

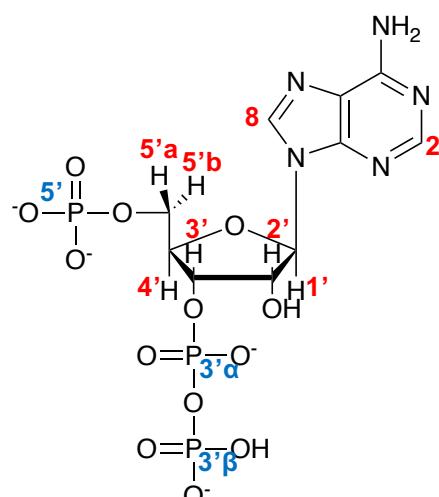
1 **Supplementary information**2
3 **Supplementary Table 1. X-ray data collection, phase and structure refinement**
4 **statistics.**

| | Tas1_{tox}-Tis1 complex (6OX6) | PurF^{EC}-ppApp complex (6OTT) |
|--|---|---|
| Data Collection | | |
| Space group | P4 ₃ 2 ₁ 2 | C222 ₁ |
| Cell dimensions | | |
| a, b, c (Å) | 66.4, 66.4, 147.9 | 115.2, 156.8, 107.5 |
| α, β, γ (°) | 90.0, 90.0, 90.0 | 90.0, 90.0, 90.0 |
| Wavelength (Å) | 0.979 | 0.979 |
| Resolution (Å) | 50.00 – 2.17 | 47.6–2.55 |
| $R_{\text{merge}}^{\text{a}}$ | 0.083 (0.243)* | 0.106 (1.27) |
| $I / \sigma(I)$ | 33.4 (6.2) | 9.0 (1.0) |
| Completeness (%) | 98.0 (96.4) | 99.8 (100) |
| Redundancy | 9.2 (4.1) | 5.8 (6.2) |
| Refinement | | |
| Resolution (Å) | 49.6 – 2.17 | 47.6 – 2.55 |
| No. reflections | | |
| Total | 18200 | 186253 |
| Unique | 17596 | 32044 |
| Free (%) | 5.00 | 6.23 |
| $R_{\text{work}} / R_{\text{free}} (%)^{\text{b}}$ | 18.5/23.1 | 22.3/26.1 |
| No. atoms | | |
| Protein | 2119 | 15508 |
| Ligand/ion | 28 | 28 |
| Water | 316 | 0 |
| B -factors (Å ²) | | |
| Protein | 40.8 | 74.6 |
| Ligand/ion | 72.0 | 61 |
| Water | 53.7 | - |
| r.m.s deviations | | |
| Bond lengths (Å) | 0.006 | 0.02 |
| Bond angles (°) | 0.993 | 0.547 |

6 Single crystals were used to collect data for each structure. *values in brackets value refer
7 to highest resolution shells.8 ^a $R_{\text{merge}} = \sum_{\text{hkl}} \sum_j |I_{\text{hkl},j} - \langle I_{\text{hkl}} \rangle| / \sum_{\text{hkl}} \sum_j I_{\text{hkl},j}$, where $I_{\text{hkl},j}$ and $\langle I_{\text{hkl}} \rangle$ are the j th and mean measurement
9 of the intensity of reflection j .10 ^b $R = \sum |F_p^{\text{obs}} - F_p^{\text{calc}}| / \sum F_p^{\text{obs}}$, where F_p^{obs} and F_p^{calc} are the observed and calculated structure
11 factor amplitudes, respectively.

12

Supplementary Table 2. ^1H and ^{31}P NMR assignments for pApp. The chemical shifts (in ppm) for pApp. α -proton of the pyrophosphate group was identified from its ddd pattern with a characteristic J-coupling constant with the proximal ^{31}P nuclei at 8.5Hz (0.014 ppm for protons or 0.036ppm for ^{31}P). This proton was assigned to the 3' location by comparing proton chemical shifts to those observed for adenosine-5'-phosphate (AMP, BMRB database ID: BMSE000837) and a chemically synthesized adenosine-3', 5'-bisdiphosphate (ppApp) under the same condition¹. Note that compared to a free hydroxyl group, pyrophosphorylation increases the chemical shift of the α -proton by 0.3-0.4 ppm.



| | Base (¹ H) | | Ribose (¹ H) | | | | | Phosphate (³¹ P) | | | |
|-------|----------------------------|-----------|--------------------------|-----------|------------|-------------|-----------|------------------------------|-----------|------------|-----------|
| | Position | 2 | 8 | 1' | 2' | 3' | 4' | 5'a / 5'b | 5'a | 3'a | 3'b |
| pApp | Chemical Shift (ppm) | 8.15 | 8.59 | 6.16 | 4.88 | 4.93 | 4.55 | 4.02 | 3.75 | -10.00 | -5.06 |
| | Integration (multiplicity) | 1H (s) | 1H (s) | 1H (d) | 1H (dd) | 1H (ddd) | 1H (m) | 2H (m) | 1P (s) | 1P (dd) | 1P (d) |
| ppApp | Chemical Shift (ppm) | 8.19 | 8.48 | 6.12 | 4.79 | 4.90 | 4.54 | 4.16 | - | - | - |
| | Integration (multiplicity) | 1H (s) | 1H (s) | 1H (d) | 1H (dd) | 1H (ddd) | 1H (m) | 2H (m) | - | - | - |
| AMP | Chemical Shift (ppm) | 8.13 | 8.48 | 6.09 | 4.75 | 4.51 | 4.38 | 4.10 | - | - | - |
| | Integration (multiplicity) | 1H (s) | 1H (s) | 1H (d) | 1H (dd) | 1H (dd) | 1H (m) | 2H (m) | - | - | - |

* “-” indicates assignment not clear or data not available.

1 **Supplementary Table 3. Absolute nucleotide quantification of *P. aeruginosa* cells**
 2 **undergoing Tas1-mediated intoxication.**

3

| Metabolite | Concentration in Tis1 depleted cells (nmol/OD) | | | | Concentration in Tis1 expressing cells (nmol/OD) | | |
|------------|---|-------|-------|-------|---|-------|-------|
| | time (min) | | | | time (min) | | |
| | 0 | 30 | 60 | 120 | 0 | 30 | 60 |
| AMP | | | | | | | |
| | R1 | 0.179 | 0.193 | 0.238 | 0.315 | 0.253 | 0.323 |
| | R2 | 0.151 | 0.172 | 0.207 | 0.236 | 0.369 | 0.326 |
| | R3 | 0.218 | 0.213 | 0.170 | 0.284 | 0.365 | 0.350 |
| ADP | | | | | | | |
| | R1 | 0.547 | 0.441 | 0.387 | 0.330 | 0.631 | 0.747 |
| | R2 | 0.507 | 0.407 | 0.379 | 0.232 | 0.700 | 0.634 |
| | R3 | 0.731 | 0.537 | 0.340 | 0.346 | 0.707 | 0.681 |
| ATP | | | | | | | |
| | R1 | 2.716 | 2.091 | 1.462 | 0.939 | 2.648 | 2.484 |
| | R2 | 3.212 | 2.054 | 1.299 | 0.614 | 2.042 | 2.281 |
| | R3 | 3.191 | 1.992 | 1.264 | 0.916 | 2.495 | 2.221 |
| ppApp | | | | | | | |
| | R1 | 0.128 | 1.020 | 1.186 | 1.154 | n.d. | n.d. |
| | R2 | 0.126 | 1.095 | 1.073 | 0.773 | n.d. | n.d. |
| | R3 | 0.154 | 1.223 | 1.105 | 1.226 | n.d. | n.d. |
| pppApp | | | | | | | |
| | R1 | 0.355 | 3.588 | 3.964 | 3.768 | n.d. | n.d. |
| | R2 | 0.576 | 3.986 | 3.763 | 2.609 | n.d. | n.d. |
| | R3 | 0.588 | 4.628 | 3.184 | 3.909 | n.d. | n.d. |
| GMP | | | | | | | |
| | R1 | 0.077 | 0.021 | 0.032 | 0.034 | 0.072 | 0.093 |
| | R2 | 0.048 | 0.019 | 0.022 | 0.003 | 0.090 | 0.094 |
| | R3 | 0.050 | 0.074 | 0.031 | 0.048 | 0.098 | 0.076 |
| GDP | | | | | | | |
| | R1 | 0.132 | 0.113 | 0.096 | 0.102 | 0.197 | 0.233 |
| | R2 | 0.141 | 0.115 | 0.085 | 0.065 | 0.203 | 0.219 |
| | R3 | 0.171 | 0.141 | 0.082 | 0.093 | 0.210 | 0.226 |
| GTP | | | | | | | |
| | R1 | 0.874 | 0.748 | 0.490 | 0.340 | 1.095 | 0.972 |
| | R2 | 1.060 | 0.742 | 0.410 | 0.249 | 0.785 | 0.907 |
| | R3 | 0.928 | 0.703 | 0.433 | 0.335 | 0.916 | 0.799 |
| UTP | | | | | | | |
| | R1 | 1.548 | 1.140 | 0.719 | 0.393 | 2.111 | 2.235 |
| | R2 | 1.780 | 1.192 | 0.590 | 0.284 | 1.745 | 2.046 |
| | R3 | 1.498 | 1.134 | 0.642 | 0.411 | 1.870 | 1.774 |
| dATP | | | | | | | |
| | R1 | 0.147 | 0.126 | 0.088 | 0.063 | 0.114 | 0.108 |
| | R2 | 0.164 | 0.120 | 0.078 | 0.041 | 0.093 | 0.090 |

| | | | | | | | | |
|-------------------|----|-------|-------|-------|-------|-------|-------|-------|
| | R3 | 0.163 | 0.116 | 0.073 | 0.057 | 0.102 | 0.088 | 0.065 |
| dCTP | R1 | 0.233 | 0.152 | 0.100 | 0.057 | 0.206 | 0.189 | 0.146 |
| | R2 | 0.258 | 0.159 | 0.082 | 0.040 | 0.167 | 0.168 | 0.143 |
| | R3 | 0.224 | 0.157 | 0.087 | 0.057 | 0.189 | 0.158 | 0.108 |
| dTTP | R1 | 0.264 | 0.220 | 0.168 | 0.090 | 0.241 | 0.228 | 0.222 |
| | R2 | 0.306 | 0.224 | 0.150 | 0.072 | 0.202 | 0.199 | 0.216 |
| | R3 | 0.297 | 0.222 | 0.143 | 0.106 | 0.228 | 0.195 | 0.178 |
| NAD ⁺ | R1 | 0.676 | 0.896 | 0.814 | 0.746 | 0.572 | 0.577 | 0.554 |
| | R2 | 0.675 | 0.837 | 0.841 | 0.628 | 0.542 | 0.531 | 0.562 |
| | R3 | 0.760 | 0.868 | 0.787 | 0.677 | 0.583 | 0.543 | 0.598 |
| NADP ⁺ | R1 | 0.272 | 0.359 | 0.309 | 0.331 | 0.241 | 0.216 | 0.196 |
| | R2 | 0.292 | 0.328 | 0.310 | 0.264 | 0.213 | 0.222 | 0.210 |
| | R3 | 0.308 | 0.355 | 0.310 | 0.338 | 0.223 | 0.199 | 0.185 |
| UDP-GlcNAc | R1 | 0.313 | 0.215 | 0.154 | 0.121 | 0.364 | 0.394 | 0.372 |
| | R2 | 0.353 | 0.204 | 0.135 | 0.117 | 0.326 | 0.390 | 0.354 |
| | R3 | 0.432 | 0.187 | 0.140 | 0.108 | 0.343 | 0.404 | 0.389 |
| IMP | R1 | 0.010 | 0.008 | 0.007 | 0.004 | 0.024 | 0.024 | 0.017 |
| | R2 | 0.011 | 0.010 | 0.007 | n.d. | 0.031 | 0.016 | 0.014 |
| | R3 | 0.011 | 0.015 | 0.008 | n.d. | 0.028 | 0.015 | 0.007 |
| dGTP/pApp | R1 | 0.129 | 0.156 | 0.130 | 0.108 | 0.169 | 0.096 | 0.089 |
| | R2 | 0.140 | 0.159 | 0.119 | 0.084 | 0.105 | 0.088 | 0.119 |
| | R3 | 0.105 | 0.203 | 0.137 | 0.132 | 0.124 | 0.086 | 0.108 |

*NADH and NADPH were not detected due to instability during extraction procedure and storage conditions for extracts.

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1 **Supplementary Table 4. Genotypes of *P. aeruginosa* and *E. coli* strains used in this**
 2 **study.**

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| Organism | Genotype | Description | Reference |
|---------------------------|--|--|------------|
| <i>P. aeruginosa</i> PAO1 | wild-type | | 2 |
| | ΔPA4856 | <i>retS</i> deletion strain | This study |
| | ΔPA4856 attB:: <i>lacZ</i> | <i>retS</i> deletion strain, constitutive <i>lacZ</i> expression strain, Tet ^R | This study |
| | ΔPA4856 ΔPA0093 | <i>retS tse6</i> deletion strain | This study |
| <i>P. aeruginosa</i> PA14 | wild-type | | 3 |
| | ΔPA14_ | | |
| | ΔPA14_52570 | <i>rsmA</i> deletion strain | This study |
| | ΔPA14_52570 ΔPA14_68450 | <i>rsmA rsmF</i> deletion strain | This study |
| | ΔPA14_52570 ΔPA14_68450 attB:: <i>lacZ</i> | <i>rsmA rsmF</i> deletion strain, constitutive <i>lacZ</i> expression strain, Tet ^R | This study |
| | ΔPA14_52570 ΔPA14_68450 | <i>rsmA rsmF clpV1</i> deletion strain | This study |
| | ΔPA14_01100 ΔPA14_68450 | <i>rsmA rsmF tas1</i> deletion strain | This study |
| | ΔPA14_52570 ΔPA14_68450 | <i>rsmA rsmF tsi6</i> deletion strain | This study |
| | ΔPA14_01140 ΔPA14_68450 | <i>rsmA rsmF tas1 tis1</i> deletion strain | This study |
| | ΔPA14_52570 ΔPA14_68450 | <i>rsmA rsmF tas1 tis1</i> deletion strain | This study |
| | ΔPA14_01140 ΔPA14_01130 attB:: <i>lacZ</i> | <i>rsmA rsmF tas1 tis1 tsi6</i> deletion strain | This study |
| | ΔPA14_52570 ΔPA14_68450 | <i>rsmA rsmF eagT6</i> deletion strain | This study |
| | ΔPA14_01150 ΔPA14_68450 | <i>rsmA rsmF vgrG1</i> deletion strain | This study |
| | ΔPA14_52570 ΔPA14_68450 | <i>rsmA rsmF vgrG2</i> deletion strain | This study |
| | ΔPA14_01160 ΔPA14_68450 | <i>rsmA rsmF vgrG4</i> deletion strain | This study |
| | ΔPA14_52570 ΔPA14_64230 | <i>retS</i> deletion strain | This study |

| | | | |
|---------------------------|--|--|-------------|
| Δ PA14_64230 | Δ PA14_57520 | <i>retS sspB</i> deletion strain | This study |
| Δ PA14_64230 | Δ PA14_57520 PA14_01130-DAS+4 | <i>retS sspB</i> deletion strain expressing Tis1 with a C-terminal DAS+4 tag | This study |
| Δ PA14_52570 | Δ PA14_68450 PA14_01140-VSV-G | <i>rsmA rsmF</i> deletion strain expressing Tas1 with a C-terminal VSV-G tag | This study |
| Δ PA14_52570 | Δ PA14_68450 PA14_01140-E382A | <i>rsmA rsmF</i> deletion strain expressing Tas1 ^{E382A} | This study |
| <i>E. coli</i> | wild-type | | 4 |
| MG1655 | | | |
| <i>E. coli</i> SM10 | Km ^R , <i>thi-1 thr leu tonA lac Y supE recA::RP4-2-Tc::Mu, pir</i> | Conjugation strain | BioMedal |
| λ pir | | | LifeScience |
| <i>E. coli</i> XL-1 Blue | <i>recA1 endA1 gyrA96 thi-1 hsdR17 supE44 relA1 lac [F' proAB lacI^q ZΔM15 Tn10 (Tet^R)]</i> | Cloning strain | Agilent |
| <i>E. coli</i> BL21 (DE3) | F ⁻ <i>ompT gal dcm lon hsdSB(rB⁻ mB⁻)</i> | Protein expression strain | Novagen |
| CodonPlus | λ (DE3) pLysS(Cm ^R) | | |

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1 **Supplementary Table 5. Plasmids used for allelic exchange, protein expression and**
 2 **toxicity assays.**

3

| Plasmid | Relevant features | Reference |
|--------------------------|--|---------------|
| pEXG2 | Allelic replacement vector containing <i>sacB</i> , Gm ^R | ⁵ |
| pPSV38-CV | Expression vector with <i>lacI</i> , <i>lacUV5</i> promoter, C-terminal VSV-G tag, Gm ^R | ⁶ |
| pPSV39-CV | Expression vector with <i>lacI</i> , <i>lacUV5</i> promoter, C-terminal VSV-G tag, Gm ^R | ⁷ |
| pSCRhaB2-CV | Expression vector with <i>PrhaB</i> , Tmp ^R | ⁸ |
| pETDuet-1 | Co-expression vector with <i>lacI</i> , T7 promoter, N-terminal His ₆ tag in MCS-1, Amp ^R | Novagen |
| pET28b | Expression vector with <i>lacI</i> , T7 promoter, C-terminal His ₆ tag, Amp ^R | Novagen |
| pCfa | Expression vector with <i>lacI</i> , T7 promoter, C-terminal Cfa-His ₆ intein tag, Amp ^R | ⁹ |
| pKSV45-Amp ^R | Expression vector with <i>tetR</i> and <i>ptet</i> promoter, replaced <i>kan</i> ^R with <i>amp</i> ^R | ¹⁰ |
| pALS13 | Expression vector for (p)ppGpp synthetase fragment of RelA 1-455 (RelA') | ¹¹ |
| pSW196 | MiniCTX1 plasmid, Tet ^R | ¹² |
| pEXG2::ΔPA0905 | <i>rsmA</i> deletion construct for PAO1 | This study |
| pEXG2::ΔPA5182 | <i>rsmF</i> deletion construct for PAO1 | This study |
| pEXG2::ΔPA0093 | <i>tse6</i> deletion construct | This study |
| pEXG2::ΔPA0090 | <i>clpV1</i> deletion construct for PAO1 | This study |
| pEXG2::ΔPA14_52570 | <i>rsmA</i> deletion construct for PA14 | This study |
| pEXG2::ΔPA14_68450 | <i>rsmF</i> deletion construct for PA14 | This study |
| pEXG2::ΔPA14_01100 | <i>clpV1</i> deletion construct for PA14 | This study |
| pEXG2::ΔPA14_01140 | <i>tas1</i> deletion construct | This study |
| pEXG2::ΔPA14_01120 | <i>tsi6</i> deletion construct | This study |
| pEXG2::ΔPA14_01140-01130 | <i>tas1 tis1</i> deletion construct | This study |
| pEXG2::ΔPA14_01140-01120 | <i>tas1 tis1 tsi6</i> deletion construct | This study |
| pEXG2::ΔPA14_01150 | <i>eagT6</i> deletion construct | This study |
| pEXG2::ΔPA14_01110 | <i>vgrG1</i> deletion construct | This study |
| pEXG2::ΔPA14_01160 | <i>vgrG2</i> deletion construct | This study |
| pEXG2::ΔPA14_29390 | <i>vgrG4</i> deletion construct | This study |

| | | |
|---|---|------------|
| pEXG2::ΔPA14_64230 | <i>retS</i> deletion construct | This study |
| pEXG2::ΔPA14_57520 | <i>sspB</i> deletion construct | This study |
| pEXG2::PA14_01130_DAS+4 | For generating strains encoding Tis1 fused to a C-terminal DAS+4 tag (AANDENYSENYADAS) | This study |
| pEXG2::PA14_01140-VSV-G | For generating strains encoding Tas1 fused to a C-terminal VSV-G epitope tag (YTDIEMNRLGK) | This study |
| pEXG2::PA14_01140_E382A | For generating strains encoding Tas1 ^{E382A} | This study |
| pPSV39-CV::PA14_01130 | Expression vector for <i>tas1</i> | This study |
| pPSV39-CV::PA14_01120 | Expression vector for <i>tsi6</i> | This study |
| pPSV39-CV::PA14_57520 | Expression vector for <i>sspB</i> | This study |
| pPSV39-CV::PA5338 | Expression vector for <i>spoT</i> | This study |
| pSCRhaB2-CV::PA14_01140_251-CT | Expression vector for the C-terminal toxin domain of Tas1 (Tas1 _{tox}) | This study |
| pSCRhaB2-CV::PA14_01140_251-CT_K305A | Expression vector for Tas1 _{tox} ^{K305A} | This study |
| pSCRhaB2-CV::PA14_01140_251-CT_K313A | Expression vector for Tas1 _{tox} ^{K313A} | This study |
| pSCRhaB2-CV::PA14_01140_251-CT_K326A | Expression vector for Tas1 _{tox} ^{K326A} | This study |
| pSCRhaB2-CV::PA14_01140_251-CT_D327A | Expression vector for Tas1 _{tox} ^{D327A} | This study |
| pSCRhaB2-CV::PA14_01140_251-CT_R330A | Expression vector for Tas1 _{tox} ^{R330A} | This study |
| pSCRhaB2-CV::PA14_01140_251-CT_E382A | Expression vector for Tas1 _{tox} ^{E382A} | This study |
| pSCRhaB2-CV::PA14_01140_251-CT_E382D | Expression vector for Tas1 _{tox} ^{E382D} | This study |
| A1-2 | Expression vector for Tas1 _{tox} with N-terminal His ₆ and C-terminal VSV-G epitope | This study |
| pETDuet-1::His ₆ _PA14_01140_251-CT :: PA14_01130 | Co-expression vector for Tas1 _{tox} with N-terminal His ₆ and Tis1 | This study |
| pETDuet-1::His ₆ _PA14_01140_251-CT_VSV-G:: PA14_01130 | Co-expression vector for Tas1 _{tox} with N-terminal His ₆ and C-terminal VSV-G and Tis1 | This study |
| pETDuet-1::PA5338_1-387_His ₆ | Expression vector for the SpoT ₁₋₃₈₇ fragment with C-terminal His ₆ | This study |
| pET28b:: D8B36_07150 | Expression vector for PurF ^{EC} | 9 |
| pET28b:: D8B36_21350 | Expression vector for PurD ^{EC} | 9 |
| pCfa:: D8B36_07150 | Expression vector for PurF ^{EC} | 9 |
| pCfa:: D8B36_07150_R62A | Expression vector for PurF ^{EC R62A} | 9 |

pCfa:: PA14_23290
pSW196:: *lacZ*

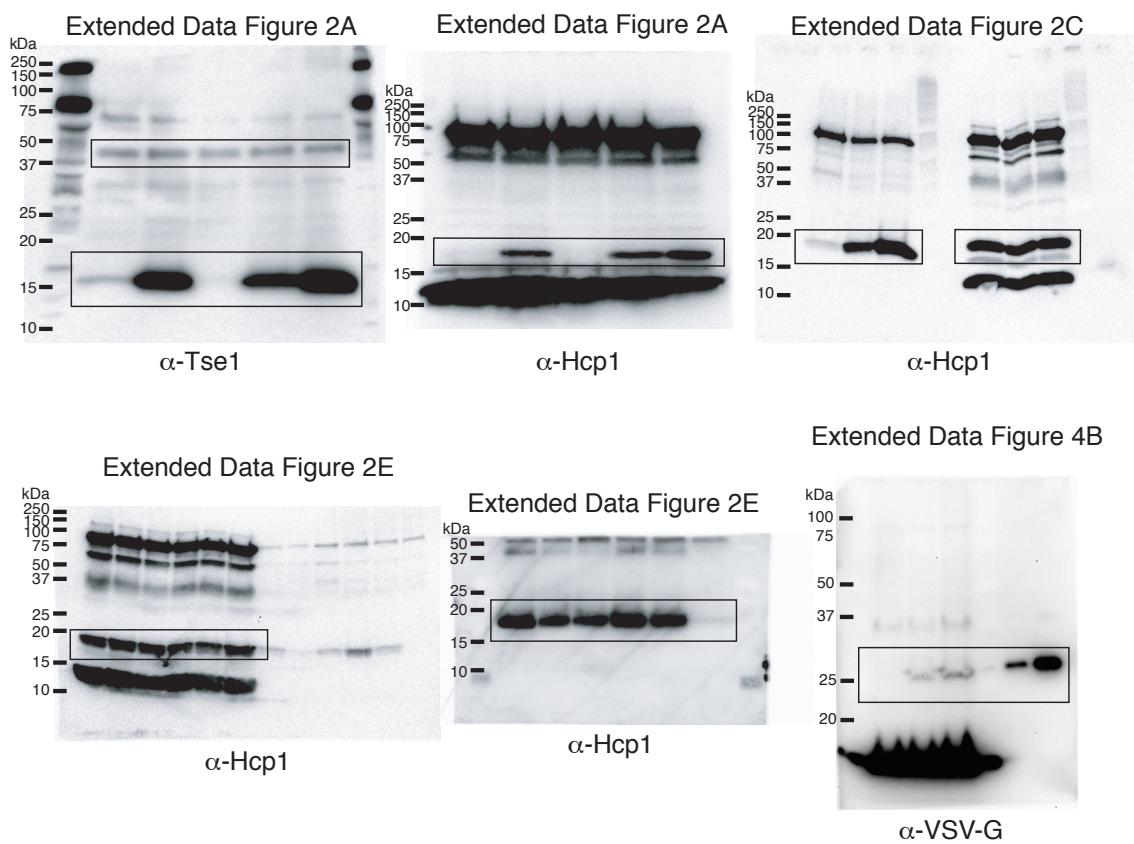
Expression vector for PurF^{PA}
lacZ in miniCTX1 plasmid

This study
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1 **Supplementary Figure 1. Source gels.** Uncropped blots for Extended Data figures.

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1 **Supplemental References**

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