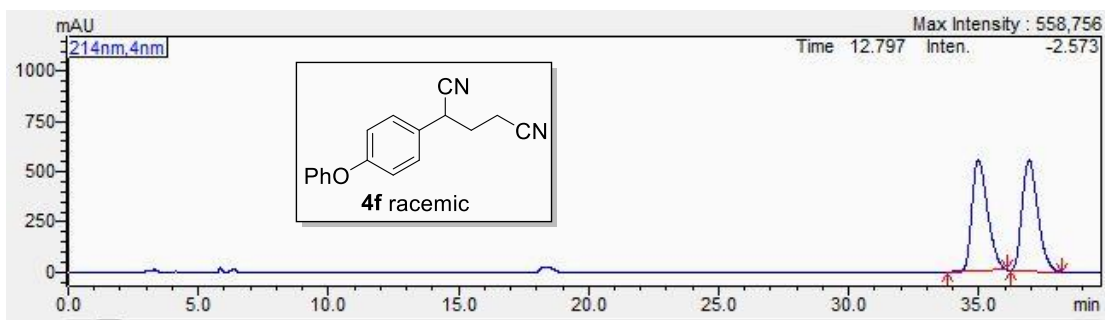
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 18.794 | 49.328 | 800897 | 24073404 |
| 2 | 20.810 | 50.672 | 702330 | 24728972 |
| Total: | | 100.000 | 1503226 | 48802376 |

Supplementary Figure 221. HPLC data of rac-**4e**



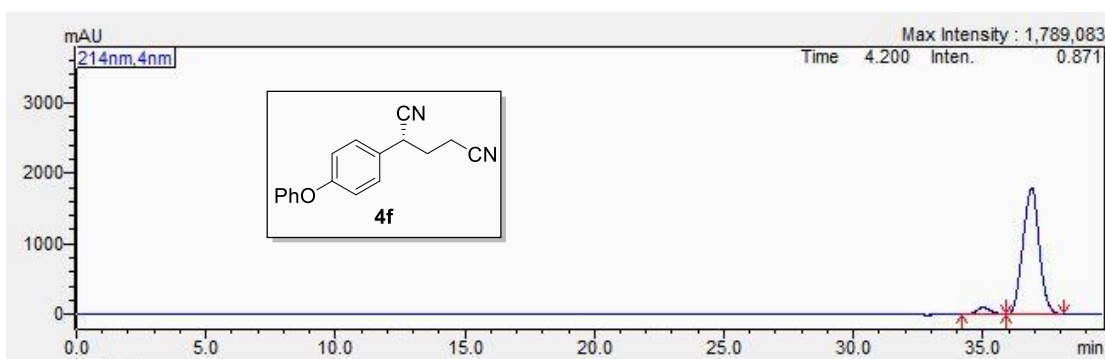
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|---------|
| 1 | 18.223 | 5.131 | 13393 | 332771 |
| 2 | 20.046 | 94.869 | 181767 | 6152745 |
| Total: | | 100.000 | 195160 | 6485516 |

Supplementary Figure 222. HPLC data of **4e**



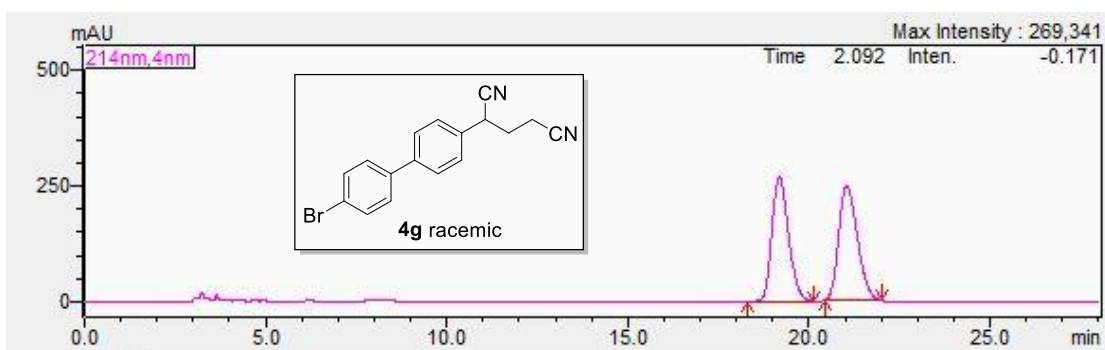
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 34.985 | 50.131 | 553878 | 23348401 |
| 2 | 36.931 | 49.869 | 552318 | 23226652 |
| Total: | | 100.000 | 1106197 | 46575053 |

Supplementary Figure 223. HPLC data of rac-4f



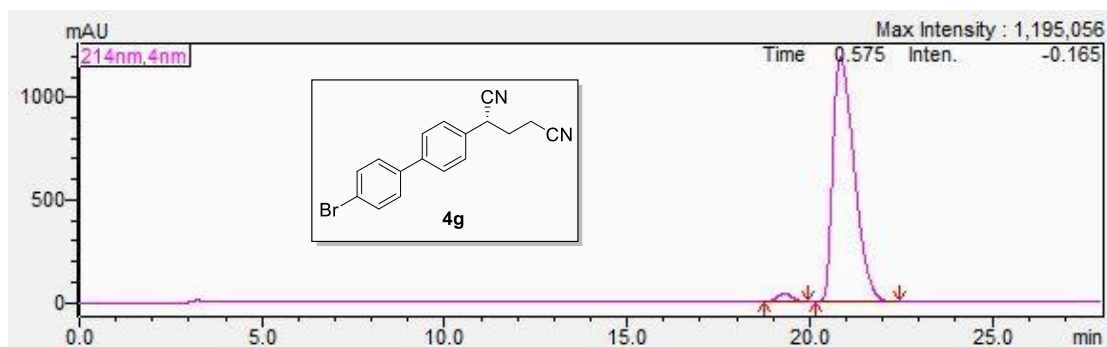
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 35.016 | 4.388 | 92178 | 3619634 |
| 2 | 36.894 | 95.612 | 1784119 | 78876356 |
| Total: | | 100.000 | 1876297 | 82495990 |

Supplementary Figure 224. HPLC data of 4f



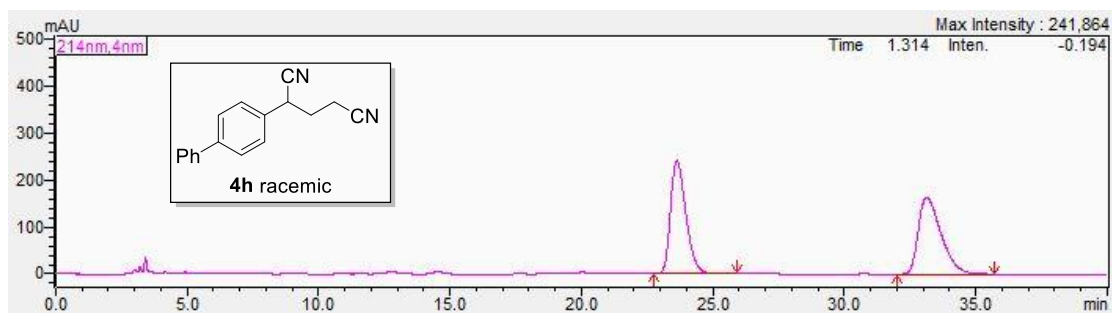
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 19.181 | 49.056 | 268396 | 8545400 |
| 2 | 21.045 | 50.944 | 248521 | 8874453 |
| Total: | | 100.000 | 516917 | 17419853 |

Supplementary Figure 225. HPLC data of rac-4g



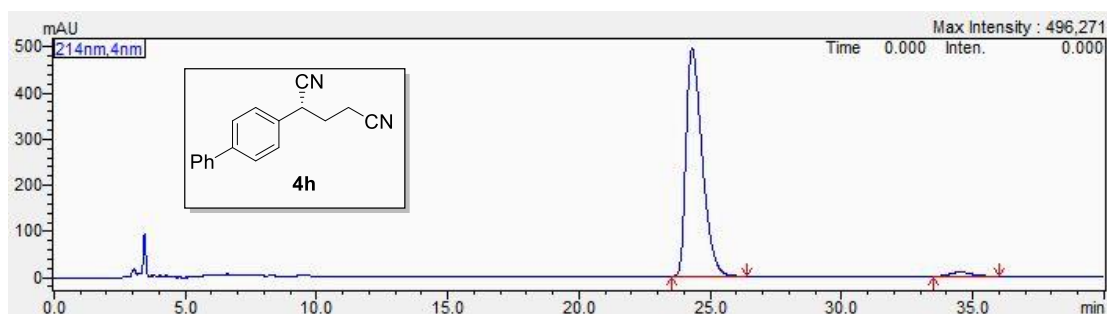
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 19.309 | 2.219 | 38118 | 1108224 |
| 2 | 20.850 | 97.781 | 1193211 | 48834361 |
| Totol: | | 100.000 | 1231329 | 49942586 |

Supplementary Figure 226. HPLC data of 4g



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 23.629 | 49.996 | 241600 | 9824775 |
| 2 | 33.134 | 50.004 | 164087 | 9826156 |
| Totol: | | 100.000 | 405687 | 19650931 |

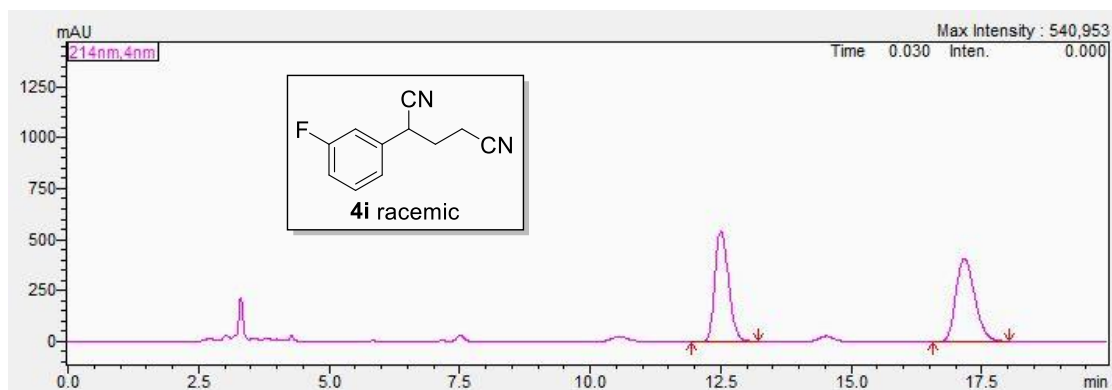
Supplementary Figure 227. HPLC data of rac-4h



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 23.629 | 49.996 | 496271 | 9824775 |
| 2 | 33.134 | 50.004 | 164087 | 9826156 |
| Totol: | | 100.000 | 660358 | 19650931 |

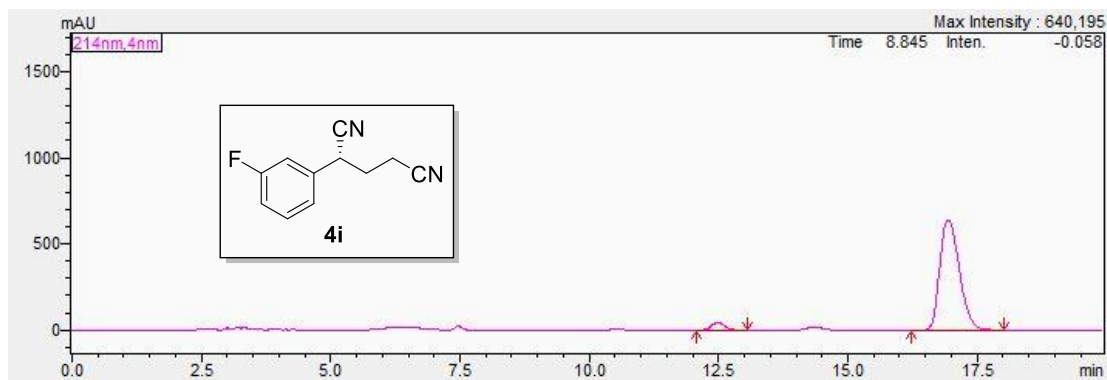
| | | | | |
|--------|--------|---------|--------|----------|
| 1 | 24.306 | 97.709 | 493055 | 21018489 |
| 2 | 34.523 | 2.291 | 8674 | 492805 |
| Totol: | | 100.000 | 501730 | 21511295 |

Supplementary Figure 228. HPLC data of **4h**



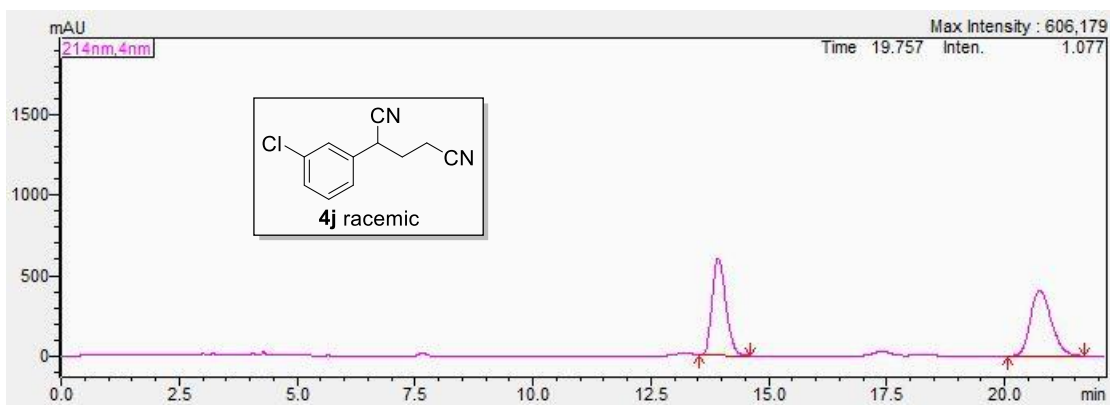
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 12.505 | 49.143 | 540240 | 9831993 |
| 2 | 17.163 | 50.857 | 405770 | 10175057 |
| Totol: | | 100.000 | 946010 | 20007050 |

Supplementary Figure 229. HPLC data of rac-**4i**



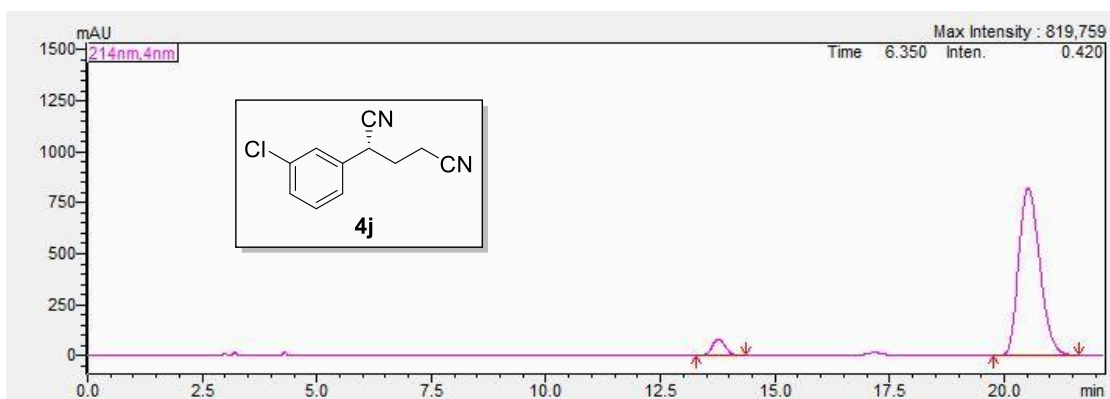
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 12.483 | 4.360 | 44777 | 770893 |
| 2 | 16.942 | 95.640 | 639849 | 16909681 |
| Totol: | | 100.000 | 684626 | 17680574 |

Supplementary Figure 230. HPLC data of **4i**



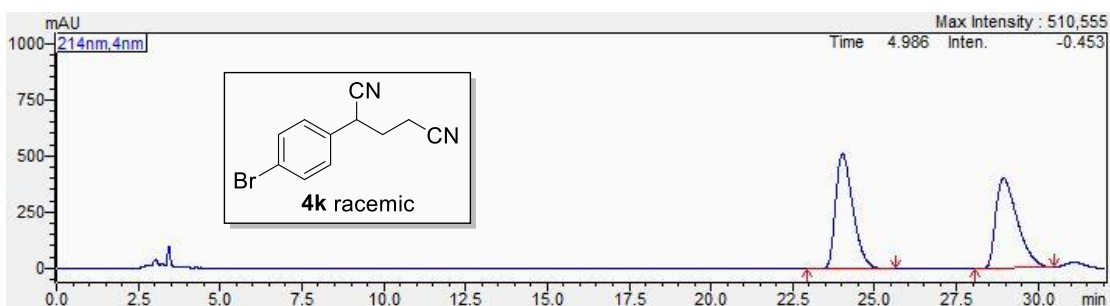
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 13.928 | 49.282 | 600345 | 12090757 |
| 2 | 20.743 | 50.718 | 407201 | 12443113 |
| Total: | | 100.000 | 1007546 | 24533870 |

Supplementary Figure 231. HPLC data of rac-4j



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 13.762 | 5.493 | 79956 | 1531043 |
| 2 | 20.523 | 94.507 | 819054 | 26343527 |
| Total: | | 100.000 | 899010 | 27874570 |

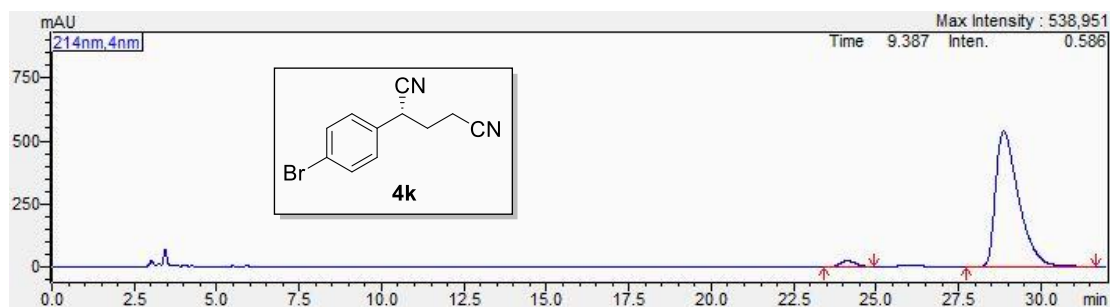
Supplementary Figure 232. HPLC data of 4j



| Peak | Ret. time | Area % | Height | Area |
|------|-----------|--------|--------|------|
| 1 | 24.986 | | | |
| 2 | 29.453 | | | |

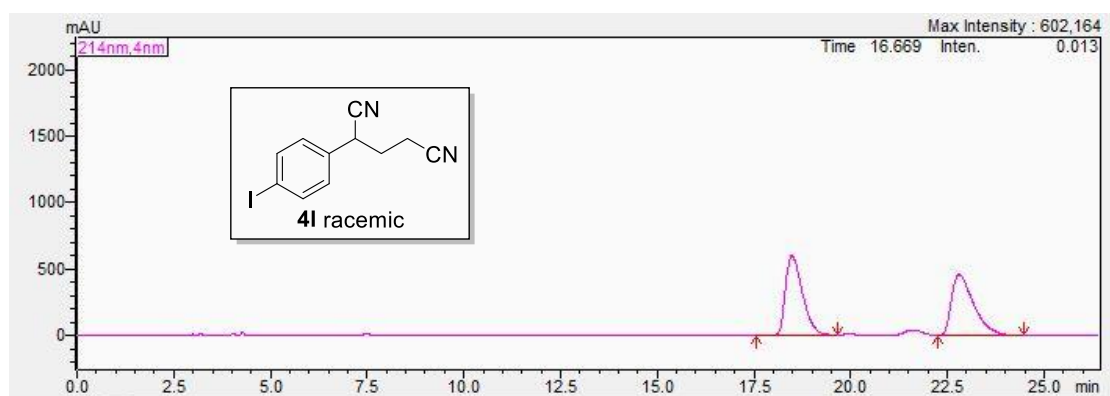
| | | | | |
|--------|--------|---------|--------|----------|
| 1 | 24.020 | 50.609 | 512053 | 18861078 |
| 2 | 28.941 | 49.391 | 401610 | 18406820 |
| Totol: | | 100.000 | 913663 | 37267898 |

Supplementary Figure 233. HPLC data of rac-4k



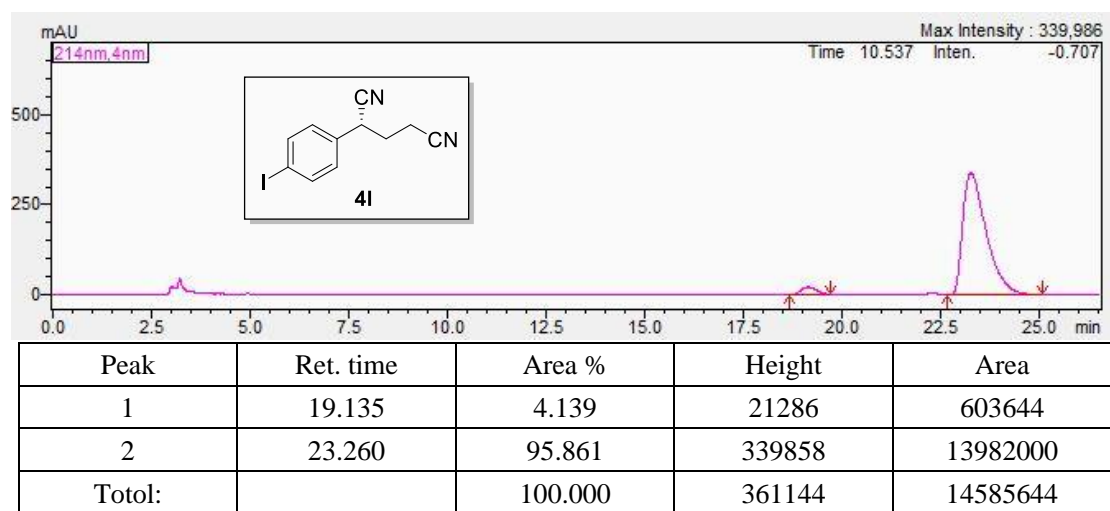
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 24.129 | 3.190 | 25159 | 870019 |
| 2 | 28.871 | 96.810 | 538977 | 26404801 |
| Totol: | | 100.000 | 564136 | 27274820 |

Supplementary Figure 234. HPLC data of 4k

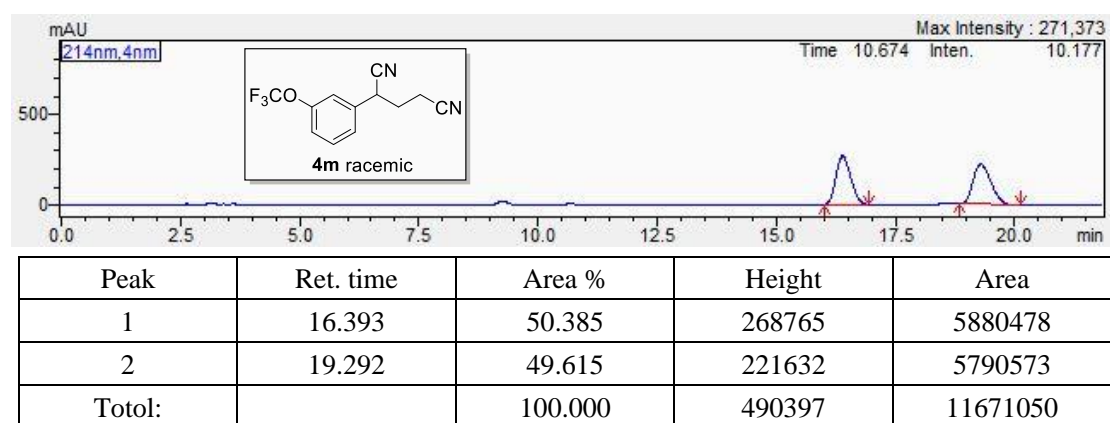


| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 18.484 | 49.673 | 598553 | 18095779 |
| 2 | 22.806 | 50.327 | 457300 | 18333829 |
| Totol: | | 100.000 | 1055853 | 36429608 |

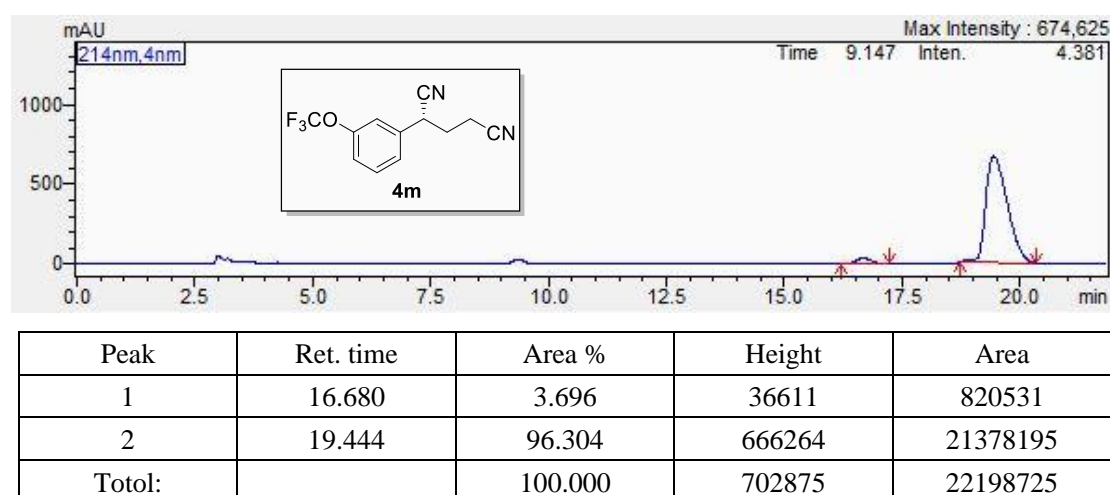
Supplementary Figure 235. HPLC data of rac-4l



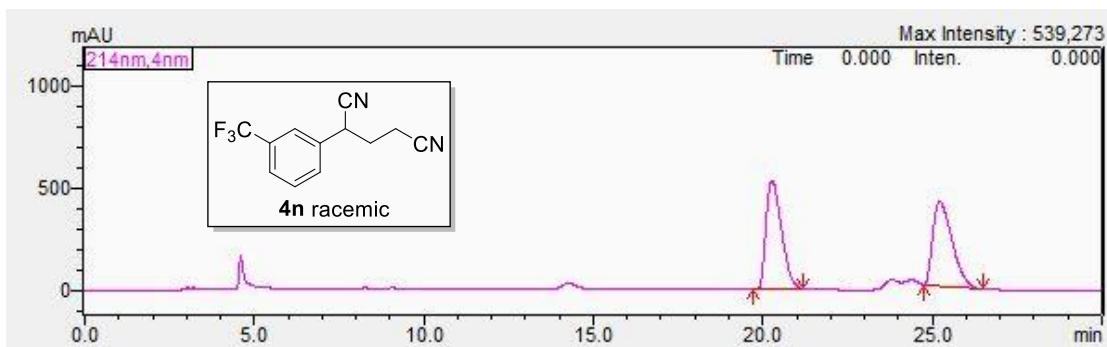
Supplementary Figure 236. HPLC data of **4l**



Supplementary Figure 237. HPLC data of rac-**4m**

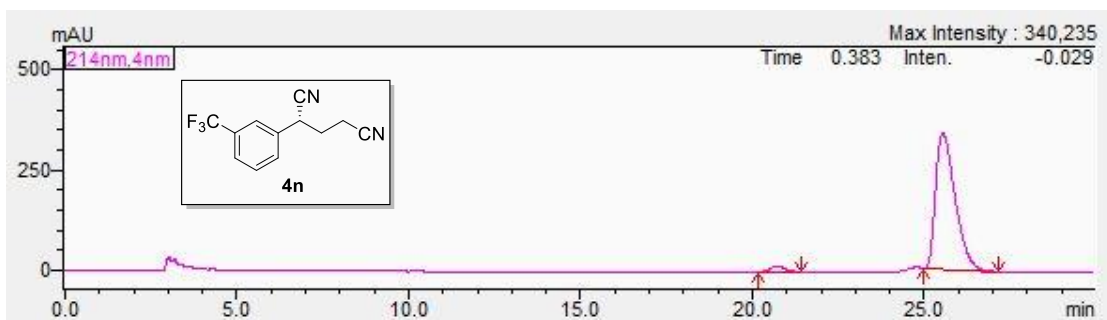


Supplementary Figure 238. HPLC data of **4m**



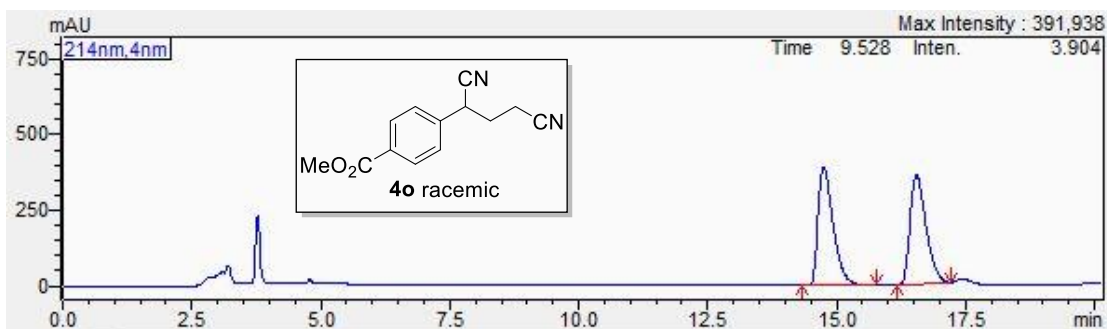
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 20.272 | 50.968 | 531718 | 16962366 |
| 2 | 25.226 | 49.032 | 414151 | 16317902 |
| Totol: | | 100.000 | 945869 | 33280268 |

Supplementary Figure 239. HPLC data of rac-4n



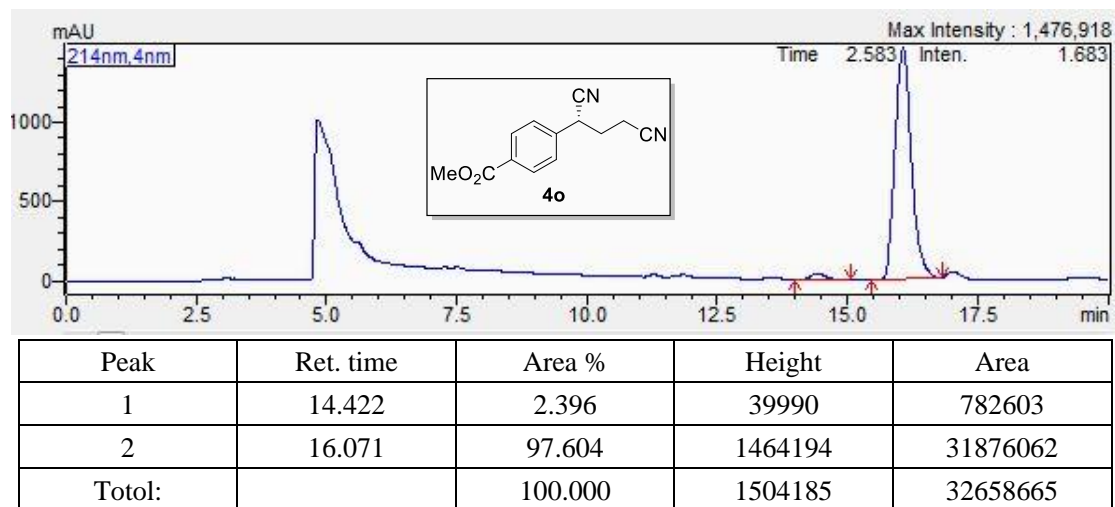
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 20.727 | 2.969 | 14274 | 408977 |
| 2 | 25.545 | 97.031 | 337800 | 13365589 |
| Totol: | | 100.000 | 352074 | 13774566 |

Supplementary Figure 240. HPLC data of 4n

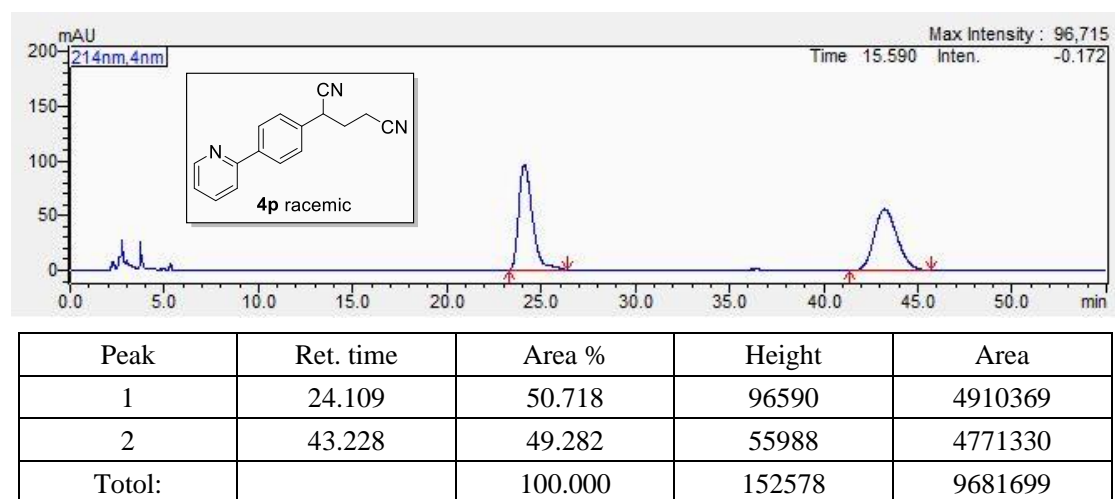


| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 14.747 | 50.337 | 388466 | 7865032 |
| 2 | 16.547 | 49.663 | 358321 | 7759843 |
| Totol: | | 100.000 | 746787 | 15624875 |

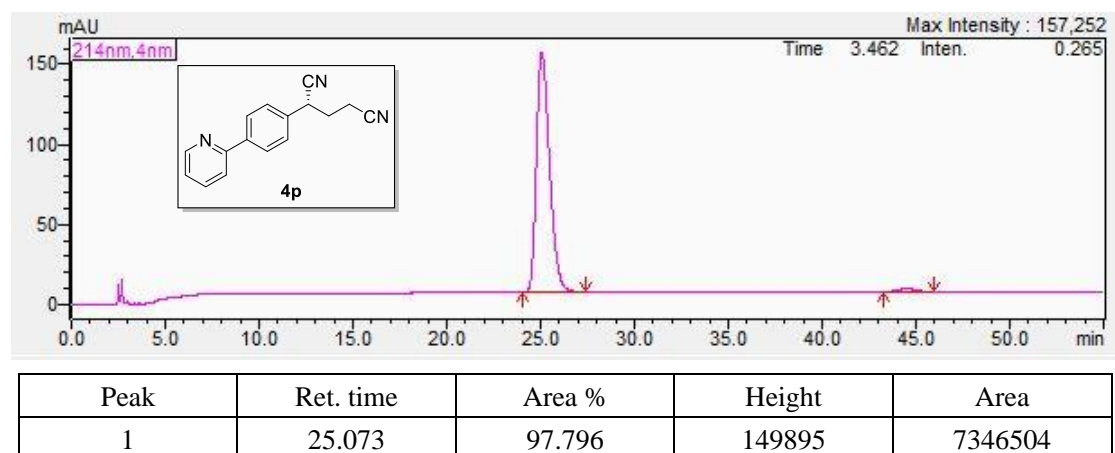
Supplementary Figure 241. HPLC data of rac-4o



Supplementary Figure 242. HPLC data of 4o

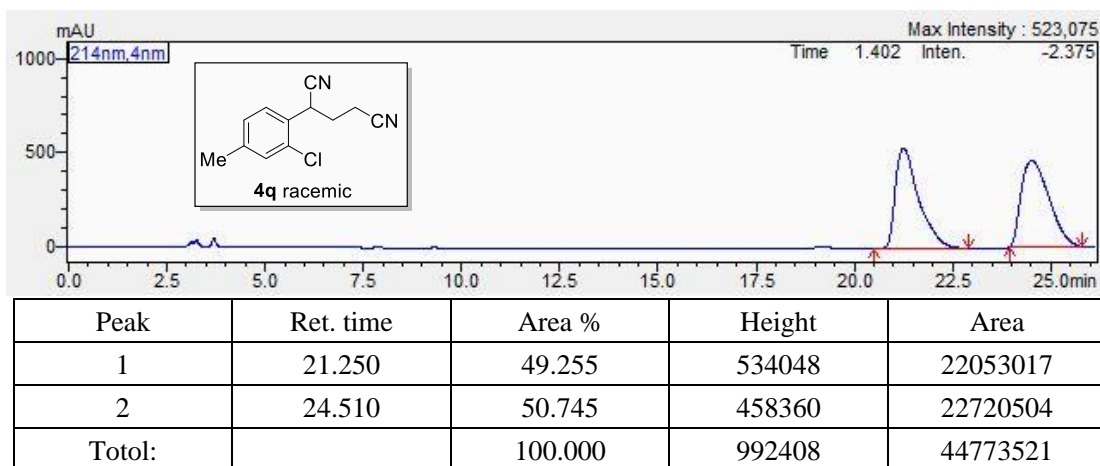


Supplementary Figure 243. HPLC data of rac-4p

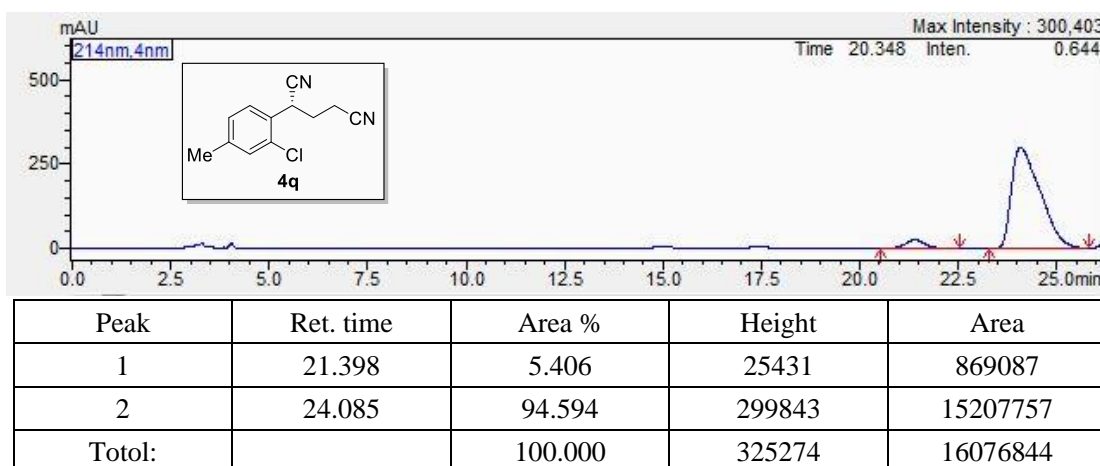


| | | | | |
|--------|--------|---------|--------|---------|
| 2 | 44.488 | 2.204 | 2160 | 165578 |
| Total: | | 100.000 | 152055 | 7512081 |

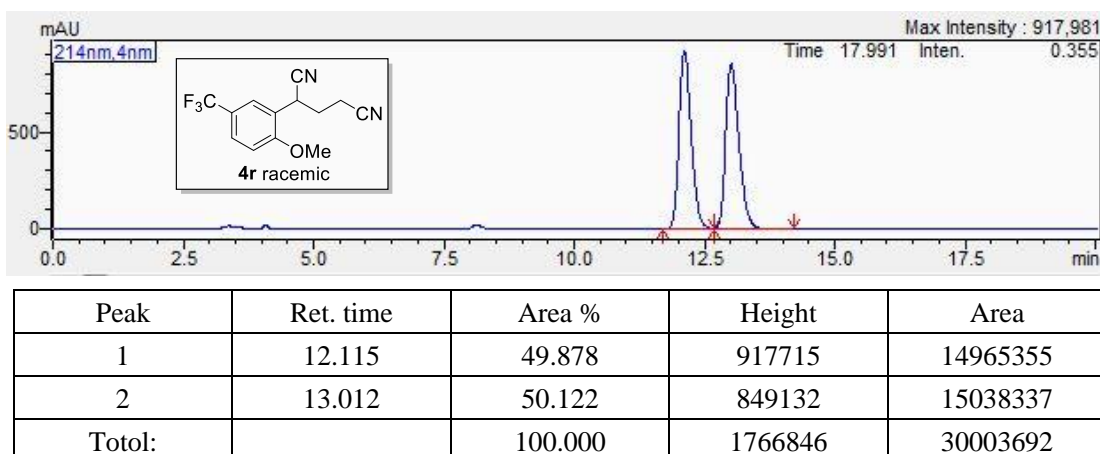
Supplementary Figure 244. HPLC data of **4p**



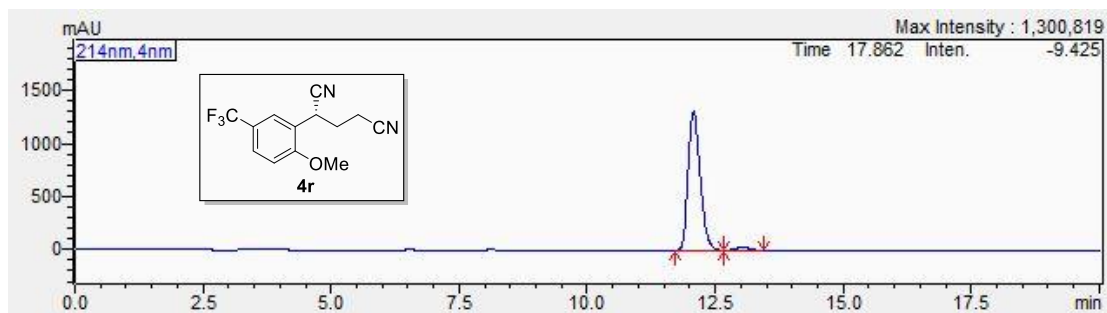
Supplementary Figure 245. HPLC data of rac-**4q**



Supplementary Figure 246. HPLC data of **4q**

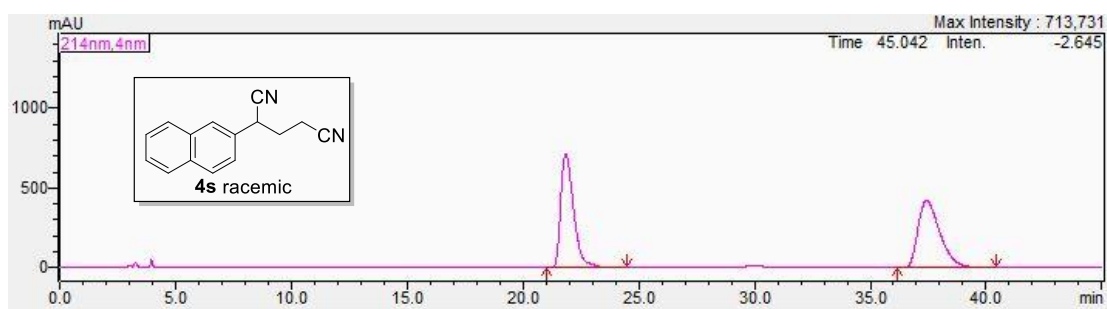


Supplementary Figure 247. HPLC data of rac-4r



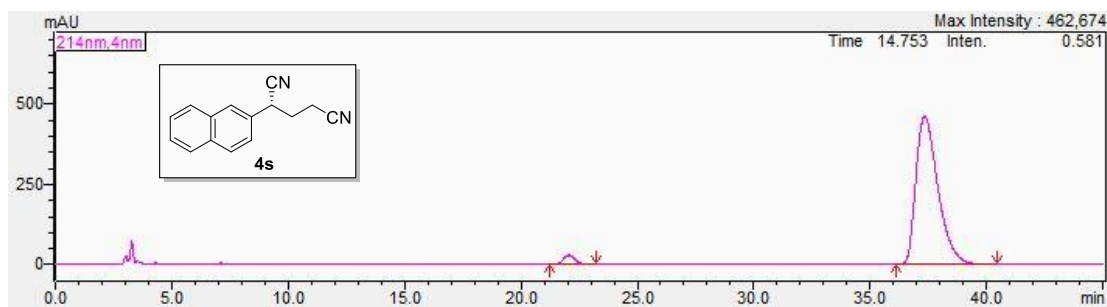
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 12.077 | 97.455 | 1308217 | 21516109 |
| 2 | 13.038 | 2.545 | 33887 | 561934 |
| Total: | | 100.000 | 1342104 | 22078043 |

Supplementary Figure 248. HPLC data of 4r



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 21.852 | 50.067 | 716411 | 27801563 |
| 2 | 37.433 | 49.933 | 421585 | 27727379 |
| Total: | | 100.000 | 1137996 | 55528942 |

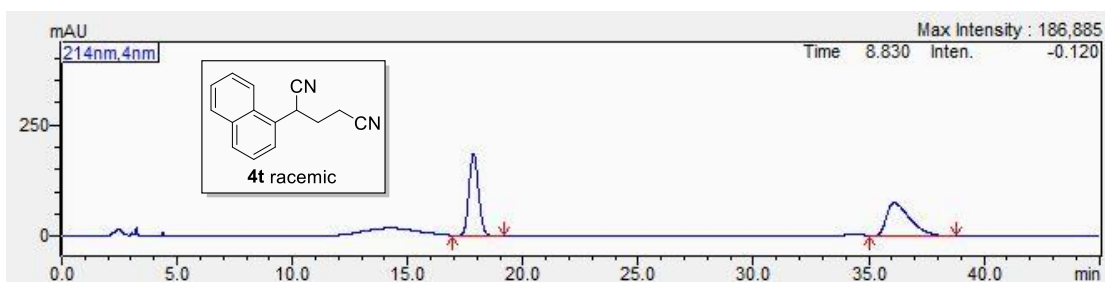
Supplementary Figure 249. HPLC data of rac-4s



| Peak | Ret. time | Area % | Height | Area |
|------|-----------|--------|--------|----------|
| 1 | 22.058 | 3.277 | 28647 | 1026689 |
| 2 | 37.341 | 96.723 | 461715 | 30301874 |

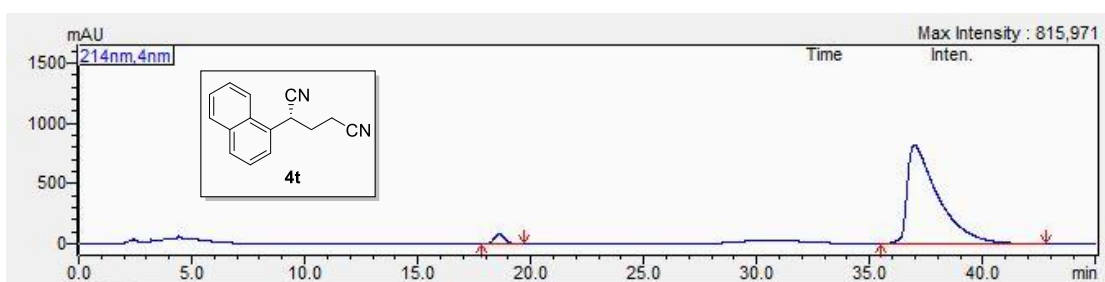
| | | | | |
|--------|--|---------|--------|----------|
| Total: | | 100.000 | 490361 | 31328563 |
|--------|--|---------|--------|----------|

Supplementary Figure 250. HPLC data of **4s**



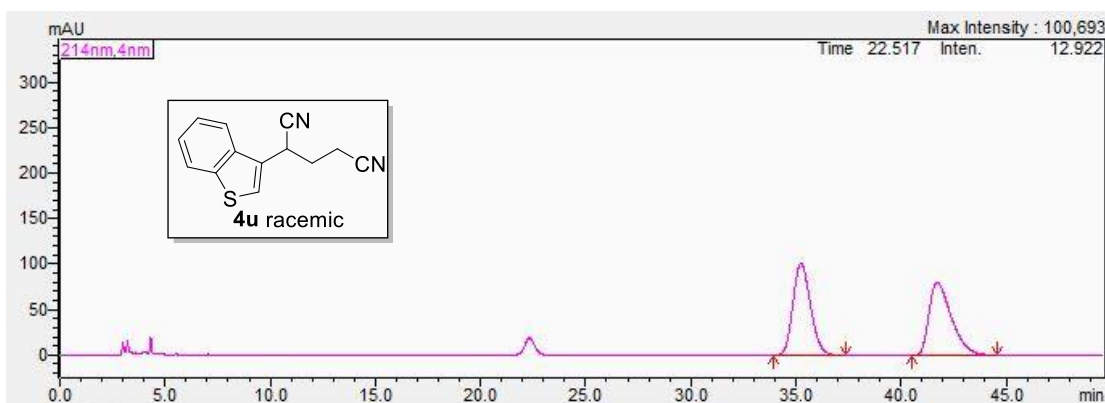
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 17.858 | 50.147 | 185718 | 5318123 |
| 2 | 36.123 | 49.853 | 74182 | 5286890 |
| Total: | | 100.000 | 259900 | 10605013 |

Supplementary Figure 251. HPLC data of rac-**4t**



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 18.610 | 3.057 | 78011 | 2437679 |
| 2 | 36.986 | 96.943 | 816502 | 77292310 |
| Total: | | 100.000 | 894513 | 79729990 |

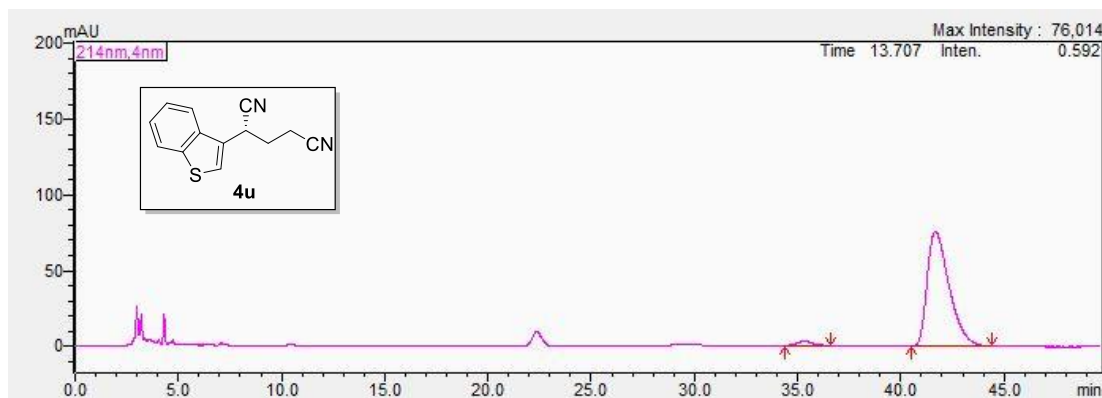
Supplementary Figure 252. HPLC data of **4t**



| Peak | Ret. time | Area % | Height | Area |
|------|-----------|--------|--------|---------|
| 1 | 35.244 | 49.950 | 101262 | 5907657 |

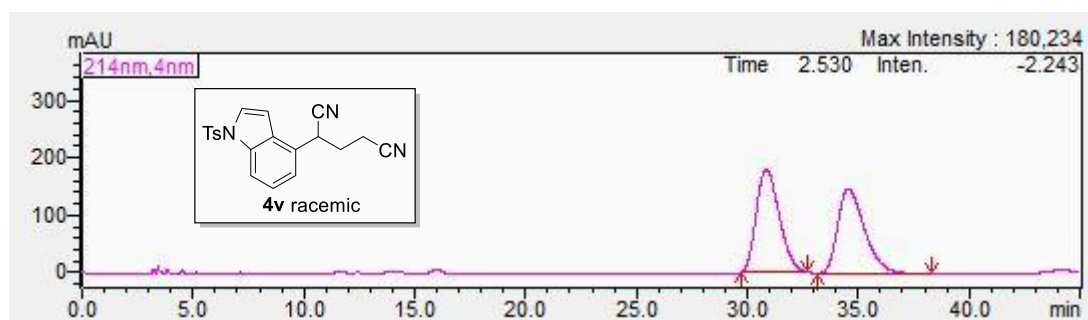
| | | | | |
|--------|--------|---------|--------|----------|
| 2 | 41.727 | 50.050 | 79936 | 5919500 |
| Totol: | | 100.000 | 181199 | 11827157 |

Supplementary Figure 253. HPLC data of rac-4u



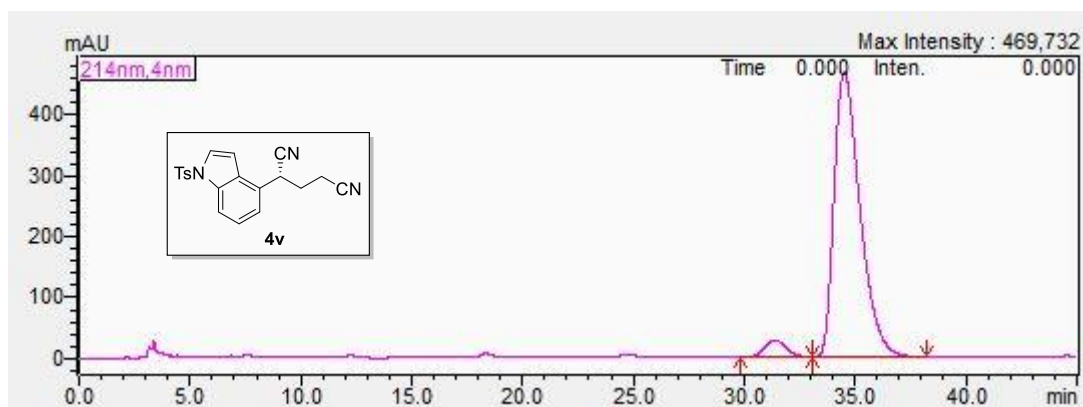
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|---------|
| 1 | 35.346 | 2.616 | 2676 | 150677 |
| 2 | 41.678 | 97.384 | 75912 | 5609811 |
| Totol: | | 100.000 | 78588 | 5760488 |

Supplementary Figure 254. HPLC data of 4u



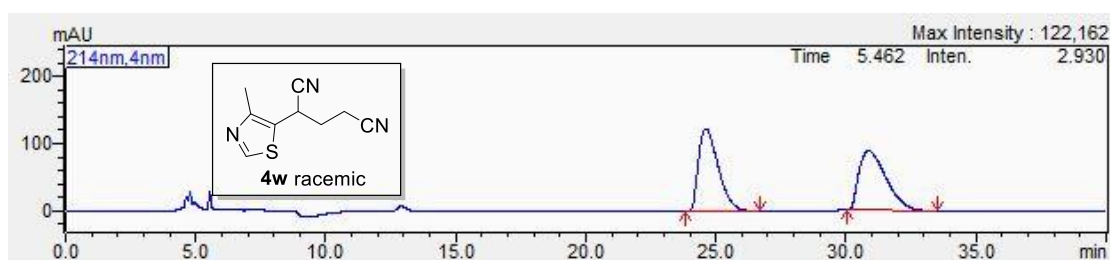
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 30.858 | 50.997 | 178733 | 12868867 |
| 2 | 34.569 | 49.003 | 147287 | 12365858 |
| Totol: | | 100.000 | 326020 | 25234724 |

Supplementary Figure 255. HPLC data of rac-4v



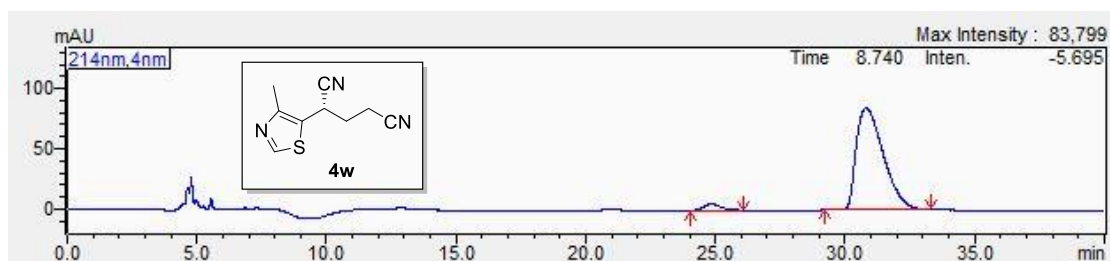
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 31.388 | 4.920 | 28587 | 1971149 |
| 2 | 34.516 | 95.080 | 467567 | 38090871 |
| Total: | | 100.000 | 496154 | 40062019 |

Supplementary Figure 256. HPLC data of **4v**



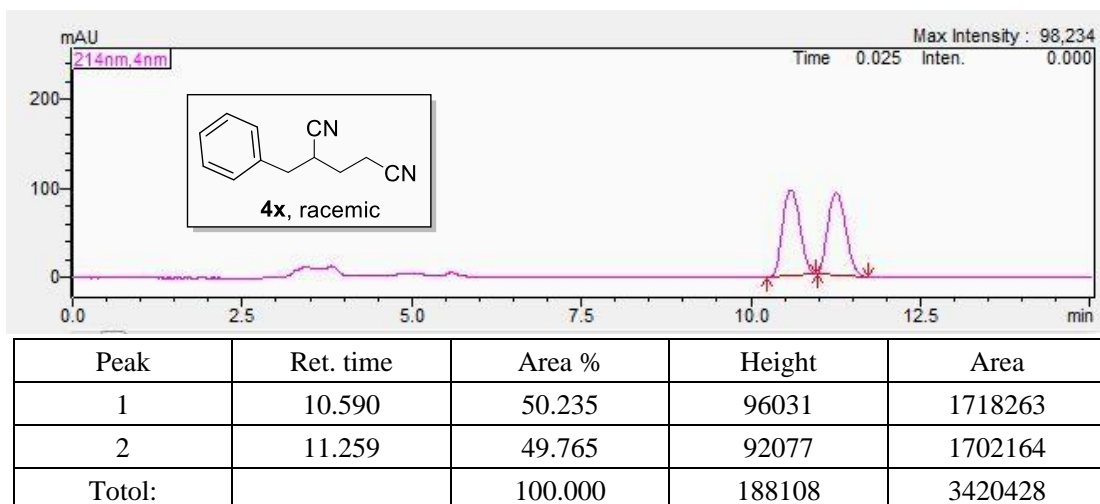
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|----------|
| 1 | 24.615 | 50.297 | 123269 | 6537809 |
| 2 | 30.879 | 49.703 | 88748 | 6460641 |
| Total: | | 100.000 | 212017 | 12998450 |

Supplementary Figure 257. HPLC data of rac-**4w**

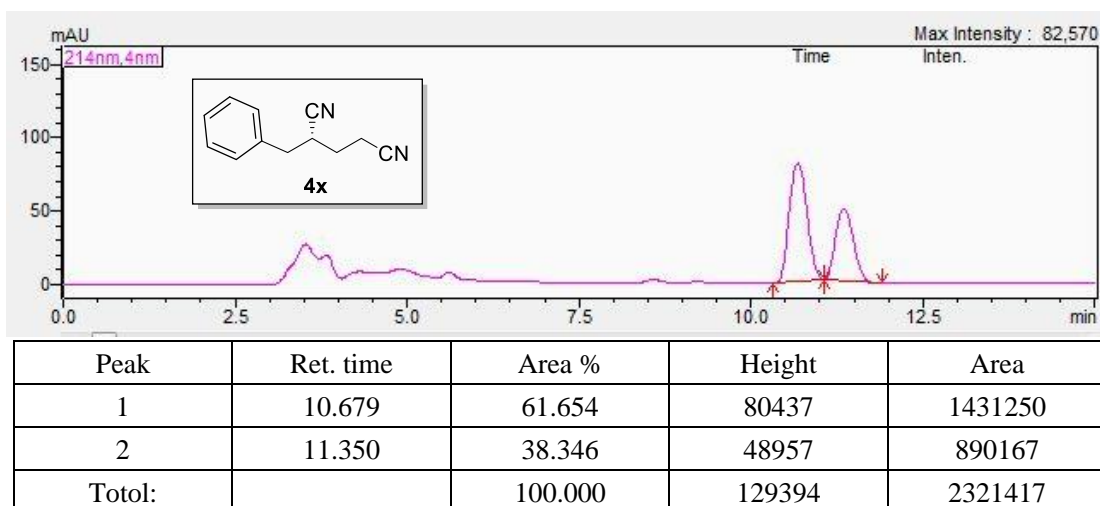


| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|--------|---------|
| 1 | 24.838 | 3.715 | 5158 | 239019 |
| 2 | 30.815 | 96.285 | 84569 | 6194066 |
| Total: | | 100.000 | 89728 | 6433085 |

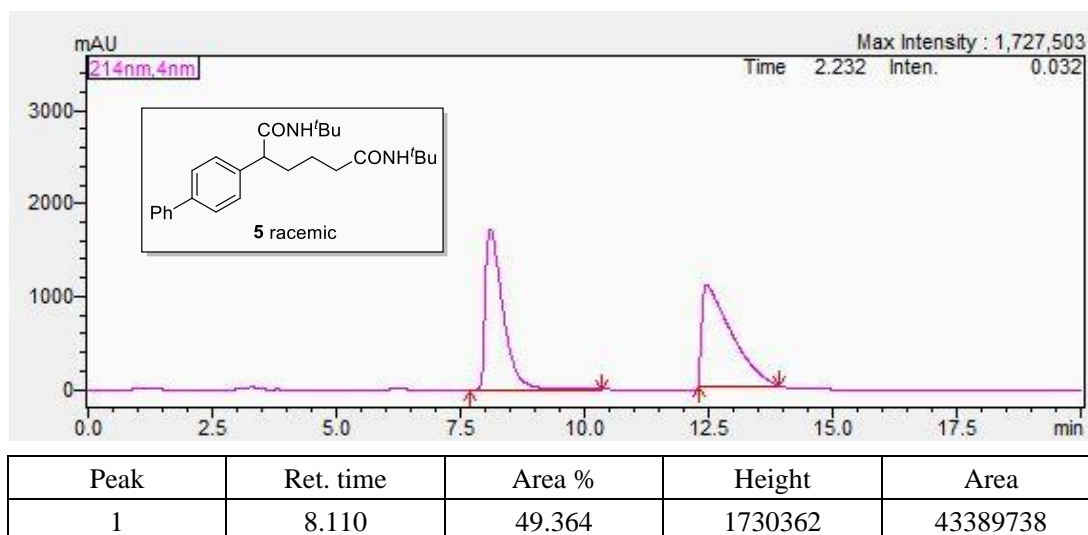
Supplementary Figure 258. HPLC data of **4w**



Supplementary Figure 259. HPLC data of rac-4x

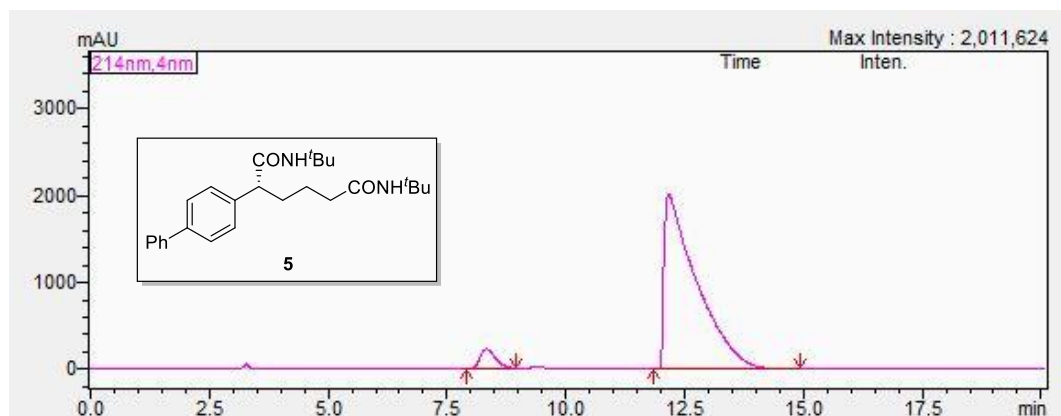


Supplementary Figure 260. HPLC data of 4x



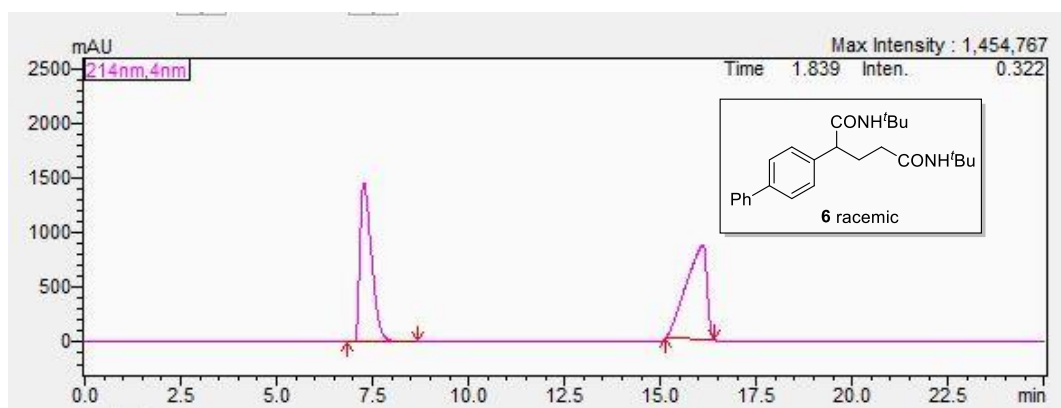
| | | | | |
|--------|--------|---------|---------|----------|
| 2 | 12.472 | 50.636 | 1080572 | 44508003 |
| Totol: | | 100.000 | 2810934 | 87897742 |

Supplementary Figure 261. HPLC data of rac-5



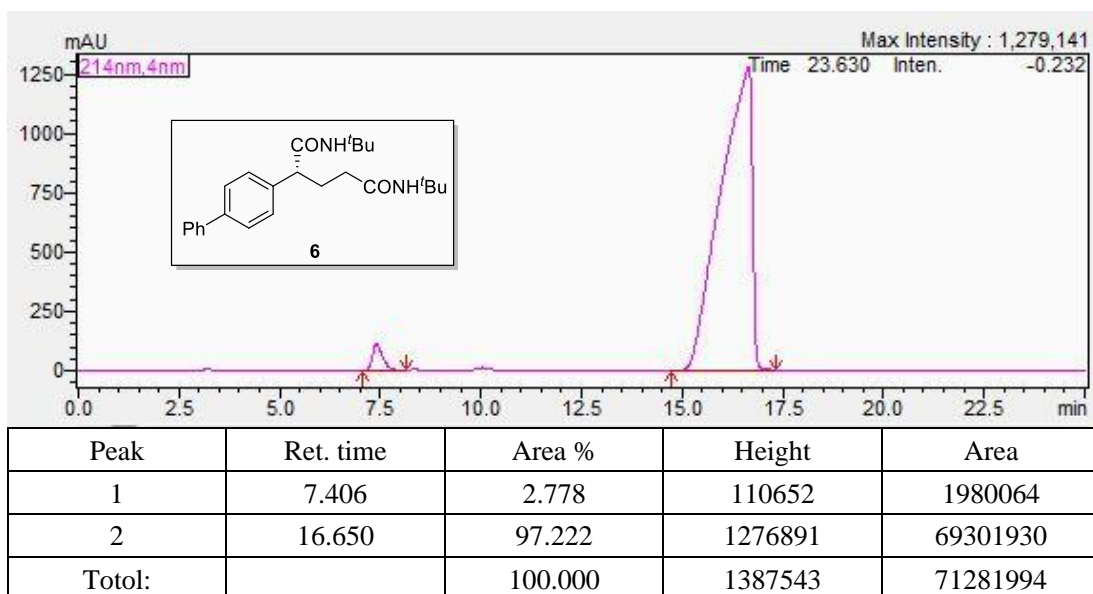
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|-----------|
| 1 | 8.334 | 5.139 | 226876 | 5191848 |
| 2 | 12.164 | 94.861 | 2009410 | 95833648 |
| Totol: | | 100.000 | 2236286 | 101025496 |

Supplementary Figure 262. HPLC data of 5

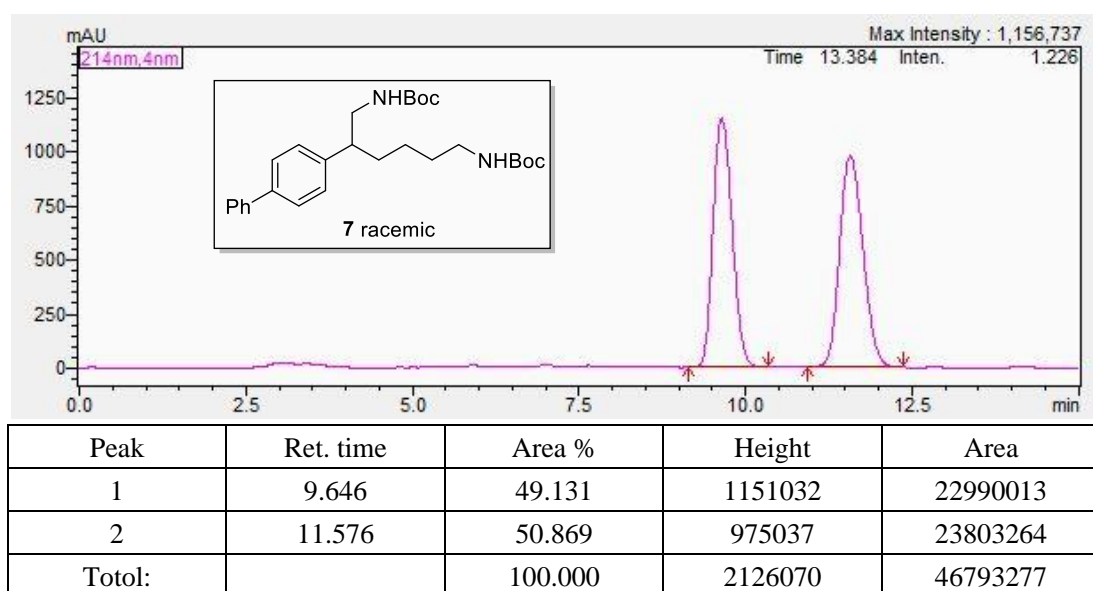


| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 7.276 | 49.208 | 1454214 | 31065110 |
| 2 | 16.101 | 50.792 | 861532 | 32065715 |
| Totol: | | 100.000 | 2315746 | 63130826 |

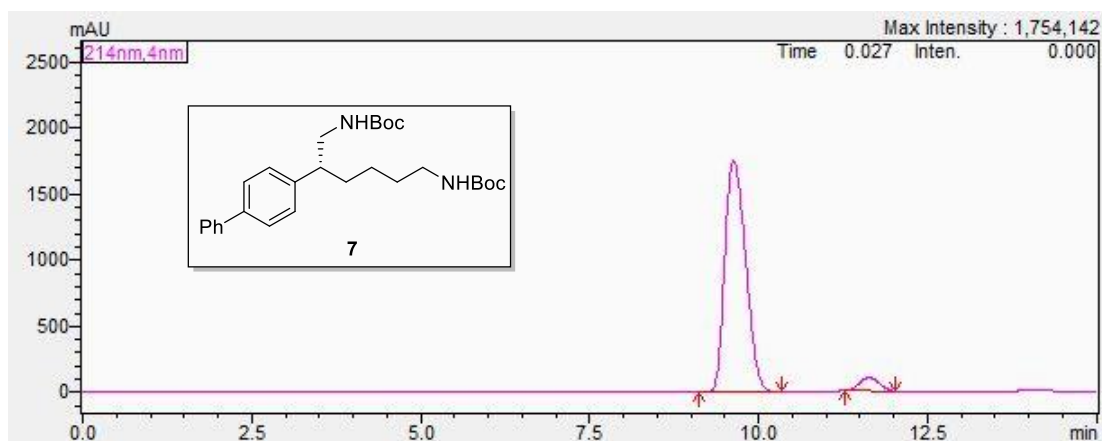
Supplementary Figure 263. HPLC data of rac-6



Supplementary Figure 264. HPLC data of 6

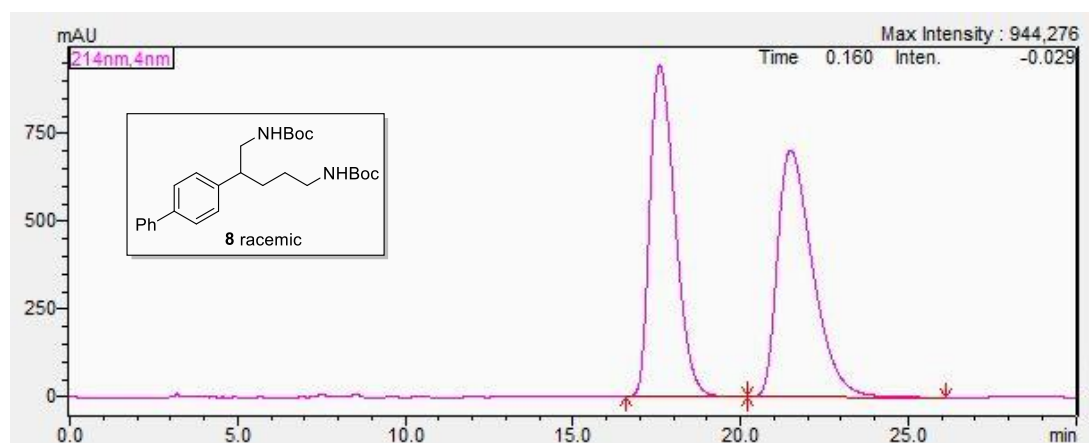


Supplementary Figure 265. HPLC data of rac-7



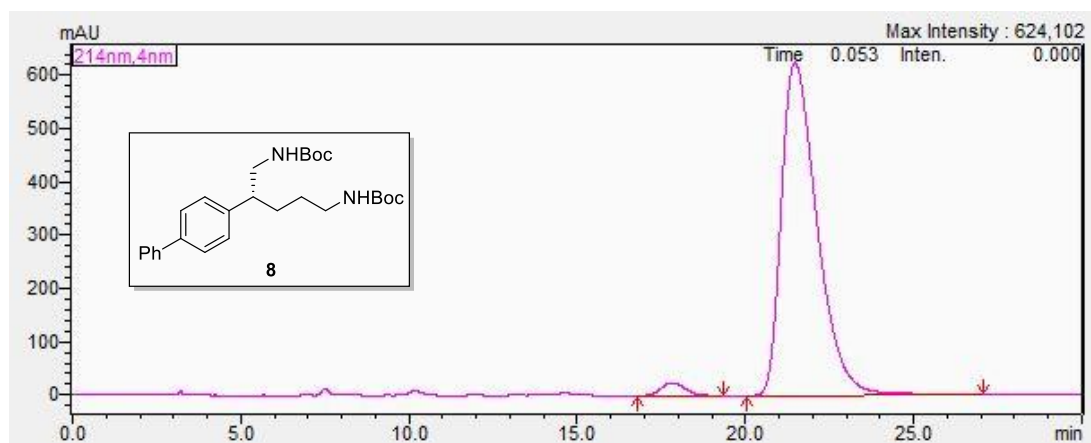
| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|----------|
| 1 | 9.629 | 94.779 | 1751637 | 37675479 |
| 2 | 11.635 | 5.221 | 100773 | 2075260 |
| Total: | | 100.000 | 1852410 | 39750738 |

Supplementary Figure 266. HPLC data of 7



| Peak | Ret. time | Area % | Height | Area |
|--------|-----------|---------|---------|-----------|
| 1 | 17.592 | 49.058 | 943880 | 49556085 |
| 2 | 21.489 | 50.942 | 703573 | 51459994 |
| Total: | | 100.000 | 1647453 | 101016079 |

Supplementary Figure 267. HPLC data of rac-8

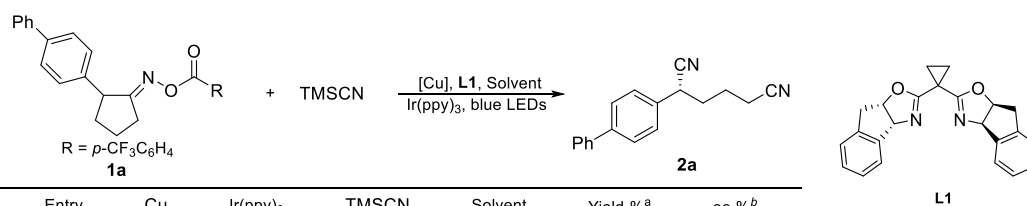


| Peak | Ret. time | Area % | Height | Area |
|---------------|-----------|----------------|---------------|-----------------|
| 1 | 17.833 | 2.526 | 24174 | 1200380 |
| 2 | 21.480 | 97.474 | 625083 | 46315576 |
| Total: | | 100.000 | 649257 | 47515956 |

Supplementary Figure 268. HPLC data of **8**

Supplementary Tables

Supplementary Table 1 Optimization of Reaction Conditions

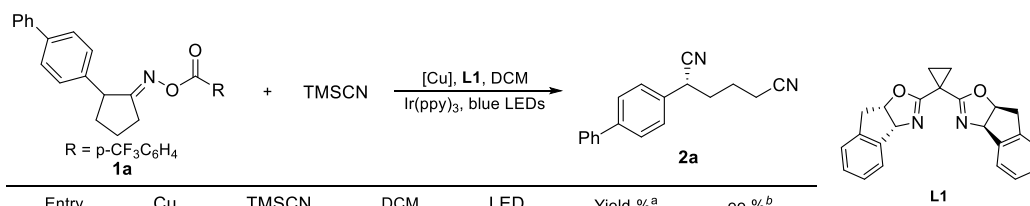


| Entry | Cu | Ir(ppy) ₃ | TMSCN | Solvent | Yield % ^a | ee % ^b |
|-------|----------|----------------------|---------|---------|----------------------|-------------------|
| 1 | 10 mol % | 1 mol % | 1.5 eq. | DMF/DCM | 38 | 87 |
| 2 | 5 mol % | 1 mol % | 1.5 eq. | DMF/DCM | 48 | 87 |
| 3 | 5 mol % | 0.5 mol % | 1.5 eq. | DMF/DCM | 50 | 89 |
| 4 | 5 mol % | 1 mol % | 3 eq. | DMF/DCM | 55 | 88 |
| 5 | 5 mol % | 1 mol % | 1.5 eq. | DCM | 49 | 88 |
| 6 | 5 mol % | 0.5 mol % | 3 eq. | DCM | 53 | 90 |
| 7 | 3 mol % | 0.5 mol % | 3 eq. | DCM | 62 | 90 |
| 8 | 2 mol % | 0.5 mol % | 3 eq. | DCM | 72 | 90 |
| 9 | 1 mol % | 0.5 mol % | 3 eq. | DCM | - ^c | - |

Conditions: x mol % Cu(CH₃CN)₄PF₆, 1.2x mol % **L1**, y mol % Ir(ppy)₃, 1.5 or 3.0 eq. TMSCN, 1.0 mL solvent was used for 0.1 mmol scale reaction, 24 W blue LEDs, RT, 24 h, 4/6 DMF/DCM.

^aisolated yield. ^bdetected by HPLC. ^clow conversion.

Supplementary Table 2 Optimization of Reaction Conditions

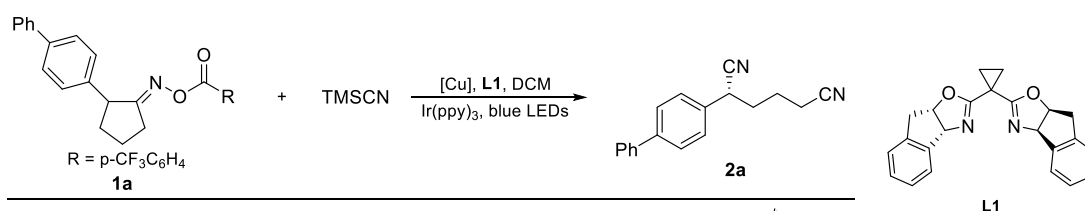


| Entry | Cu | TMSCN | DCM | LED | Yield % ^a | ee % ^b |
|-------|-----------|-------|--------|------|----------------------|-------------------|
| 1 | 2 mol % | 3 eq. | 1 mL | 24 W | 69 | 90 |
| 2 | 2 mol % | 2 eq. | 1 mL | 24 W | 68 | 90 |
| 3 | 2 mol % | 2 eq. | 0.5 mL | 24 W | 50 | 90 |
| 4 | 2 mol % | 2 eq. | 2 mL | 24 W | - ^c | - |
| 5 | 1.8 mol % | 3 eq. | 1 mL | 24 W | 53 | 90 |
| 6 | 1.5 mol % | 3 eq. | 1 mL | 24 W | 62 | 89 |
| 7 | 2 mol % | 3 eq. | 1 mL | 5 W | 71 | 90.6 |

Conditions: x mol % $\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6$, 1.5x mol % **L1**, 0.5 mol % $\text{Ir}(\text{ppy})_3$, y eq. TMSCN was used for 0.1 mmol scale reaction, 24 or 5 W blue LEDs. RT, 36 h.

^aisolated yield. ^bdetected by HPLC. ^clow conversion.

Supplementary Table 3 Screening of Copper Catalyst

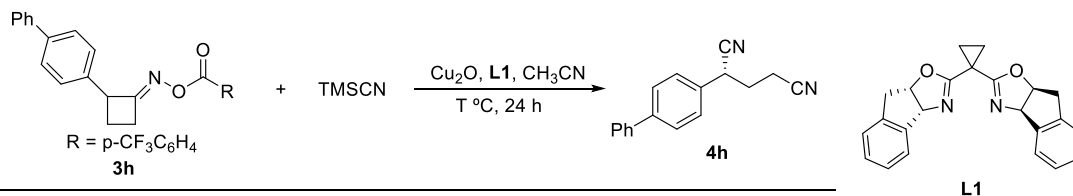


| Entry | Cu | Yield % ^a | ee % ^b |
|-------|--|----------------------|-------------------|
| 1 | CuBr | 48 | 90 |
| 2 | CuCN | 42 | 90.6 |
| 3 | CuOAc | 51 | 91 |
| 4 | $\text{Cu}(\text{CH}_3\text{CN})_4\text{BF}_4$ | 64 | 91 |
| 5 | CuCl | 43 | 91 |
| 6 | CuSCN | 49 | 90.6 |
| 7 | $\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6$ | 71 | 91 |

Conditions: 2 mol % [Cu], 3 mol % **L1**, 0.5 mol % $\text{Ir}(\text{ppy})_3$, 3 eq. TMSCN, 1 mL DCM was used for 0.1 mmol scale reaction, 5 W blue LEDs, RT, 36 h.

^aisolated yield. ^bdetected by HPLC.

Supplementary Table 4 Screening of Temperature

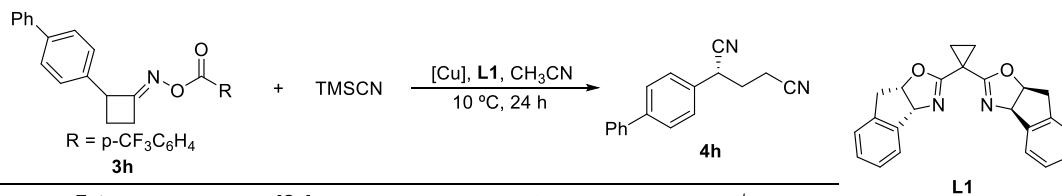


| Entry | [Cu] | T/°C | Yield % ^a | ee % ^b |
|-------|-------------------|--------|----------------------|-------------------|
| 1 | Cu ₂ O | -10 °C | 47 | 94 |
| 2 | Cu ₂ O | 0 °C | 68 | 94 |
| 3 | Cu ₂ O | 10 °C | 91 | 94 |
| 4 | Cu ₂ O | 25 °C | 98 | 92.5 |

Conditions: 1 mol% Cu₂O, 4 mol % L1, 1.1 eq. TMSCN, 1.0 mL CH₃CN was used for 0.1 mmol scale reaction, T °C, 24h.

^aisolated yield. ^bdetected by HPLC.

Supplementary Table 5 Screening of Copper Catalyst

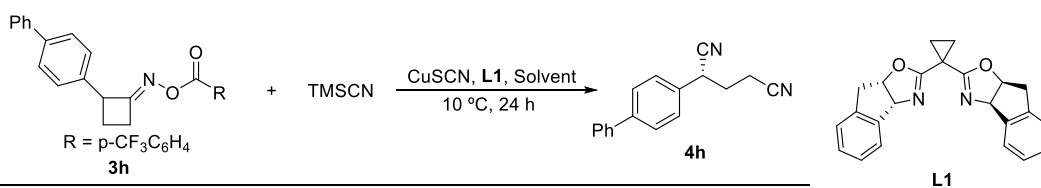


| Entry | [Cu] | Yield % ^a | ee % ^b |
|-------|---|----------------------|-------------------|
| 1 | CuCl | 88 | 94 |
| 2 | CuBr | 78 | 94 |
| 3 | CuI | 91 | 94 |
| 4 | CuSCN | 97 | 94 |
| 5 | CuCN | 88 | 94 |
| 6 | Cu(acac) ₂ | 81 | 94 |
| 7 | Cu(CH ₃ CN) ₄ PF ₆ | trace | - |
| 8 | CuF ₂ | 76 | 94 |
| 9 | Cu(OAc) ₂ | 74 | 94 |

Conditions: 2 mol % [Cu], 4 mol % L1, 1.1 eq. TMSCN, 1.0 mL CH₃CN was used for 0.1 mmol scale reaction, 10 °C, 24h .

^aisolated yield. ^bdetected by HPLC.

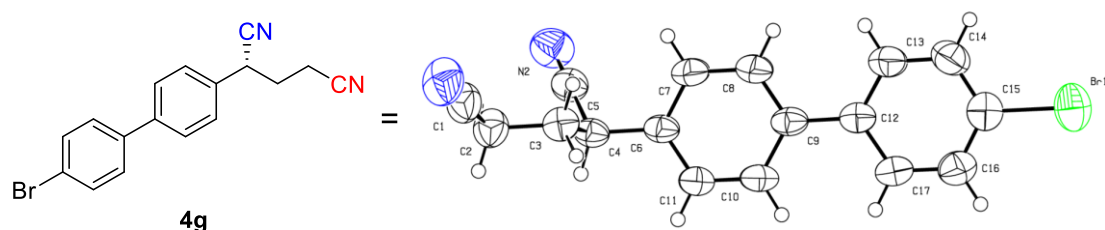
Supplementary Table 6 Screening of Solvent



| Entry | [Cu] | solvent | Yield % ^a | ee % ^b |
|----------------|----------------|-------------------|----------------------|-------------------|
| 1 | CuSCN | DCM | 72 | 94 |
| 2 | CuSCN | Et ₂ O | 56 | 93 |
| 3 | CuSCN | DMF | 71 | 93 |
| 4 | CuSCN | toluene | 78 | 94 |
| 5 | CuSCN | MeOH | 75 | 89 |
| 6 | CuSCN | Acetone | 70 | 95 |
| 7 ^c | CuSCN(3 mol %) | Acetone | 93 | 95 |

Conditions: 2 mol % CuSCN, 4 mol % L1, 1.1 eq. TMSCN, 1.0 mL Solvent was used for 0.1 mmol scale reaction, 10 °C, 24h.

^aisolated yield. ^bdetected by HPLC. ^c3 mol % Cu, 3.6 mol % L1, 1.5 eq. TMSCN, 1.0 mL degassed Acetone.



Supplementary Figure 269. X-ray crystallography of **4g**

Supplementary Table 7 Crystal data and structure refinement for **4g**.

| | |
|---------------------|--|
| Identification code | 4g |
| Empirical formula | C ₁₇ H ₁₃ BrN ₂ |
| Formula weight | 325.20 |
| Temperature/K | 293(2) |
| Crystal system | monoclinic |
| Space group | P2 ₁ |
| a/Å | 7.98210(10) |
| b/Å | 5.77100(10) |
| c/Å | 16.0567(3) |
| α/° | 90 |

| | |
|--|---|
| $\beta/^\circ$ | 93.240(2) |
| $\gamma/^\circ$ | 90 |
| Volume/ \AA^3 | 738.46(2) |
| Z | 2 |
| $\rho_{\text{calc}}/\text{g/cm}^3$ | 1.463 |
| μ/mm^{-1} | 3.706 |
| F(000) | 328.0 |
| Crystal size/ mm^3 | $0.3 \times 0.22 \times 0.2$ |
| Radiation | CuK α ($\lambda = 1.54184$) |
| 2 θ range for data collection/ $^\circ$ | 11.038 to 147.586 |
| Index ranges | $-9 \leq h \leq 8, -6 \leq k \leq 6, -19 \leq l \leq 19$ |
| Reflections collected | 6082 |
| Independent reflections | 2726 [$R_{\text{int}} = 0.0195, R_{\text{sigma}} = 0.0166$] |
| Data/restraints/parameters | 2726/1/181 |
| Goodness-of-fit on F^2 | 1.367 |
| Final R indexes [$I \geq 2\sigma(I)$] | $R_1 = 0.0402, wR_2 = 0.1493$ |
| Final R indexes [all data] | $R_1 = 0.0405, wR_2 = 0.1504$ |
| Largest diff. peak/hole / $e \text{\AA}^{-3}$ | 0.31/-0.61 |
| Flack parameter | 0.024(15) |

Supplementary Table 8 Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for **4g**. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

| Atom | x | y | z | $U(\text{eq})$ |
|------|----------|------------|-----------|----------------|
| Br1 | 240.9(6) | 3265.2(16) | 2337.5(3) | 81.7(3) |
| C12 | 2534(4) | 3632(8) | 5089(3) | 49.0(9) |
| C5 | 5078(6) | 6564(9) | 8880(3) | 58.4(10) |
| N2 | 5053(6) | 8376(11) | 9137(3) | 80.9(12) |
| C9 | 3206(4) | 3779(7) | 5975(3) | 50.0(9) |
| C4 | 5164(4) | 4181(7) | 8549(3) | 49.4(8) |
| C3 | 7014(5) | 3332(11) | 8559(3) | 56.3(8) |
| C15 | 1195(5) | 3403(11) | 3455(3) | 61.0(10) |
| C6 | 4420(5) | 4085(7) | 7650(2) | 48.8(9) |
| C2 | 7864(5) | 3338(15) | 9429(3) | 67.0(11) |
| N1 | 11113(5) | 3097(17) | 9336(4) | 93.6(16) |
| C14 | 2200(8) | 5243(11) | 3707(3) | 74.4(13) |
| C1 | 9689(6) | 3173(14) | 9380(3) | 71.3(11) |
| C17 | 1537(6) | 1768(9) | 4803(3) | 64.0(11) |
| C13 | 2845(7) | 5337(9) | 4515(3) | 66.8(12) |

| | | | | |
|-----|---------|----------|---------|----------|
| C11 | 3454(7) | 2255(10) | 7381(4) | 71.2(14) |
| C16 | 882(6) | 1642(10) | 3992(4) | 67.1(12) |
| C10 | 2877(8) | 2065(11) | 6556(4) | 77.5(16) |
| C8 | 4164(8) | 5629(10) | 6251(3) | 71.7(14) |
| C7 | 4774(8) | 5793(10) | 7085(4) | 73.3(14) |

Supplementary Table 9 Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for **4g**. The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^{*2}U_{11}+2hka^*b^*U_{12}+\dots]$.

| Atom | U_{11} | U_{22} | U_{33} | U_{23} | U_{13} | U_{12} |
|------|----------|----------|----------|----------|----------|-----------|
| Br1 | 81.4(4) | 98.9(5) | 64.5(4) | -9.1(3) | 2.6(2) | 20.4(3) |
| C12 | 40.4(15) | 45(2) | 63(2) | 0.6(16) | 12.6(14) | 2.5(13) |
| C5 | 55(2) | 57(3) | 64(2) | 3.8(19) | 9.0(17) | 0.7(17) |
| N2 | 99(3) | 61(3) | 85(3) | -5(3) | 16(2) | 10(2) |
| C9 | 37.7(14) | 43(2) | 70(2) | 4.8(15) | 9.5(14) | -0.8(12) |
| C4 | 42.0(16) | 46(2) | 61(2) | 7.6(15) | 9.8(14) | -0.9(13) |
| C3 | 47.3(16) | 56(2) | 66(2) | -1(2) | 8.4(14) | 3.0(19) |
| C15 | 54.5(18) | 68(3) | 61(2) | -5(2) | 9.5(15) | 8(2) |
| C6 | 40.2(13) | 45(2) | 62(2) | 7.2(15) | 10.1(13) | -3.3(13) |
| C2 | 56(2) | 78(3) | 67(2) | -1(3) | -0.6(17) | -1(2) |
| N1 | 54(2) | 124(5) | 101(3) | 4(4) | -6(2) | 4(3) |
| C14 | 101(4) | 63(3) | 60(3) | 9(2) | 11(2) | -7(3) |
| C1 | 60(2) | 82(3) | 70(2) | 3(3) | -7.7(18) | 3(3) |
| C17 | 67(3) | 52(3) | 73(3) | 3.6(19) | 4(2) | -16.2(19) |
| C13 | 80(3) | 50(3) | 71(3) | 5(2) | 6(2) | -10.9(19) |
| C11 | 77(3) | 59(3) | 76(3) | 21(2) | -13(2) | -23(2) |
| C16 | 63(2) | 65(3) | 74(3) | -1(2) | 8(2) | -10(2) |
| C10 | 87(3) | 56(3) | 88(4) | 24(3) | -12(3) | -29(3) |
| C8 | 86(3) | 66(3) | 63(3) | 14(2) | 6(2) | -31(3) |
| C7 | 90(3) | 62(3) | 67(3) | 7(2) | 5(2) | -36(3) |

Supplementary Table 10 Bond Lengths for **4g**

| Atom | Atom | Length/ \AA | Atom | Atom | Length/ \AA |
|------|------|----------------------|------|------|----------------------|
| Br1 | C15 | 1.911(4) | C15 | C14 | 1.377(8) |
| C12 | C9 | 1.494(6) | C15 | C16 | 1.364(8) |
| C12 | C17 | 1.400(6) | C6 | C11 | 1.364(6) |
| C12 | C13 | 1.381(7) | C6 | C7 | 1.380(6) |
| C5 | N2 | 1.124(9) | C2 | C1 | 1.466(6) |
| C5 | C4 | 1.478(7) | N1 | C1 | 1.143(7) |
| C9 | C10 | 1.395(6) | C14 | C13 | 1.370(8) |

| | | | | | |
|----|----|----------|-----|-----|----------|
| C9 | C8 | 1.372(7) | C17 | C16 | 1.378(8) |
| C4 | C3 | 1.555(5) | C11 | C10 | 1.381(8) |
| C4 | C6 | 1.530(6) | C8 | C7 | 1.401(8) |
| C3 | C2 | 1.519(6) | | | |

Supplementary Table 11 Bond Angles for **4g**

| Atom | Atom | Atom | Angle/° | Atom | Atom | Atom | Angle/° |
|------|------|------|----------|------|------|------|----------|
| C17 | C12 | C9 | 121.5(4) | C11 | C6 | C4 | 120.6(4) |
| C13 | C12 | C9 | 121.8(4) | C11 | C6 | C7 | 118.5(4) |
| C13 | C12 | C17 | 116.7(4) | C7 | C6 | C4 | 120.8(4) |
| N2 | C5 | C4 | 178.4(5) | C1 | C2 | C3 | 110.1(4) |
| C10 | C9 | C12 | 121.7(4) | C13 | C14 | C15 | 119.3(5) |
| C8 | C9 | C12 | 121.0(4) | N1 | C1 | C2 | 178.4(9) |
| C8 | C9 | C10 | 117.3(5) | C16 | C17 | C12 | 121.9(5) |
| C5 | C4 | C3 | 110.7(4) | C14 | C13 | C12 | 122.2(5) |
| C5 | C4 | C6 | 110.5(4) | C6 | C11 | C10 | 121.2(4) |
| C6 | C4 | C3 | 108.5(3) | C15 | C16 | C17 | 119.0(5) |
| C2 | C3 | C4 | 112.6(4) | C11 | C10 | C9 | 121.3(5) |
| C14 | C15 | Br1 | 120.1(4) | C9 | C8 | C7 | 121.2(4) |
| C16 | C15 | Br1 | 119.0(4) | C6 | C7 | C8 | 120.5(4) |
| C16 | C15 | C14 | 120.9(4) | | | | |

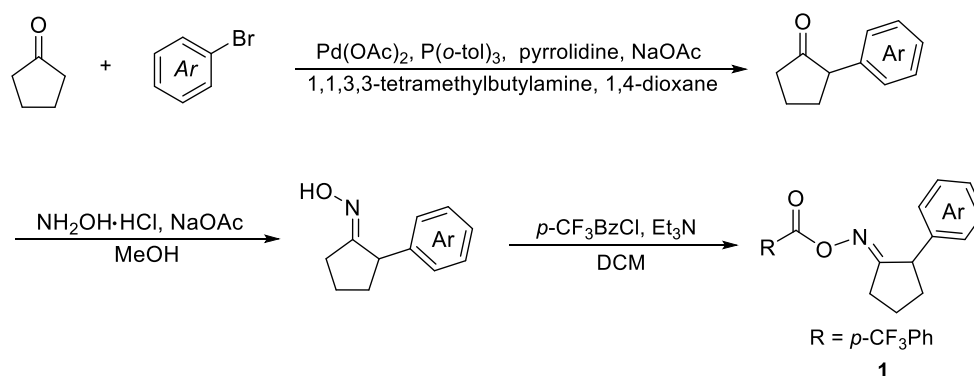
Supplementary Table 12 Hydrogen Atom Coordinates ($\text{\AA}\times 10^4$) and Isotropic Displacement Parameters ($\text{\AA}^2\times 10^3$) for **4g**.

| Atom | <i>x</i> | <i>y</i> | <i>z</i> | U(eq) |
|------|----------|----------|----------|-------|
| H4 | 4524.68 | 3144.47 | 8895.43 | 59 |
| H3A | 7038.37 | 1772.68 | 8334.28 | 68 |
| H3B | 7638.71 | 4325.65 | 8200.46 | 68 |
| H2A | 7584.47 | 4752.89 | 9716.14 | 80 |
| H2B | 7463.42 | 2037.29 | 9745.02 | 80 |
| H14 | 2438.98 | 6408.79 | 3331.45 | 89 |
| H17 | 1309.55 | 580.01 | 5171.73 | 77 |
| H13 | 3514.81 | 6590.71 | 4682.02 | 80 |
| H11 | 3179.1 | 1115.15 | 7758.92 | 85 |
| H16 | 236.47 | 376.94 | 3812.85 | 80 |
| H10 | 2257.31 | 768.22 | 6385.18 | 93 |
| H8 | 4412.29 | 6796.1 | 5878.29 | 86 |
| H7 | 5422.63 | 7061.83 | 7257.88 | 88 |

Supplementary Methods

General Information ^1H NMR spectra were recorded at ambient temperature on Bruker-400 (400 MHz) spectrometers and are referenced relative to the residual protons in CDCl_3 at δ 7.26 ppm or $(\text{CD}_3)_2\text{SO}-d_6$ at δ 2.50 ppm. Data for ^1H NMR are reported as follows: chemical shift (ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, ap = apparent), integration, and coupling constant (Hz). ^{13}C NMR spectra were recorded at ambient temperature on Bruker-400 (100 MHz) spectrometers and are referenced relative to CDCl_3 at δ 77.16 ppm or $(\text{CD}_3)_2\text{SO}-d_6$ at δ 39.25 ppm. The ^{13}C NMR spectra were obtained with ^1H decoupling. Data for ^{13}C NMR are reported in terms of chemical shift and multiplicity where appropriate. High resolution mass spectra were recorded on P-SIMS-Gly of Bruker Daltonics Inc. using ESI-TOF (electrospray ionization-time of flight). High performance liquid chromatography was performed on shimadzu Series HPLC, using AD-H, OD-H, ID chiral column eluted with a mixture of hexane and isopropyl alcohol. TMSCN was purchased from energy-chemical, $\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6$ and CuSCN was purchased from TCI. $\text{Ir}(\text{ppy})_3$ was purchased from Laajoo Reagent. DMAc was purchased from J&K Chemical Reagent. And acetone was purchased from Sinopharm Chemical Reagent Co. Ltd.

General procedure (A) for synthesis of the substrates 1^{1,2,3}.



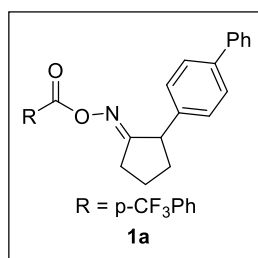
The 2-arylcyclopentan-1-ones were synthesized according to the reported procedure³. An 100 mL Schlenk flask was charged with Pd(OAc)₂ (67.6 mg, 0.3 mmol, 0.05 equiv), P(*o*-tol)₃ (182.8 mg, 0.6 mmol, 0.1 equiv), NaOAc (492.4 mg, 6.0 mmol, 1.0 equiv) and aryl bromides (7.8 mmol, 1.3 equiv), and then the flask was evacuated and backfilled with N₂ (repeated for 3 times). 1,4-dioxane (30.0 mL) was added in, followed by cyclopentanone (531 uL, 6.0 mmol, 1.0 equiv), pyrrolidine (148 μL, 1.8 mmol, 0.3 equiv) and 1,1,3,3-tetramethylbutylamine (289 uL, 1.8 mmol, 0.3 equiv). The flask was then heated in an oil bath at 110 °C under stirring for 16 hours, before cooled to room temperature. The mixture was filtered through a small plug of silica gel, eluted with ethyl acetate. The solvent was removed in vacuo and flash column chromatography of the residue gave the arylation product.

The ketones (1.0 equiv, ~5 mmol), hydroxylamine hydrochloride (1.2 equiv) and sodium acetate (1.5 equiv) were added to a round bottom flask containing a stirrer bar. MeOH (0.56 M) was added and the reaction was stirred for 12 h at 75 °C. Then methanol was removed under vacuum and the resulting mixture was extracted with DCM. The organic layer was washed with water and dried over Na₂SO₄. The solvent was removed under reduced pressure and the crude material was subjected to column chromatography to afford oximes.

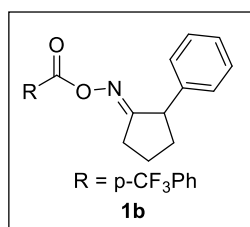
To a mixture of oxime (1.0 equiv, ~4 mmol), triethylamine (1.5 equiv) and DCM (0.5 M) in a flask was added 4-(trifluoromethyl)benzoyl chlorides (1.1 equiv) slowly at 0 °C. After 1 h, a saturated solution of aqueous NaHCO₃ (10 mL) was added to the above

solution, and the mixture was diluted with DCM. The organic layer was washed with brine (20 mL) and dried over Na₂SO₄. The solvent was removed under vacuum and the residue was subjected to column chromatography on silica gel with EtOAc–petroleum ether as an eluent to give the substrate.

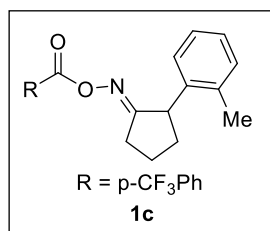
Characterization data for cyclopentanone oxime esters **1**



Prepared according to general procedure (A) from cyclopentanone and 4-bromo-1,1'-biphenyl to provide the title compound **1a** as a yellowish solid¹. ¹H NMR (CDCl₃, 400 MHz) δ 8.19 (d, *J* = 8.2 Hz, 2H), 7.74 (d, *J* = 8.4 Hz, 2H), 7.64 – 7.54 (m, 4H), 7.48 – 7.38 (m, 4H), 7.34 (t, *J* = 7.4 Hz, 1H), 4.11 (t, *J* = 7.6 Hz, 1H), 3.01 – 2.94 (m, 1H), 2.88 – 2.81 (m, 1H), 2.41 – 2.34 (m, 1H), 2.14 – 2.03 (m, 2H), 1.92 – 1.87 (m, 1H); ¹³C NMR (CDCl₃, 101 MHz) δ 178.2, 162.6, 140.8, 139.8, 139.1, 134.6 (q, *J* = 32.8 Hz), 132.5, 129.9, 128.7, 128.3, 127.4, 127.2, 127.0, 125.5 (q, *J* = 3.6 Hz), 123.5 (q, *J* = 272.8 Hz), 49.0, 34.7, 30.0, 22.5.

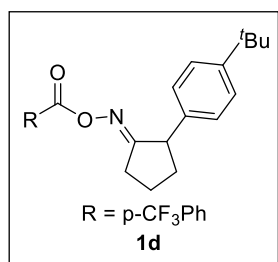


Prepared according to general procedure (A) from cyclopentanone and bromobenzene to provide the title compound **1b** as a yellowish solid¹. ¹H NMR (400 MHz, CDCl₃) 8.17 (d, *J* = 8.2 Hz, 2H), 7.73 (d, *J* = 8.3 Hz, 2H), 7.34 (d, *J* = 6.6 Hz, 4H), 7.26 – 7.21 (m, 1H), 4.07 (t, *J* = 7.2 Hz, 1H), 3.01 – 2.74 (m, 2H), 2.38 – 2.29 (m, 1H), 2.13 – 1.94 (m, 2H), 1.93 – 1.79 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 178.2, 162.6, 140.1, 134.6 (q, *J* = 32.7 Hz), 132.5, 129.9, 128.6, 127.8, 126.9, 125.5 (q, *J* = 3.5 Hz), 123.5 (q, *J* = 272.8 Hz), 49.2, 34.7, 30.0, 22.5.



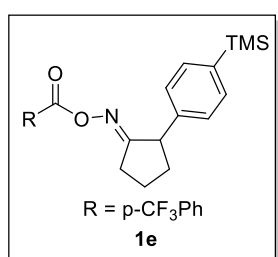
Prepared according to general procedure (A) from cyclopentanone and 1-bromo-2-methylbenzene to provide the title compound **1c** as a yellowish solid¹. ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.1 Hz, 2H), 7.73 (d, *J* = 8.3 Hz, 2H), 7.21 – 7.08 (m, 4H), 4.22 (t, *J* = 7.3 Hz, 1H), 3.02 – 2.83 (m, 2H), 2.40 (s, 3H), 2.36 – 2.26 (m, 1H), 2.07 – 2.01 (m, 1H), 1.94 – 1.83 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 178.7, 162.6, 139.4, 135.9, 134.6 (q, *J* = 32.8 Hz), 132.5, 130.6,

129.9, 127.1, 126.8, 126.2, 125.5 (q, $J = 3.6$ Hz), 123.5 (q, $J = 270.4$ Hz), 46.5, 34.0, 30.5, 22.5, 19.9.



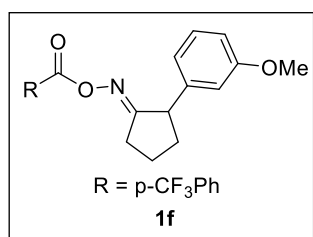
Prepared according to general procedure (A) from cyclopentanone and 1-bromo-4-(*tert*-butyl)benzene to provide the title compound **1d** as a white solid¹. ¹H NMR (CDCl₃, 400 MHz) δ 8.18 (d, $J = 8.1$ Hz, 2H), 7.73 (d, $J = 8.3$ Hz, 2H), 7.37 (d, $J = 8.4$ Hz, 2H), 7.27 (d, $J = 8.0$ Hz, 2H), 4.04 (t, $J = 7.3$ Hz,

1H), 2.97 – 2.91 (m, 1H), 2.85 – 2.78 (m, 1H), 2.40 – 2.22 (m, 1H), 2.15 – 1.97 (m, 2H), 1.93 – 1.78 (m, 1H), 1.31 (s, 9H); ¹³C NMR (CDCl₃, 101 MHz) δ 178.4, 162.6, 149.6, 136.9, 134.5 (q, $J = 32.7$ Hz), 132.5, 129.9, 127.4, 125.5 (q, $J = 3.3$ Hz), 123.5 (q, $J = 272.7$ Hz), 48.8, 34.5, 34.3, 31.3, 29.9, 22.5.



Prepared according to general procedure (A) from cyclopentanone and (4-bromophenyl)trimethylsilane to provide the title compound **1e** as a yellowish oil². ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, $J = 8.1$ Hz, 2H), 7.73 (d, $J = 8.3$ Hz, 2H), 7.50 (d, $J = 8.1$ Hz, 2H), 7.31 (d, $J = 7.9$ Hz, 2H), 4.05 (t, $J = 7.2$ Hz,

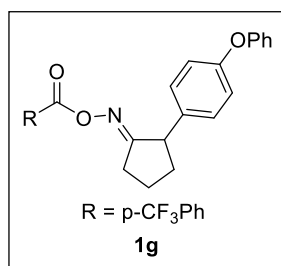
1H), 2.98 – 2.92 (m, 1H), 2.85 – 2.78 (m, 1H), 2.37 – 2.30 (m, 1H), 2.10 – 2.00 (m, 2H), 1.93 – 1.80 (m, 1H), 0.25 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.3, 162.6, 140.6, 138.8, 134.6 (q, $J = 32.7$ Hz), 133.7, 132.5, 129.9, 127.2, 125.5 (q, $J = 3.6$ Hz), 123.5 (q, $J = 272.9$ Hz), 49.3, 34.6, 30.1, 22.6, -1.6.



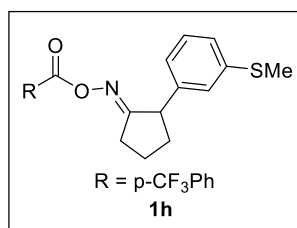
Prepared according to general procedure (A) from cyclopentanone and 1-bromo-3-methoxybenzene to provide the title compound **1f** as a yellowish liquid (37% total yield).

¹H NMR (CDCl₃, 400 MHz) δ 8.17 (d, $J = 8.1$ Hz, 2H), 7.73 (d, $J = 8.2$ Hz, 2H), 7.26 (t, $J = 7.9$ Hz, 1H), 6.94 – 6.88 (m, 2H), 6.82 – 6.76 (m, 1H), 4.05 (t, $J = 7.1$ Hz, 1H), 3.81 (s, 3H), 3.00 – 2.87 (m, 1H), 2.87 – 2.73 (m, 1H), 2.39 – 2.26 (m, 1H), 2.13 – 1.95 (m, 2H), 1.94 – 1.80 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 178.2, 162.7, 159.8, 141.8, 134.7 (q, $J = 32.7$ Hz), 130.1, 129.7, 125.6 (q, $J = 3.7$ Hz), 123.7 (q, $J = 272.8$ Hz), 120.3, 114.1, 112.1, 55.3, 49.2, 34.7, 30.1, 22.6. HRMS (ESI) calcd. for C₂₀H₁₈O₃NF₃

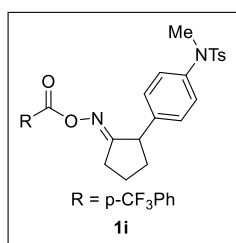
$[M+Na]^+$ m/z 400.1136, found 400.1138.



Prepared according to general procedure (A) from cyclopentanone and 1-bromo-4-phenoxybenzene to provide the title compound **1g** as a yellowish liquid (44% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.16 (d, J = 8.1 Hz, 2H), 7.71 (d, J = 8.2 Hz, 2H), 7.37 – 7.25 (m, 4H), 7.11 – 7.04 (m, 1H), 7.03 – 6.93 (m, 4H), 4.02 (t, J = 7.6 Hz, 1H), 3.02 – 2.87 (m, 1H), 2.86 – 2.70 (m, 1H), 2.39 – 2.23 (m, 1H), 2.08 – 1.93 (m, 2H), 1.92 – 1.78 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 178.2, 162.6, 157.2, 156.1, 134.9, 134.6 (q, J = 32.7 Hz), 132.6, 130.0, 129.7, 129.2, 125.6 (q, J = 3.7 Hz), 123.6 (q, J = 272.8 Hz), 123.3, 118.9, 48.7, 34.9, 30.0, 22.5. HRMS (ESI) calcd. for C₂₅H₂₀O₃NF₃ $[M+Na]^+$ m/z 462.1293, found 462.1291.

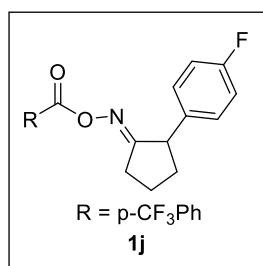


Prepared according to general procedure (A) from cyclopentanone and (3-bromophenyl)(methyl)sulfane to provide the title compound **1h** as a yellowish solid (43% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, J = 8.1 Hz, 2H), 7.72 (d, J = 8.3 Hz, 2H), 7.31 – 7.21 (m, 2H), 7.17 – 7.05 (m, 2H), 4.02 (t, J = 7.4 Hz, 1H), 3.04 – 2.88 (m, 1H), 2.87 – 2.73 (m, 1H), 2.48 (s, 3H), 2.39 – 2.25 (m, 1H), 2.10 – 1.93 (m, 2H), 1.92 – 1.77 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 177.9, 162.6, 140.9, 138.8, 134.6 (q, J = 32.7 Hz), 132.5, 130.0, 129.1, 126.3, 125.6 (q, J = 3.7 Hz), 125.1, 124.7, 123.6 (q, J = 272.8 Hz), 49.2, 34.8, 30.1, 22.6, 15.9. HRMS (ESI) calcd. for C₂₀H₁₈O₂NF₃S $[M+Na]^+$ m/z 416.0908, found 416.0901.

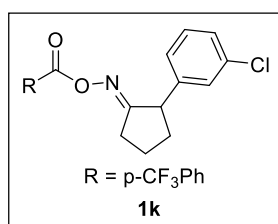


Prepared according to general procedure (A) from cyclopentanone and N-(4-bromophenyl)-N,4-dimethylbenzenesulfonamide to provide the title compound **1i** as a yellowish solid (11% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.14 (d, J = 8.1 Hz, 2H), 7.70 (d, J = 8.3 Hz, 2H), 7.41 (d, J = 8.2 Hz, 2H), 7.26 – 7.20 (m, 4H), 7.04 (d, J = 8.5 Hz, 2H), 4.01 (t, J = 7.6 Hz, 1H), 3.10 (s, 3H), 3.00 – 2.87 (m, 1H), 2.86 – 2.72 (m, 1H), 2.37 (s, 3H), 2.36 – 2.26 (m, 1H), 2.06 – 1.92 (m, 2H), 1.91 – 1.78 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 177.9, 162.6, 143.6, 140.2, 139.3, 134.5 (q, J = 32.7 Hz), 133.4, 132.4, 129.9, 129.4, 128.4, 127.8, 126.7, 125.5 (q, J = 3.6 Hz), 123.5 (q, J = 272.8 Hz), 48.8,

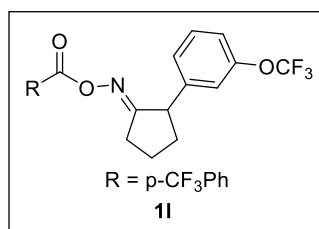
38.0 , 34.7 , 30.0 , 22.5 , 21.5. HRMS (ESI) calcd. for C₂₇H₂₅O₄N₂F₃S [M+Na]⁺ *m/z* 553.1385, found 553.1385.



Prepared according to general procedure (A) from cyclopentanone and 1-bromo-4-fluorobenzene to provide the title compound **1j** as a yellowish solid¹. ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, *J* = 8.1 Hz, 2H), 7.73 (d, *J* = 8.3 Hz, 2H), 7.29 (dd, *J* = 8.6, 5.3 Hz, 2H), 7.03 (t, *J* = 8.7 Hz, 2H), 4.03 (t, *J* = 7.3 Hz, 1H), 2.98 – 2.92 (m, 1H), 2.83 – 2.76 (m, 1H), 2.37 – 2.31 (m, 1H), 2.05 – 1.97 (m, 2H), 1.91 – 1.83 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 178.0, 162.7 (d, *J* = 20.7 Hz), 160.8, 135.7 (d, *J* = 3.3 Hz), 134.6 (q, *J* = 32.8 Hz), 132.4, 129.9, 129.4 (d, *J* = 8.0 Hz), 125.5 (q, *J* = 3.7 Hz), 123.5 (q, *J* = 272.8 Hz), 115.4 (d, *J* = 21.4 Hz), 48.6, 34.8, 29.9, 22.4.

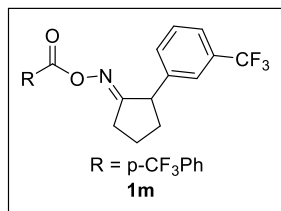


Prepared according to general procedure (A) from cyclopentanone and 1-bromo-3-chlorobenzene to provide the title compound **1k** as a yellowish solid (43% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.1 Hz, 2H), 7.74 (d, *J* = 8.3 Hz, 2H), 7.37 – 7.17 (m, 4H), 4.03 (t, *J* = 7.5 Hz, 1H), 3.04 – 2.90 (m, 1H), 2.89 – 2.74 (m, 1H), 2.43 – 2.29 (m, 1H), 2.10 – 1.95 (m, 2H), 1.95 – 1.79 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 177.6, 162.7, 142.3, 134.8 (q, *J* = 32.8 Hz), 134.5, 132.5, 130.1, 130.0, 128.1, 127.3, 126.4, 125.7 (q, *J* = 3.7 Hz), 123.7 (q, *J* = 272.9 Hz), 49.1, 34.9, 30.2, 22.6. HRMS (ESI) calcd. for C₁₉H₁₅O₂NF₃Cl [M+Na]⁺ *m/z* 404.0641, found 404.0638.



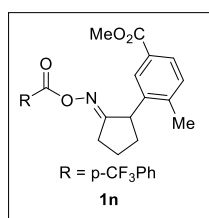
Prepared according to general procedure (A) from cyclopentanone and 1-bromo-3-(trifluoromethoxy)benzene to provide the title compound **1l** as a yellowish solid (33% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.1 Hz, 2H), 7.74 (d, *J* = 8.2 Hz, 2H), 7.37 (t, *J* = 7.9 Hz, 1H), 7.28 (d, *J* = 7.9 Hz, 1H), 7.17 (s, 1H), 7.15 – 7.08 (m, 1H), 4.07 (t, *J* = 7.2 Hz, 1H), 3.06 – 2.91 (m, 1H), 2.91 – 2.73 (m, 1H), 2.45 – 2.28 (m, 1H), 2.12 – 1.96 (m, 2H), 1.96 – 1.81 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 177.4, 162.7, 149.6, 142.6, 134.9

(q, $J = 32.7$ Hz), 132.6, 130.12, 130.08, 126.6, 125.7 (q, $J = 3.7$ Hz), δ 123.7 (q, $J = 272.8$ Hz), 120.7, 120.6 (q, $J = 257.2$ Hz), 119.4, 49.1, 35.0, 30.2, 22.6. HRMS (ESI) calcd. for $C_{20}H_{15}O_3NF_6$ $[M+Na]^+$ m/z 454.0854, found 454.0856.



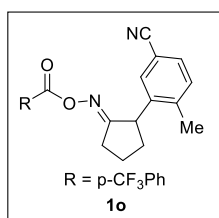
Prepared according to general procedure (A) from cyclopentanone and 1-bromo-3-(trifluoromethyl)benzene to provide the title compound **1m** as a yellowish solid (24% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, $J = 8.6$ Hz, 2H),

7.73 (d, $J = 8.6$ Hz, 2H), 7.63 – 7.41 (m, 4H), 4.10 (t, $J = 7.4$ Hz, 1H), 3.08 – 2.94 (m, 1H), 2.92 – 2.77 (m, 1H), 2.48 – 2.33 (m, 1H), 2.14 – 1.96 (m, 2H), 1.96 – 1.80 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 177.6, 162.7, 141.2, 134.9 (q, $J = 32.8$ Hz), 132.5, 131.7, 131.0 (q, $J = 32.2$ Hz), 130.1, 129.3, 125.7 (q, $J = 3.7$ Hz), 124.8 (q, $J = 3.8$ Hz), 124.2 (q, $J = 272.3$ Hz), 124.0 (q, $J = 3.8$ Hz), 123.7 (q, $J = 272.7$ Hz), 49.3, 35.1, 30.2, 22.7. HRMS (ESI) calcd. for $C_{20}H_{15}O_2NF_6$ $[M+Na]^+$ m/z 438.0905, found 438.0904.



Prepared according to general procedure (A) from cyclopentanone and methyl 3-bromo-4-methylbenzoate to provide the title compound **1n** as a white solid (42% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, $J = 8.1$ Hz, 2H), 7.85 – 7.78 (m, 2H), 7.73 (d, $J =$

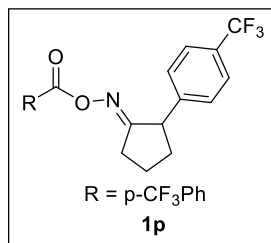
8.3 Hz, 2H), 7.30 – 7.21 (m, 1H), 4.22 (t, $J = 8.2$ Hz, 1H), 3.87 (s, 3H), 3.08 – 2.86 (m, 2H), 2.47 (s, 3H), 2.42 – 2.27 (m, 1H), 2.19 – 2.02 (m, 1H), 2.03 – 1.80 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 178.1, 167.2, 162.7, 142.0, 139.7, 134.8 (q, $J = 32.7$ Hz), 132.6, 130.9, 130.1, 128.5, 128.3, 128.2, 125.7 (q, $J = 3.7$ Hz), 123.7 (q, $J = 272.6$ Hz), 52.1, 46.5, 34.0, 30.6, 22.8, 20.3. HRMS (ESI) calcd. for $C_{22}H_{20}O_4NF_3$ $[M+Na]^+$ m/z 442.1242, found 442.1240.



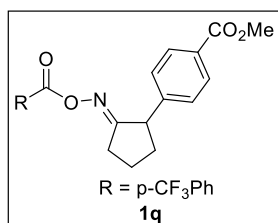
Prepared according to general procedure (A) from cyclopentanone and 3-bromo-4-methylbenzonitrile to provide the title compound **1o** as a yellowish solid (5% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, $J = 8.1$ Hz, 2H), 7.74 (d, $J = 8.3$ Hz, 2H), 7.46 – 7.40 (m,

2H), 7.27 (d, $J = 7.4$ Hz, 1H), 4.20 (t, $J = 7.6$ Hz, 1H), 3.09 – 2.83 (m, 2H), 2.46 (s, 3H), 2.42 – 2.29 (m, 1H), 2.14 – 1.98 (m, 1H), 1.98 – 1.80 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 177.4, 162.6, 142.2, 140.9, 134.8 (q, $J = 32.8$ Hz), 132.4, 131.5, 131.0,

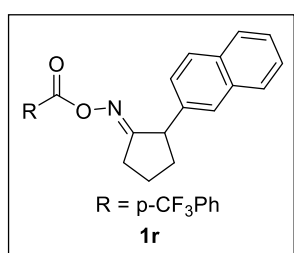
130.5 , 130.1, 125.7 (q, $J = 3.7$ Hz), 123.6 (q, $J = 272.9$ Hz), 119.2 , 110.2, 46.3, 33.9 , 30.5, 22.7 , 20.4. HRMS (ESI) calcd. for $C_{21}H_{17}O_2N_2F_3$ $[M+Na]^+$ m/z 409.1140, found 409.1129.



Prepared according to general procedure (A) from cyclopentanone and 1-bromo-4-(trifluoromethyl)benzene to provide the title compound **1p** as a yellowish solid (36% total yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.17 (d, $J = 8.2$ Hz, 2H), 7.73 (d, $J = 8.2$ Hz, 2H), 7.59 (d, $J = 8.2$ Hz, 2H), 7.45 (d, $J = 8.1$ Hz, 2H), 4.09 (t, $J = 7.6$ Hz, 1H), 3.08 – 2.93 (m, 1H), 2.91 – 2.74 (m, 1H), 2.47 – 2.31 (m, 1H), 2.12 – 1.96 (m, 2H), 1.95 – 1.80 (m, 1H); ¹³C NMR (101 MHz, Chloroform-*d*) δ 177.6, 162.7, 144.3, 134.8 (q, $J = 32.7$ Hz), 132.5, 130.1, 129.3 (q, $J = 32.5$ Hz), 128.5, 125.7 (q, $J = 3.8$ Hz), 124.3 (q, $J = 270.3$ Hz), 123.6 (q, $J = 272.8$ Hz), 49.4, 35.0, 30.2, 22.7. HRMS (ESI) calcd. for $C_{20}H_{15}O_2NF_6$ $[M+H]^+$ m/z 416.1085, found 416.1072.

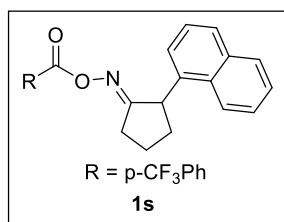


Prepared according to general procedure (A) from cyclopentanone and methyl 4-bromobenzoate to provide the title compound **1q** as a yellowish solid². ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, $J = 8.1$ Hz, 2H), 7.73 (d, $J = 8.3$ Hz, 2H), 7.25 (d, $J = 8.7$ Hz, 2H), 6.88 (d, $J = 8.7$ Hz, 2H), 4.01 (t, $J = 7.2$ Hz, 1H), 3.79 (s, 3H), 3.00 – 2.87 (m, 1H), 2.84 – 2.73 (m, 1H), 2.36 – 2.23 (m, 1H), 2.07 – 1.96 (m, 2H), 1.90 – 1.80 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 178.4, 162.6, 158.4, 134.6 (q, $J = 32.7$ Hz), 132.5, 131.9, 129.9, 128.8, 125.5 (q, $J = 3.7$ Hz), 123.5 (q, $J = 272.8$ Hz), 114.0, 55.2, 48.5, 34.6, 29.9, 22.4. HRMS (ESI) calcd. for $C_{21}H_{18}O_4NF_3$ $[M+H]^+$ m/z 406.1266, found 406.1254.



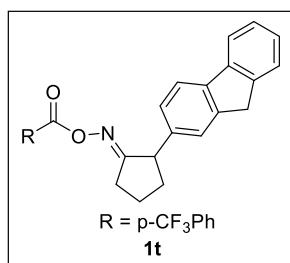
Prepared according to general procedure (A) from cyclopentanone and 2-bromonaphthalene to provide the title compound **1r** as a yellowish solid¹. ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, $J = 8.1$ Hz, 2H), 7.88 – 7.79 (m, 3H), 7.77 – 7.72 (m, 3H), 7.50 – 7.42 (m, 3H), 4.24 (t, $J = 7.7$ Hz, 1H),

3.02 – 2.96 (m, 1H), 2.91 – 2.84 (m, 1H), 2.44 – 2.37 (m, 1H), 2.22 – 2.15 (m, 1H), 2.11 – 2.03 (m, 1H), 1.97 – 1.86 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 178.1, 162.6, 137.4, 134.6 (q, *J* = 32.9 Hz), 133.4, 132.5, 132.4, 130.0, 128.4, 127.8, 127.6, 126.4, 126.1, 126.2, 125.7, 125.5 (q, *J* = 3.7 Hz), 123.5 (q, *J* = 272.8 Hz), 49.4, 34.6, 30.1, 22.6.



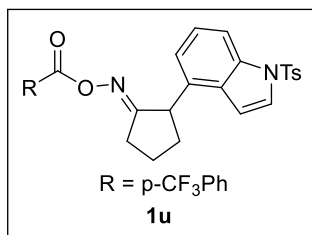
Prepared according to general procedure (A) from cyclopentanone and 1-bromonaphthalene to provide the title compound **1s** as a yellowish solid (20% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.22 (d, *J* = 8.1 Hz, 2H), 8.06 (d, *J* = 8.3

Hz, 1H), 7.88 (d, *J* = 7.8 Hz, 1H), 7.81 – 7.71 (m, 3H), 7.60 – 7.48 (m, 2H), 7.44 (t, *J* = 7.7 Hz, 1H), 7.34 (d, *J* = 7.1 Hz, 1H), 4.86 (t, *J* = 7.0 Hz, 1H), 3.03 (t, *J* = 7.5 Hz, 2H), 2.54 – 2.35 (m, 1H), 2.14 – 2.03 (m, 1H), 2.03 – 1.87 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 178.7, 162.6, 137.4, 134.6 (q, *J* = 32.7 Hz), 134.1, 132.6, 131.3, 130.0, 129.0, 127.6, 126.1, 125.7, 125.6 (q, *J* = 3.7 Hz), 125.4, 124.7, 123.6 (q, *J* = 272.7 Hz), 123.5, 46.0, 34.7, 30.6, 22.6. HRMS (ESI) calcd. for C₂₃H₁₈O₂NF₃ [M+Na]⁺ *m/z* 420.1187, found 420.1186.



Prepared according to general procedure (A) from cyclopentanone and 2-bromo-9H-fluorene to provide the title compound **1t** as a yellowish solid (38% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, *J* = 8.1 Hz, 2H), 7.75 (dt, *J* = 8.8, 4.8 Hz, 4H), 7.57 – 7.48 (m, 2H), 7.40 – 7.28 (m, 3H), 4.15 (t,

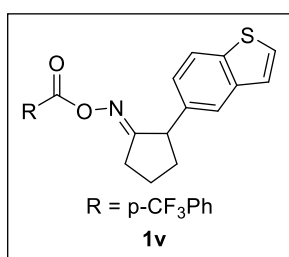
J = 7.8 Hz, 1H), 3.90 (s, 2H), 3.08 – 2.78 (m, 2H), 2.47 – 2.33 (m, 1H), 2.20 – 2.01 (m, 2H), 1.99 – 1.83 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 178.6, 162.8, 143.8, 143.4, 141.5, 140.7, 138.9, 134.7 (q, *J* = 32.9 Hz), 132.7, 130.1, 126.8, 126.7, 126.6, 125.7 (q, *J* = 3.7 Hz), 125.1, 124.7, 123.7 (q, *J* = 272.8 Hz), 120.1, 119.9, 49.5, 37.0, 35.0, 30.2, 22.7. HRMS (ESI) calcd. for C₂₆H₂₀O₂NF₃ [M+Na]⁺ *m/z* 458.1344, found 458.1342.



Prepared according to general procedure (A) from cyclopentanone and 4-bromo-1-tosyl-1*H*-indole to provide the title compound **1u** as a yellowish solid (28% total yield).

¹H NMR (400 MHz, CDCl₃) δ 8.16 (d, *J* = 8.2 Hz, 2H), 7.88 (d, *J* = 8.3 Hz, 1H), 7.75 (d, *J* = 8.5 Hz, 2H), 7.73 (d, *J* = 8.9

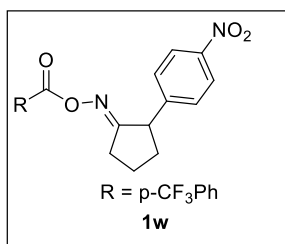
Hz, 2H), 7.60 (d, *J* = 3.7 Hz, 1H), 7.32 – 7.19 (m, 3H), 7.06 (d, *J* = 7.5 Hz, 1H), 6.71 (d, *J* = 3.7 Hz, 1H), 4.34 (t, *J* = 7.2 Hz, 1H), 3.06 – 2.84 (m, 2H), 2.42 – 2.28 (m, 4H), 2.09 – 1.85 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 177.9, 162.7, 145.2, 135.3, 135.1, 134.8 (q, *J* = 32.6 Hz), 133.9, 132.6, 130.1, 130.1, 129.7, 127.0, 126.3, 125.7 (q, *J* = 3.6 Hz), 124.9, 123.7 (q, *J* = 272.7 Hz), 121.8, 112.5, 107.2, 47.1, 34.8, 30.5, 22.9, 21.7. HRMS (ESI) calcd. for C₂₈H₂₃O₄N₂F₃S [M+Na]⁺ *m/z* 563.1228, found 563.1224.



Prepared according to general procedure (A) from cyclopentanone and 5-bromobenzo[b]thiophene to provide the title compound **1v** as a yellowish solid¹. ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.1 Hz, 2H), 7.85 (d, *J* = 8.4 Hz, 1H),

7.80 – 7.69 (m, 3H), 7.43 (d, *J* = 5.4 Hz, 1H), 7.35 – 7.28 (m,

2H), 4.19 (t, *J* = 7.9 Hz, 1H), 3.02 – 2.94 (m, 1H), 2.91 – 2.78 (m, 1H), 2.43 – 2.35 (m, 1H), 2.20 – 2.02 (m, 2H), 1.95 – 1.83 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 178.3, 162.6, 139.9, 138.4, 136.2, 134.6 (q, *J* = 32.7 Hz), 132.5, 129.9, 126.8, 125.5 (q, *J* = 3.7 Hz), 124.4, 123.8, 123.5 (q, *J* = 272.8 Hz), 122.7, 49.2, 34.9, 30.0, 22.5.

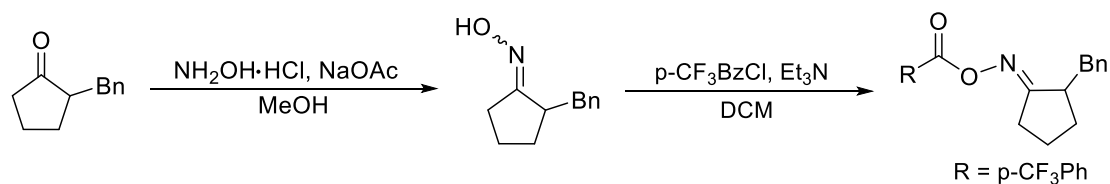


Prepared according to general procedure (A) from cyclopentanone and 1-bromo-4-nitrobenzene to provide the title compound **1w** as a yellow oil (18% total yield). ¹H NMR

(400 MHz, Chloroform-*d*) δ 8.23 – 8.07 (m, 4H), 7.71 (d, *J* = 8.3 Hz, 2H), 7.48 (d, *J* = 8.7 Hz, 2H), 4.11 (t, *J* = 8.3 Hz, 1H),

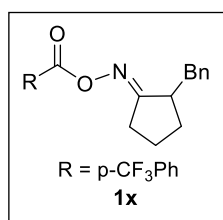
3.12 – 2.94 (m, 1H), 2.93 – 2.77 (m, 1H), 2.51 – 2.33 (m, 1H), 2.14 – 1.81 (m, 3H); ¹³C NMR (101 MHz, Chloroform-*d*) δ 177.1, 162.5, 147.8, 146.9, 134.7 (q, *J* = 32.8 Hz), 132.3, 130.0, 129.1, 125.6 (q, *J* = 3.7 Hz), 123.8, 123.5 (q, *J* = 272.8 Hz), 49.4, 35.0, 30.2, 22.7. HRMS (ESI) calcd. for C₁₉H₁₅O₄N₂F₃ [M+H]⁺ *m/z* 393.1062, found 393.1057.

The procedure for synthesis of the substrate **1x**.



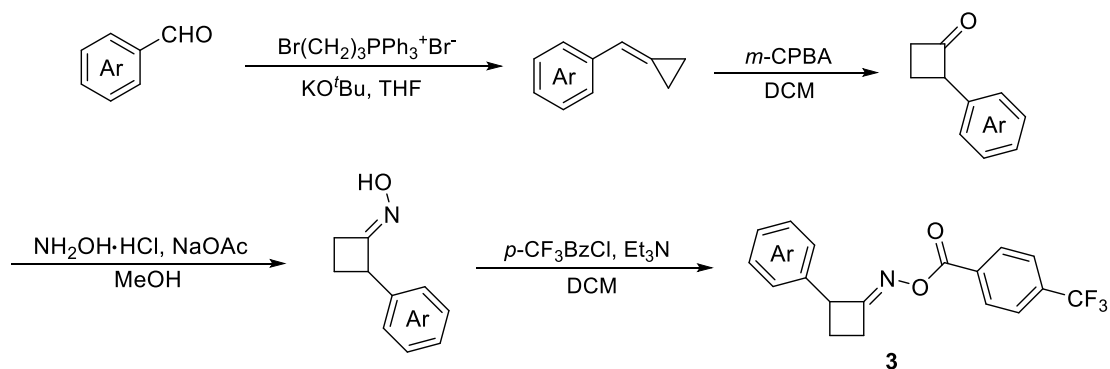
The 2-benzylcyclopentan-1-one were synthesized according to the reported procedure⁴. 2-benzylcyclopentan-1-one (1.0 equiv, 12.6 mmol), hydroxylamine hydrochloride (1.2 equiv) and sodium acetate (1.5 equiv) were added to a round bottom flask containing a stirrer bar. MeOH (0.56 M) was added and the reaction was stirred for 12 h at 75 °C. Then methanol was removed under vacuum and the resulting mixture was extracted with DCM. The organic layer was washed with water and dried over Na₂SO₄. The solvent was removed under reduced pressure and the crude material was subjected to column chromatography to afford oximes.

To a mixture of oxime (1.0 equiv, 6.7 mmol), triethylamine (1.5 equiv) and DCM (0.5 M) in a flask was added 4-(trifluoromethyl)benzoyl chlorides (1.1 equiv) slowly at 0 °C. After 1 h, a saturated solution of aqueous NaHCO₃ (10 mL) was added to the above solution, and the mixture was diluted with DCM. The organic layer was washed with brine (20 mL) and dried over Na₂SO₄. The solvent was removed under vacuum and the residue was subjected to column chromatography on silica gel with EtOAc–petroleum ether as an eluent to give the substrate.



Prepared from 2-benzylcyclopentan-1-one to provide the title compound **1x** as a white solid (77% total yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.20 (d, *J* = 8.1 Hz, 2H), 7.74 (d, *J* = 8.2 Hz, 2H), 7.38 – 7.18 (m, 5H), 3.39 (dd, *J* = 13.7, 4.2 Hz, 1H), 3.14 – 3.01 (m, 1H), 2.93 – 2.78 (m, 1H), 2.77 – 2.64 (m, 2H), 1.97 – 1.83 (m, 2H), 1.77 – 1.63 (m, 1H), 1.62 – 1.50 (m, 1H); ¹³C NMR (101 MHz, Chloroform-*d*) δ 178.1, 162.9, 139.8, 134.8 (q, *J* = 32.7 Hz), 132.7, 130.1, 129.2, 128.6, 126.4, 125.7 (q, *J* = 3.7 Hz), 123.7 (q, *J* = 272.8 Hz), 45.8, 38.1, 30.9, 29.9, 22.3. HRMS (ESI) calcd. for C₂₀H₁₈O₂NF₃ [M+H]⁺ *m/z* 362.1368, found 362.1361.

General procedure (B) for synthesis of the substrates 3⁵⁻⁹.



A solution of KO^tBu (3.0 equiv) in THF (1.3 M) was slowly added to a solution of (3-bromopropyl)triphenylphosphonium bromide (1.5 equiv) in dry THF (0.5 M) and stirred at 70 °C for 1 h. Then a THF solution of benzaldehyde (~20 mmol, 2.0 M in THF, 1.0 equiv) was added dropwise and the mixture was refluxed for 6 h. After cooling, the suspension was filtered and the solvent of the filtrate was removed under vacuum, the products were purified by column chromatography on silica gel to afford (phenylmethylene)cyclopropanes.

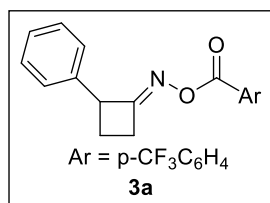
To the solution of (phenylmethylene)cyclopropanes (~15 mmol, 1.0 equiv) in DCM (0.15 M) was added a solution of *m*-CPBA (1.0 equiv) in DCM (0.38 M) dropwise at 0 °C and stirred at 0 °C (as for synthesis of substrates **3m**, **3n**, **3o**, **3p**, temperature should be elevated to RT) for ~1 h (detected by TLC). Then, the solution was diluted with a saturated solution of aqueous Na_2SO_3 (30 mL) and extracted with DCM (3*20 mL). The organic phase was washed successively with a saturated solution of aqueous NaHCO_3 (2*30 mL), and brine (30 mL), then dried over Na_2SO_4 and concentrated in vacuum. The crude material was then purified by column chromatography on silica gel with a mixture of petroleum ether and ethyl acetate to give various cyclobutanones.

To a mixture of hydroxylamine hydrochloride (1.2 equiv), sodium acetate (1.5 equiv), methanol (0.56 M) in a flask was added cyclobutanone (~5 mmol, 1.0 equiv) and the mixture was stirred at 75 °C for 12 h. The reaction mixture was cooled to room

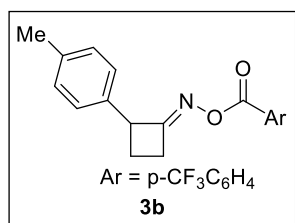
temperature and then methanol was removed under vacuum and the resulting mixture was extracted with DCM. The organic layer was washed with water and dried over Na₂SO₄. The solvent was removed under reduced pressure and the crude material was subjected to column chromatography to afford cyclobutanone oximes.

To a mixture of cyclobutanone oxime (~3 mmol, 1.0 equiv), triethylamine (1.5 equiv) and DCM (0.5 M) in a flask was added 4-(trifluoromethyl)benzoyl chlorides (1.1 equiv) slowly at 0 °C. After 1 h, a saturated solution of aqueous NaHCO₃ (10 mL) was added to the above solution, and the mixture was diluted with DCM. The organic layer was washed with brine and dried over Na₂SO₄. The solvent was removed under vacuum and the residue was subjected to column chromatography on silica gel with EtOAc–petroleum ether as an eluent to give cyclobutanone *O*-(4-(trifluoromethyl)benzoyl) oximes.

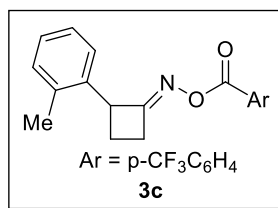
Characterization data for cyclobutanone oxime esters **3**



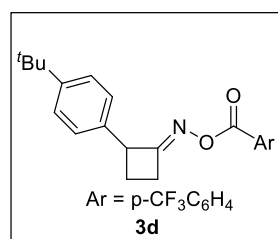
Prepared according to general procedure (B) from benzaldehyde to provide the title compound **3a** as a yellowish solid⁵. ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, *J* = 8.1 Hz, 2H), 7.74 (d, *J* = 8.2 Hz, 2H), 7.44 – 7.26 (m, 5H), 4.70 (ddd, *J* = 10.0, 7.1, 2.8 Hz, 1H), 3.31 – 3.07 (m, 2H), 2.65 (dtd, *J* = 11.3, 9.6, 6.1 Hz, 1H), 2.31 (dtd, *J* = 11.2, 9.9, 7.3 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.6, 162.7, 138.5, 134.7 (q, *J* = 33.3 Hz), 132.4, 130.0, 128.7, 127.2, 127.1, 125.5 (q, *J* = 4.0 Hz), 123.5 (q, *J* = 274.7 Hz), 49.7, 29.6, 23.2.



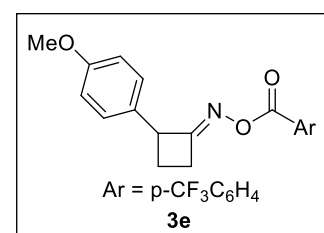
Prepared according to general procedure (B) from 4-methylbenzaldehyde to provide the title compound **3b** as a yellowish solid⁵. ¹H NMR (400 MHz, CDCl₃) δ 8.21–8.13 (m, 2H), 7.73 (d, *J* = 8.2 Hz, 2H), 7.29 (d, *J* = 7.8 Hz, 2H), 7.17 (d, *J* = 7.9 Hz, 2H), 4.66 (ddd, *J* = 10.0, 6.9, 2.9 Hz, 1H), 3.34–3.04 (m, 2H), 2.62 (dtd, *J* = 11.3, 9.7, 6.1 Hz, 1H), 2.34 (s, 3H), 2.33–2.22 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.8, 162.7, 136.8, 135.5, 134.7 (q, *J* = 32.3 Hz), 132.4, 130.0, 129.3, 127.0, 125.5 (q, *J* = 4.0 Hz), 123.5 (q, *J* = 273.7 Hz), 49.4, 29.5, 23.2, 21.1.



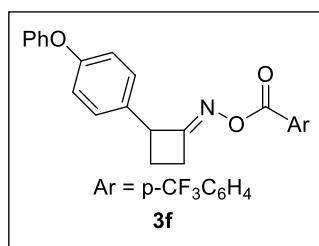
Prepared according to general procedure (B) from 2-methylbenzaldehyde to provide the title compound **3c** as a white solid⁵. ¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, *J* = 8.1 Hz, 2H), 7.74 (d, *J* = 8.2 Hz, 2H), 7.60–7.53 (m, 1H), 7.27–7.13 (m, 3H), 4.80 (ddd, *J* = 10.0, 7.1, 3.0 Hz, 1H), 3.33–3.03 (m, 2H), 2.65 (dtd, *J* = 11.2, 9.6, 6.1 Hz, 1H), 2.33 (s, 3H), 2.22–2.05 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.6, 162.7, 136.9, 135.5, 134.7 (q, *J* = 33.3 Hz), 132.4, 130.4, 130.0, 127.2, 126.2, 126.1, 125.5 (q, *J* = 4.0 Hz), 123.5 (q, *J* = 243.4 Hz), 47.9, 29.3, 23.4, 19.5.



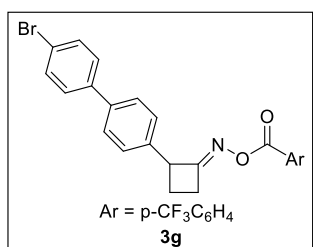
Prepared according to general procedure (B) from 4-(tert-butyl)benzaldehyde to provide the title compound **3d** as a white solid (5% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, *J* = 8.1 Hz, 2H), 7.74 (d, *J* = 8.3 Hz, 2H), 7.43 – 7.30 (m, 4H), 4.70 – 4.61 (m, 1H), 3.31 – 3.04 (m, 2H), 2.69 – 2.52 (m, 1H), 2.38 – 2.24 (m, 1H), 1.32 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 172.1, 162.9, 150.2, 135.5, 134.8 (q, *J* = 32.7 Hz), 132.6, 130.1, 127.0, 125.73, 125.67 (q, *J* = 3.7 Hz), 123.7 (q, *J* = 272.7 Hz), 49.5, 34.6, 31.4, 29.7, 23.2. HRMS (ESI) calcd. for C₂₂H₂₂O₂NF₃ [M+Na]⁺ *m/z* 412.1500, found 412.1498.



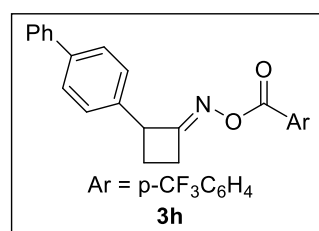
Prepared according to general procedure (B) from 4-methoxybenzaldehyde to provide the title compound **3e** as a yellowish solid⁵. ¹H NMR (400 MHz, CDCl₃) δ 7.63–7.40 (m, 4H), 7.22 (d, *J* = 8.7 Hz, 2H), 6.87 (d, *J* = 8.7 Hz, 2H), 4.50 (ddd, *J* = 10.2, 7.4, 2.8 Hz, 1H), 3.77 (s, 3H), 3.28–3.08 (m, 2H), 2.65–2.49 (m, 1H), 2.23–2.08 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.5, 162.5, 158.8, 134.3 (q, *J* = 32.3 Hz), 131.9, 131.0, 129.8, 128.4, 125.1 (q, *J* = 3.0 Hz), 123.4 (q, *J* = 273.7 Hz), 114.1, 55.2, 50.8, 29.1, 24.3.



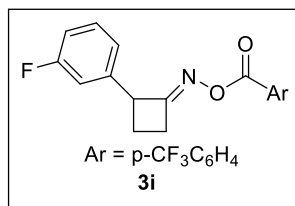
Prepared according to general procedure (B) from 4-phenoxybenzaldehyde to provide the title compound **3f** as a yellowish solid (3% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, *J* = 8.0 Hz, 2H), 7.74 (d, *J* = 8.2 Hz, 2H), 7.42 – 7.30 (m, 4H), 7.13 – 7.06 (m, 1H), 7.05 – 6.97 (m, 4H), 4.73 – 4.60 (m, 1H), 3.31 – 3.08 (m, 2H), 2.71 – 2.57 (m, 1H), 2.37 – 2.21 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.9, 162.8, 157.3, 156.4, 134.8 (q, *J* = 32.7 Hz), 133.5, 132.5, 130.1, 129.8, 128.7, 125.7 (q, *J* = 3.7 Hz), 123.7 (q, *J* = 272.9 Hz), 123.4, 119.2, 118.9, 49.2, 29.6, 23.5. HRMS (ESI) calcd. for C₂₄H₁₈O₃NF₃ [M+Na]⁺ *m/z* 448.1136, found 448.1138.



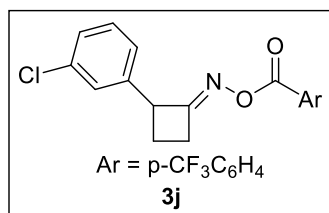
Prepared according to general procedure (B) from 4'-bromo-[1,1'-biphenyl]-4-carbaldehyde to provide the title compound **3g** as a yellowish solid (3% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.1 Hz, 2H), 7.74 (d, *J* = 8.2 Hz, 2H), 7.58 – 7.51 (m, 4H), 7.51 – 7.40 (m, 4H), 4.78 – 4.67 (m, 1H), 3.33 – 3.13 (m, 2H), 2.74 – 2.60 (m, 1H), 2.41 – 2.26 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.6, 162.9, 139.7, 139.0, 138.1, 134.8 (q, *J* = 32.6 Hz), 132.4, 132.0, 130.1, 128.8, 127.7, 127.3, 125.7 (q, *J* = 3.6 Hz), 123.7 (q, *J* = 272.7 Hz), 121.7, 49.5, 29.8, 23.3. HRMS (ESI) calcd. for C₂₄H₁₇O₂NF₃Br [M+Na]⁺ *m/z* 510.0292, found 510.0292.



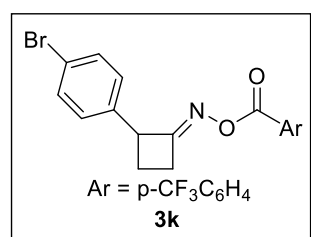
Prepared according to general procedure (B) from [1,1'-biphenyl]-4-carbaldehyde to provide the title compound **3h** as a yellowish solid (8% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.21 – 8.15 (m, 2H), 7.75 (d, *J* = 8.2 Hz, 2H), 7.62 – 7.56 (m, 4H), 7.51 – 7.41 (m, 4H), 7.39 – 7.32 (m, 1H), 4.80 – 4.69 (m, 1H), 3.37 – 3.11 (m, 2H), 2.75 – 2.62 (m, 1H), 2.43 – 2.27 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.7, 162.7, 140.7, 140.1, 137.6, 134.6 (q, *J* = 32.8 Hz), 132.4, 130.0, 128.8, 127.6, 127.4, 127.3, 127.0, 125.5, 123.6 (q, *J* = 272.8 Hz), 49.4, 29.6, 23.2. HRMS (ESI) calcd. for C₂₄H₁₈O₂NF₃ [M+Na]⁺ *m/z* 432.1187, found 432.1178.



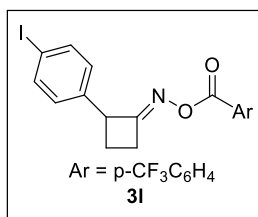
Prepared according to general procedure (B) from 3-fluorobenzaldehyde to provide the title compound **3i** as a yellowish solid⁵. ¹H NMR (400 MHz, CDCl₃): δ 7.62–7.51 (m, 4H), 7.33 (td, *J* = 7.9, 6.0 Hz, 1H), 7.10 (d, *J* = 7.7 Hz, 1H), 7.06–6.94 (m, 2H), 4.56 (ddd, *J* = 10.2, 7.2, 2.8 Hz, 1H), 3.34–3.13 (m, 2H), 2.64 (dtd, *J* = 11.5, 9.7, 6.2 Hz, 1H), 2.21 (ddt, *J* = 11.5, 9.9, 7.6 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃): δ 169.1, 163.1 (d, *J* = 247.4 Hz), 162.4, 141.3 (d, *J* = 7.1 Hz), 134.6 (d, *J* = 33.3 Hz), 131.8, 130.4 (d, *J* = 9.1 Hz), 129.8, 125.3 (q, *J* = 4.0 Hz), 123.5 (q, *J* = 273.7 Hz), 123.0 (d, *J* = 3.0 Hz), 114.5 (d, *J* = 18.2 Hz), 114.3 (d, *J* = 18.2 Hz), 50.9 (d, *J* = 2.0 Hz), 29.4, 24.2.



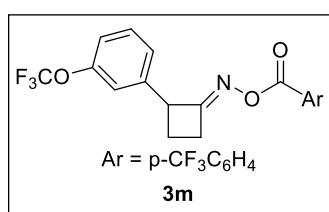
Prepared according to general procedure (B) from 3-chlorobenzaldehyde to provide the title compound **3j** as a yellowish solid (3% total yield). ¹H NMR (400 MHz, CDCl₃): δ 8.05 (d, *J* = 7.4 Hz, 2H), 7.61 (d, *J* = 8.1 Hz, 2H), 7.27 (s, 1H), 7.24 – 7.07 (m, 3H), 4.63 – 4.45 (m, 1H), 3.25 – 2.97 (m, 2H), 2.66 – 2.44 (m, 1H), 2.27 – 2.06 (m, 1H); ¹³C NMR (101 MHz, CDCl₃): δ 170.9, 162.7, 140.5, 134.7 (q, *J* = 32.6 Hz), 134.5, 132.3, 130.03, 130.00, 127.4, 127.3, 125.6 (q, *J* = 3.6 Hz), 125.5, 123.6 (q, *J* = 272.8 Hz), 49.1, 29.6, 23.2. HRMS (ESI) calcd. for C₁₈H₁₃O₂NF₃Cl [M+Na]⁺ *m/z* 390.0485, found 390.0484.



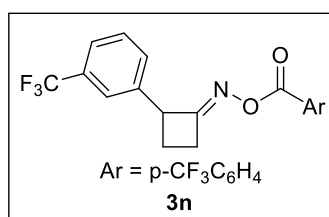
Prepared according to general procedure (B) from 4-bromobenzaldehyde to provide the title compound **3k** as a yellow solid⁵. ¹H NMR (400 MHz, CDCl₃) δ 8.16 (d, *J* = 8.1 Hz, 2H), 7.73 (d, *J* = 8.3 Hz, 2H), 7.47 (d, *J* = 8.4 Hz, 2H), 7.29 (d, *J* = 8.2 Hz, 2H), 4.64 (ddd, *J* = 10.1, 7.1, 2.9 Hz, 1H), 3.37–3.06 (m, 2H), 2.64 (dtd, *J* = 11.4, 9.6, 6.1 Hz, 1H), 2.38–2.17 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.9, 162.6, 137.4, 134.7 (q, *J* = 33.3 Hz), 132.3, 131.8, 130.0, 128.8, 125.5 (q, *J* = 3.0 Hz), 123.5 (q, *J* = 274.7 Hz), 121.1, 49.0, 29.5, 23.2.



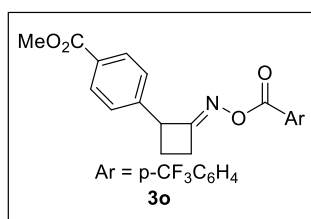
Prepared according to general procedure (B) from 4-iodobenzaldehyde to provide the title compound **3l** as a yellowish solid⁵. ¹H NMR (400 MHz, CDCl₃) δ 8.16 (d, *J* = 8.1 Hz, 2H), 7.73 (d, *J* = 8.2 Hz, 2H), 7.68 (d, *J* = 8.4 Hz, 2H), 7.16 (d, *J* = 8.0 Hz, 2H), 4.63 (ddd, *J* = 10.0, 7.1, 2.8 Hz, 1H), 3.31–3.05 (m, 2H), 2.64 (dtd, *J* = 11.4, 9.6, 6.1 Hz, 1H), 2.25 (ddt, *J* = 11.4, 10.0, 7.4 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.9, 162.6, 138.1, 137.8, 134.8 (q, *J* = 32.3 Hz), 132.3, 130.0, 129.1, 125.6 (q, *J* = 4.0 Hz), 123.5 (q, *J* = 273.7 Hz), 92.6, 49.1, 29.6, 23.1.



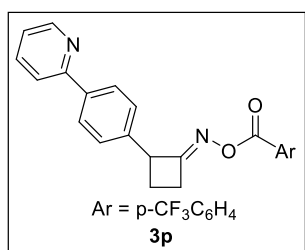
Prepared according to general procedure (B) from 3-(trifluoromethoxy)benzaldehyde to provide the title compound **3m** as a white solid (4 % total yield). ¹H NMR (CDCl₃, 400 MHz) δ 8.20 – 8.13 (m, 2H), 7.73 (dd, *J* = 8.8, 0.6 Hz, 2H), 7.40 – 7.37 (m, 2H), 7.25 – 7.22 (m, 1H), 7.17 – 7.09 (m, 1H), 4.75 – 4.65 (m, 1H), 3.32 – 3.09 (m, 2H), 2.75 – 2.59 (m, 1H), 2.36 – 2.20 (m, 1H); ¹³C NMR (CDCl₃, 101 MHz) δ 170.8, 162.8, 149.6 (q, *J* = 1.8 Hz), 140.9, 134.9 (q, *J* = 32.7 Hz), 132.4, 130.2, 130.2, 125.8, 125.7 (q, *J* = 3.8 Hz), 123.7 (q, *J* = 272.8 Hz), 120.6 (q, *J* = 257.2 Hz), 119.9, 119.7, 49.2, 29.7, 23.4. HRMS (ESI) calcd. for C₁₉H₁₃O₃NF₆ [M+Na]⁺ *m/z* 440.0697, found 440.0700.



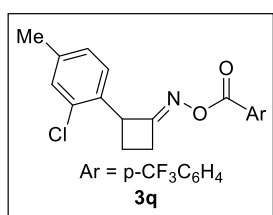
Prepared according to general procedure (B) from 3-(trifluoromethyl)benzaldehyde to provide the title compound **3n** as a yellowish liquid (4% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.16 (d, *J* = 8.7 Hz, 2H), 7.72 (d, *J* = 8.6 Hz, 2H), 7.68 – 7.58 (m, 2H), 7.56 – 7.42 (m, 2H), 4.84 – 4.66 (m, 1H), 3.40 – 3.11 (m, 2H), 2.78 – 2.61 (m, 1H), 2.40 – 2.21 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.8, 162.8, 139.6, 134.9 (q, *J* = 32.7 Hz), 132.3, 131.1 (q, *J* = 32.3 Hz), 130.9, 130.1, 129.4, 125.69 (q, *J* = 3.7 Hz), 124.2 (q, *J* = 3.8 Hz), 124.16 (q, *J* = 272.4 Hz), 123.99 (q, *J* = 3.8 Hz), 123.65 (q, *J* = 272.7 Hz), 49.3, 29.7, 23.4. HRMS (ESI) calcd. for C₁₉H₁₃O₂NF₆ [M+Na]⁺ *m/z* 423.0748, found 424.0753.



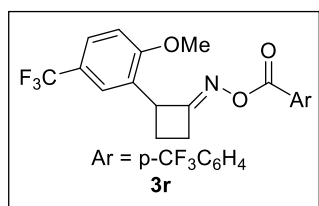
Prepared according to general procedure (B) from methyl 4-formylbenzoate to provide the title compound **3o** as a yellowish solid (2% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.14 (d, *J* = 8.2 Hz, 2H), 8.00 (d, *J* = 8.2 Hz, 2H), 7.71 (d, *J* = 8.1 Hz, 2H), 7.47 (d, *J* = 8.2 Hz, 2H), 4.79 – 4.63 (m, 1H), 3.89 (s, 3H), 3.32 – 3.08 (m, 2H), 2.75 – 2.59 (m, 1H), 2.36 – 2.20 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.9, 166.8, 162.7, 143.7, 134.8 (q, *J* = 32.8 Hz), 132.3, 130.06, 130.08, 129.1, 127.2, 125.6 (q, *J* = 3.6 Hz), 123.6 (q, *J* = 272.8 Hz), 52.1, 49.5, 29.7, 23.3. HRMS (ESI) calcd. for C₂₀H₁₆O₄NF₃ [M+Na]⁺ *m/z* 414.0929, found 414.0927.



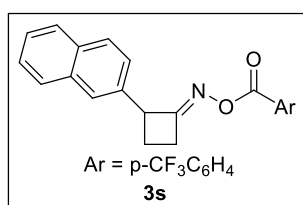
Prepared according to general procedure (B) from 4-(pyridin-2-yl)benzaldehyde to provide the title compound **3p** as a yellowish solid (14% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.67 (d, *J* = 4.6 Hz, 1H), 8.16 (d, *J* = 8.1 Hz, 2H), 7.98 (d, *J* = 8.3 Hz, 2H), 7.76 – 7.66 (m, 4H), 7.50 (d, *J* = 8.2 Hz, 2H), 7.23 – 7.17 (m, 1H), 4.79 – 4.67 (m, 1H), 3.32 – 3.09 (m, 2H), 2.73 – 2.57 (m, 1H), 2.38 – 2.24 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 171.5, 162.8, 157.0, 149.7, 139.4, 138.3, 136.9, 134.7 (q, *J* = 32.7 Hz), 132.4, 130.1, 127.6, 127.3, 125.6 (q, *J* = 3.7 Hz), 123.6 (q, *J* = 272.7 Hz), 122.2, 120.5, 49.4, 29.7, 23.3. HRMS (ESI) calcd. for C₂₃H₁₇O₂N₂F₃ [M+H]⁺ *m/z* 411.1320, found 411.1320.



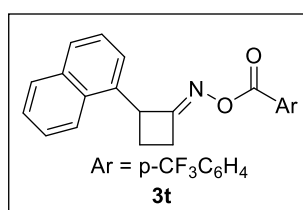
Prepared according to general procedure (B) from 2-chloro-4-methylbenzaldehyde to provide the title compound **3q** as a white solid (5% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.7 Hz, 2H), 7.72 (d, *J* = 8.6 Hz, 2H), 7.54 (d, *J* = 7.9 Hz, 1H), 7.19 (s, 1H), 7.05 (d, *J* = 7.9 Hz, 1H), 4.95 – 4.83 (m, 1H), 3.29 – 3.05 (m, 2H), 2.78 – 2.64 (m, 1H), 2.30 (s, 3H), 2.17 – 2.03 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.0, 162.7, 138.8, 134.8 (q, *J* = 32.7 Hz), 133.3, 132.9, 132.4, 130.2, 130.1, 128.2, 127.9, 125.6 (q, *J* = 3.7 Hz), 123.6 (q, *J* = 272.8 Hz), 47.7, 29.4, 24.1, 20.8. HRMS (ESI) calcd. for C₁₉H₁₅O₂NF₃Cl [M+Na]⁺ *m/z* 404.0641, found 404.0634.



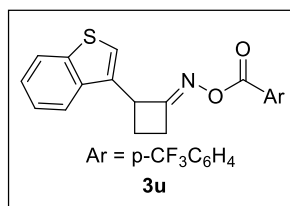
Prepared according to general procedure (B) from 2-methoxy-5-(trifluoromethyl)benzaldehyde to provide the title compound **3r** as a yellowish solid (15% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.4 Hz, 2H), 7.73 (d, *J* = 8.2 Hz, 2H), 7.66 (s, 1H), 7.52 (d, *J* = 8.6 Hz, 1H), 6.94 (d, *J* = 8.6 Hz, 1H), 4.84 – 4.72 (m, 1H), 3.90 (s, 3H), 3.35 – 3.20 (m, 1H), 3.20 – 3.06 (m, 1H), 2.67 – 2.53 (m, 1H), 2.28 – 2.14 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.6, 162.8, 159.6, 134.8 (q, *J* = 32.7 Hz), 132.6, 130.2, 128.0, 126.2 (q, *J* = 3.8 Hz), 125.9 – 125.6 (m), 124.5 (q, *J* = 271.3 Hz), 123.7 (q, *J* = 272.8 Hz), 122.9 (q, *J* = 32.7 Hz), 110.7, 55.9, 45.9, 29.7, 23.2. HRMS (ESI) calcd. for C₂₀H₁₅O₃NF₆ [M+Na]⁺ *m/z* 454.0854, found 454.0854.



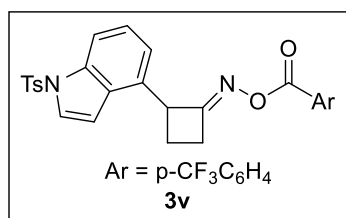
Prepared according to general procedure (B) from 2-naphthaldehyde to provide the title compound **3s** as a yellowish solid⁵. ¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, *J* = 8.1 Hz, 2H), 7.84 (ddd, *J* = 12.3, 8.4, 3.6 Hz, 4H), 7.74 (d, *J* = 8.2 Hz, 2H), 7.59–7.38 (m, 3H), 4.86 (ddd, *J* = 10.0, 6.8, 2.9 Hz, 1H), 3.41–3.13 (m, 2H), 2.71 (dtd, *J* = 11.3, 9.7, 6.1 Hz, 1H), 2.40 (ddt, *J* = 11.2, 9.9, 7.1 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.4, 162.7, 135.8, 134.7 (q, *J* = 32.3 Hz), 133.3, 132.6, 132.4, 130.0, 128.6, 127.9, 127.6, 126.3, 125.9, 125.6 (q, *J* = 3.0 Hz), 125.3, 123.5 (d, *J* = 273.7 Hz), 49.8, 29.6, 23.2.



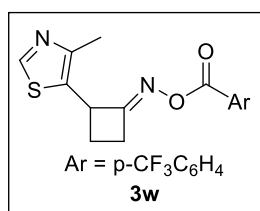
Prepared according to general procedure (B) from 1-naphthaldehyde to provide the title compound **3t** as a yellowish solid (6% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.22 (d, *J* = 8.1 Hz, 2H), 7.91 – 7.82 (m, 3H), 7.81 – 7.73 (m, 3H), 7.58 – 7.43 (m, 2H), 5.29 (t, *J* = 8.1 Hz, 1H), 3.30 – 3.21 (m, 2H), 2.91 – 2.79 (m, 1H), 2.26 – 2.12 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.3, 162.8, 134.8 (q, *J* = 32.7 Hz), 134.3, 134.1, 132.4, 130.7, 130.1, 129.1, 128.0, 126.3, 125.9, 125.7 (q, *J* = 3.3 Hz), 125.6, 123.7, 123.7 (q, *J* = 272.8 Hz), 123.4, 47.7, 29.6, 24.5. HRMS (ESI) calcd. for C₂₂H₁₆O₂NF₃ [M+Na]⁺ *m/z* 406.1031, found 406.1028.



Prepared according to general procedure (B) from benzo[b]thiophene-3-carbaldehyde to provide the title compound **3u** as a yellowish solid (2% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.20 (d, *J* = 8.0 Hz, 2H), 7.91 – 7.85 (m, 1H), 7.78 – 7.68 (m, 3H), 7.65 (d, *J* = 1.3 Hz, 1H), 7.44 – 7.35 (m, 2H), 5.00 – 4.88 (m, 1H), 3.39 – 3.17 (m, 2H), 2.85 – 2.68 (m, 1H), 2.36 – 2.20 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.8, 162.9, 141.0, 137.6, 134.9 (q, *J* = 32.8 Hz), 132.7, 130.7, 130.2, 125.7 (q, *J* = 3.7 Hz), 124.7, 124.3, 123.7 (q, *J* = 272.8 Hz), 123.2, 122.9, 121.8, 44.7, 30.0, 22.8. HRMS (ESI) calcd. for C₂₀H₁₄O₂NSF₃ [M+H]⁺ *m/z* 390.0776, found 390.0780.

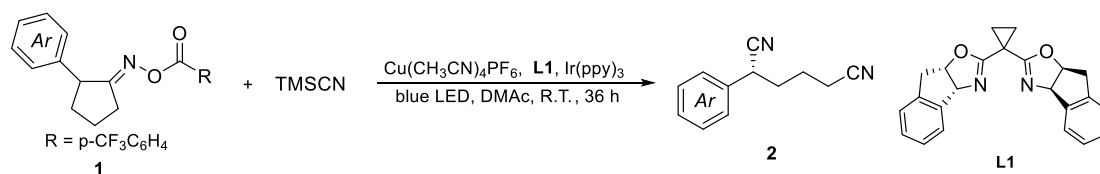


Prepared according to general procedure (B) from 1-tosyl-1*H*-indole-4-carbaldehyde to provide the title compound **3v** as a yellowish solid (8% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, *J* = 8.1 Hz, 2H), 7.91 (d, *J* = 8.3 Hz, 1H), 7.80 – 7.71 (m, 4H), 7.61 (d, *J* = 3.7 Hz, 1H), 7.40 (dt, *J* = 7.5, 0.8 Hz, 1H), 7.29 (t, 1H), 7.25 – 7.21 (m, 2H), 6.69 (dd, *J* = 3.7, 0.8 Hz, 1H), 4.96 – 4.87 (m, 1H), 3.35 – 3.13 (m, 2H), 2.75 – 2.62 (m, 1H), 2.35 (s, 3H), 2.40 – 2.25 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 171.2, 162.7, 145.2, 135.2, 135.0, 134.7 (q, *J* = 32.5 Hz), 132.4, 131.3, 130.1, 130.0, 129.0, 126.9, 126.3, 125.6 (q, *J* = 3.7 Hz), 124.8, 123.6 (q, *J* = 272.8 Hz), 121.2, 112.7, 107.0, 29.8, 23.4, 21.6. HRMS (ESI) calcd. for C₂₇H₂₁O₄N₂SF₃ [M+Na]⁺ *m/z* 549.1072, found 549.1074.



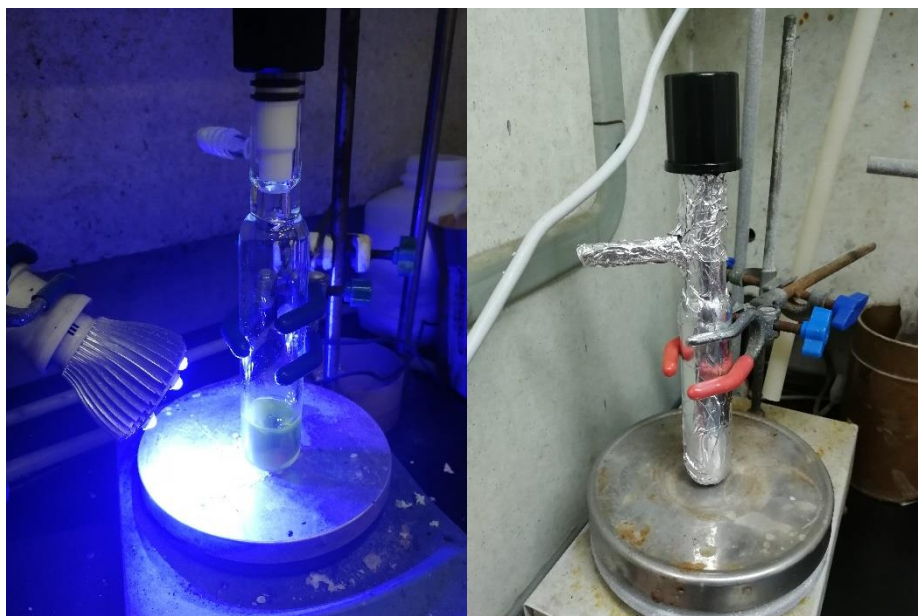
Prepared according to general procedure (B) from 4-methylthiazole-5-carbaldehyde to provide the title compound **3w** as a yellowish solid (9% total yield). ¹H NMR (400 MHz, CDCl₃) δ 8.60 (s, 1H), 8.12 (d, *J* = 8.1 Hz, 2H), 7.68 (d, *J* = 8.2 Hz, 2H), 4.87 – 4.76 (m, 1H), 3.35 – 3.22 (m, 1H), 3.20 – 3.07 (m, 1H), 2.77 – 2.64 (m, 1H), 2.41 (s, 3H), 2.20 – 2.09 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.8, 162.5, 150.3, 149.8, 134.8 (q, *J* = 32.7 Hz), 132.1, 130.1, 128.9, 125.6 (q, *J* = 3.7 Hz), 123.5 (q, *J* = 272.8 Hz), 42.2, 29.8, 25.6, 15.4. HRMS (ESI) calcd. for C₁₆H₁₃O₂N₂SF₃ [M+H]⁺ *m/z* 355.0728, found 355.0726.

General Procedure (C) for the asymmetric ring-opening cyanation of cyclopentanone oxime esters 1:



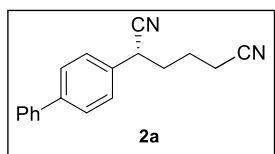
To a 25 mL Schlenk tube, $\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6$ (3.7 mg, 0.01 mmol), chiral bisoxazoline ligand **L1** (5.3 mg, 0.015 mmol), $\text{Ir}(\text{ppy})_3$ (1.6 mg, 0.0025 mmol) were added in super dry DMAc (5 mL) under N_2 atmosphere. The tube was sealed with a Teflon-lined cap, then the mixture was stirred at room temperature for 0.5 h. The **catalyst solution I** was prepared firstly and used in the next step.

To a 25 mL Schlenk tube containing substrate **1** (0.1 mmol, 1.0 equiv), **catalyst solution I** (1 mL) and TMSCN (0.15 mmol, 1.5 equiv) were sequentially added under N_2 atmosphere. The tube was sealed with a Teflon-lined cap, and the mixture was stirred at a distance of ~5 cm from a 5 W blue LEDs at room temperature for 36 h. The reaction mixture was diluted with EA (10 mL). The organic layer was washed with brine (3×5 mL) and dried over anhydrous Na_2SO_4 . After filtration and concentration, the residue was purified by silica gel chromatography with petroleum ether and ethyl acetate (PE/EA = 9:1~3:1) to afford the product **2**.



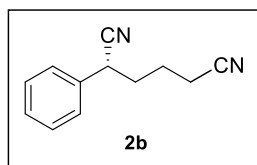
Supplementary Figure 270. photoreactions' set-up and the reaction performed in dark

Characterization data for 2-arylhexanedinitriles



Prepared according to general procedure (C) using **1a** to provide the title compound **2a** as a yellowish solid (78% yield, 92% ee; 1mmol scale: 68% yield, 90% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.68 – 7.55 (m, 4H), 7.51 – 7.44 (m, 2H), 7.44 – 7.34 (m, 3H), 3.92 (t, $J = 7.2$ Hz, 1H), 2.52 – 2.33 (m, 2H), 2.19 – 2.04 (m, 2H), 2.01 – 1.77 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 141.7, 140.2, 133.7, 129.0, 128.1, 127.9, 127.7, 127.2, 120.1, 118.8, 36.6, 34.5, 22.9, 17.0. HRMS (ESI) calcd. for $\text{C}_{18}\text{H}_{16}\text{N}_2$ $[\text{M}+\text{Na}]^+$ m/z 283.1211, found 283.1206.

HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 21.23 min (major) and 27.83 min (minor). $[\alpha]_D^{20.0} = 3.77$ (c 1.0, CHCl_3).

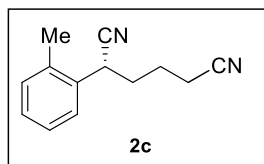


Prepared according to general procedure (C) using **1b** to provide the title compound **2b** as a yellow oil (64% yield, 91% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.46 – 7.31 (m, 5H), 3.87 (t, $J = 7.2$ Hz, 1H), 2.51 – 2.31 (m, 2H), 2.18 – 1.99 (m, 2H), 1.94 – 1.73 (m, 2H); ^{13}C NMR (101

MHz, CDCl_3) δ 141.7, 140.2, 133.7, 129.0, 128.1, 127.9, 127.7, 127.2, 120.1, 118.8, 36.6, 34.5, 22.9, 17.0.

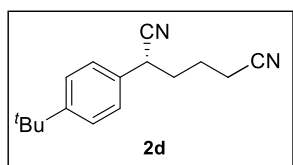
MHz, CDCl₃) δ 134.8, 129.5, 128.6, 127.3, 120.1, 118.8, 36.9, 34.6, 22.9, 16.9. HRMS (ESI) calcd. for C₁₂H₁₂N₂ [M+Na]⁺ m/z 207.0898, found 207.0889.

HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 18.83 min (minor) and 21.38 min (major). $[\alpha]_D^{20.0}$ = 37.0 (c 0.39, CHCl₃).



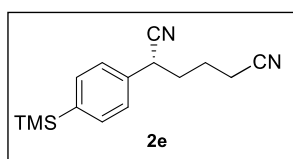
Prepared according to general procedure (C) using **1c** to provide the title compound **2c** as a yellow oil (81% yield, 90% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.46 – 7.39 (m, 1H), 7.31 – 7.18 (m, 3H), 4.07 – 3.97 (m, 1H), 2.47 – 2.39 (m, 2H), 2.36 (s, 3H), 2.09 – 1.77 (m, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 135.0, 133.2, 131.4, 128.6, 127.5, 127.2, 120.4, 118.8, 33.7, 33.1, 23.1, 19.3, 16.9. HRMS (ESI) calcd. for C₁₃H₁₄N₂ [M+Na]⁺ m/z 221.1055, found 221.1050.

HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 20.60 min (minor) and 29.09 min (major). $[\alpha]_D^{20.0}$ = 49.0 (c 0.99, CHCl₃).



Prepared according to general procedure (C) using **1d** to provide the title compound **2d** as a yellowish oil (91% yield, 88% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.46 – 7.37 (m, 2H), 7.30 – 7.23 (m, 2H), 3.84 (t, J = 7.1 Hz, 1H), 2.46 – 2.37 (m, 2H), 2.13 – 2.01 (m, 2H), 1.94 – 1.73 (m, 2H), 1.32 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 151.7, 131.7, 127.0, 126.4, 120.3, 118.8, 36.4, 34.7, 34.5, 31.4, 22.9, 16.9. HRMS (ESI) calcd. for C₁₆H₂₀N₂ [M+Na]⁺ m/z 263.1524, found 263.1522.

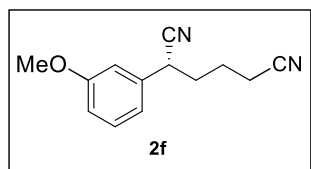
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 16.39 min (minor) and 17.83 min (major). $[\alpha]_D^{20.0}$ = 9.95 (c 1.24, CHCl₃).



Prepared according to general procedure (C) using **1e** to provide the title compound **2e** as a yellow oil (70% yield, 90% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.55 (d, J = 8.1 Hz, 2H), 7.32 (d, J = 7.9 Hz, 2H), 3.86 (t, J = 7.1 Hz, 1H), 2.51 – 2.32 (m, 2H), 2.14 – 2.00 (m, 2H), 1.95 – 1.73 (m, 2H), 0.28 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 141.3, 135.1,

134.4, 126.6, 120.0, 118.8, 36.8, 34.5, 22.9, 16.9, -1.1. HRMS (ESI) calcd. for $C_{15}H_{20}N_2Si$ $[M+Na]^+$ m/z 279.1293, found 279.1292.

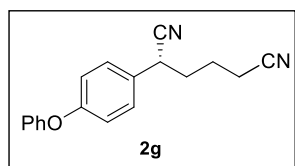
HPLC (AS-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 19/1, flow 1.0 mL/min, detection at 214 nm) retention time = 18.20 min (major) and 22.01 min (minor). $[\alpha]_D^{20.0}$ = 7.66 (c 1.34, $CHCl_3$).



Prepared according to general procedure (C) using **1f** to provide the title compound **2f** as a yellow oil (52% yield, 90% ee). 1H NMR (400 MHz, $CDCl_3$) δ 7.32 (t, J = 7.9 Hz, 1H),

6.94 – 6.85 (m, 3H), 3.84 (t, J = 7.1 Hz, 1H), 3.83 (s, 3H), 2.50 – 2.33 (m, 2H), 2.16 – 2.01 (m, 2H), 1.93 – 1.75 (m, 2H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 160.3, 136.2, 130.5, 120.0, 119.5, 118.8, 113.9, 113.2, 55.5, 36.8, 34.4, 22.8, 16.9. HRMS (ESI) calcd. for $C_{13}H_{14}N_2O$ $[M+Na]^+$ m/z 237.1004, found 237.0998.

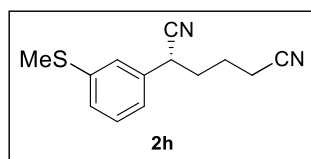
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 30.34 min (minor) and 36.25 min (major). $[\alpha]_D^{20.0}$ = 18.0 (c 0.43, $CHCl_3$).



Prepared according to general procedure (C) using **1g** to provide the title compound **2g** as a yellow oil (63% yield, 84% ee). 1H NMR (400 MHz, $CDCl_3$) δ 7.41 – 7.34 (m, 2H), 7.31

– 7.25 (m, 3H), 7.18 – 7.12 (m, 1H), 7.07 – 6.99 (m, 4H), 3.85 (t, J = 7.2 Hz, 1H), 2.50 – 2.34 (m, 2H), 2.14 – 2.01 (m, 2H), 1.97 – 1.75 (m, 2H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 157.8, 156.6, 130.0, 129.2, 128.7, 124.0, 120.1, 119.5, 119.3, 118.8, 36.2, 34.6, 22.9, 16.9. HRMS (ESI) calcd. for $C_{18}H_{16}N_2O$ $[M+Na]^+$ m/z 299.1160, found 299.1158.

HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 19.77 min (major) and 25.61 min (minor). $[\alpha]_D^{20.0}$ = -3.41 (c 0.99, $CHCl_3$).

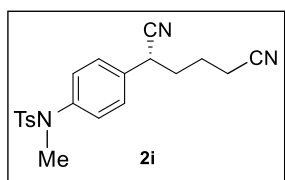


Prepared according to general procedure (C) using **1h** to provide the title compound **2h** as a yellow oil (81% yield, 90% ee). 1H NMR (400 MHz, $CDCl_3$) δ 7.36 – 7.26 (m, 1H),

7.26 – 7.16 (m, 2H), 7.09 (dt, J = 7.7, 1.4 Hz, 1H), 3.83 (t, J = 7.2 Hz, 1H), 2.50 (s, 3H), 2.47 – 2.36 (m, 2H), 2.14 – 2.00 (m, 2H), 1.95 – 1.73 (m, 2H); ^{13}C NMR (101 MHz,

CDCl₃) δ 140.4, 135.5, 129.8, 126.3, 125.0, 123.8, 119.8, 118.7, 36.8, 34.4, 22.9, 16.9, 15.7. HRMS (ESI) calcd. for C₁₃H₁₄N₂S [M+Na]⁺ m/z 253.0775, found 253.0775.

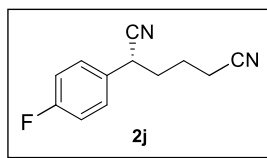
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 25.41 min (minor) and 27.72 min (major). $[\alpha]_D^{20.0}$ = 22.7 (c 1.04, CHCl₃).



Prepared according to general procedure (C) using **1i** to provide the title compound **2i** as a yellowish oil (79% yield, 90% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, J = 8.3 Hz, 2H), 7.31 – 7.23 (m, 4H), 7.14 (d, J = 8.5 Hz, 2H), 3.87 (t, J = 7.2

Hz, 1H), 3.15 (s, 3H), 2.49 – 2.36 (m, 5H), 2.12 – 1.98 (m, 2H), 1.94 – 1.74 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 144.0, 142.0, 133.6, 133.3, 129.6, 127.9, 127.8, 127.3, 119.8, 118.8, 38.0, 36.4, 34.4, 22.9, 21.7, 16.9. HRMS (ESI) calcd. for C₂₀H₂₁N₃O₂S [M+Na]⁺ m/z 390.1252, found 390.1253.

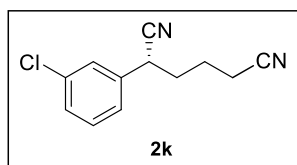
HPLC (AD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 85/15, flow 0.5 mL/min, detection at 214 nm) retention time = 88.43 min (minor) and 93.25 min (major). $[\alpha]_D^{20.0}$ = 4.15 (c 1.37, CHCl₃).



Prepared according to general procedure (C), with modifications of 1.5 mol% Cu(CH₃CN)₄PF₆ and 2.25 mol% **L1**, using **1j** to provide the title compound **2j** as a yellow oil (71% yield, 87% ee).

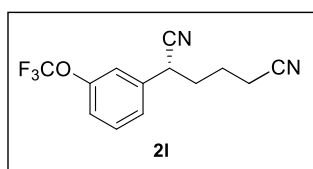
¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.29 (m, 2H), 7.15 – 7.06 (m, 2H), 3.86 (t, J = 7.2 Hz, 1H), 2.50 – 2.34 (m, 2H), 2.12 – 1.99 (m, 2H), 1.97 – 1.73 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 162.7 (d, J = 248.3 Hz), 130.6 (d, J = 3.4 Hz), 129.0 (d, J = 8.4 Hz), 119.9, 118.7, 116.5 (d, J = 21.9 Hz), 36.2, 34.6, 22.8, 16.9. HRMS (ESI) calcd. for C₁₂H₁₁N₂F [M+Na]⁺ m/z 225.0804, found 225.0806.

HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 28.40 min (major) and 32.39 min (minor). $[\alpha]_D^{20.0}$ = 18.5 (c 1.49, CHCl₃).



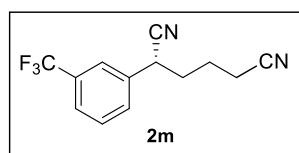
Prepared according to general procedure (C) using **1k** to provide the title compound **2k** as a yellow oil (64% yield, 91% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.31 (m, 3H), 7.26 – 7.20 (m, 1H), 3.86 (t, *J* = 7.2 Hz, 1H), 2.53 – 2.34 (m, 2H), 2.14 – 1.99 (m, 2H), 1.95 – 1.73 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 136.7, 135.4, 130.8, 129.0, 127.5, 125.5, 119.4, 118.7, 36.6, 34.4, 22.8, 16.9. HRMS (ESI) calcd. for C₁₂H₁₁N₂Cl [M+Na]⁺ *m/z* 241.0508, found 241.0503.

HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 15.20 min (minor) and 18.67 min (major). [α]_D^{20.0} = 22.0 (c 0.81, CHCl₃).



Prepared according to general procedure (C), with modifications of 1.5 mol% Cu(CH₃CN)₄PF₆ and 2.25 mol% **L1**, using **1l** to provide the title compound **2l** as a yellow oil (79% yield, 90% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.46 (t, *J* = 8.0 Hz, 1H), 7.31 (d, *J* = 7.8 Hz, 1H), 7.26 – 7.18 (m, 2H), 3.91 (t, *J* = 7.2 Hz, 1H), 2.55 – 2.36 (m, 2H), 2.17 – 2.01 (m, 2H), 1.98 – 1.76 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 149.9, 137.0, 131.0, 125.7, 121.1, 120.5 (q, *J* = 258.0 Hz), 120.0, 119.3, 118.6, 36.6, 34.4, 22.8, 16.9. HRMS (ESI) calcd. for C₁₃H₁₁N₂OF₃ [M+Na]⁺ *m/z* 291.0721, found 291.0720.

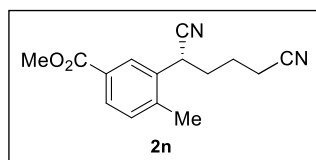
HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 92/8, flow 1.0 mL/min, detection at 214 nm) retention time = 26.14 min (minor) and 28.11 min (major). [α]_D^{20.0} = 18.8 (c 0.66, CHCl₃).



Prepared according to general procedure (C) using **1m** to provide the title compound **2m** as a yellow oil (72% yield, 90% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.70 – 7.52 (m, 4H), 3.95 (t, *J* = 7.3 Hz, 1H), 2.59 – 2.36 (m, 2H), 2.16 – 2.03 (m, 2H), 1.99 – 1.77 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 135.9, 132.0 (q, *J* = 32.8 Hz), 130.7, 130.2, 125.7 (q, *J* = 3.7 Hz), 124.1 (q, *J* = 3.7 Hz), 123.7 (q, *J* = 272.5 Hz), 119.3, 118.6, 36.8, 34.5, 22.9, 16.9. HRMS (ESI) calcd. for C₁₃H₁₁N₂F₃ [M+Na]⁺ *m/z* 275.0772, found 275.0765.

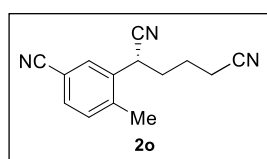
HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 92/8, flow 1.0 mL/min, detection at 214 nm) retention time = 26.14 min (minor) and 28.11 min (major). [α]_D^{20.0} = 18.8 (c 0.66, CHCl₃).

HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 22.71 min (minor) and 24.10 min (major). $[\alpha]_D^{20.0}$ = 19.9 (c 1.12, CHCl₃).



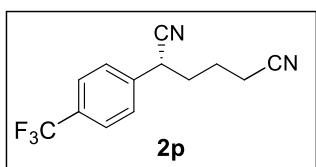
Prepared according to general procedure (C) using **1n** to provide the title compound **2n** as a yellowish oil (79% yield, 89% ee). ¹H NMR (400 MHz, CDCl₃) δ 8.08 (d, J = 1.6 Hz, 1H), 7.92 (dd, J = 7.9, 1.7 Hz, 1H), 7.30 (d, J = 8.0 Hz, 1H), 4.04 (dd, J = 8.8, 5.6 Hz, 1H), 3.93 (s, 3H), 2.55 – 2.38 (m, 5H), 2.17 – 1.80 (m, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 166.4, 140.5, 133.6, 131.6, 129.8, 129.3, 128.6, 119.8, 118.7, 52.4, 33.7, 32.9, 23.2, 19.6, 16.9. HRMS (ESI) calcd. for C₁₅H₁₆N₂O₂ [M+Na]⁺ m/z 279.1109, found 279.1107.

HPLC (ID, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 37.33 min (major) and 42.41 min (minor). $[\alpha]_D^{20.0}$ = 45.7 (c 0.94, CHCl₃).



Prepared according to general procedure (C) using **1o** to provide the title compound **2o** as a yellow solid (85% yield, 89% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.72 (d, J = 1.5 Hz, 1H), 7.56 (dd, J = 7.9, 1.6 Hz, 1H), 7.35 (d, J = 7.9 Hz, 1H), 4.04 (dd, J = 8.3, 6.0 Hz, 1H), 2.58 – 2.39 (m, 5H), 2.13 – 1.82 (m, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 140.9, 134.9, 132.4, 132.2, 131.1, 119.2, 118.5, 118.2, 111.4, 33.7, 32.8, 23.1, 19.7, 16.9. HRMS (ESI) calcd. for C₁₄H₁₃N₃ [M+Na]⁺ m/z 246.1007, found 246.1003.

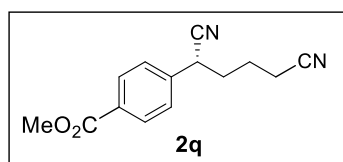
HPLC (ID, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 30.24 min (major) and 35.87 min (minor). $[\alpha]_D^{20.0}$ = 40.7 (c 0.91, CHCl₃).



Prepared according to general procedure (C) using **1p** to provide the title compound **2p** as a yellow oil (50% yield, 85% ee). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.69 (d, J = 8.1 Hz, 2H), 7.49 (d, J = 8.1 Hz, 2H), 3.96 (t, J = 7.2 Hz, 1H), 2.55 – 2.36 (m, 2H), 2.18 – 2.01 (m, 2H), 1.98 – 1.76 (m, 2H); ¹³C NMR (101 MHz, Chloroform-*d*) δ 138.8, 131.1 (q, J = 32.8 Hz), 127.8, 126.5 (q, J = 3.7 Hz), 123.8 (q, J = 270.7 Hz), 119.3, 118.6,

36.8, 34.4, 22.8, 16.9. HRMS (ESI) calcd. for $C_{13}H_{11}N_2F_3$ $[M+H]^+$ m/z 253.0953, found 253.0946.

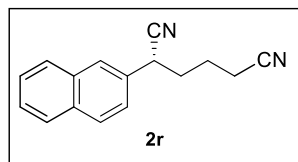
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 95/5, flow 1.0 mL/min, detection at 214 nm) retention time = 51.07 min (minor) and 53.43 min (major). $[\alpha]_D^{20.0}$ = 10.1 (c 0.73, $CHCl_3$).



Prepared according to general procedure (C) using **1q** to provide the title compound **2q** as a yellowish oil (74% yield, 76% ee). 1H NMR (400 MHz, Chloroform-*d*) δ 8.08

(d, J = 8.4 Hz, 2H), 7.43 (d, J = 8.3 Hz, 2H), 4.03 – 3.87 (m, 4H), 2.52 – 2.34 (m, 2H), 2.17 – 2.02 (m, 2H), 1.96 – 1.74 (m, 2H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 166.4, 139.6, 130.7, 130.6, 127.4, 119.4, 118.6, 52.5, 36.9, 34.3, 22.8, 16.9. HRMS (ESI) calcd. for $C_{14}H_{14}N_2O_2$ $[M+H]^+$ m/z 243.1134, found 243.1124.

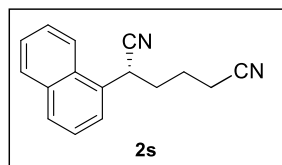
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 21.47 min (major) and 24.00 min (minor). $[\alpha]_D^{20.0}$ = 7.2 (c 1.17, $CHCl_3$).



Prepared according to general procedure (C) using **1r** to provide the title compound **2r** as a yellowish solid (79% yield, 90% ee). 1H NMR (400 MHz, $CDCl_3$) δ 7.95 – 7.81 (m, 4H),

7.60 – 7.50 (m, 2H), 7.40 (dd, J = 8.5, 1.9 Hz, 1H), 4.04 (t, J = 7.1 Hz, 1H), 2.55 – 2.29 (m, 2H), 2.23 – 2.08 (m, 2H), 1.97 – 1.74 (m, 2H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 133.4, 133.1, 132.0, 129.6, 128.0, 127.9, 127.1, 126.9, 126.5, 124.5, 120.1, 118.8, 37.0, 34.4, 22.9, 17.0. HRMS (ESI) calcd. for $C_{16}H_{14}N_2$ $[M+Na]^+$ m/z 257.1055, found 257.1054.

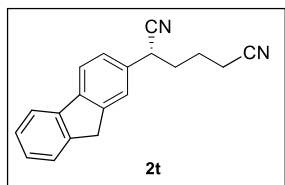
HPLC (AD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 19/1, flow 1.0 mL/min, detection at 214 nm) retention time = 31.36 min (minor) and 33.61 min (major). $[\alpha]_D^{20.0}$ = 11.7 (c 1.08, $CHCl_3$).



Prepared according to general procedure (C) using **1s** to provide the title compound **2s** as a yellow oil (89% yield, 93% ee). 1H NMR (400 MHz, $CDCl_3$) δ 7.98 – 7.83 (m, 3H), 7.69

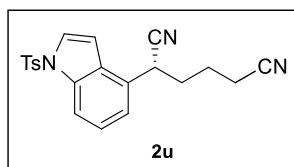
(d, J = 6.8 Hz, 1H), 7.65 – 7.47 (m, 3H), 4.64 (dd, J = 8.3, 5.5 Hz, 1H), 2.51 – 2.32 (m,

2H), 2.30 – 2.12 (m, 2H), 2.01 – 1.87 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 134.2, 130.4, 129.9, 129.6, 129.5, 127.4, 126.5, 125.8, 125.6, 121.9, 120.4, 118.8, 33.9, 33.3, 23.2, 16.9. HRMS (ESI) calcd. for $\text{C}_{16}\text{H}_{14}\text{N}_2$ $[\text{M}+\text{Na}]^+$ m/z 257.1055, found 257.1050. HPLC (AD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 17.89 min (minor) and 21.75 min (major). $[\alpha]_{\text{D}}^{20.0}$ = 98.3 (c 1.01, CHCl_3).



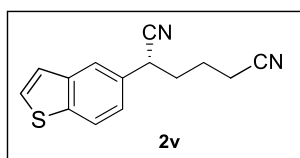
Prepared according to general procedure (C) using **1t** to provide the title compound **2t** as a yellowish solid (88% yield, 90% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.85 – 7.76 (m, 2H), 7.61 – 7.50 (m, 2H), 7.40 (t, J = 7.1 Hz, 1H), 7.37 – 7.29 (m, 2H), 4.00 – 3.87 (m, 3H), 2.49 – 2.34 (m, 2H), 2.17 – 2.06 (m, 2H), 1.97 – 1.77 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 144.5, 143.4, 142.2, 140.8, 133.0, 127.4, 127.1, 126.0, 125.3, 124.0, 120.6, 120.3, 120.2, 118.8, 36.98, 36.96, 34.8, 22.9, 16.9. HRMS (ESI) calcd. for $\text{C}_{19}\text{H}_{16}\text{N}_2$ $[\text{M}+\text{Na}]^+$ m/z 295.1211, found 295.1205.

HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 23.48 min (major) and 28.48 min (minor). $[\alpha]_{\text{D}}^{20.0}$ = 11.4 (c 1.17, CHCl_3).



Prepared according to general procedure (C) using **1u** to provide the title compound **2u** as a yellowish oil (84% yield, 89% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.99 (d, J = 8.3 Hz, 1H), 7.78 (d, J = 8.4 Hz, 2H), 7.67 (d, J = 3.8 Hz, 1H), 7.37 – 7.28 (m, 1H), 7.29 – 7.21 (m, 3H), 6.75 (dd, J = 3.8, 0.9 Hz, 1H), 4.11 (t, J = 7.4 Hz, 1H), 2.47 – 2.29 (m, 5H), 2.20 – 2.04 (m, 2H), 1.94 – 1.70 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 145.5, 135.2, 135.1, 130.2, 128.4, 127.4, 127.2, 127.0, 125.1, 122.2, 119.7, 118.7, 113.9, 105.6, 34.7, 33.3, 23.0, 21.7, 16.8. HRMS (ESI) calcd. for $\text{C}_{21}\text{H}_{19}\text{N}_3\text{O}_2\text{S}$ $[\text{M}+\text{Na}]^+$ m/z 400.1096, found 400.1096.

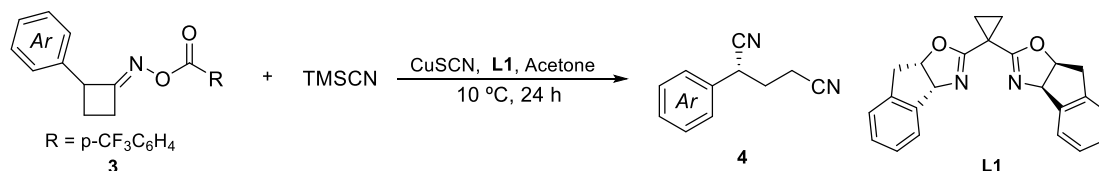
HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 39.69 min (minor) and 43.44 min (major). $[\alpha]_{\text{D}}^{20.0}$ = 9.13 (c 1.14, CHCl_3).



Prepared according to general procedure (C) using **1v** to provide the title compound **2v** as a yellow solid (67% yield, 80% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.91 (d, *J* = 8.3 Hz, 1H), 7.82 (d, *J* = 1.5 Hz, 1H), 7.54 (d, *J* = 5.4 Hz, 1H), 7.36 (d, *J* = 5.4 Hz, 1H), 7.29 (dd, *J* = 8.4, 1.7 Hz, 1H), 4.01 (t, *J* = 7.1 Hz, 1H), 2.51 – 2.34 (m, 2H), 2.21 – 2.08 (m, 2H), 1.98 – 1.77 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 140.2, 139.9, 130.9, 128.3, 123.8, 123.6, 123.1, 122.4, 120.2, 118.8, 36.9, 34.8, 22.8, 16.9. HRMS (ESI) calcd. for C₁₄H₁₂N₂S [M+Na]⁺ *m/z* 263.0619, found 263.0618.

HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 35.62 min (minor) and 38.44 min (major). [α]_D^{20.0} = 15.8 (c 0.96, CHCl₃).

General procedure (D) for the asymmetric ring-opening cyanation of cyclobutanone oxime esters **3**:

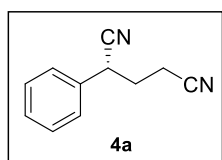


To a 5 mL Schlenk tube, CuSCN (1.8 mg, 0.015 mmol), chiral bisoxazoline ligand **L1** (6.4 mg, 0.018 mmol) were added in degassed Acetone (1 mL) under N₂ atmosphere. The tube was sealed with a Teflon-lined cap, then the mixture was stirred at room temperature for 0.5 h. The **catalyst solution II** was prepared firstly and used in the next step.

To a 25 mL Schlenk tube containing substrate **3**(0.1 mmol, 1.0 equiv), 0.8 mL Acetone, **catalyst solution II**(200 uL) and TMSCN (0.15 mmol, 1.5 equiv) were sequentially added under N₂ atmosphere. The tube was sealed with a Teflon-lined cap, and the mixture was stirred at 10 °C for 24 h . Then solvent was removed under vacuum

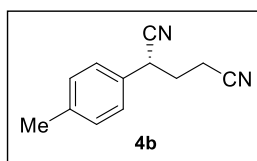
and the residue was purified by silica gel chromatography with petroleum ether and ethyl acetate (PE/EA = 10:1~5:1) to afford the product **4**.

Characterization data for 2-arylpentanedinitriles



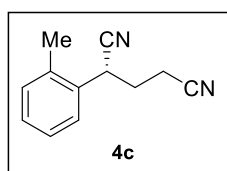
Prepared according to general procedure (D) using **3a** to provide the title compound **4a** as a colorless oil (73% yield, 94% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.48 – 7.32 (m, 5H), 3.99 (t, $J = 7.5$ Hz, 1H), 2.64 – 2.49 (m, 1H), 2.49 – 2.16 (m, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 133.5, 129.7, 129.1, 127.4, 119.3, 117.9, 36.2, 31.5, 15.1. HRMS (ESI) calcd. for $\text{C}_{11}\text{H}_{10}\text{N}_2$ $[\text{M}+\text{H}]^+$ m/z 171.0922, found 171.0926.

HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 86/14, flow 1.0 mL/min, detection at 214 nm) retention time = 19.77 min (minor) and 28.07 min (major). $[\alpha]_{\text{D}}^{20.0} = -18.7$ (c 0.52, CHCl_3).



Prepared according to general procedure (D) using **3b** to provide the title compound **4b** as a yellowish oil (64% yield, 89% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.23 (s, 4H), 3.94 (t, $J = 7.6$ Hz, 1H), 2.59 – 2.47 (m, 1H), 2.46 – 2.33 (m, 1H), 2.37 (s, 3H), 2.33 – 2.15 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 139.1, 130.5, 130.3, 127.3, 119.5, 117.9, 35.8, 31.5, 21.2, 15.1. HRMS (ESI) calcd. for $\text{C}_{12}\text{H}_{12}\text{N}_2$ $[\text{M}+\text{Na}]^+$ m/z 207.0898, found 207.0891.

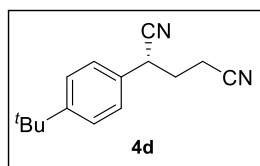
HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 86/14, flow 1.0 mL/min, detection at 214 nm) retention time = 17.97 min (minor) and 20.59 min (major). $[\alpha]_{\text{D}}^{20.0} = -20.0$ (c 0.36, CHCl_3).



Prepared according to general procedure (D) using **3c** to provide the title compound **4c** as a yellowish oil (95% yield, 93% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.43 – 7.37 (m, 1H), 7.32 – 7.20 (m, 3H), 4.16 (dd, $J = 8.8, 6.2$ Hz, 1H), 2.67 – 2.47 (m, 2H), 2.39 (s, 3H), 2.29 – 2.14 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 135.3, 131.9, 131.6, 129.1, 127.5, 127.3, 119.5, 118.0, 33.1, 30.2, 19.2, 15.4. HRMS (ESI) calcd. for $\text{C}_{12}\text{H}_{12}\text{N}_2$ $[\text{M}+\text{Na}]^+$ m/z 207.0898, found 207.0893.

HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 8/2, flow 1.0 mL/min,

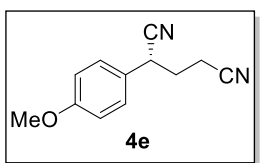
detection at 214 nm) retention time = 16.98 min (minor) and 31.80 min (major). $[\alpha]_D^{20.0}$ = 14.3 (c 0.66, CHCl₃).



Prepared according to general procedure (D) using **3d** to provide the title compound **4d** as a yellowish oil (89% yield, 89% ee).

¹H NMR (400 MHz, CDCl₃) δ 7.44 (d, *J* = 8.4 Hz, 2H), 7.27 (d, *J* = 8.2 Hz, 2H), 3.95 (t, *J* = 7.4 Hz, 1H), 2.60 – 2.49 (m, 1H), 2.47 – 2.36 (m, 1H), 2.35 – 2.17 (m, 2H), 1.32 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 152.3, 130.4, 127.1, 126.6, 119.5, 118.0, 35.7, 34.8, 31.4, 31.3, 15.1. HRMS (ESI) calcd. for C₁₅H₁₈N₂ [M+Na]⁺ *m/z* 249.1368, found 249.1359.

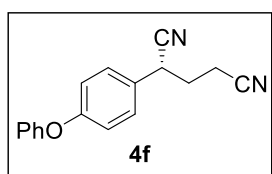
HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 86/14, flow 1.0 mL/min, detection at 214 nm) retention time = 11.31 min (minor) and 13.75 min (major). $[\alpha]_D^{20.0}$ = -25.9 (c 0.93, CHCl₃).



Prepared according to general procedure (D) using **3e** to provide the title compound **4e** as a yellowish oil (62% yield, 90% ee). ¹H

NMR (400 MHz, CDCl₃) δ 7.26 (d, *J* = 8.6 Hz, 2H), 6.94 (d, *J* = 8.7 Hz, 2H), 3.93 (t, *J* = 7.5 Hz, 1H), 3.82 (s, 3H), 2.61 – 2.47 (m, 1H), 2.47 – 2.34 (m, 1H), 2.35 – 2.13 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 160.1, 128.6, 125.3, 119.6, 118.0, 115.0, 55.5, 35.4, 31.5, 15.0. HRMS (ESI) calcd. for C₁₂H₁₂N₂O [M+Na]⁺ *m/z* 223.0847, found 223.0853.

HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 18.22 min (minor) and 20.05 min (major). $[\alpha]_D^{20.0}$ = -31.0 (c 0.56, CHCl₃).

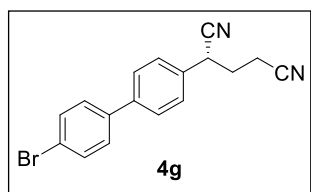


Prepared according to general procedure (D) using **3f** to provide the title compound **4f** as a yellowish oil (82% yield, 91% ee).

¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.34 (m, 2H), 7.33 – 7.27 (m, 2H), 7.20 – 7.13 (m, 1H), 7.07 – 7.00 (m, 4H), 3.97 (t, *J* = 7.5 Hz, 1H), 2.62 – 2.51 (m, 1H), 2.50 – 2.39 (m, 1H), 2.37 – 2.15 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 158.3, 156.4, 130.1, 128.9, 127.8, 124.2, 119.6, 119.4, 117.9, 35.5, 31.5, 15.1. HRMS (ESI) calcd. for C₁₇H₁₄N₂O [M+Na]⁺ *m/z* 285.1004, found 285.0998.

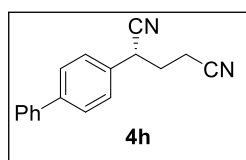
HPLC (AD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 97/3, flow 1.0 mL/min,

detection at 214 nm) retention time = 35.02 min (minor) and 36.89 min (major). $[\alpha]_D^{20.0} = -40.7$ (c 0.97, CHCl_3).



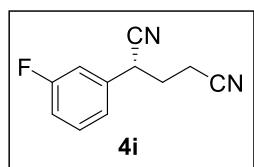
Prepared according to general procedure (D) using **3g** to provide the title compound **4g** as a yellowish solid (71% yield, 96% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.64 – 7.56 (m, 4H), 7.47 – 7.40 (m, 4H), 4.04 (t, $J = 8.8$, 1H), 2.65 – 2.54 (m, 1H), 2.52 – 2.41 (m, 1H), 2.40 – 2.21 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 141.0, 138.9, 132.8, 132.2, 128.8, 128.2, 128.0, 122.4, 119.2, 117.8, 35.9, 31.4, 15.2. HRMS (ESI) calcd. for $\text{C}_{17}\text{H}_{13}\text{N}_2\text{Br}$ $[\text{M}+\text{Na}]^+$ m/z 347.0160, found 347.0157.

HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 19.31 min (minor) and 20.85 min (major). $[\alpha]_D^{20.0} = -38.5$ (c 1.0, CHCl_3).



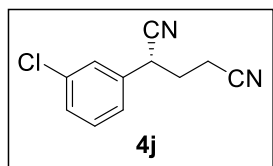
Prepared according to general procedure (D) using **3h** to provide the title compound **4h** as a yellowish solid (93% yield, 95% ee; 1 mmol scale: 87% yield, 95% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.69 – 7.63 (m, 2H), 7.62 – 7.56 (m, 2H), 7.52 – 7.35 (m, 5H), 4.04 (t, $J = 7.5$ Hz, 1H), 2.66 – 2.53 (m, 1H), 2.53 – 2.41 (m, 1H), 2.41 – 2.21 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 142.1, 140.0, 132.4, 129.1, 128.3, 128.0, 127.8, 127.2, 119.3, 117.9, 35.9, 31.4, 15.2. HRMS (ESI) calcd. for $\text{C}_{17}\text{H}_{14}\text{N}_2$ $[\text{M}+\text{Na}]^+$ m/z 269.1055, found 269.1052.

HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 24.31 min (major) and 34.52 min (minor). $[\alpha]_D^{20.0} = -40.7$ (c 1.0, CHCl_3).



Prepared according to general procedure (D) using **3i** to provide the title compound **4i** as a yellowish oil (86% yield, 91% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.47 – 7.38 (m, 1H), 7.20 – 7.06 (m, 3H), 4.00 (dd, $J = 8.1, 6.9$ Hz, 1H), 2.67 – 2.52 (m, 1H), 2.52 – 2.40 (m, 1H), 2.39 – 2.18 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.3 (d, $J = 249.1$ Hz), 135.8 (d, $J = 7.4$ Hz), 131.5 (d, $J = 8.4$ Hz), 123.2 (d, $J = 3.1$ Hz), 118.7, 117.7, 116.3 (d, $J = 21.0$ Hz), 114.7 (d, $J = 23.0$ Hz), 35.92, 35.9, 31.4, 15.2. HRMS (ESI) calcd. for $\text{C}_{11}\text{H}_9\text{N}_2\text{F}$ $[\text{M}+\text{H}]^+$ m/z 189.0828, found 189.0839.

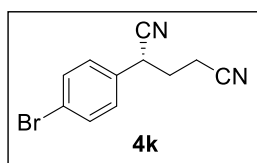
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 12.48 min (minor) and 16.94 min (major). $[\alpha]_D^{20.0} = -16.2$ (c 0.64, CHCl_3).



Prepared according to general procedure (D) using **3j** to provide the title compound **4j** as a yellowish oil (77% yield, 89% ee).

^1H NMR (400 MHz, CDCl_3) δ 7.42 – 7.34 (m, 3H), 7.29 – 7.23 (m, 1H), 3.98 (dd, $J = 8.2, 6.8$ Hz, 1H), 2.67 – 2.52 (m, 1H), 2.52 – 2.39 (m, 1H), 2.38 – 2.17 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 135.7, 135.5, 131.0, 129.5, 127.6, 125.6, 118.7, 117.6, 35.87, 31.37, 15.20. HRMS (ESI) calcd. for $\text{C}_{11}\text{H}_9\text{N}_2\text{Cl}$ $[\text{M}+\text{Na}]^+$ m/z 227.0352, found 227.0353.

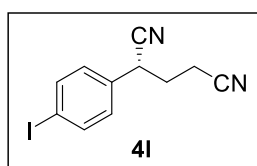
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 13.76 min (minor) and 20.52 min (major). $[\alpha]_D^{20.0} = -15.5$ (c 0.67, CHCl_3).



Prepared according to general procedure (D) using **3k** to provide the title compound **4k** as a yellowish oil (89% yield, 94% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.68 – 7.46 (m, 2H), 7.35 – 7.13 (m,

2H), 3.97 (t, $J = 8.8$ Hz, 1H), 2.65 – 2.52 (m, 1H), 2.50 – 2.37 (m, 1H), 2.35 – 2.15 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 132.9, 132.6, 129.1, 123.3, 118.8, 117.7, 35.7, 31.3, 15.2. HRMS (ESI) calcd. for $\text{C}_{11}\text{H}_9\text{N}_2\text{Br}$ $[\text{M}+\text{Na}]^+$ m/z 270.9847, found 270.9865.

HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/6/14, flow 1.0 mL/min, detection at 214 nm) retention time = 24.13 min (minor) and 28.87 min (major). $[\alpha]_D^{20.0} = -29.2$ (c 0.97, CHCl_3).

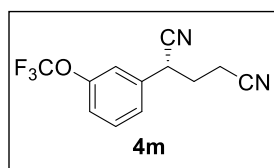


Prepared according to general procedure (D) using **3l** to provide the title compound **4l** as a yellowish oil (83% yield, 92% ee). ^1H NMR (400 MHz, CDCl_3) δ 7.77 (d, $J = 8.4$ Hz, 2H), 7.11 (d, $J =$

8.4 Hz, 2H), 3.95 (t, $J = 7.6$ Hz, 1H), 2.65 – 2.51 (m, 1H), 2.50 – 2.37 (m, 1H), 2.36 – 2.16 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 138.9, 133.2, 129.2, 118.7, 117.7, 94.9, 35.9, 31.4, 15.2. HRMS (ESI) calcd. for $\text{C}_{11}\text{H}_9\text{N}_2\text{I}$ $[\text{M}+\text{H}]^+$ m/z 296.9889, found 296.9890.

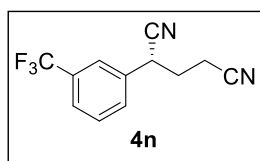
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min,

detection at 214 nm) retention time = 19.14 min (minor) and 23.26 min (major). $[\alpha]_D^{20.0}$ = -27.5 (c 1.0, CHCl₃).



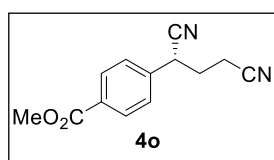
Prepared according to general procedure (D), with modifications of 5 mol% CuSCN and 6 mol% **L1**, using **3m** to provide the title compound **4m** as a yellow oil (91% yield, 93% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.49 (t, *J* = 8.0 Hz, 1H), 7.32 (d, *J* = 7.9 Hz, 1H), 7.29 – 7.24 (m, 1H), 7.23 (s, 1H), 4.03 (dd, *J* = 8.2, 6.9 Hz, 1H), 2.66 – 2.54 (m, 1H), 2.53 – 2.42 (m, 1H), 2.38 – 2.19 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 150.1, 135.8, 131.3, 125.8, 121.5, 120.5 (q, *J* = 258.3 Hz), 120.1, 118.5, 117.6, 36.0, 31.4, 15.2. HRMS (ESI) calcd. for C₁₂H₉N₂OF₃ [M+Na]⁺ *m/z* 277.0565, found 277.0555.

HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 16.68 min (minor) and 19.44 min (major). $[\alpha]_D^{20.0}$ = -8.67 (c 1.24, CHCl₃).



Prepared according to general procedure (D) using **3n** to provide the title compound **4n** as a yellowish oil (75% yield, 94% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.78 – 7.52 (m, 4H), 4.08 (dd, *J* = 8.6, 6.6 Hz, 1H), 2.69 – 2.57 (m, 1H), 2.56 – 2.45 (m, 1H), 2.40 – 2.21 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 134.8, 132.3 (q, *J* = 32.8 Hz), 130.8, 130.4, 126.1 (q, *J* = 3.7 Hz), 124.2 (q, *J* = 3.8 Hz), 123.6 (q, *J* = 272.6 Hz), 118.5, 117.5, 36.1, 31.4, 15.3. HRMS (ESI) calcd. for C₁₂H₉N₂F₃ [M+H]⁺ *m/z* 239.0796, found 239.0806.

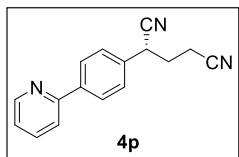
HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 20.73 min (minor) and 25.55 min (major). $[\alpha]_D^{20.0}$ = -6.31 (c 1.18, CHCl₃).



Prepared according to general procedure (D) using **3o** to provide the title compound **4o** as a yellowish oil (89% yield, 95% ee, ee of the crude product. 28% ee of **4o** was obtained after purification by silica gel chromatography). ¹H NMR (400 MHz, CDCl₃) δ 8.10 (d, *J* = 8.4 Hz, 2H), 7.45 (d, *J* = 8.3 Hz, 2H), 4.06 (dd, *J* = 8.2, 6.8 Hz, 1H), 3.94 (s, 3H), 2.64 – 2.55 (m, 1H), 2.51 – 2.40 (m, 1H), 2.38 – 2.19 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 166.2, 138.3, 131.1, 130.9, 127.5, 118.6, 117.6, 52.5, 36.2, 31.3, 15.2. HRMS (ESI)

calcd. for C₁₃H₁₂N₂O₂ [M+H]⁺ *m/z* 229.0977, found 229.0971.

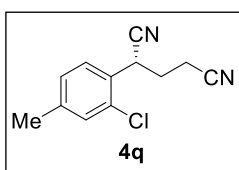
HPLC (AD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 85/15, flow 1.0 mL/min, detection at 214 nm) retention time = 14.42 min (minor) and 16.07 min (major).



Prepared according to general procedure (D) using **3p** to provide the title compound **4p** as a white solid (92% yield, 91% ee). ¹H NMR (400 MHz, CDCl₃) δ 8.75 – 8.67 (m, 1H), 8.06 (d, *J* = 8.2

Hz, 2H), 7.83 – 7.71 (m, 2H), 7.47 (d, *J* = 8.4 Hz, 2H), 7.30 – 7.24 (m, 1H), 4.06 (t, *J* = 7.4 Hz, 1H), 2.65 – 2.52 (m, 1H), 2.50 – 2.22 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 156.3, 150.0, 140.4, 137.1, 134.1, 128.2, 127.9, 122.8, 120.7, 119.2, 117.8, 36.0, 31.5, 15.1. HRMS (ESI) calcd. for C₁₆H₁₃N₃ [M+H]⁺ *m/z* 248.1188, found 248.1187.

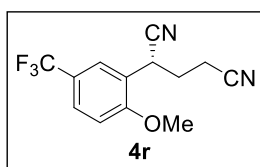
HPLC (OD-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 7/3, flow 1.2 mL/min, detection at 214 nm) retention time = 24.10 min (major) and 43.66 min (minor). [α]_D^{20.0} = -46.4 (c 0.95, CHCl₃).



Prepared according to general procedure (D) using **3q** to provide the title compound **4q** as a yellowish oil (87% yield, 89% ee). ¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 7.9 Hz, 1H), 7.26 (s, 1H),

7.19 – 7.13 (m, 1H), 4.41 (dd, *J* = 8.6, 5.8 Hz, 1H), 2.66 – 2.47 (m, 2H), 2.36 (s, 3H), 2.35 – 2.16 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 141.1, 132.6, 131.0, 128.9, 128.8, 128.3, 118.8, 117.8, 33.6, 29.8, 21.0, 15.2. HRMS (ESI) calcd. for C₁₂H₁₁N₂Cl [M+Na]⁺ *m/z* 241.0508, found 241.0506.

HPLC (AS-H, 0.46*25 cm, 5 μm, hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 21.40 min (minor) and 24.09 min (major). [α]_D^{20.0} = 27.3 (c 0.94, CHCl₃).

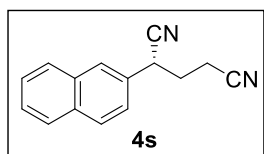


Prepared according to general procedure (D) using **3r** to provide the title compound **4r** as a yellowish oil (96% yield, 95% ee).

¹H NMR (400 MHz, CDCl₃) δ 7.74 – 7.56 (m, 2H), 7.02 (d, *J* = 9.2 Hz, 1H), 4.37 (dd, *J* = 7.9, 6.7 Hz, 1H), 3.96 (s, 3H), 2.66 – 2.44 (m, 2H), 2.37 – 2.18 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 158.7, δ 128.0 (q, *J* = 3.8 Hz), 125.7 (q, *J* = 3.6 Hz), 123.9 (q, *J* = 269.9 Hz), 123.8 (q, *J* = 33.3 Hz), 122.8, 118.8, 117.9, 111.3, 56.2, 31.0, 29.3, 15.4. HRMS (ESI) calcd. for C₁₃H₁₁N₂OF₃

$[M+Na]^+$ m/z 291.0721, found 291.0719.

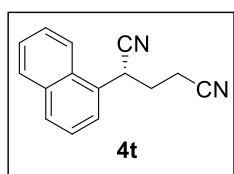
HPLC (ID, 0.46*25 cm, 5 μ m, hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 12.08 min (major) and 13.04 min (minor). $[\alpha]_D^{20.0} = 17.2$ (c 0.94, $CHCl_3$).



Prepared according to general procedure (D) using **3s** to provide the title compound **4s** as a yellowish oil (94% yield, 93% ee). 1H

NMR (400 MHz, $CDCl_3$) δ 7.93 (d, $J = 8.5$ Hz, 1H), 7.87 (dd, $J = 6.2, 3.8$ Hz, 3H), 7.60 – 7.52 (m, 2H), 7.41 (dd, $J = 8.5, 1.9$ Hz, 1H), 4.17 (t, $J = 7.2$ Hz, 1H), 2.65 – 2.53 (m, 1H), 2.50 – 2.28 (m, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 133.4, 133.2, 130.7, 129.9, 128.1, 128.0, 127.3, 127.2, 126.9, 124.3, 119.3, 117.9, 36.3, 31.4, 15.2. HRMS (ESI) calcd. for $C_{15}H_{12}N_2$ $[M+Na]^+$ m/z 243.0898, found 243.0892.

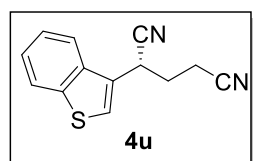
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 77/23, flow 1.0 mL/min, detection at 214 nm) retention time = 22.06 min (minor) and 37.34 min (major). $[\alpha]_D^{20.0} = -51.2$ (c 0.79, $CHCl_3$).



Prepared according to general procedure (D) using **3t** to provide the title compound **4t** as a yellowish oil (83% yield, 94% ee). 1H

NMR (400 MHz, $CDCl_3$) δ 7.98 – 7.88 (m, 3H), 7.70 (dd, $J = 7.2, 1.1$ Hz, 1H), 7.64 (ddd, $J = 8.6, 6.9, 1.4$ Hz, 1H), 7.58 (ddd, $J = 7.9, 6.9, 1.2$ Hz, 1H), 7.52 (dd, $J = 8.2, 7.2$ Hz, 1H), 4.77 (dd, $J = 8.7, 5.5$ Hz, 1H), 2.76 – 2.62 (m, 1H), 2.60 – 2.32 (m, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 134.3, 130.0, 129.8, 129.7, 129.2, 127.7, 126.7, 126.0, 125.6, 121.7, 119.5, 118.0, 33.4, 30.5, 15.5. HRMS (ESI) calcd. for $C_{15}H_{12}N_2$ $[M+H]^+$ m/z 221.1079, found 221.1075.

HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 7/3, flow 1.0 mL/min, detection at 214 nm) retention time = 18.61 min (minor) and 36.99 min (major). $[\alpha]_D^{20.0} = 31.4$ (c 0.94, $CHCl_3$).

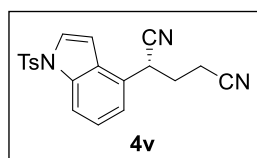


Prepared according to general procedure (D) using **3u** to provide the title compound **4u** as a yellowish oil (50% yield, 95% ee). 1H

NMR (400 MHz, $CDCl_3$) δ 7.95 – 7.90 (m, 1H), 7.82 – 7.77 (m, 1H), 7.56 (s, 1H), 7.51 – 7.42 (m, 2H), 4.45 – 4.38 (m, 1H), 2.70 – 2.60 (m, 1H), 2.55 – 2.39 (m, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 141.1, 135.9, 127.3, 125.8, 125.5, 125.2,

123.7, 120.9, 118.6, 117.9, 30.6, 29.3, 15.3. HRMS (ESI) calcd. for $C_{13}H_{10}N_2S$ $[M+Na]^+$ m/z 249.0462, found 249.0455.

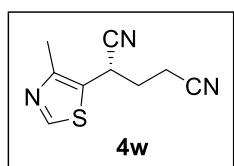
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 77/23, flow 1.0 mL/min, detection at 214 nm) retention time = 35.35 min (minor) and 41.68 min (major). $[\alpha]_D^{20.0}$ = 37.3 (c 0.22, $CHCl_3$).



Prepared according to general procedure (D), with modifications of 5 mol% $CuSCN$ and 6 mol% **L1**, using **3v** to provide the title compound **4v** as a yellowish oil (73% yield, 90% ee). 1H NMR

(400 MHz, $CDCl_3$) δ 8.03 (d, J = 8.3 Hz, 1H), 7.79 (d, J = 8.4 Hz, 2H), 7.70 (d, J = 3.8 Hz, 1H), 7.35 (t, J = 8.0 Hz, 1H), 7.30 – 7.22 (m, 3H), 6.78 (d, J = 3.8 Hz, 1H), 4.25 (t, J = 7.3 Hz, 1H), 2.63 – 2.48 (m, 1H), 2.37 (s, 3H), 2.46 – 2.21 (m, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 145.6, 135.3, 135.0, 130.2, 128.4, 127.7, 127.0, 125.8, 125.2, 122.3, 119.0, 117.9, 114.4, 105.4, 34.0, 30.4, 21.7, 15.3. HRMS (ESI) calcd. for $C_{20}H_{17}N_3O_2S$ $[M+Na]^+$ m/z 386.0939, found 386.0934.

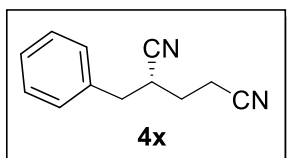
HPLC (OD-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 77/23, flow 1.0 mL/min, detection at 214 nm) retention time = 31.39 min (minor) and 34.52 min (major). $[\alpha]_D^{20.0}$ = -22.6 (c 1.23, $CHCl_3$).



Prepared according to general procedure (D) using **3w** to provide the title compound **4w** as a yellowish oil (57% yield, 93% ee). 1H NMR

(400 MHz, $CDCl_3$) δ 8.76 (s, 1H), 4.29 (t, J = 7.8 Hz, 1H), 2.67 – 2.57 (m, 1H), 2.56 – 2.46 (m, 1H), 2.51 (s, 3H), 2.46 – 2.36 (m, 1H), 2.31 – 2.19 (m, 1H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 152.1, 152.0, 123.6, 117.8, 117.4, 31.3, 28.2, 15.5, 15.3. HRMS (ESI) calcd. for $C_9H_9N_3S$ $[M+H]^+$ m/z 192.0595, found 192.0592.

HPLC (AS-H, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 24.84 min (minor) and 30.82 min (major). $[\alpha]_D^{20.0}$ = -34.0 (c 0.44, $CHCl_3$).

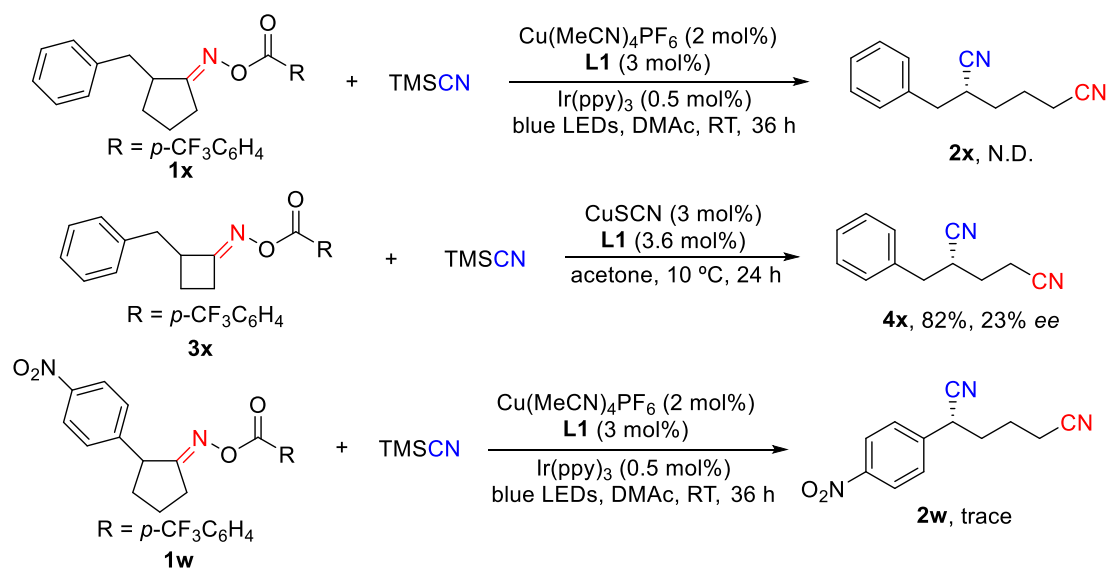


Prepared according to general procedure (D) using **3x** (prepared according to the reported literature⁵) to provide the title compound **4x** as a yellowish oil (82% yield, 23% ee). ¹H

NMR (400 MHz, Chloroform-*d*) δ 7.40 – 7.29 (m, 3H), 7.26 – 7.22 (m, 2H), 3.09 – 2.87 (m, 3H), 2.71 – 2.49 (m, 2H), 2.08 – 1.88 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 135.7, 129.12, 129.09, 127.9, 120.0, 118.0, 38.0, 32.8, 27.7, 15.5. HRMS (ESI) calcd. for C₁₂H₁₂N₂ [M+H]⁺ *m/z* 185.1079, found 185.1070.

HPLC (ID, 0.46*25 cm, 5 μ m, hexane/isopropanol = 8/2, flow 1.0 mL/min, detection at 214 nm) retention time = 10.68 min (major) and 11.35 min (minor). $[\alpha]_D^{20.0} = 5.1$ (c 1.06, CHCl₃).

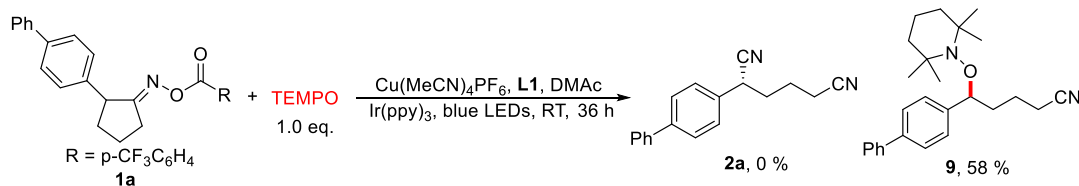
The ring-opening reactions of several limited substrates:



Supplementary Figure 271. The ring-opening reactions of several limited substrates

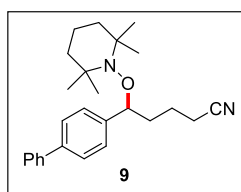
Mechanism Details

(1) Procedure of the radical trapping experiment with TEMPO:

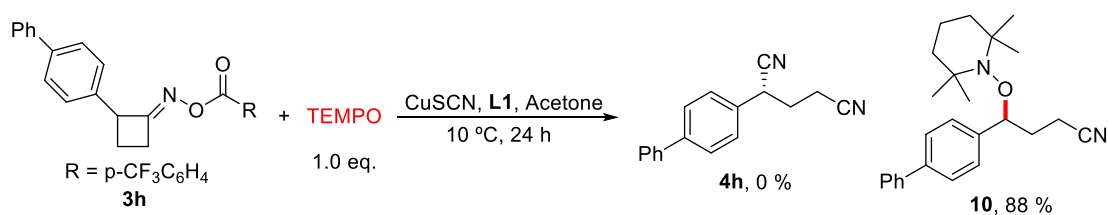


To a 25 mL Schlenk tube, Cu(CH₃CN)₄PF₆ (3.7 mg, 0.01 mmol), chiral bisoxazoline ligand **L1** (5.3 mg, 0.015 mmol), Ir(ppy)₃ (1.6 mg, 0.0025 mmol) were added in super dry DMAc (5 mL) under N₂ atmosphere. The tube was sealed with a Teflon-lined cap, then the mixture was stirred at room temperature for 0.5 h. The **catalyst solution I** was prepared firstly and used in the next step.

To a 25 mL Schlenk tube containing substrate **1a** (0.1 mmol, 1.0 equiv) and TEMPO (0.1 mmol, 1.0 equiv), **catalyst solution I** (1 mL) and TMSCN (0.15 mmol, 1.5 equiv) were sequentially added under N₂ atmosphere. The tube was sealed with a Teflon-lined cap, and the mixture was stirred at a distance of ~5 cm from a 5 W blue LEDs at room temperature for 36 h. The reaction mixture was diluted with EA (10 mL). The organic layer was washed with brine (3×5 mL) and dried over anhydrous Na₂SO₄. After filtration and concentration, the residue was purified by silica gel chromatography to afford product **9** as a yellowish oil (58 %).

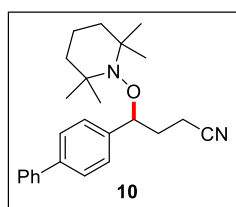


¹H NMR (400 MHz, CDCl₃) δ 7.65 – 7.59 (m, 2H), 7.59 – 7.55 (m, 2H), 7.48 – 7.41 (m, 2H), 7.38 – 7.31 (m, 3H), 4.71 (dd, *J* = 9.1, 4.0 Hz, 1H), 2.32 – 2.15 (m, 3H), 2.11 – 1.94 (m, 1H), 1.60 – 0.54 (m, 20H); ¹³C NMR (101 MHz, CDCl₃) δ 141.8, 140.9, 140.2, 128.9, 128.0, 127.4, 127.1, 126.9, 119.7, 86.1, 59.9, 40.5, 35.0, 34.6, 34.3, 21.5, 20.5, 17.4, 17.3. HRMS (ESI) calcd. for C₂₆H₃₄N₂O [M+H]⁺ *m/z* 391.2749, found 391.2743.



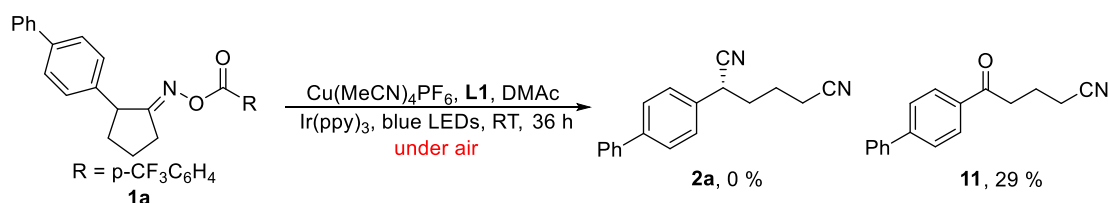
To a 5 mL Schlenk tube, CuSCN (1.8 mg, 0.015 mmol), chiral bisoxazoline ligand **L1** (6.4 mg, 0.018 mmol) were added in degassed Acetone (1 mL) under N₂ atmosphere. The tube was sealed with a Teflon-lined cap, then the mixture was stirred at room temperature for 0.5 h. The **catalyst solution II** was prepared firstly and used in the next step.

To a 25 mL Schlenk tube containing substrate **3h** (0.1 mmol, 1.0 equiv) and TEMPO (0.1 mmol, 1.0 equiv), 0.8 mL Acetone, **catalyst solution II** (200 μL), TMSCN (0.15 mmol, 1.5 equiv) were sequentially added under N₂ atmosphere. The tube was sealed with a Teflon-lined cap, and the mixture was stirred at 10 °C for 24 h. Then Solvent was removed under vacuum and the crude product was purified by flash chromatography on silica gel directly to give the product **10** as a white solid.



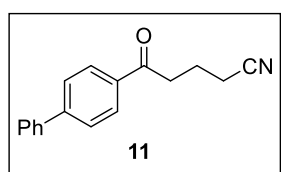
¹H NMR (400 MHz, CDCl₃) δ 7.66 – 7.55 (m, 4H), 7.49 – 7.41 (m, 2H), 7.40 – 7.31 (m, 3H), 4.85 (dd, $J = 8.8, 3.9$ Hz, 1H), 2.54 – 2.41 (m, 1H), 2.34 – 2.13 (m, 2H), 2.08 – 1.94 (m, 1H), 1.73 – 0.58 (m, 18H); ¹³C NMR (101 MHz, CDCl₃) δ 140.7, 140.7, 140.3, 128.9, 127.8, 127.5, 127.2, 127.1, 119.7, 84.9, 60.1, 40.5, 34.6, 34.2, 31.6, 20.5, 17.2, 13.4. HRMS (ESI) calcd. for C₂₅H₃₂N₂O [M+H]⁺ m/z 377.2593, found 377.2594.

(2) Procedure of the radical trapping experiment with O₂:

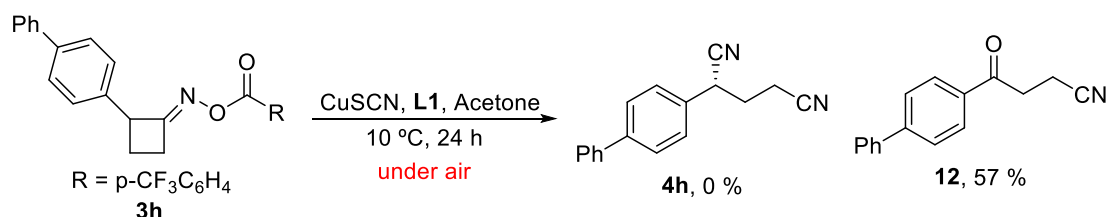


To a 25 mL screw-cap sealed tube containing substrate **1a** (0.1 mmol, 1.0 equiv), **catalyst solution I** (1 mL) and TMSCN (0.15 mmol, 1.5 equiv) were sequentially added under air atmosphere. The tube was sealed with a Teflon screw-cap, and the mixture

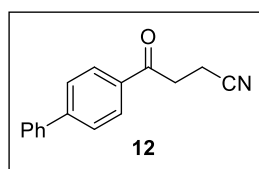
was stirred at a distance of ~5 cm from a 5 W blue LEDs at room temperature for 36 h. The reaction mixture was diluted with EA (10 mL). The organic layer was washed with brine (3×5 mL) and dried over anhydrous Na₂SO₄. After filtration and concentration, the residue was purified by silica gel chromatography to afford product **11** as a white solid.



¹H NMR (400 MHz, CDCl₃) δ 8.08 – 8.02 (m, 2H), 7.74 – 7.68 (m, 2H), 7.66 – 7.61 (m, 2H), 7.52 – 7.45 (m, 2H), 7.45 – 7.38 (m, 1H), 3.22 (t, *J* = 6.8 Hz, 2H), 2.55 (t, *J* = 7.0 Hz, 2H), 2.15 (p, *J* = 6.9 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 197.9, 146.3, 139.9, 135.3, 129.1, 128.7, 128.5, 127.5, 127.4, 119.5, 36.5, 19.9, 16.8. HRMS (ESI) calcd. for C₁₇H₁₅NO [M+Na]⁺ *m/z* 272.1051, found 272.1049.



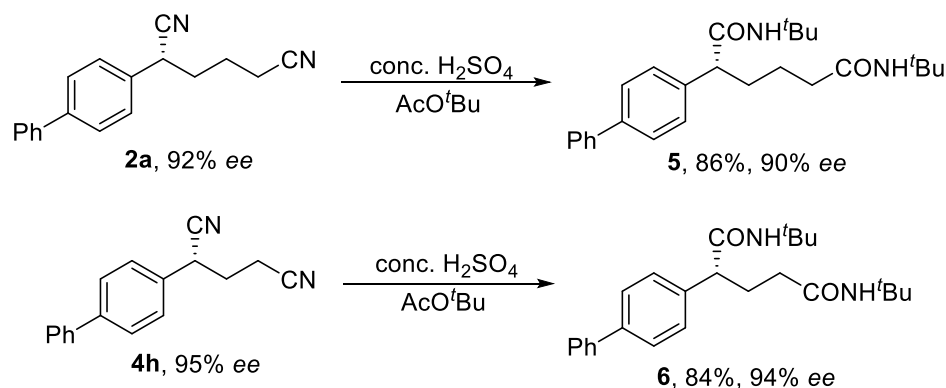
To a 25 mL screw-cap sealed tube containing substrate **3h** (0.1 mmol, 1.0 equiv), 0.8 mL Acetone, **catalyst solution II** (200 uL) and TMS-CN (0.15 mmol, 1.5 equiv) were sequentially added under air atmosphere. The tube was sealed with a Teflon screw-cap, and the mixture was stirred at 10 °C for 24 h. Then Solvent was removed under vacuum and the crude product was purified by flash chromatography on silica gel directly to give the product **12** as a white solid.



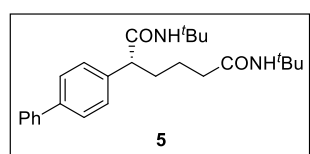
¹H NMR (400 MHz, CDCl₃) δ 8.08 – 8.00 (m, 2H), 7.76 – 7.69 (m, 2H), 7.68 – 7.61 (m, 2H), 7.53 – 7.45 (m, 2H), 7.46 – 7.40 (m, 1H), 3.42 (t, *J* = 7.2 Hz, 2H), 2.81 (t, *J* = 7.2 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 195.0, 146.7, 139.7, 134.4, 129.2, 128.8, 128.6, 127.6, 127.4, 119.4, 34.4, 12.0. HRMS (ESI) calcd. for C₁₆H₁₃NO [M+H]⁺ *m/z* 236.1075, found 236.1070.

Further transformations of the products

(1) Procedure for the synthesis of diamides **5** and **6**.¹⁰

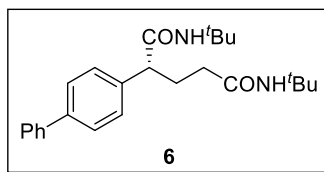


To a stirred solution of **2a** or **4h** (1.0 equiv, 0.1 mmol) in *tert*-butyl acetate (0.4 mL) in a 25 mL screw-cap sealed tube was added slowly conc. H₂SO₄ (10 μ L). The tube was sealed with a Teflon screw-cap, and the mixture was stirred at 42 °C for 2 h. The reaction mixture was diluted with EA (10 mL). The organic layer was washed with sat. aq. NaHCO₃ (2 \times 5 mL) and dried over anhydrous Na₂SO₄. After filtration and concentration, the residue was purified by silica gel chromatography to afford product **5** or **6** as a white solid.



¹H NMR (400 MHz, CDCl₃) δ 7.61 – 7.50 (m, 4H), 7.43 (t, *J* = 7.6 Hz, 2H), 7.38 – 7.29 (m, 3H), 5.48 (s, 1H), 5.45 (s, 1H), 3.28 (t, *J* = 7.4 Hz, 1H), 2.21 – 2.02 (m, 3H), 1.81 – 1.65 (m, 1H), 1.66 – 1.54 (m, 2H), 1.33 (s, 9H), 1.30 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 172.7, 172.2, 140.8, 140.0, 139.7, 128.9, 128.2, 127.5, 127.3, 127.1, 53.6, 51.4, 51.2, 37.4, 33.3, 28.9, 28.8, 24.0. HRMS (ESI) calcd. for C₂₆H₃₆O₂N₂ [M+H]⁺ *m/z* 409.2855, found 409.2849.

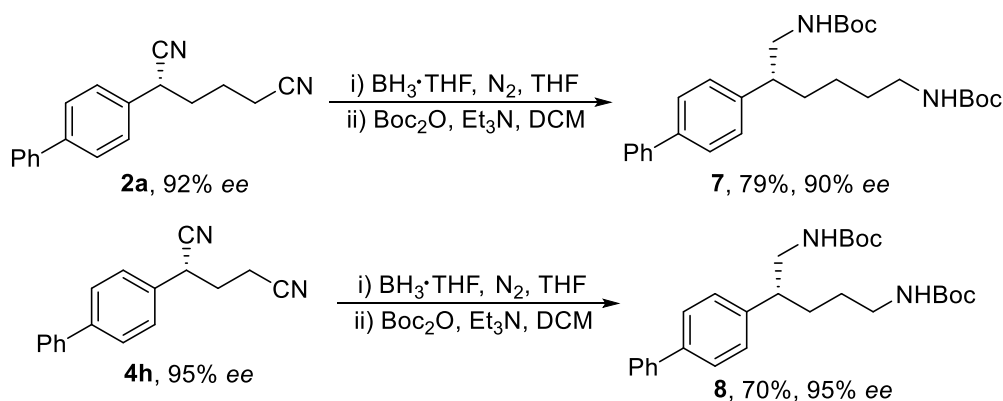
HPLC (AD-H, 0.46 \times 25 cm, 5 μ m, hexane/isopropanol = 97/3, flow 1.0 mL/min, detection at 214 nm) retention time = 8.33 min (minor) and 12.16 min (major). $[\alpha]_D^{20.0}$ = -11.6 (c 1.0, CHCl₃).



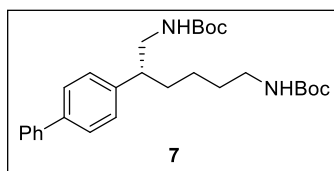
^1H NMR (400 MHz, CDCl_3) δ 7.64 – 7.51 (m, 4H), 7.48 – 7.40 (m, 2H), 7.39 – 7.30 (m, 3H), 5.56 (s, 1H), 5.41 (s, 1H), 3.44 (dd, $J = 8.0, 6.7$ Hz, 1H), 2.45 – 2.27 (m, 1H), 2.25 – 1.96 (m, 3H), 1.34 (s, 9H), 1.30 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.5, 172.0, 140.7, 140.1, 139.1, 128.9, 128.4, 127.5, 127.4, 127.1, 52.2, 51.4, 51.2, 35.2, 29.9, 28.9, 28.8. HRMS (ESI) calcd. for $\text{C}_{25}\text{H}_{34}\text{O}_2\text{N}_2$ [$\text{M}+\text{H}$] $^+$ m/z 395.2699, found 395.2692.

HPLC (AD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 97/3, flow 1.0 mL/min, detection at 214 nm) retention time = 7.41 min (minor) and 16.65 min (major). $[\alpha]_{\text{D}}^{20.0} = -16.5$ (c 1.0, CHCl_3).

(2) Procedure for the synthesis of diamines **7** and **8**.¹¹



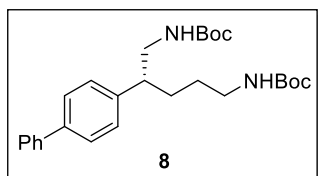
To the solution of **2a** or **4h** (0.1 mmol) in THF (0.4 mL) was added $\text{BH}_3 \cdot \text{THF}$ (0.6 mmol, 1M in THF) at room temperature under N_2 atmosphere, then the mixture was refluxed for 3 h. The reaction was quenched by the dropwise addition of MeOH. Then solvent was removed under vacuum. And 6 M aqueous HCl (~3 mL) was added followed by refluxing another 2 h. After cooling to room temperature, the solution was made basic with 6 M aqueous NaOH. Then it was extracted with DCM (5×10 mL), dried over Na_2SO_4 , filtered. After DCM was removed under vacuum, 2 mL DCM, 2.4 eq. Et_3N (0.24 mmol) and 2.4 eq. Boc_2O (0.24 mmol) were added. The mixture was stirred for 3 h at room temperature, and purified by column chromatography directly to afford the product **7** or **8** as a colorless oil.



^1H NMR (400 MHz, CDCl_3) δ 7.67 – 7.49 (m, 4H), 7.44 (t, $J = 7.6$ Hz, 2H), 7.34 (t, $J = 7.3$ Hz, 1H), 7.22 (d, $J = 8.0$ Hz, 2H), 4.52 (s, 1H), 4.46 (s, 1H), 3.63 – 3.44 (m, 1H), 3.26 – 3.11 (m, 1H), 3.10 – 2.96 (m, 2H), 2.78 (s, 1H), 1.74

– 1.53 (m, 3H), 1.52 – 1.35 (m, 19H), 1.33 – 1.14 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 156.1, 141.9, 140.9, 139.7, 128.9, 128.3, 127.5, 127.3, 127.1, 79.3, 79.1, 46.3, 45.9, 40.4, 33.3, 30.2, 28.52, 28.49, 24.6. HRMS (ESI) calcd. for $\text{C}_{28}\text{H}_{40}\text{O}_4\text{N}_2$ $[\text{M}+\text{Na}]^+$ m/z 491.2886, found 491.2885.

HPLC (AD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 9/1, flow 1.0 mL/min, detection at 214 nm) retention time = 9.63 min (major) and 11.64 min (minor). $[\alpha]_{\text{D}}^{20.0} = 32.3$ (c 1.0, CHCl_3).



^1H NMR (400 MHz, Chloroform- d) δ 7.57 (dd, $J = 14.5, 7.8$ Hz, 4H), 7.44 (t, $J = 7.6$ Hz, 2H), 7.34 (t, $J = 7.3$ Hz, 1H), 7.23 (d, $J = 8.0$ Hz, 2H), 4.49 (s, 1H), 3.62 – 3.42 (m, 1H), 3.30 – 3.13 (m, 1H), 3.15 – 2.96 (m, 2H), 2.80 (s, 1H), 1.77

– 1.67 (m, 1H), 1.66 – 1.52 (m, 2H), 1.52 – 1.31 (m, 19H); ^{13}C NMR (101 MHz, CDCl_3) δ 156.1, 141.6, 140.9, 139.8, 128.9, 128.3, 127.5, 127.3, 127.1, 79.3, 79.2, 46.3, 45.6, 40.5, 30.6, 28.5, 28.5, 28.0. HRMS (ESI) calcd. for $\text{C}_{27}\text{H}_{38}\text{O}_4\text{N}_2$ $[\text{M}+\text{Na}]^+$ m/z 477.2729, found 477.2723.

HPLC (OD-H, 0.46*25 cm, 5 μm , hexane/isopropanol = 19/1, flow 1.0 mL/min, detection at 214 nm) retention time = 17.83 min (minor) and 21.48 min (major). $[\alpha]_{\text{D}}^{20.0} = 32.6$ (c 1.0, CHCl_3).

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